

Supplementary Online Content

Volesky-Avellaneda KD, Morais S, Walter SD, et al. Cancers attributable to infections in the US in 2017: a meta-analysis. *JAMA Oncol*. Published online October 19, 2023.

doi:10.1001/jamaoncol.2023.4273

eAppendix 1. Acknowledgements

eAppendix 2. Acronyms and Abbreviations

eAppendix 3. Cancers and Associated *ICD-O-3* Codes

eTable 1. Search Performed in MEDLINE 1946–January 6, 2023

eAppendix 4. Cancer Incidence

eAppendix 5. Multiple Imputation

eAppendix 6. Hepatitis B and C Viruses

eTable 2. Estimated Prevalence of HBsAg Infection in the US, NHANES Data Collected 1999–2010

eTable 3. Estimated Prevalence of HCV-RNA Infection in the US, NHANES Data Collected 1999–2010

eAppendix 7. Hepatocellular Carcinoma

eTable 4. Characteristics of Case-Control Studies on the Association Between HBV or HCV Infection and HCC

eFigure 1. Pooled ORs for the Association Between Each (1) HBV and (2) HCV and HCC

eTable 5. HBV and HCV Associated PAFs (%) for HCC, by Age Group and Sex

eAppendix 8. Non-Hodgkin Lymphoma

eTable 6. The Association Between HCV Infection and NHL Subtypes as Reported in the InterLymph Study

eAppendix 9. Intrahepatic Bile Duct Cancer

eTable 7. Characteristics of Case-Control Studies on the Association Between HBV or HCV Infection and Intrahepatic Bile Duct Cancer

eFigure 2. Pooled ORs for the Association Between Each (1) HBV and (2) HCV and Intrahepatic Bile Duct Cancer

eAppendix 10. *Helicobacter pylori*

eTable 8. Estimated *H. pylori* Prevalence in the US and PAFs for NCGC

eAppendix 11. Gastric Cancer (Non-Cardia)

eTable 9. Characteristics of Studies on the Association Between *H. pylori* Infection Detected Using ELISA or EIA and NCGC

eTable 10. Characteristics of Studies on the Association Between *H. pylori* Infection Detected Using Immunoblot and NCGC

eFigure 3. Pooled Corrected (1) and Uncorrected (2) ORs for the Association Between *H. pylori* and NCGC

eAppendix 12. Gastric MALT and DLBCL

eAppendix 13. Esophageal Adenocarcinoma

eTable 11. Characteristics of Studies on the Association Between *H. pylori* Infection and Esophageal Adenocarcinoma
eFigure 4. Forest Plot of the Association Between *H. pylori* Infection and Esophageal Adenocarcinoma (Fixed Effects)
eAppendix 14. Epstein-Barr Virus
eAppendix 15. Burkitt Lymphoma
eTable 12. Characteristics of Studies on EBV Prevalence in BLs From Individuals Aged 0-19
eFigure 5. Forest Plot of EBV Prevalence (%) in BL Tumor Tissues Collected From Individuals Aged 0-19
eAppendix 16. Hodgkin Lymphoma
eTable 13. Characteristics of Studies Reporting on EBV Prevalence in HLs
eFigure 6. Forest Plot of EBV Prevalence (%) in HL Tumor Tissues Collected From Individuals Aged 0–19
eFigure 7. Forest Plot of EBV Prevalence (%) in HL Tumor Tissues
eAppendix 17. Nasopharyngeal Carcinoma
eTable 14. Characteristics of Studies Reporting on EBV Prevalence in NPC Cases
eFigure 8. Forest plot of EBV Prevalence (%) in NPC Tumor Tissues Collected From Adults
eAppendix 18. Extranodal Natural Killer T-Cell Lymphoma – Nasal Type
eAppendix 19. Diffuse Large B-Cell Lymphoma
eTable 15. Characteristics of Studies Reporting on EBV Prevalence in DLBCL Cases
eFigure 9. Forest Plot of EBV Prevalence (%) in DLBCL Tumor Tissues
eAppendix 20. Gastric Carcinoma
eTable 16. Characteristics of Studies Reporting on EBV Prevalence in GC Cases
eFigure 10. Forest Plot of EBV Prevalence (%) in GC, by Sex
eAppendix 21. Human Papillomavirus
eAppendix 22. Anal SCC
eTable 17. Characteristics of Studies Reporting on HR-HPV Prevalence in Invasive Anal SCCs, by Sex and HIV Status
eFigure 11. Forest Plot of the Prevalence (%) of HR-HPV in Anal SCC, by Sex
eAppendix 23. Penile Cancer
eFigure 12. Forest Plot of HR-HPV Prevalence (%) in Penile Cancer
eTable 18. Characteristics of Studies Reporting on HR-HPV Prevalence in Penile Cancers
eAppendix 24. Vaginal Cancer
eTable 19. Characteristics of Studies Reporting on HR-HPVa Prevalence in Vaginal Cancers
eAppendix 25. Vulvar Cancer
eTable 20. Characteristics of Studies Reporting on the Prevalence of HR- HPV in Vulvar Cancer Cases, by Age-Group
eFigure 13. Forest Plot for HR-HPV Prevalence (%) in Vulvar Cancer, by Age Group
eAppendix 26. Head and Neck Cancers
eTable 21. Characteristics of Studies Reporting on HPV16 Prevalence Detected via E6 and/or E7 in HNCs
eFigure 14. Forest Plot of HPV16 E6/E7 Prevalence (%) in Cancer of the Oropharynx

eFigure 15. Forest Plot of HPV16 E6/E7 Prevalence (%) in Cancer of the Oral Cavity

eFigure 16. Forest Plot of HPV16 E6/E7 Prevalence (%) in Cancer of the Larynx

eAppendix 27. Merkel Cell Polyomavirus

eAppendix 28. Merkel Cell Carcinoma of the Skin

eTable 22. Characteristics of Studies Reporting on MCPyV Prevalence in Merkel Cell Carcinoma of the Skin

eFigure 17. Forest Plot of MCPyV Prevalence (%) in Merkel Cell Carcinoma of the Skin

eReferences.

This supplementary material has been provided by the authors to give readers additional information about their work.

eAppendix 1. Acknowledgements

We are grateful to the following individuals for sharing additional data with us:

Dr. Jon H. Chung (Clinical development, Foundation Medicine, Cambridge, Massachusetts); **Dr. Janet Daling** (Program in Epidemiology, Fred Hutchinson Cancer Research Center Seattle, Washington); **Dr. Cecilie Dupont Harwood** (Department of Otorhinolaryngology, Head and Neck Surgery, and Audiology, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark); **Dr. Terence Dunn** (College of Medicine, The University of Oklahoma, Norman, Oklahoma); **Dr. Julia Gargano** (Division of Viral Diseases, HPV Team, Centers for Disease Control and Prevention, Atlanta, Georgia); **Dr. Michael Herfs** (Laboratory of Experimental Pathology, GIGA-Cancer, University of Liège, Liège, Belgium); **Ms. Elysha Kolitz** (The University of Texas Southwestern Medical Center, Dallas, Texas); **Dr. Christina S. Kong** (Stanford Cancer Center & Lucile Packard Children's Hospital, Palo Alto, California); **Dr. Sam Mbulaiteye** (Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda, Maryland); **Dr. Margaret Madeleine** (Program in Epidemiology, Fred Hutchinson Cancer Research Center, Seattle, Washington); **Dr. Edyta C. Pirog** (Weill Cornell Medicine, Cornell University, Ithaca, New York).

eAppendix 2. Acronyms and Abbreviations

AIDS	acquired immunodeficiency syndrome
anti-HBc	total hepatitis B core antibody
BL/L	Burkitt lymphoma/leukemia
CI	confidence interval
CLL/SLL	chronic/small lymphocytic leukemia/lymphoma
DLBCL	diffuse large B-cell lymphoma
EBER ISH	EBV-encoded RNA <i>in situ</i> hybridization
EBV	Epstein-Barr virus
EIA	enzyme immunoassay
ELISA	enzyme-linked immunosorbent assay
ENKTL	extranodal natural killer T-cell lymphoma
ES	effect size
FFPE	formalin-fixed paraffin-embedded
GC	gastric carcinoma/cancer
GI	gastrointestinal
HBsAg	hepatitis B surface antigen
HBV	hepatitis B virus
HCV	hepatitis C virus
HCC	hepatocellular carcinoma
HIV	human immunodeficiency virus
HL	Hodgkin lymphoma
HNC	head and neck cancer
<i>H. pylori</i>	<i>Helicobacter pylori</i>
HIV	human immunodeficiency virus
HPV	human papillomavirus
HR	high-risk
I^2	index of consistency
IARC	International Agency for Research on Cancer
ICC	intrahepatic cholangiocarcinoma
ICCC	International Classification for Childhood Cancer
ICD	International Classification of Diseases
KSHV	Kaposi sarcoma-associated herpesvirus
LMP-1	latent membrane protein 1
LPL/WM	lymphoplasmacytic lymphoma/Waldenström's macroglobulinemia
MALT	mucosa-associated lymphoid tissue
MCPyV	Merkel cell polyomavirus
MZL	marginal zone lymphoma
NA	not applicable
NCGC	non-cardia gastric cancer
NHANES	National Health and Nutrition Examination Survey
NOS	not otherwise specified
NPC	nasopharyngeal carcinoma
NS	not specified
OR	odds ratio
PAF	population attributable fraction
PWH	people with HIV
Pos	positive
RR	relative risk
RSE	relative standard error
SCC	squamous cell carcinoma
SD	standard deviation
SE	standard error
SEER	Surveillance, Epidemiology, and End Results Program
SES	socioeconomic status
US	United States
WHO	World Health Organization
yrs	years

eAppendix 3. Cancers and Associated ICD-O-3 Codes

Cancer	ICD-O-3 codes	Recode used (if applicable) / exclusions
Lymphomas		
Burkitt lymphoma	9687	Children only: ICCC site recode extended
Chronic/small lymphocytic leukemia/lymphoma	9823	Lymphoma subtype recode/WHO 2008
DLBCL NOS	9680	Lymphoma subtype recode/WHO 2008
Lymphoplasmacytic lymphoma	9671	Lymphoma subtype recode/WHO 2008
Marginal zone lymphoma	9699	Lymphoma subtype recode/WHO 2008
Gastric, mucosa-associated lymphoid tissue	C16; 9699	NA
Gastric, DLBCL NOS	C16; 9680	NA
Hodgkin lymphoma	C81	Lymphoma subtype recode/WHO 2008
ENKTL, nasal type	9719	NA
Primary effusion lymphoma	9678	NA
Adult T-cell leukemia/lymphoma	9827	Lymphoma subtype recode/WHO 2008
Head and neck cancers		
Nasopharyngeal carcinoma	C11.0–9; 8010, 8020–1, 8070–3, 8082–3	Children only: ICCC site recode extended
Oropharynx	C01.9, C02.4, C02.8, C05.1–2, C09, C10, C14.2	
Oral cavity	C00.3–5, C00.9, C02.0–3, C02.9, C03, C04, C05.0, C05.8–9, C06, C14.8	Excluded: 9050–9055 (mesothelioma), 9140 (Kaposi sarcoma), 9590–9992 (malignant lymphomas)
Larynx	C32	
Gastrointestinal tract cancers		
Esophageal adenocarcinoma	C15; 8050–83	NA
Hepatocellular carcinoma	C22.0; 8170–5	NA
Intrahepatic bile duct	C22.1	Excluded: 9050–9055, 9140, 9590–9992
Gastric, non-cardia	C16.1–6, proportion of C16.8–9 ^a	NA
Gastric carcinoma	C16	Excluded: 9050–9055, 9140, 9590–9992
Anal squamous cell carcinoma	C21.0–2, C21.8; 8050–8076, 8083–4, 8123–4	NA
Genital cancers		
Cervix	C53	
Penis	C60	
Vagina	C52	Excluded: 9050–9055, 9140, 9590–9992
Vulva	C51	
Skin cancers		
Kaposi sarcoma	9140	NA
Merkel cell carcinoma of the skin	C44.0, C44.2–9; 8247	NA

DLBCL = diffuse large B-cell lymphoma, ENKTL = extranodal natural killer T-cell lymphoma, ICCC = International Classification for Childhood Cancer, ICD = International Classification of Diseases, NA = not applicable, NOS = not otherwise specified, WHO = World Health Organization

^a Non-cardia gastric counts were adjusted by reassigning a proportion of 'overlapping lesion' and 'NOS' gastric carcinoma to non-cardia gastric cancer.

LITERATURE SEARCH

The search shown in **eTable 1** was designed to capture knowledge syntheses (i.e., systematic reviews with or without meta-analyses, scoping reviews, etc.). It included MeSH terms and keywords related to infections and cancers included in this study, and knowledge syntheses, and was limited to records published in English. This search captured 3,604 records of which 353 underwent full-text review.

eTable 1. Search Performed in MEDLINE 1946–January 6, 2023

Individual infections	<ol style="list-style-type: none"> 1. exp Hepatitis B virus/ or exp Hepatitis B/ or exp Hepatitis C/ or exp Hepacivirus/ or (hepatitis virus* or hepatitis B or hepatitis C or HBV or HCV or hep B or hep C).tw,kf. 2. exp Herpesvirus 4, Human/ or exp Epstein-Barr Virus Infections/ or (herpesvirus type 4 or herpesvirus 4 or ebv or hhv4 or hhv-4).tw,kf. or ((epstein-barr or epstein barr) adj2 (virus* or viral*)).tw,kf. 3. exp HTLV-1 Infections/ or exp Human T-lymphotropic virus 1/ or (human T-cell lymphotropic virus or Human T-lymphotropic virus or HTLV-1 or HTLV1).tw,kf. 4. exp Herpesvirus 8, Human/ or (human herpesvirus 8 or human herpesvirus type 8 or sarcoma-associated herpesvirus or Kaposi sarcoma-associated herpesvirus or HHV-8 or HHV8 or KSHV).tw,kf. or (Kaposi* adj3 (virus* or viral*)).tw,kf. 5. exp Helicobacter/ or exp Helicobacter infection/ or (helicobacter or pylori or pyloridis or HP or campylobacter, H* pylori).tw,kf. 6. exp Papillomavirus Infections/ or exp Papillomaviridae/ or (human papillomavirus* or human papilloma virus* or hpv).tw,kf. 7. exp HIV Infections/ or exp HIV/ or (hiv or hiv-1 or hiv-2 or hiv1 or hiv2 or hiv infect* or deficiency virus).tw,kf. or (human immun* adj2 (virus* or viral*)).tw,kf. 8. exp Merkel cell polyomavirus/ or (merkel cell polyomavirus or MCV or MCPyV).tw,kf. or (merkel adj3 polyomavirus).tw,kf. 9. 1 or 2 or 3 or 4 or 5 or 6 or 7 or 8
Cancer	<ol style="list-style-type: none"> 10. exp Neoplasms/ or (cancer* or neoplas* or tumor* or tumour* or malignan* or carcinoma* or metasta* or oncolog* or leukemi* or leukaemi* or lymphoma* or myeloma* or sarcoma* or squamous cell* or adenocarcinoma*).tw,kf.
Knowledge syntheses ^a	<ol style="list-style-type: none"> 11. (meta-analysis or systematic review).pt. or meta-analysis/ or systematic review/ or exp meta-analysis as topic/ or ((systematic* adj3 (review* or overview*)) or (methodologic* adj3 (review* or overview*))).ti,ab,kf. or ((quantitative adj3 (review* or overview* or syntheses*)) or (research adj3 (integrati* or overview*))).ti,ab,kf. or ((integrative adj3 (review* or overview*)) or (collaborative adj3 (review* or overview*)) or (pool* adj3 analy*)).ti,ab,kf. or (data syntheses* or data extraction* or data abstraction*).ti,ab,kf. or (handsearch* or hand search*).ti,ab,kf. or (met analy* or metanaly*).ti,ab,kf. or (meta regression* or metaregression*).ti,ab,kf. or (meta-analy* or metaanaly* or systematic review*).mp,hw. or (medline or cochrane or pubmed or medlars or embase or cinahl).ti,ab,hw. or (mantel haenszel or peto or der simonian or dersimonian or fixed effect* or latin square*).ti,ab,kf.
Limits	<ol style="list-style-type: none"> 12. 9 and 10 and 11 13. limit 12 to English 14. limit 13 to humans

^a. The knowledge syntheses search terms were adapted from the Canadian Agency for Drugs and Technologies in Health (CADTH) database search filters. Ottawa: CADTH; 2016. [Available from: <https://searchfilters.cadth.ca/>]

eAppendix 4. Cancer Incidence

Cancer incidence data covering 100% of the US population, including the 50 states, the District of Columbia and Puerto Rico were obtained (due to Hurricane Maria, Puerto Rico's incidence counts are restricted to the first six months of 2017). Cancer was categorized according to ICD-O-3. We used the following coding classifications: ICD-O-3/WHO 2008 for primary sites, lymphoma subsite recode/WHO 2008 for lymphomas, and the ICC site extended or recode ICD-O-3/WHO 2008 for children. However, the recode could not be used to obtain all the sites required (e.g., subsites of oropharynx or oral cavity). Cancers that did not have a specific histology associated with them (e.g., NCGC, oropharynx, etc.) had the following histologies excluded from their counts: 9050–9055 (mesothelioma), 9140 (Kaposi sarcoma), 9590–9992 (malignant lymphomas).

For the main analysis, the NCGC incidence counts were adjusted by reassigning a proportion of 'overlapping lesion' and NOS GC to NCGC. This proportion was determined by calculating the distribution of cardia (C16.0) versus NCGC (C16.1–16.6) by sex and 5-year age groups and multiplying the proportion that was NCGC by the counts of overlapping lesion and NOS, then adding those counts to the existing NCGC counts. We applied the PAFs to unadjusted NCGC (C16.1–16.6) incidence as a sensitivity analysis. For adults, we reclassified B-cell NOS lymphomas based on distribution of B-cell lymphomas of known histology by sex and 5-year age groups, then applied PAFs for EBV to Burkitt lymphoma, EBV and HCV to DLBCL, and HCV to other non-Hodgkin lymphomas.

eAppendix 5. Multiple Imputation

The imputation model included variables known to be associated with both the infection and missingness, as applicable (i.e., for all three infections: sex, age [missing age at medical examination was imputed using age at interview–last observation carried forward method], education, race, and primary sampling units and strata; HBV infection also included country of birth, intravenous drug use, men who have sex with men, and number of lifetime sexual partners; HCV infection also included injection drug use, receiving a blood transfusion before 1992, HIV diagnosis and anti-HCV antibody result; *H. pylori* infection also included time living in the US, number of people living in the household and family income).¹ We then estimated the prevalence of HBV, HCV and *H. pylori* infection; analyses included the sampling weights provided by NHANES to account for unequal probabilities of selection resulting from the sample design. The recommended variance estimation of Taylor series linearization for variance estimation was used to calculate 95% CIs for the prevalence estimates.²

eAppendix 6. Hepatitis B and C Viruses

The general population prevalence estimates for the hepatitis viruses are displayed in eTables 2 and 3.

eTable 2. Estimated Prevalence of HBsAg Infection in the US, NHANES Data Collected 1999–2010

Sex-age group (yrs)	Sample				Weighted			Imputed + Weighted	
	Pos no.	Pos %	RSE ^a %	Missing ^b %	Pos %	RSE ^a %	Missing ^b %	Pos %	RSE ^a %
Males									
15–29	12	0.21	28.8	7.8	0.33	32.3	7.9	0.34	32.7
30–39	13	0.58	27.7	6.6	0.51	32.3	5.4	0.55	32.2
40–49	12	0.50	28.8	4.7	0.26	30.3	4.0	0.28	32.2
50–59	22	1.08	21.2	5.1	0.93	22.4	3.6	0.96	21.8
≥60	14	0.32	26.7	5.3	0.28	40.1	4.2	0.31	38.3
Overall	73	0.44	11.7	6.2	0.44	14.8	7.0	0.46	6.8
Females									
15–29	4	0.07	50.0	8.0	0.06	75.4	8.1	0.08	65.1
30–39	10	0.38	31.6	6.0	0.34	30.5	5.2	0.35	30.5
40–49	11	0.43	30.1	5.0	0.28	36.4	4.4	0.30	36.3
50–59	6	0.30	40.8	5.6	0.20	42.4	4.9	0.28	42.4
≥60	13	0.30	27.7	6.3	0.19	33.5	5.3	0.21	33.4
Overall	44	0.25	15.1	6.6	0.20	19.2	7.5	0.23	5.4

HBsAg = hepatitis B surface antigen, NHANES = National Health and Nutrition Examination Survey, Pos = positive, RSE = relative standard error, US = United States, yrs = years

a. The RSE, which is calculated by dividing the estimate's standard error by the estimate itself, RSE's of <30% should be indicated in reporting.³
 b. Missing refers to individuals who attended the interview and medical examination but do not have a test result for HBsAg infection.

eTable 3. Estimated Prevalence of HCV-RNA Infection in the US, NHANES Data Collected 1999–2010

Sex-age group (yrs)	Sample				Weighted			Imputed + Weighted	
	Pos no.	Pos %	RSE ^a %	Missing ^b %	Pos %	RSE ^a %	Missing ^b %	Pos %	RSE ^a %
Males									
15–29	7	0.12	37.8	7.9	0.24	45.1	7.9	0.26	44.2
30–34	7	0.65	37.7	7.5	0.43	33.5	6.6	0.81	38.1
35–39	18	1.54	23.4	6.4	1.19	23.6	5.2	1.26	22.8
40–44	34	2.78	16.9	6.1	2.67	18.8	5.8	3.47	18.1
45–49	48	4.21	14.1	5.6	3.83	17.3	4.3	4.45	16.8
50–54	45	3.86	14.6	6.6	2.87	17.7	5.5	4.22	18.2
55–59	20	2.38	22.1	6.5	1.62	31.6	4.4	2.14	26.5
60–64	26	2.16	19.4	5.7	1.40	26.5	3.7	1.55	24.6
≥65	15	0.48	25.8	5.8	0.27	29.8	4.8	0.48	26.6
Overall	220	1.32	6.7	6.8	1.36	9.2	7.5	1.76	8.7
Females									
15–29	6	0.10	40.8	8.1	0.09	59.7	8.2	0.10	54.6
30–34	5	0.37	44.6	7.0	0.43	53.9	6.3	0.54	46.6
35–39	12	0.95	28.7	5.5	0.75	34.3	4.8	0.97	32.4
40–44	14	1.08	26.6	5.8	0.83	30.2	5.6	1.34	30.4
45–49	25	2.05	19.8	6.2	1.58	22.2	5.1	2.43	21.4
50–54	19	1.71	22.7	7.2	1.05	30.9	6.5	1.61	29.1
55–59	7	0.81	37.6	5.4	0.35	38.7	4.5	0.73	41.1
60–64	8	0.62	35.2	5.9	0.46	45.3	4.5	0.56	39.0
≥65	11	0.41	27.3	7.1	0.22	48.4	5.9	0.38	33.9
Overall	107	0.62	9.6	7.0	0.55	12.4	7.9	0.82	11.7

HCV = hepatitis C virus, NHANES = National Health and Nutrition Examination Survey, Pos = positive, RSE = relative standard error, US = United States, yrs = years

a. The RSE, which is calculated by dividing the estimate's standard error by the estimate itself, RSE's of <30% should be indicated in reporting.³
 b. Missing refers to individuals who attended the interview and medical examination but do not have a test result for HCV RNA infection (note, this can include those who tested anti-HCV positive but did not have sufficient volume of sera to be tested for HCV RNA).

eAppendix 7. Hepatocellular Carcinoma

Through inflammation of the liver (cirrhosis), HBV and HCV can cause the major liver cancer histological type – HCC.⁴ Additionally, HBV can cause HCC directly through chromosomal integration.⁵ Studies reporting a measure of association are detailed in **eTable 4**.

eTable 4. Characteristics of Case-Control Studies on the Association Between HBV or HCV Infection and HCC

Study ^a	Study population	Matching variables	Characteristics of participants	Detection method	Cases		Controls		OR (95% CI) ^b	Adjustment variables & remarks
					n/N	Pos %	n/N	Pos %		
Hassan 2009 ⁶	Cases: diagnosed HCC GI outpatient clinics at M.D. Anderson Cancer Center Controls: three healthy controls/case non-blood family members of patients recruited from radiology clinic; similar in age, sex, race/ethnicity, education level	Sex, age group, race	Males: 245 cases; 615 controls Mean age (SE): 62 (0.7) for cases; 60 (NS) for controls	Anti-HCV (3 rd gen. ELISA)	79/347	22.8	6/1075	0.6	79.2 (30.6–204.8)	Age, sex, race, educational level, cigarette smoking, alcohol consumption, diabetes mellitus, family history of cancer, HBsAg, anti-HBc
Ognjanovic 2009 ⁷	Recruited: 2000–2006 Diagnosed: 2000–2008 Cases: Los Angeles HCC Study (HCC cases were identified through the Los Angeles County Cancer Surveillance Program, a population-based cancer registry) Controls: two neighbourhood controls/case and from Health Care Financing Administration files	Sex, age (±5), race	Males: 82 cases; 139 controls Mean age (SD): 60.5 (10.3) for cases; 59.5 (10.7) for controls, range: 18–74 yrs	Anti-HCV via ELISA v2 kit, confirmed with RIBA	58/120	48.3	1/230	0.4	211.0 (40.01–4368)	None OR calculated in OpenEpi
Davila 2005 ⁸	Diagnosed: 1984–2001 Sera collection: 1992–NS Cases: HCC in SEER registries also enrolled in Medicare (aged ≥65 yrs) Controls: population-based non-cancer controls aged ≥65 yrs, matched 3:1 to cases on time of diagnosis Recruited: 1994–1999	Frequency matched	Males: 1352 cases; 2248 controls Minimum age for study: 65 Age ≥75: 1139 cases; 3260 controls	ICD-9 codes for HBV ICD-09 codes for HCV or unspecified hepatitis diagnosed before 1992	182/2061	8.8	14/6183	0.2	23.94 (13.65–41.99)	Age, sex, race, SEER registry, Medicare/Medicaid dual enrolment; HCV OR without diabetes

anti-HBc = total hepatitis B core antibody, CI = confidence interval, gen. = generation, GI = gastrointestinal, HBV = hepatitis B virus, HBsAg = hepatitis B surface antigen, HCC = hepatocellular carcinoma, HCV = hepatitis C virus, EIA = enzyme immunoassay, ELISA = enzyme-linked immunoassay, NS = not specified, OR = odds ratio, Pos = positive, RIBA = Recombinant ImmunoBlot Assay, SD = standard deviation, SE = standard error, SEER = Surveillance, Epidemiology, and End Results Program, US = United States, yrs = years

^a Inclusion criteria: hepatitis infection confirmed by serology (HBsAg, anti-HCV [with or without RIBA confirmation], HCV RNA), 10 or more HCC cases, controls without liver disease, US study population.

^b After the normalizing transformation was performed, the CIs listed in the table may not match those in the forest plot.

eTable 4. Characteristics of case-control studies on the association between HBV or HCV infection and HCC (continued)

Study ^a	Study population	Matching variables	Characteristics of participants	Detection method	Cases		Controls		OR (95% CI) ^b	Adjustment variables & remarks
					n/N	Pos %	n/N	Pos %		
Hassan 2002 ⁹	Cases: HCC patients diagnosed at The University of Texas M. D. Anderson Cancer Center hospital Controls: histologically confirmed malignant neoplasms other than HCC, which included primary tumors of the GI tract (44.3%), urogenital tract (18.7%), respiratory tract (17.8%), and skin (19.1%) Diagnosed: 1994–1995	2 controls matched for sex, age (5 yrs), year of diagnosis to 1 case	Males: 87 cases; 174 controls Mean age (SD): 59.5 (10.7) for cases; 59.1 (10.9) for controls	HBsAg via ELISA	17/115	14.8	2/230	0.9	23.8 (3.9–141.6)	Alcohol consumption, cigarette smoking, diabetes mellitus, anti-HCV (for HBV only), HBsAg (for HCV only)
				Anti-HCV (2 nd gen. ELISA) confirmed with RIBA	26/115	22.6	5/230	2.2	14.1 (4.0–49.7)	
Nomura 1996 ¹⁰	Cases: American men of Japanese ancestry with HCC, born between 1900-1919 living in Hawaii Controls: males without cancer selected from the cohort Recruited/diagnosed: NS	Age at examination, date of serum collection	All male	HBsAg	15/24	62.5	2/72	2.8	43.0 (5.7–325.5)	Not adjusted
				Anti-HCV via EIA and confirmed with RIBA (1 st gen.)	0/23	0.0	0/67	0.0	Not computed due to a lack of exposure in cases and controls	
Di Bisceglie 1991 ¹¹	Cases: consecutive HCC patients at Johns Hopkins Oncology Center Controls: patients with other malignant tumors (20% GI tract, 34% respiratory tract, 20% urogenital tract, and 16% breast, 10% neurological or hematological) at same institution Diagnosed: 1987–1988	Not matched	Males: 67 cases; 53 controls Mean (range) age at diagnosis: 52 (10–86) for cases; 55 (18–70) for controls	HBsAg	7/99	7.1	0/98 (0.5 added to empty cell)	0.0	11.31 (1.39–335.3)	Not adjusted; 0.5 added to empty cell (HBsAg controls)
				Anti-HCV	13/99	13.1	2/98	2.0	7.20 (1.78–48.22)	

anti-HBc = total hepatitis B core antibody, CI = confidence interval, gen. = generation, GI = gastrointestinal, HBV = Hepatitis B virus, HBsAg = hepatitis B surface antigen, HCC = hepatocellular carcinoma, HCV = hepatitis C virus, EIA = enzyme immunoassay, ELISA = enzyme-linked immunoassay, NS = not specified, OR = odds ratio, Pos = positive, RIBA = Recombinant ImmunoBlot Assay, SD = standard deviation, SE = standard error, SEER = Surveillance, Epidemiology, and End Results Program, US = United States of America, yrs = years

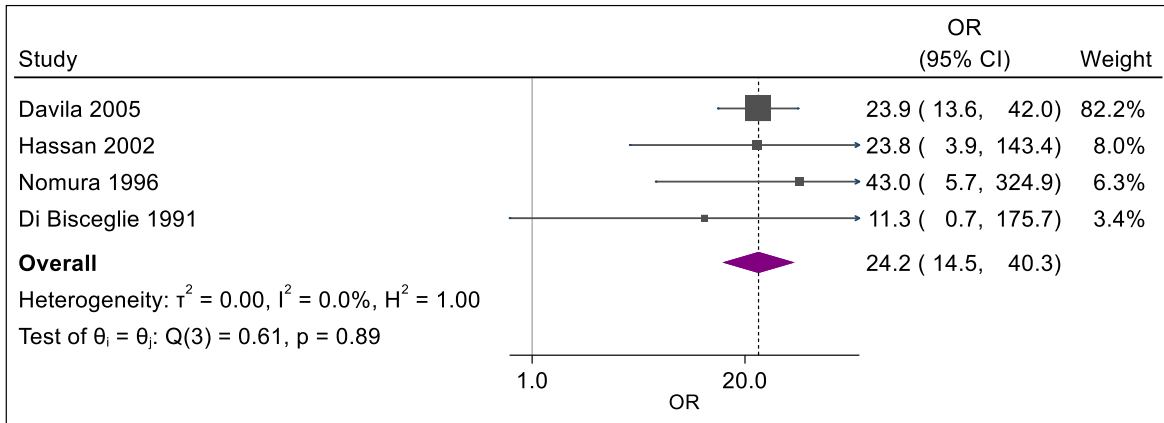
^a. Inclusion criteria: hepatitis infection confirmed by serology (HBsAg, anti-HCV [with or without RIBA confirmation], HCV RNA), 10 or more HCC cases, controls without liver disease, US study population.

^b. After the normalizing transformation was performed, the CIs listed in the table may not match those in the forest plot.

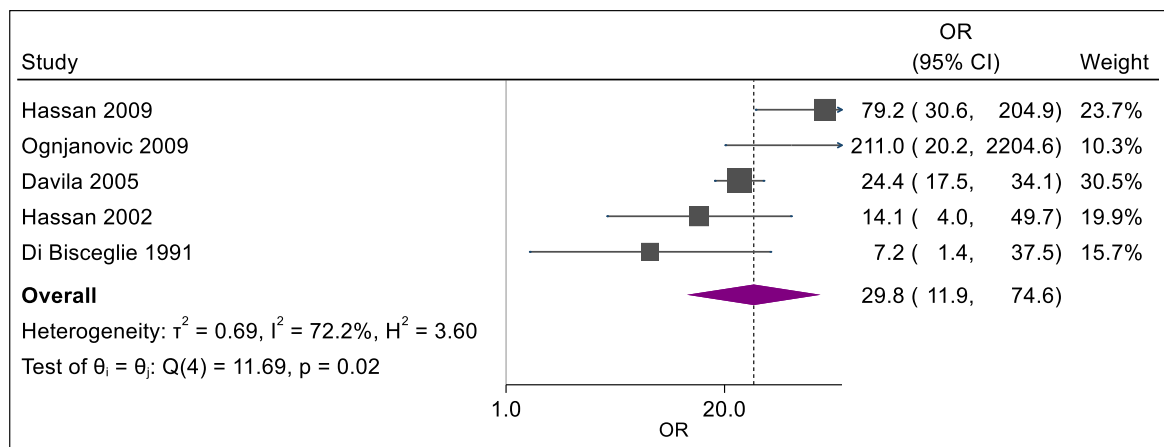
Pooling four studies reporting on HBV and five studies reporting on HCV gave a pooled OR of 24.2 (CI: 14.5–40.3) for HBV and 29.8 (CI: 11.9–74.6) for HCV (eFig. 1).

eFigure 1. Pooled ORs for the Association Between Each (1) HBV and (2) HCV and HCC

(1) Hepatitis B virus (fixed effects)



(2) Hepatitis C virus



CI = confidence interval, HCC = hepatocellular carcinoma, I^2 = index of consistency, OR = odds ratio

The partitioned PAFs for HBV ranged from 1.8–16.0% and for HCV from 2.8–53.1% (eTable 5).

eTable 5. HBV and HCV Associated PAFs (%) for HCC, by Age Group and Sex

HCC sex-age group incidence (yrs)	HBV		HCV		Combined HBV-HCV PAF for 2017 % ^a	Partitioned PAFs ^a	
	Prevalence from age group (yrs)	Individual PAF %	Prevalence from age group (yrs)	Individual PAF %		HBV %	HCV %
Males							
20–24	15–29	7.3	15–29	6.9	13.7	7.0	6.6
25–29	15–29	7.3	15–29	6.9	13.7	7.0	6.6
30–34	15–29	7.3	15–29	6.9	13.7	7.0	6.6
35–39	15–29	7.3	15–29	6.9	13.7	7.0	6.6
40–44	30–39	11.4	30–34	19.0	28.2	10.5	17.6
45–49	30–39	11.4	35–39	26.6	35.0	10.5	24.5
50–54	40–49	6.1	40–44	50.0	53.1	5.8	47.3
55–59	40–49	6.1	45–49	56.2	58.9	5.8	53.1
60–64	50–59	18.3	50–54	54.9	63.2	15.8	47.4
65–69	50–59	18.3	55–59	38.1	49.4	16.0	33.4
70–74	≥60	6.7	60–64	30.9	35.5	6.4	29.2
≥75	≥60	6.7	≥60	12.1	17.9	6.4	11.6
Females							
20–24	15–29	1.9	15–29	2.8	4.6	1.8	2.8
25–29	15–29	1.9	15–29	2.8	4.6	1.8	2.8
30–34	15–29	1.9	15–29	2.8	4.6	1.8	2.8
35–39	15–29	1.9	15–29	2.8	4.6	1.8	2.8
40–44	30–39	7.4	30–34	13.6	20.0	7.1	12.9
45–49	30–39	7.4	35–39	21.8	27.6	7.0	20.6
50–54	40–49	6.6	40–44	27.8	32.6	6.2	26.3
55–59	40–49	6.6	45–49	41.2	45.1	6.2	38.9
60–64	50–59	6.1	50–54	31.7	35.9	5.8	30.1
65–69	50–59	6.1	55–59	17.4	22.4	5.8	16.6
70–74	≥60	4.6	60–64	13.9	17.9	4.5	13.4
≥75	≥60	4.6	≥60	9.8	14.0	4.5	9.5

HBV = hepatitis B virus, HCV = hepatitis C virus, HCC = hepatocellular carcinoma, PAF = population attributable fraction, yrs = years

^a. The PAFs for HBV and HCV in HCC were combined with this equation: $1 - (1 - \text{HBV PAF}) * (1 - \text{HCV PAF})$ then partitioned by determining the proportion of the summed number of attributable cases.¹²

eAppendix 8. Non-Hodgkin Lymphoma

Since NHLs are a heterogeneous group of cancers and studies show that the magnitude of the association between HCV and NHL varies by subtype,¹³⁻¹⁵ we utilized subtype specific measures of association. Data arising from the InterLymph Non-Hodgkin Lymphoma Subtypes Project, which pooled data from 11 mostly population-based case-control studies conducted in Australia, Europe and North America were used in the PAF calculations.^{13,16} Of the 11 InterLymph studies, six¹ assessed HCV seropositivity via third-generation enzyme-linked immunosorbent assay (ELISA).¹⁶ The overall OR for the association for HCV and NHL was 1.81 (CI: 1.39–2.37). Subtypes that HCV demonstrated a statistically significant association with were included (**eTable 6**). Notably, there were few cases of Burkitt lymphoma/leukemia (BL/L); however, since a similar magnitude of association (OR = 5.2, CI: 1.6–16.8) was also found in another large (33,940 NHL cases overall, 197 BL cases) study conducted in the US, we retained BL/L in the analysis.¹⁴ An OR for HCV and BL/L in those aged <50 years was not calculated by the original study authors; we imputed 0.5 persons to the empty cell and calculated an OR of 1.47 (CI: 0.07–8.03). Since it was not statistically significant, we did not include BL/L among those <50 years in the PAF calculations. With only three cases of Waldenström’s macroglobulinemia (the remaining 204 cases were lymphoplasmacytic lymphoma [LPL]), we applied the resulting PAF from LPL/WM to LPL incidence only.

eTable 6. The Association Between HCV Infection and NHL Subtypes as Reported in the InterLymph Study

NHL subtype	Cases		Controls		Adjusted OR (95% CI)	Adjustment variables
	n/N	Pos %	n/N	Pos %		
BL/L: age <50 yrs ¹⁷	0/31	0.0	42/1933	2.2	--	
BL/L: age ≥50 yrs ¹⁷	3/33	9.1	109/4562	2.4	4.1 (1.1–15.4)	Age, sex, race/ethnicity, & study
CLL/SLL ¹⁸	21/994	2.1	95/5354	1.8	2.08 (1.23–3.49)	
DLBCL ¹⁹	63/1654	3.8	152/6898	2.2	Males: 2.17 (1.44–3.26) Females: 1.98 (1.18–3.34)	Age, sex, race/ethnicity, study, SES, history of autoimmune disease, any atopic disorder, blood transfusion, year of first OC use, age at first HT use, 1 st degree family history – NHL, BMI as young adult, usual adult BMI, lifetime alcohol consumption, recreational sun exposure, field crop vegetable farmer, sewer & embroiderer, women’s hairdresser, driver/material handling equipment operator
LPL/WM ²⁰	6/207 ^b	2.9	95/5354	1.8	2.51 (1.03–6.17)	Age, sex, race/ethnicity, study, Sjögren syndrome, systemic lupus erythematosus, hay fever, usual adult weight, smoking duration, family history of hematological malignancy, & medical occupation
MZL ²¹	14/368	3.8	95/5354	1.8	3.04 (1.65–5.60)	Age, sex, race/ethnicity, & study

BL/L = Burkitt lymphoma/leukemia, BMI = body mass index, CLL/SLL = chronic/small lymphocytic leukemia/lymphoma, DLBCL = diffuse large B-cell lymphoma, HCV = hepatitis C virus, HT = hormone therapy, LPL/WM = lymphoplasmacytic lymphoma/Waldenström’s macroglobulinemia, MZL = marginal zone lymphoma, NHL = non-Hodgkin lymphoma, Pos = positive, OC = oral contraceptive, SES = socioeconomic status, yrs = years

^{a.} Inclusion criteria: hepatitis infection confirmed by serology (anti-HCV [with or without RIBA confirmation], HCV RNA), 10 or more cases, study population from Western countries.

^{b.} Among the 374 cases enrolled, only three were diagnosed with WM and the remainder LPL.

¹ The six studies were four population-based (region of recruitment and years cases diagnosed): British Columbia (Vancouver & Victoria, Canada; 2000–2004), UCSF1 (San Francisco, US; 1988–1995), SCALE (Denmark & Sweden, 1999–2002), New South Wales (Australian Capital Territory, 2000–2001); one mixed population-based and/or hospital-based: EpiLymph (Spain, France, Germany, Italy, Ireland, Czech Republic; 1998–2004); Italy and Germany were population-based – the remainder were hospital-based, and one hospital-based: Italy – Aviano-Milan (1983–1992).

eAppendix 9. Intrahepatic Bile Duct Cancer

Pooling four studies reporting on intrahepatic bile duct cancer for each HBV and HCV (**eTable 7**) gave a pooled OR of 3.4 for HBV and 4.5 for HCV (**eFig. 2**).

eTable 7. Characteristics of Case-Control Studies on the Association Between HBV or HCV Infection and Intrahepatic Bile Duct Cancer

Study ^a	Study population	Matching variables	Characteristics of participants, age in yrs (SD)	Detection method	Cases		Controls		OR ^b (95% CI)	Adjustment variables & remarks
					n/N	Pos %	n/N	Pos %		
Petrick 2017 ²²	SEER-Medicare database	None	Cases: 78.0 (6.5) Controls: 76.6 (7.7)	ICD-9 codes for HBV	25/2092	1.2	1200/323,615	0.4	2.97 (1.97–4.46)	Age, race/ethnicity, geographic region, state buy-in status
	Cases: Medicare beneficiaries enrolled continuously in Medicare Parts A and B for a minimum of three yrs prior to cancer diagnosis with ICC			ICD-9 codes for HCV	58/2092	2.8	2161/323,615	0.7	4.67 (3.57–6.11)	
Choi 2016 ²³	Cases: patients seen at the Mayo Clinic from 2000–2014	Frequency matched 1:2 for age (±5 yrs), sex, race, residence	Cases: 60.6 (13.1) Controls: 61.6 (13.5)	HBsAg	10/1169	0.9	8/4769	0.2	12.9 (2.69–61.61)	Propensity score adjustment: age, sex, race, obesity, hypertension, diabetes, cerebrovascular accident etc, ^c
	Controls: recruited from the Mayo Clinic Biobank from 2009–2015, which comprises a collection of blood samples & health information from Mayo Clinic patients and other community volunteers (without a history of cancer other than nonmelanoma skin cancer)			Anti-HCV	23/1169	2.0	17/4769	0.4	1.95 (0.75–5.11)	
Shaib 2007 ²⁴	Cases: cholangiocarcinoma patients referred to the M.D. Anderson Cancer Center between 1992 and 2002	Frequency matched by gender, ethnicity and age (±5 yrs)	Cases: 59.8 (11.4) Controls: 58.1 (11.4)	HBsAg	1/83	1.2	1/236	0.4	2.9 (0.1–236.8)	Race, age, gender, HCV & HBV markers, heavy drinking Lower bound CI was imputed
	Controls: randomly selected from an existing database of healthy individuals (genetically unrelated family members, spouses and friends of patients who had cancer other than gastrointestinal cancer) interviewed between 1999–2004 at M.D. Anderson			Anti-HCV	5/83	6.0	2/236	0.8	7.9 (1.3–84.5)	
Shaib 2005 ²⁵	SEER-Medicare database	Yrs of search for risk factors	Cases: 78.7 (6.4) Controls: 76.5 (6.9)	ICD-9 codes for HBV	1/625	0.2	181/90,834	0.2	0.8 (0.1–5.9)	Age, sex, race, geographic region & Medicare/Medicaid enrollment
	Cases: persons diagnosed no earlier than 1993 and who had two yrs of Medicare data before the date of diagnosis and up to one year after ICC diagnosis or until death			ICD-9 codes for HCV	5/625	0.8	161/90,834	0.2	5.2 (2.1–12.8)	
	Controls: 5% random sample of Medicare-enrolled beneficiaries with no cancer of any type residing in the geographic regions of SEER registries									
	Period: 1993–1999									

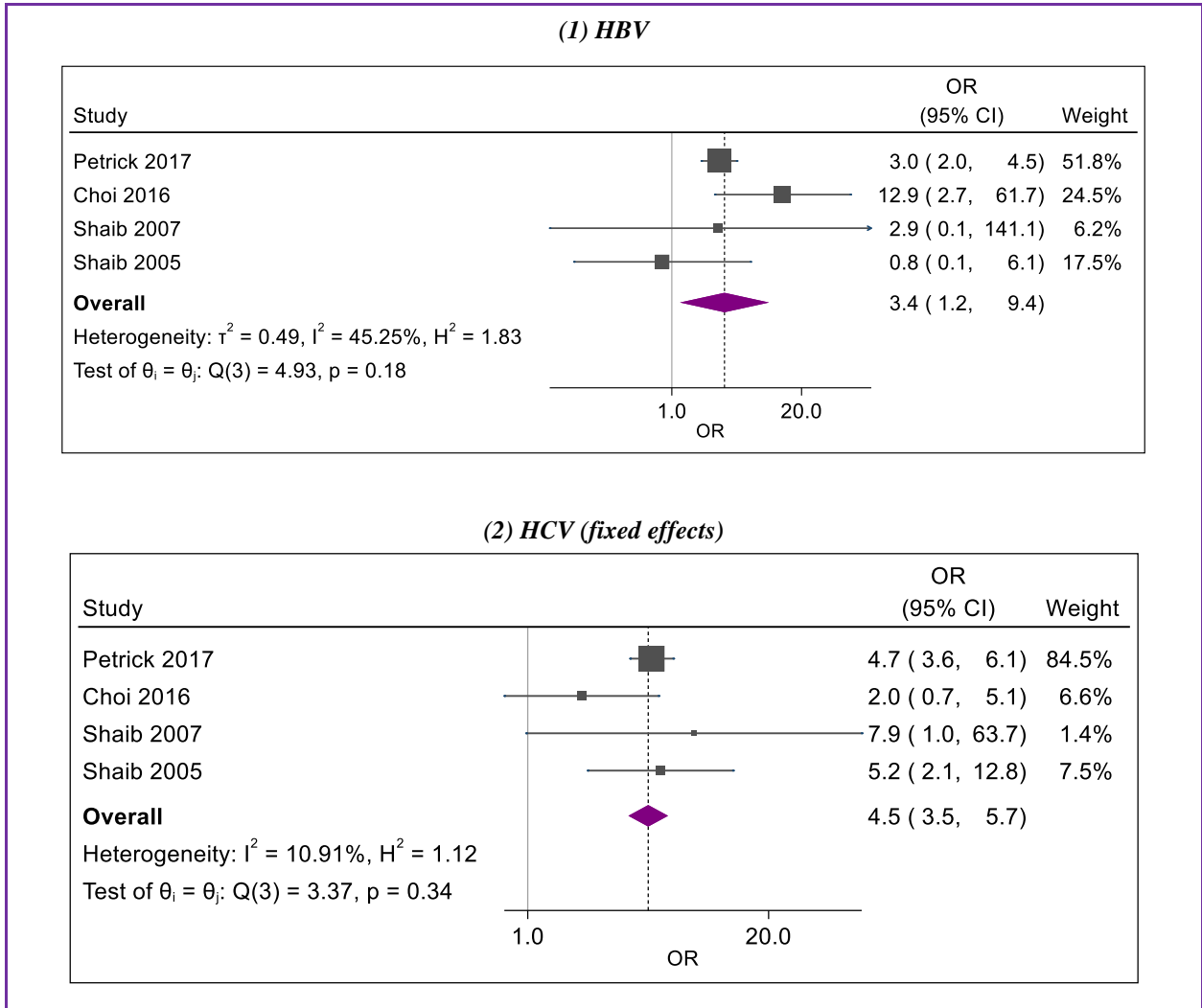
CI = confidence interval, HBV = Hepatitis B virus, HBsAg = hepatitis B surface antigen, HCV = hepatitis C virus, ICC = intrahepatic cholangiocarcinoma, Pos = positive, SD = standard deviation, SEER = Surveillance, Epidemiology, and End Results Program, yrs = years

a. Inclusion criteria: hepatitis infection confirmed by serology (HBsAg, anti-HCV [with or without RIBA confirmation], HCV RNA), 10 or more cases, US study population

b. After the normalizing transformation was performed, the CIs listed in the table may not match those in the forest plot.

c. As well as coronary artery disease, peripheral vascular disease, atrial fibrillation, non-alcoholic fatty liver disease, non-alcoholic steatohepatitis, primary sclerosing cholangitis, cirrhosis, inflammatory bowel disease and smoking status

eFigure 2. Pooled ORs for the Association Between Each (1) HBV and (2) HCV and Intrahepatic Bile Duct Cancer



CI = confidence interval, HBV = hepatitis B virus, HCV = hepatitis C virus, I^2 = index of consistency, OR = odds ratio

eAppendix 10. *Helicobacter pylori*

H. pylori is estimated to infect about 50% of the world's population, but the prevalence varies globally, likely reflecting socio-demographic and economic conditions of the regions.²⁶ Although infection is mostly acquired during childhood, *H. pylori* prevalence increases with age.²⁷ A decrease in the overall prevalence of *H. pylori* infection has been observed in recent years, with successive generations presenting lower prevalence.²⁶ The one cycle of the NHANES assessed *H. pylori* serostatus collected data from participants aged ≥ 3 years, via ELISA from 1999–2000. Equivocal results (representing <2% of results) were categorized as positive (eTable 8).

eTable 8. Estimated *H. pylori* Prevalence in the US and PAFs for NCGC

Sex-age Group (yrs) ^a	NHANES estimates from the 1999–2000 cycle									
	Sample				Weighted			Imputed + Weighted		PAF estimates for NCGC for 2017
	Pos no.	Pos %	RSE %	Missing ^b %	Pos %	RSE %	Missing %	Pos %	RSE %	
Males										
10–14	134	23.5	7.6	9.4	15.0	16.9	13.3	14.6	16.5	Not included
15–19	211	32.4	5.7	8.2	16.2	9.9	11.2	16.4	10.8	Not included
20–24	42	27.1	13.2	6.6	19.9	14.9	3.7	20.4	14.5	63.3
25–29	54	35.1	11.0	4.9	28.8	12.1	4.3	28.9	11.8	65.9
30–34	67	43.5	9.2	5.5	31.1	10.2	5.1	31.1	9.7	70.6
35–39	71	43.0	9.0	5.7	27.1	12.6	4.1	27.2	11.9	77.3
40–44	98	52.1	7.0	6.0	35.6	14.6	5.8	34.5	14.7	78.5
45–49	69	51.9	8.4	5.7	29.3	19.0	4.6	29.6	19.1	76.2
50–54	71	52.2	8.2	6.9	37.7	7.4	8.0	37.8	7.9	80.2
55–59	54	50.0	9.6	5.3	37.8	15.2	1.7	38.4	15.3	77.7
60–64	111	56.6	6.3	3.9	38.2	17.7	4.0	38.8	17.7	81.6
65–69	82	50.9	7.7	8.0	35.3	11.4	6.9	36.7	11.2	81.9
70–74	73	50.3	8.2	7.6	37.1	15.5	10.1	36.8	14.8	82.0
75–79	57	55.9	8.8	8.1	43.4	13.7	5.9	44.1	12.9	81.2
80–84	48	51.1	10.1	9.6	50.7	10.9	11.3	51.0	11.7	81.2
≥ 85	--	--	--	--	--	--	--	--	--	83.8
Overall	1242	39.9	2.2	7.3	29.2	4.0	6.4	29.2	4.0	80.8
Females										
10–14	103	17.9	8.9	12.2	9.2	19.7	14.4	9.3	18.9	Not included
15–19	172	27.8	6.5	7.4	16.5	14.4	9.8	17.1	14.5	Not included
20–24	79	33.1	9.2	7.0	20.9	18.3	7.7	20.8	18.2	52.2
25–29	59	28.4	11.0	8.4	20.0	15.1	8.7	19.8	14.8	66.8
30–34	73	34.0	9.5	4.9	27.4	13.7	4.3	27.3	13.1	71.0
35–39	76	40.9	8.8	6.1	26.5	14.6	3.9	27.1	14.8	70.0
40–44	86	47.5	7.8	5.2	26.8	14.2	6.2	27.2	13.8	76.2
45–49	74	43.8	8.7	8.2	29.9	8.7	5.7	30.9	8.7	76.1
50–54	79	48.2	8.1	5.8	37.3	10.9	5.6	38.2	11.1	76.2
55–59	64	55.7	8.3	2.5	38.8	14.1	2.7	38.8	14.0	78.4
60–64	91	50.3	7.4	9.5	38.6	15.3	7.5	39.4	14.6	81.8
65–69	94	59.5	6.6	10.2	46.1	10.4	9.1	45.6	11.0	82.0
70–74	82	57.3	7.2	5.3	49.9	11.0	5.4	49.6	11.4	82.3
75–80	53	54.1	9.3	5.8	48.2	15.3	3.9	48.3	14.9	84.3
80–84	39	40.2	12.4	7.6	30.7	10.9	8.5	32.9	12.5	85.4
≥ 85	--	--	--	--	--	--	--	--	--	85.0
Overall	1224	36.6	2.3	7.9	28.2	4.6	7.0	28.3	4.7	81.1

NHANES = National Health and Nutrition Examination Survey, NCGC = non-cardia gastric cancer, PAF = population attributable fraction, Pos = positive, RSE = relative standard error, US = United States

-- *H. pylori* prevalence among those ≥ 85 (in 1999–2000) was not calculated because it was not required after applying a latency period.

a. We did not consider *H. pylori* infection prevalence among those aged 3 to 9 (26% did not have a test result).

b. Missing refers to individuals who attended the interview and medical examination but do not have a test result for *H. pylori* infection.

eAppendix 11. Gastric Cancer (Non-Cardia)

This cancer is often classified according to its physical location within the stomach: tumors located in the upper region of the stomach, specifically within 1 to 2 centimeters proximal and 2 centimeters distal to the esophagogastric mucosal junction, are identified as cardia cancers; cancers located in the fundus, body, pyloric antrum or pylorus regions are identified as non-cardia.²⁸ The latter are the most frequent, accounting for 61% of GC cases diagnosed in the US in 2012 (males: 51.8%, females: 75.5%).²⁹ *H. pylori* infection is known to increase the risk of NCGC with a reported pooled estimate of 2.81 (CI: 2.14–3.68) considering case-control studies and case-control studies nested within prospective cohorts.³⁰ The association between *H. pylori* infection and gastric cardia adenocarcinomas remains conflicting. Studies from low gastric cancer risk settings, namely Europe, the US and Australia, generally report null or inverse associations (pooled RR = 0.78, CI: 0.63–0.97), while statistically significant associations have been observed in high-risk settings, namely China, Japan and Korea (pooled RR = 1.98, CI: 1.38–2.83).³⁰ A recent case-cohort study from China, an area of high *H. pylori* infection endemicity, obtained a statistically significant association (hazard ratio = 3.06, CI: 1.54–6.10).²⁸ These differences and null associations observed may be explained by the coexistence of two distinct types of cardia GC.³¹ One arises from non-atrophic gastric mucosa, associated with acid/bile-induced damage to the distal esophagus, resembling esophageal adenocarcinoma³² and is likely to have a higher relative frequency in settings with low overall gastric cancer risk. The other is associated with *H. pylori* induced atrophic gastritis,³² which is etiologically similar to non-cardia tumors and more frequent in populations with a high frequency of gastric cancer. It is possible that *H. pylori* infection may be associated with a small fraction of cardia gastric cancer, however it is difficult to determine the origin of these cancers to obtain an accurate estimate.

In retrospective studies, individuals with GC may test negative following the clearance of infection associated with atrophic gastritis, thus underestimating the prevalence of *H. pylori* infection among cases.²⁹ As such, only cohort studies or case-control studies nested within prospective cohorts were considered to estimate the association between *H. pylori* infection and NCGC.

The finding that immunoblot is more sensitive than ELISA/ EIA³⁰ in detecting *H. pylori* necessitated a correction for this potential error.³³ The sensitivity and specificity were extracted, and pooled from two studies that compared ELISA to immunoblot head-to-head.^{34,35} A derivation of a formula used to correct measurement error (91% sensitivity and 95% specificity) in the ORs was applied to the five nested case-controls that used EIA or ELISA (**eTable 9**).³⁶ The corrected and immunoblot studies (**eTable 10**) were pooled with fixed effects due to a lack of heterogeneity (**eFig. 3**).

eTable 9. Characteristics of Studies on the Association Between *H. pylori* Infection Detected Using ELISA or EIA and NCGC

Study ^a	Study population	Follow-up yrs, mean/median	Matching variables	Characteristics of participants, ages in yrs	Cases		Controls		Unadjusted OR (95% CI)	Corrected OR ^{b,c} (95% CI)
					n/N	Pos %	n/N	Pos %		
Persson 2011 ³⁷	Swedish cohorts (Swedish Institute for Infectious Disease Control Biobank and Malmö Microbiology Biobank)	16.5	Sex, age, sera collection year, biobank	Mean (SD; range) age at sera collection: 30.8 (6.1; 16–40) for cases; 30.9 (6.0; 16–40) for controls Mean (SD; range) age at diagnosis: 47.3 (9.4; 25–68)	35/41	85.4	30/81	37.0	9.9 (3.7–26.3)	21.5 (6.1–75.8)
Hansen 2007 ³¹	Norwegian cohort (Janus Serum Bank Cohort)	11.9	Sex, age, cohort, sera collection date & study source	Males: 91 cases; 267 controls Median (range) age at sera collection: 45.6 (23.6–63.4) Median (range) age at diagnosis: 55.8 (34.3–68.2)	116/129	89.9	247/376	65.7	4.7 (2.5–8.6)	26.6 (6.5–109.1)
Knekt 2006 ³⁸	Finnish cohort (Finnish Mobile Clinic Health Examination cohort)	Up to 24	Sex, age, municipality	Males: 120 cases; 231 controls Mean age (SD) at baseline: 68 (14) for cases	176/193	91.2	292/372	78.5	2.8 (1.6–4.9)	66.2 (4.1–1078.6)
Nomura 2002 ³⁹	US cohort of men of Japanese ancestry	12.7	Age, sera collection date	All men Mean (range) age at diagnosis: 72.5 (50.2–90.3)	231/261	88.5	193/261	73.9	2.7 (1.7–4.3)	7.9 (3.7–16.9)
Parsonnet 1993 ⁴⁰	US cohort of adult subscribers to the Kaiser Permanente Medical Care Program	15	Sex, age group, race, sera collection date & study site	Median age at sera collection: 53.6	84/98	85.7	61/98	62.2	3.6 (1.8–7.3)	7.4 (3.1–19.6)

CI = confidence interval, EIA = enzyme immunosorbent assay, ELISA = enzyme-linked immunosorbent assay, NCGC = non-cardia gastric cancer, OR = odds ratio, Pos = positive, SD = standard deviation, US = United States, yrs = years

^a. Inclusion criteria: prospective serology collection (~10 years in advance of diagnosis), ELISA or EIA, 10 or more non-cardia gastric cancer cases, North American, European, Australian or New Zealand study populations, data required to correct sensitivity and specificity.

^b. Corrected to 91% sensitivity and 95% specificity. ORs were calculated based on the condition maximum likelihood estimates, and CIs were based on Fisher exact tests.

^c. After the normalizing transformation was performed, CIs listed in the table may not match those in the forest plot.

eTable 10. Characteristics of Studies on the Association Between *H. pylori* Infection Detected Using Immunoblot and NCGC

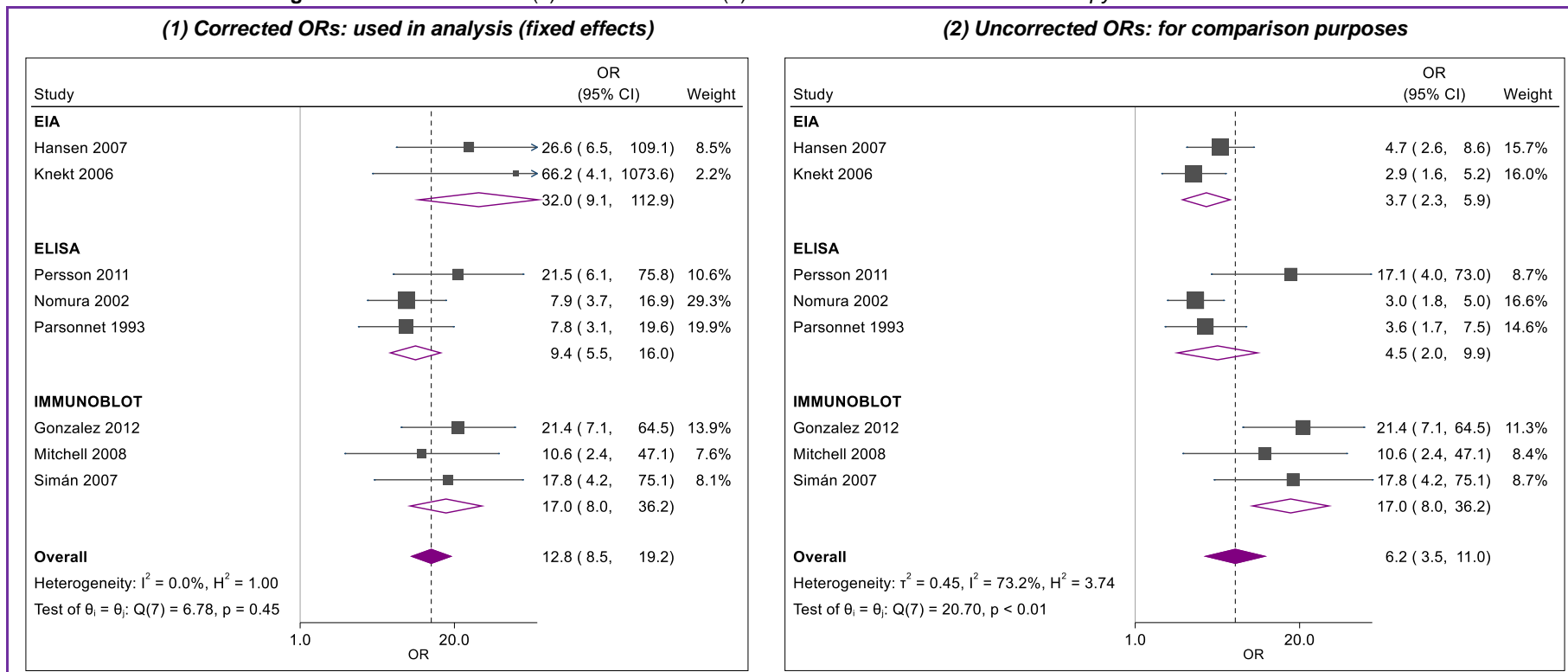
Study ^a	Study population	Follow-up yrs, mean/median	Matching variables	Characteristics of participants, ages in yrs	Cases		Controls		Adjusted OR ^b (95% CI)	Adjustment variables
					n/N	Pos %	n/N	Pos %		
Gonzalez 2012 ³⁴	10 European countries in the EPIC cohort Recruited: 1992–1998 Diagnosed: 2000–2004	10.7	Sex, age group, study center, date of blood collection	Age range at baseline: 40–65	82/88	93.2	199/338	58.9	21.4 (7.1–64.4)	Smoking status, school level, red & processed meat intake, fruit & vegetable consumption
Mitchell 2008 ³⁵	Australian cohort (Melbourne Collaborative Cohort Study) Recruited: 1990–1994 Diagnosed: 1990–2002	11.6	Sex, age, birth country, sera collection date	Males: 21 cases; 84 controls Median (range) age at baseline: 62 (42–69)	32/34	94.1	85/134	63.4	10.6 (2.4–47.4)	None
Simán 2007 ⁴¹	Swedish cohort (Malmö Preventive Medicine) Recruited: 1974–1992 Diagnosed: –2000	Range: 9.2–12.6	Sex, age, sera collection date	Males: 54 cases Mean (range) age at baseline: 50.7 (34.0–60.9)	65/67	97.0	147/250	58.8	17.8 (4.2–74.8)	Occupation, tobacco consumption

CI = confidence interval, EPIC = European Prospective Investigation into Cancer and Nutrition, NCGC = non-cardia gastric cancer, OR = odds ratio, Pos = positive, yrs = years

^a Inclusion criteria: prospective serology collection (~10 yrs in advance of diagnosis), immunoblot detection, 10 or more NCGC cases, North American, European or Australia or New Zealand study populations.

^b After the normalizing transformation was performed, the CIs listed in the table may not match those in the forest plot.

eFigure 3. Pooled Corrected (1) and Uncorrected (2) ORs for the Association Between *H. pylori* and NCGC



CI = confidence interval, EIA = enzyme immunoassay, ELISA = enzyme-linked immunosorbent assay, I^2 = index of consistency, OR = odds ratio

eAppendix 12. Gastric MALT and DLBCL

MALT lymphoma, a type of NHL, is most often diagnosed in the stomach, but can also be found in the lungs, thyroid, skin or soft tissues.⁴² A systematic review of published series found that *H. pylori* infection is present in nearly 90% of patients with gastric MALT lymphoma.⁴³ According to current guidelines, antibiotic therapy against *H. pylori* infection is the first-line of treatment in patients with gastric MALT regardless of stage of disease and prognosis factors.^{44,45} In fact, *H. pylori* eradication confers a ~74% remission rate of MALT in Western populations.⁴³ Even among patients with *H. pylori*-negative gastric MALT, complete remission following eradication therapy is nearly 30%.⁴⁶ We identified only one cohort study examining the relationship between *H. pylori* and gastric NHL. This study, conducted by Parsonnet et al., combined data from two cohort studies conducted in Norway and the US,⁴⁷ and reported a measure of association (OR = 6.3, CI: 2.0–19.9) for NHL of gastric location. This study included 33 cases matched to four controls by cohort, sex, age and sera collection date.⁴⁷ Of the 33 cases, just three cases were gastric MALTs, one case was lymphocytic lymphoma, and the remaining 29 cases were DLBCLs. We opted to utilize the OR for the US cohort (7.9, CI: 1.6–38.1) which included 20 gastric NHL cases and applied it to gastric MALT and DLBCL incidence.

eAppendix 13. Esophageal Adenocarcinoma

Esophageal cancer presents with two major histological types: SCC (morphology codes 8140–8576) that most often arises in the middle third of the esophagus, followed by the lower and the upper third, and adenocarcinoma (morphology codes: 8050–8083) that usually develops in the lower third.⁴⁸ In the US, esophageal adenocarcinoma accounted for 55% of esophageal cancer cases diagnosed between 2001 and 2015.⁴⁹ *H. pylori* infection is inversely associated with the occurrence of esophageal adenocarcinoma, regardless of other environmental and genetic exposures,^{48,50} and the decline in the prevalence of *H. pylori* infection may have contributed to an increase in esophageal adenocarcinoma incidence. The effect of *H. pylori* infection has been evaluated by several meta-analyses reporting results for both esophageal SCC and adenocarcinoma.^{51–56} All reported similar results, showing no association between *H. pylori* and esophageal SCC, while for adenocarcinoma a protective effect of *H. pylori* infection was found (OR ≈ 0.5).

The mechanism through which *H. pylori* infection reduces the risk of esophageal adenocarcinoma is not yet clear. Studies have suggested *H. pylori* infection may decrease gastric cancer secretion by acting on parietal cells via bacterial products and cytokines or through mucosal atrophy resulting from chronic inflammation. Consequently, there may be less reflux esophagitis, Barrett's esophagus, and development of esophageal adenocarcinoma.^{57,58} However, the association between the absence of *H. pylori* infection and increased gastroesophageal reflux,⁵⁹ and whether infection interacts directly with host epithelial cells and/or affects the microbial composition of the esophagus remain unclear.⁶⁰ Nevertheless, previous studies have suggested that the association between *H. pylori* infection and esophageal adenocarcinoma may be independent of CagA status and atrophy of the stomach.^{57,61,62}

Our search produced six meta-analyses that reported results for the association between *H. pylori* infection and esophageal adenocarcinoma, all reported a protective effect.^{51–56} Ten individual studies were conducted in the US.^{57,62–70} Studies that did not provide estimates for esophageal adenocarcinoma specifically (i.e., considered esophageal and gastric cardia adenocarcinoma,⁶⁴ Barrett's esophagus complicated by dysplasia or adenocarcinoma)^{63,70} and/or considered controls with gastrointestinal symptoms or undergoing endoscopy for reasons other than screening (i.e., patients undergoing endoscopy due to achalasia, familial adenomatous polyposis, chronic diarrhea, lower abdominal pain, hemocult-positive stools, unexplained nausea and vomiting, and unexplained chest pain;^{63,70} patients with benign disease and symptoms suggestive of foregut disease;⁶⁹ patients undergoing esophagogastroduodenoscopy for classic symptoms of GERD with or without complaints of dysphagia, nocturnal cough, chest pain, nausea, vomiting, or signs of acute or chronic gastrointestinal bleeding;⁶⁵ patients with intestinal metaplasia⁶⁷ were excluded.^{63,70} Four studies met the inclusion criteria (**eTable 11**) and were pooled with fixed effects due to a lack of heterogeneity (**eFig. 4**).

eTable 11. Characteristics of Studies on the Association Between *H. pylori* Infection and Esophageal Adenocarcinoma

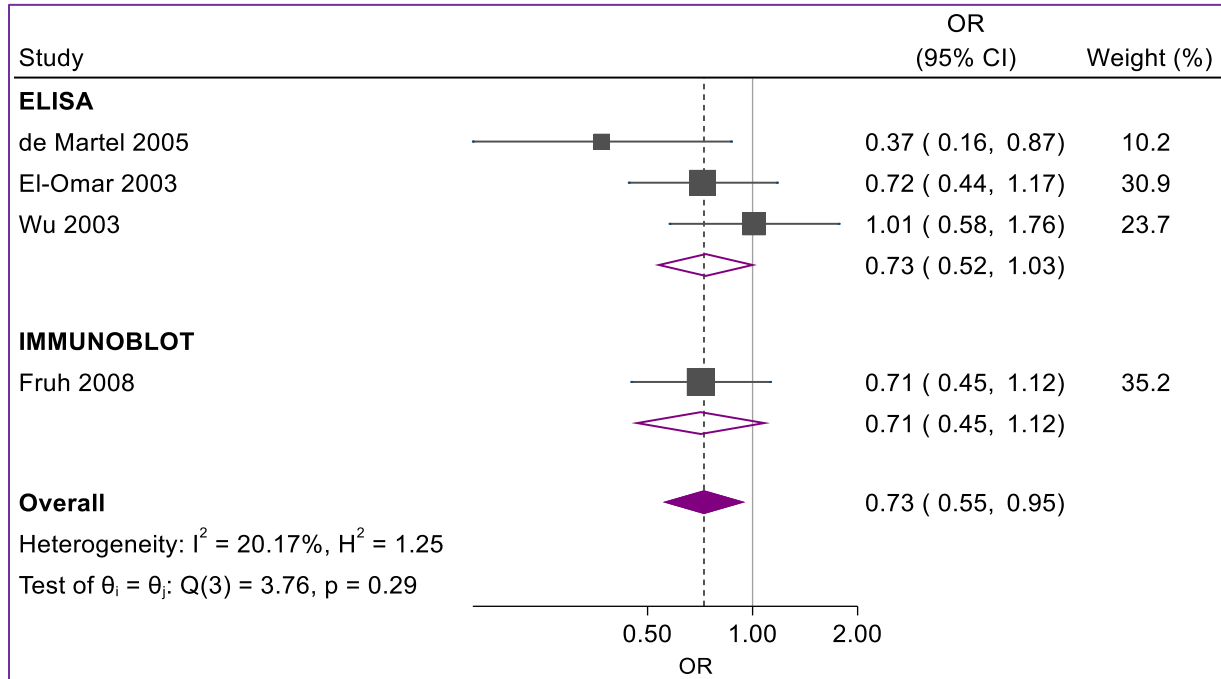
Study ^a	Study population	Matching variables	Characteristics of participants, ages in yrs	Assessment of <i>H. pylori</i> infection	Cases		Controls		Adjusted OR (95% CI) ^b	Adjustment variables
					n/N	Pos %	n/N	Pos %		
Fruh 2008 ⁶²	Case-control study Cases: histologically confirmed esophageal adenocarcinoma patients at the Massachusetts General Hospital Controls: selected from healthy GERD-free, non-blood-related family members and friends of other cancer/surgical patients Diagnosed/Recruited: not specified	Sex, age	Males: 88 cases; 88 controls Mean age (SD): 64 (8) for cases; 63 (8) for controls	Serum (Helicoblot)	36/100	36.0	43/101	42.6	0.71 (0.4–1.0)	Adult BMI, smoking status, age, sex
de Martel 2005 ⁵⁷	Nested case-control study (Kaiser Permanente Medical Care Program) Cases: esophageal adenocarcinoma patients were identified in the cohort and were confirmed by information in the SEER database Controls: randomly selected from the cohort Recruited: 1964–1969 Diagnosed: 1964–2000	Sex, age, race, date & site of sera collection	Males: 41 cases; 121 controls Mean age (SD): 47.9 (10.0) for cases; 47.7 (9.6) for controls	Serum IgG (ELISA)	19/51	37.3	74/150	51.0	0.37 (0.16–0.88)	BMI, cigarette smoking, education
Wu 2003 ⁶⁸	Case-control study Cases: esophageal adenocarcinoma patients from the Los Angeles County Cancer Surveillance Program (population-based cancer registry) Controls: selected from the neighborhood of residence of the case patient Diagnosed/Recruited: 1992–1997	Sex, age group, race	Males: 73 cases; 261 controls	Serum IgG (ELISA)	49/80	61.2	230/356	64.6	1.01 (0.58–1.77)	Sex, age, education, birthplace, ethnic group, smoking status, BMI
Ei-Omar 2003 ⁶⁶	Case-control study Cases: esophageal adenocarcinoma patients from New Jersey and western Washington Controls: population-based controls selected by random-digit dialing and from Health Care Financing Administration files Diagnosed/Recruited: 1993–1995	Sex, age group, study centre	Males: 93 cases; 178 controls Median age: 65 for cases; 66 for controls	Serum IgG (ELISA)	35/108	32.4	84/210	40.0	0.72 (0.44–1.17)	None

BMI = body mass index, CI = confidence interval, ELISA = enzyme-linked immunosorbent assay, GERD = gastroesophageal reflux disease, Pos = positive, OR = odds ratio, SD = standard deviation, SEER = Surveillance, Epidemiology, and End Results, Program, US = United States

^a. Inclusion criteria: cohort, nested case-control or case-control studies with *H. pylori* infection confirmed by serology (ELISA, enzyme immunoassay [EIA] or immunoblot), 10 or more esophageal adenocarcinoma cases, controls without gastrointestinal symptoms and not undergoing endoscopy for purposes other than screening, study population from the US.

^b. After the normalizing transformation was performed, the CIs listed in the table may not match those in the forest plot.

eFigure 4. Forest Plot of the Association Between *H. pylori* Infection and Esophageal Adenocarcinoma (Fixed Effects)^{a,b}



CI = confidence interval, ELISA = enzyme-linked immunosorbent assay, I^2 = index of consistency, OR = odds ratio

- a. The study by de Martel and colleagues published in 2005 is a nested case-control where *H. pylori* sera collection occurred prior to adenocarcinoma diagnosis, the remaining studies are case-controls.
- b. Pooling the unadjusted ORs from the four studies resulted in a pooled OR of 0.75 (CI: 0.57–0.98).

eAppendix 14. Epstein-Barr Virus

Carcinogenicity is demonstrated by the detection of the EBV viral genome within the tumor cells (i.e., where the EBV genome is translated and transcribed).⁷¹ To detect EBV within cancer tissues, EBER ISH is viewed as the gold standard assay;^{4,72} for HL, LMP-1 is comparable to EBER ISH.⁷²

eAppendix 15. Burkitt Lymphoma

BL in children (aged 0–19 yrs)

We identified seven studies conducted in the US and Europe (**eTable 12**). The pooled prevalence of EBV was 15.5% (CI: 8.1–23.0%) (**eFig. 5**).

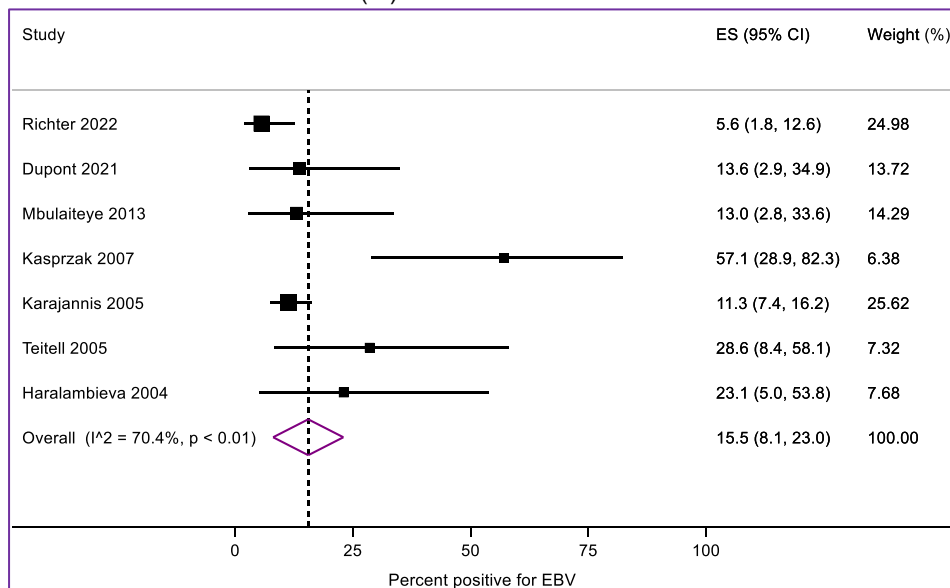
eTable 12. Characteristics of Studies on EBV Prevalence in BLs From Individuals Aged 0–19

Study ^a	Region(s)	Source of cases	Diagnosis dates	Male %	Age range (yrs)	Tested N	Pos %
Richter 2022 ⁷³	Germany	Hematopathology Section & Lymph Node Registry of the University Hospital Schleswig-Holstein, Campus Kiel	2001–2013	86.8	NS	89	5.6
Dupont 2021 ⁷⁴	Denmark	Danish Registry of Pathology	1980–2018	81.8	3–19	22	13.6
Mbulaiteye 2013 ⁷⁵	Los Angeles County, Hawaii & Iowa	Residual tissue repositories (population-based) & diagnostic referral centers	1979–2009	91.3	0–19	23	13.0
Kasprzak 2007 ⁷⁶	Poland	Department of Haematology & Paediatric Oncology	1999–2003	92.9	3–16	14	57.1
Karajannis 2005 ⁷⁷	Austria, Germany & Switzerland	NHL-BFM (Berlin-Frankfurt-Munster) data center	1990–1998	79.7	1–18	222	11.3
Teitell 2005 ⁷⁸	France & United Kingdom	Institut Gustave Roussy & Children's Hospital	NS	85.7	2–16	14	28.6
Haralambieva 2004 ⁷⁹	the Netherlands	Pathology departments & Dutch Childhood Oncology Group	NS	NS	5–13	13	23.1

BL = Burkitt lymphoma, EBV = Epstein-Barr virus, EBER ISH = Epstein-Barr encoding region *in situ* hybridization, NS = Not specified, Pos = positive, yrs = years

^a Inclusion criteria: tissue specimen tested for EBV, EBER ISH detection, European or North American cases, and 8 or more participants.

eFigure 5. Forest Plot of EBV Prevalence (%) in BL Tumor Tissues Collected From Individuals Aged 0–19



BL = Burkitt lymphoma, CI = confidence interval, EBV = Epstein-Barr virus, ES = effect size, I^2 = index of consistency

BL in adults (aged ≥ 20 yrs)

We identified two studies conducted in the US that utilized EBER ISH; they were one study by Mbulaiteye and colleagues (2014) of 40 HIV-negative or unknown HIV status cases (11 HIV+ cases excluded by us) diagnosed from 1979–2009 using SEER data collected from Los Angeles County, Hawaii and Iowa, where 27.5% tested EBV positive (including the 11 PWH, 35.3% tested positive).⁷⁵ Another study by Naeini and colleagues (2016), tested 27 BL cases of unknown HIV status sent to pathology services in California, and reported that 37.0% tested positive.⁸⁰

Pooling five studies (four conducted in Europe and one in the US)^{75,81–84} reporting on EBV prevalence in 118 BLs among PWH, provided prevalence of 50.1% (CI: 34.6–65.6; individual studies not shown). Considering individuals aged 20–59 (since the estimated proportion of BLs occurring among PWH aged ≥ 60 years was only 2.0% over 1980–2007, we did not consider this age group) an estimated 21.5% of BLs from the most recent period available (2001–2007) were diagnosed among PWH in the US.⁸⁵ Weighting the pooled prevalence by HIV status provided EBV prevalence of 35.1%, which is near identical pooled EBV prevalence reported by Mbulaiteye and colleagues that included general and PWH cases. For this reason, we instead opted to use age-group specific EBV prevalence from the Mbulaiteye study which included some HIV+ cases: 55% (aged 20–34), 33% (aged 35–59), 25% (aged ≥ 60).⁷⁵

eAppendix 16. Hodgkin Lymphoma

HL in children (aged 0–19 yrs)

Pooling six studies providing EBV prevalence for younger versus older children (**eTable 13**), resulted in EBV prevalence of 62.2% for children aged 0–9 and 22.3% for those aged 10–19 years (**eFig. 6**).

HL in adults (aged ≥ 20 yrs)

Pooling four studies reporting on EBV prevalence in two adult age groups, provided a pooled prevalence of 20.5% in adults aged 15–44 years old and 42.5% in adults aged ≥ 45 years old (**eFig. S7**). Pooling six studies (two^{86,87} conducted in the US and four⁸⁸⁻⁹¹ in Europe) reporting on EBV prevalence in 282 HL cases diagnosed among PWH, resulted in prevalence of 92.9%. Using data from the 14 SEER cancer registries (2000–2010), Shiels and colleagues estimated the proportion of HLs among PWH by sex and age group.⁹² We utilized the proportion of HL cases estimated to be among PWH by 10-year age groups from age 20 to 69 to partition HL cancer incidence; these proportions were 1.5% (age 20–29), 5.4% (age 30–39), 9.3% (age 40–49), 7.3% (age 50–59), and 1.9% (age 60–69) and applied to males HL incidence counts only.⁹²

eTable 13. Characteristics of Studies Reporting on EBV Prevalence in HLs

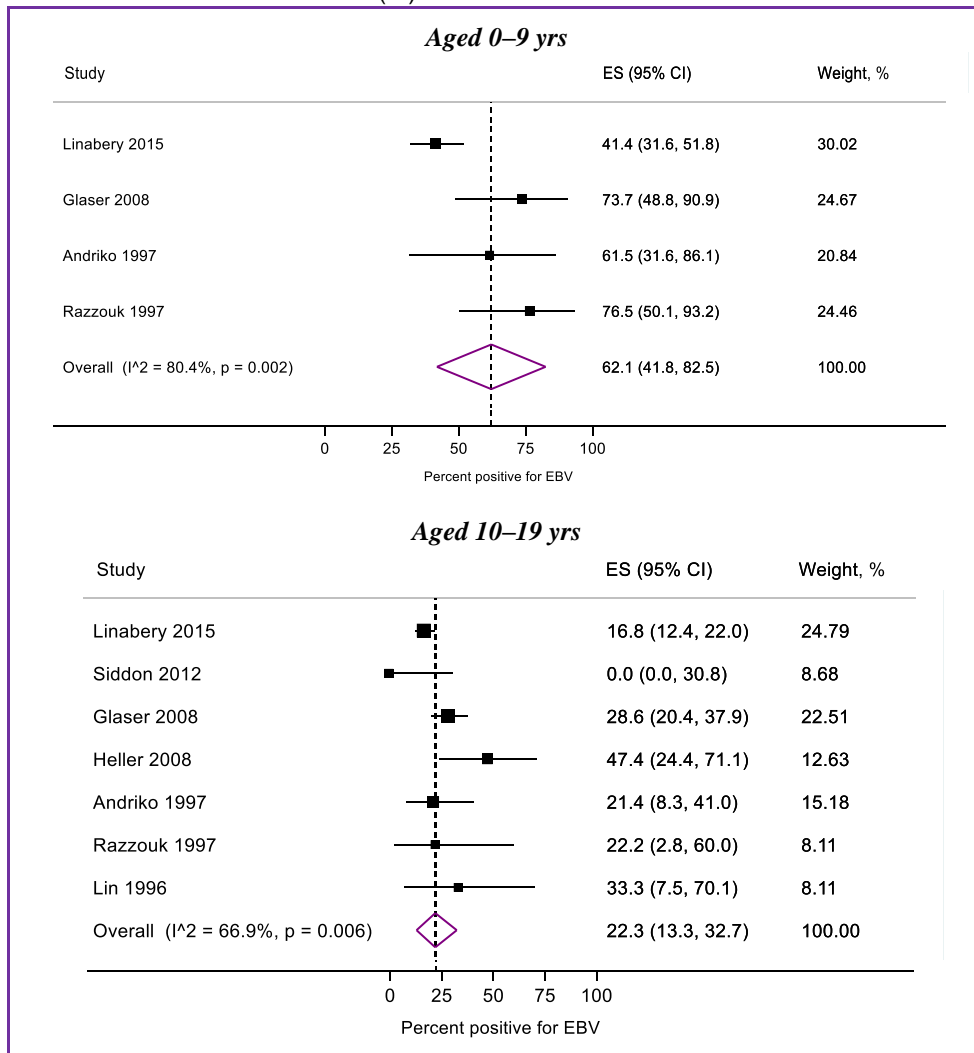
Study ^a	Region(s)	Source of cases	Diagnosis dates	Male %	Detection method(s)	HIV status	Age range (yrs)	Tested N	Pos %
Linabery 2015 ⁹³	US, Puerto Rico & Canada	Children's Oncology Group	1989–2003	NS	EBER ISH	Unknown	0–9 10–14	99 256	41.4 16.8
Siddon 2012 ⁹⁴	Connecticut	Yale-New Haven Hospital	NS	50.0	EBER ISH	HIV-	10–19 0–9	10 19	0.0 73.7
Glaser 2008 ⁹⁵	California	California Cancer Registry and non-White Los Angeles County residents	1988–1997	NS	EBER ISH, LMP-1	HIV-	10–19 20–49 ≥50	112 650 251	28.6 20.9 48.6
Heller 2008 ⁹⁶	New York	Memorial Sloan-Kettering Cancer Center	NS	45.5	EBER ISH	HIV-	10–19	19	47.4
Chang 2004 ⁹⁷	Massachusetts & Connecticut	Population-based case-control study	1997–2001	57.0	EBER ISH, LMP-1	HIV-	15–44 ≥45	291 108	18.9 38.0
Vasef 2004 ⁹⁸	Iowa	Pathology department	NS	58.6	EBER ISH, LMP-1	Unknown	15–44	24	25.0
Andriko 1997 ⁹⁹	Washington, D.C.	Lymphatic Pathology Registry, Armed Force Institute of Pathology	1984–1996	90.9	LMP-1	Unknown	0–9 10–19	13 28	61.5 21.4
Razzouk 1997 ¹⁰⁰	Tennessee	St. Jude Children's Research Hospital	NS	42.3	EBER ISH	Unknown	0–9 10–19	17 9	76.5 22.2
Elenitoba-Johnson 1996 ¹⁰¹	Rhode Island	Pathology departments	NS	42.9	LMP-1	Unknown	15–44 ≥45	18 10	33.3 30.0
Lin 1996 ¹⁰²	Maryland	Clinical Center of National Institutes of Health	1971–1992	NS	EBER ISH	Unknown	10–19	9	33.3
Studies conducted among adults living with HIV									
Besson 2015 ⁸⁸	France	22 centres: French Cohort of HIV-related lymphomas—French National Agency for Research on AIDS and Viral Hepatitis ANRS-CO16 Lymphovir cohort	2008–2014	86.8	EBER-1 ISH, LMP-1	HIV+	38–48	42	92.9
Hentrich 2012 ⁸⁹	Austria & Germany	42 institutions in Austria & Germany	2004–2010	92.6	LMP (81%), EBER ISH (4%), PCR (3%), LMP & EBER (4%), method NS (9%)	HIV+	27–70	103	92.2
Glaser 2003 ⁸⁶	California	California Cancer Registry non-White Los Angeles County residents	1988–1998	100.0	EBER ISH, LMP-1	HIV+	NS	59	89.8
Thompson 2004 ⁸⁷	D.C.	AIDS Registry of the Armed Forces Institute of Pathology ^b	1984–2000	97.8	LMP	HIV+	21–75	33	97.0
Carbone 1999 ⁹⁰	Italy	NS	NS	NS	EBER ISH	HIV+	NS	27	92.6
Tirelli 1995 ⁹¹	Italy	Division of Pathology at the Centro di Riferimento Oncologico	NS	NS	EBER-1 & EBER-2 ISH, Southern blotting	HIV+	NS	18	77.8

AIDS = acquired immunodeficiency syndrome, D.C. = District of Columbia, EBV = Epstein-Barr virus, EBER ISH = Epstein-Barr encoding region *in situ* hybridization, HIV = human immunodeficiency virus, LMP = latent membrane protein, HL = Hodgkin lymphoma, NS = Not specified, PCR = polymerase chain reaction, Pos = positive, US = United States, yrs = years

^a Inclusion criteria: tissue specimen tested for EBV, EBER ISH detection, North American cases (for HIV+ cases from Europe were also eligible), and 8 or more cases (children) or 10 or more cases (adults), EBV prevalence reported by age-group.

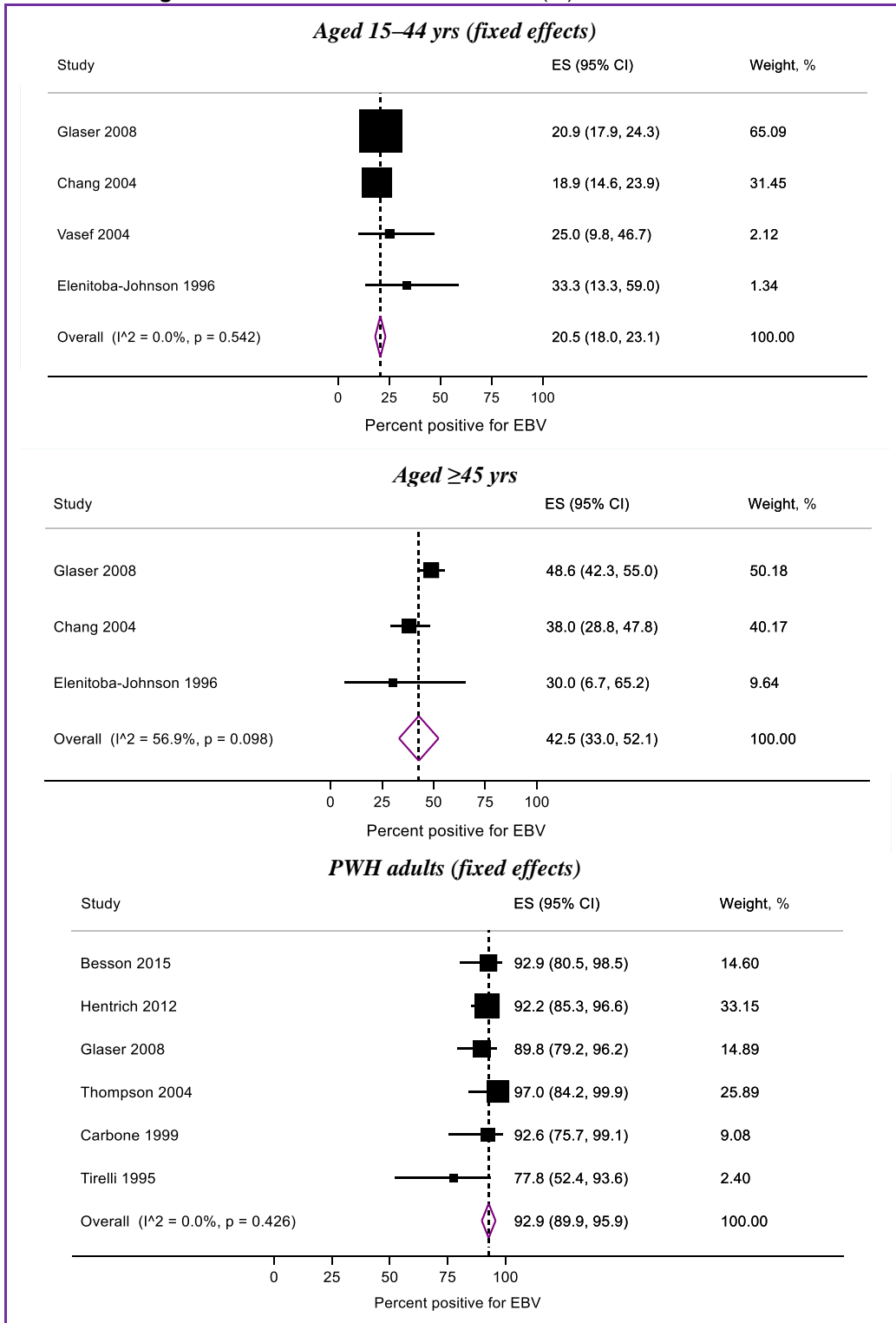
^b 26 cases from civilian sources, 15 cases from Veterans Administration medical centers, four cases from military hospitals.

eFigure 6. Forest Plot of EBV Prevalence (%) in HL Tumor Tissues Collected From Individuals Aged 0–19



CI = confidence interval, EBV = Epstein-Barr virus, ES = effect size, HL = Hodgkin lymphoma, I^2 = index of consistency, US = United States, yrs = years

eFigure 7. Forest Plot of EBV Prevalence (%) in HL Tumor Tissues



CI = confidence interval, EBV = Epstein-Barr virus, ES = effect size, HL = Hodgkin lymphoma, I^2 = index of consistency, PWH = people with human immunodeficiency virus, US = United States, yrs = years

eAppendix 17. Nasopharyngeal Carcinoma

A tumor of the epithelial tissues, NPC, is classified into three main types, keratinizing SCC accounting for 20% of all NPCs and non-keratinizing type accounting for the remaining 80% (further divided into differentiated and undifferentiated).¹⁰³ The pooled prevalence of the seven included studies reporting on EBV in NPC was 61.2% (CI: 45.1–77.2%) (eTable 14, eFig. 8). For individuals aged 0–19 years old, we identified two eligible studies (Table S14)^{104,105} each with eight NPC cases – all EBV positive. Since this was insufficient to perform a meta-analysis, we calculated exact CIs in OpenEpi¹⁰⁶ using a numerator and denominator of eight patients for a prevalence of 100.0% (CI: 63.1–100.0%).

eTable 14. Characteristics of Studies Reporting on EBV Prevalence in NPC Cases

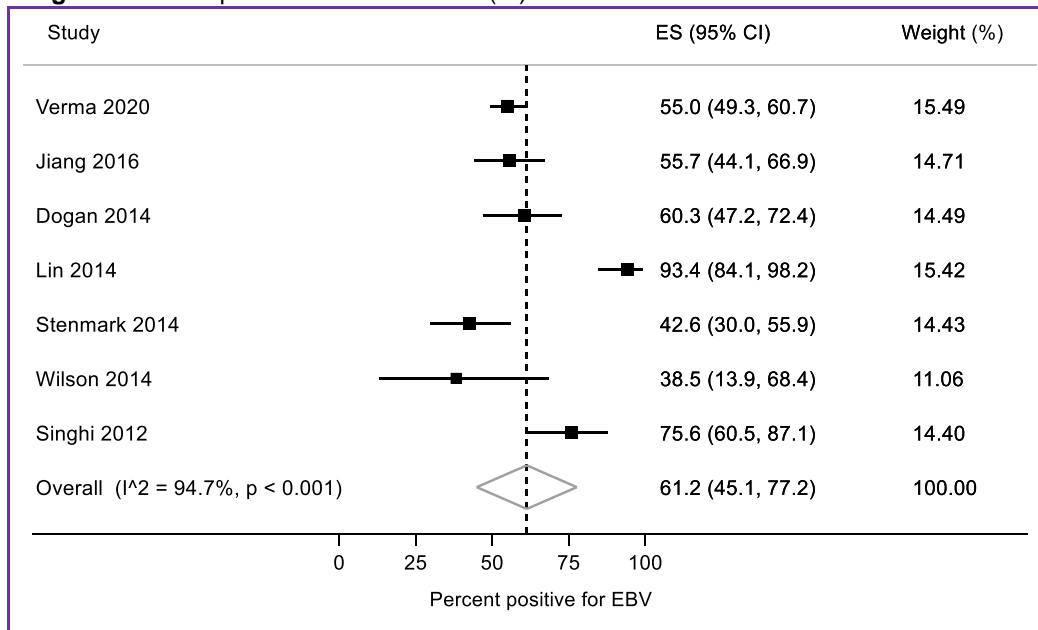
Study ^a	Region	Source of cases	Diagnosis dates	Male %	Mean/median age	Non-keratinizing %	Tested N	Pos %
Studies conducted among adults (aged ≥20 yrs)								
Verma 2020 ¹⁰⁷	New York	Memorial Sloan-Kettering Cancer Center	1998–2017	72.0	52.0	86.0	307	55.0
Jiang 2016 ¹⁰⁸	Texas	M.D. Anderson Cancer Center	2000–2014	70.9	51.4	79.7	79	55.7
Dogan 2014 ¹⁰⁹	Pennsylvania & Washington	University of Pittsburgh Medical Center, Virginia Mason Medical Center	1981–2012	69.8	53.0	85.7	63	60.3
Lin 2014 ¹¹⁰	California	Stanford University	1993–2010	75.4	45	NS	61	93.4
Stenmark 2014 ¹¹¹	Michigan	University of Michigan	1985–2011	65.6	54.3	72.1	61	42.6
Wilson 2014 ¹¹²	Virginia	University of Virginia	2002–2013	NS	NS	76.9	13	38.5
Singhi 2012 ¹¹³	Maryland	Johns Hopkins Hospital	1985–2010	80.0	42.0	100.0	45	75.6
Studies conducted among children (aged 0–19 yrs)								
Polychronopoulou 2004 ¹⁰⁵	Greece	Aghia Sophia Children's Hospital	1987–2001	NS	NS	100.0	8	100.0
Mertens 1997 ¹⁰⁴	Germany	Institute of Pathology, University of Kiel	1992–NS	NS	NS	100.0	8	100.0

EBER ISH = Epstein-Barr encoding region *in situ* hybridization, EBV = Epstein-Barr virus, NPC = nasopharyngeal carcinoma, NS = not specified, yrs = years

^a. Inclusion criteria: tissue specimens from 10 or more cases tested for EBV, EBER ISH detection, conducted in the US, cases aged 15 and older.

^b. EBV positivity was reported for two periods of diagnoses: 1956–1977 and 1981–2012, cases from the first diagnoses period were excluded because they occurred 61 to 40 yrs before year the PAFs were applied to (2017).

eFigure 8. Forest plot of EBV Prevalence (%) in NPC Tumor Tissues Collected From Adults



CI = confidence interval, EBV = Epstein-Barr virus, ES = effect size, I^2 = index of consistency, US = United States

eAppendix 18. Extranodal Natural Killer T-Cell Lymphoma – Nasal Type

EBV is detected in virtually all cases of ENKTL – nasal type and considered part of the diagnostic criteria for that cancer.¹¹⁴⁻¹¹⁷ A study conducted at the University of Texas M.D. Anderson Cancer Center, reported that all 73 ENKTL – nasal type cases identified and tested, were EBER ISH positive.¹¹⁶ All 186 ENKTL – nasal type cases diagnosed in the US in 2017 were attributed to EBV.

eAppendix 19. Diffuse Large B-Cell Lymphoma

DLBCL, the most common subtype of NHL, has an average age of onset of mid-60s.¹¹⁸ Studies meeting the inclusion criteria were published from 1996–2021, and all but one study reported on the HIV or the general immune status of cases thereby allowing us to calculate separate PAFs by HIV status (**eTable 15**). Pooling 13 studies conducted in HIV negative populations and one study (Naeini 2016,⁸⁰ where HIV status was not reported) yielded EBV prevalence of 4.9% (**eFig. 9**). The pooled prevalence of EBV in DLBCLs diagnosed among PWH was substantially higher at 45.7%. Utilizing estimated proportions of DLBCLs occurring in males with HIV (10.4% among those aged 0–29, 15.7% among those 30–59),⁸⁵ we partitioned the cancer incidence data and applied the pooled PAFs (4.9% and then 45.7% for PWH).

eTable 15. Characteristics of Studies Reporting on EBV Prevalence in DLBCL Cases

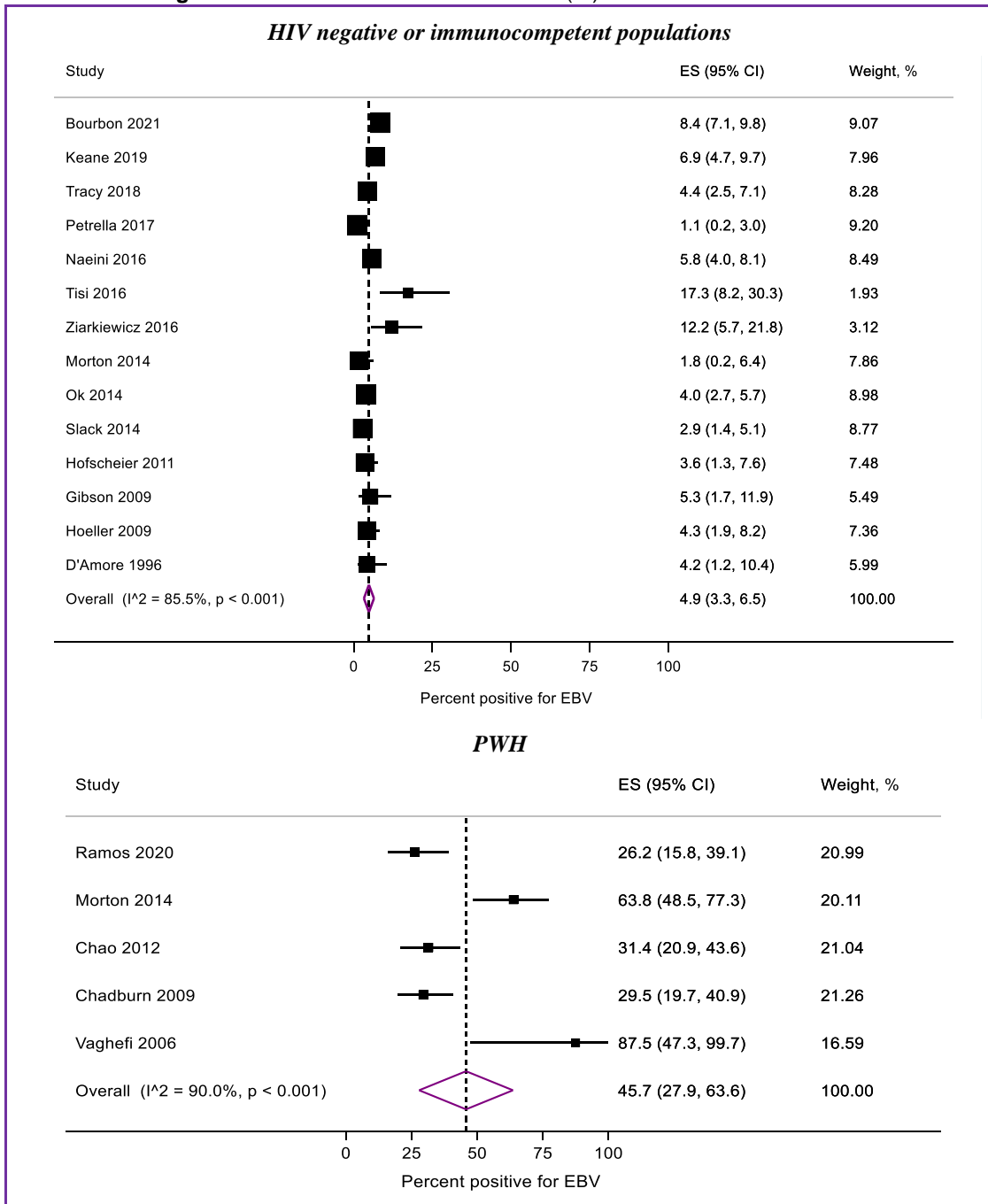
Study ^a	Region(s)	Source of cases	Diagnosis dates	Male %	Mean/median age (yrs, range)	HIV status ^b	EBV positivity cut-off, %	Tested N	Pos %
Studies conducted among immunocompetent or HIV negative populations									
Bourbon 2021 ¹¹⁹	France	Hematopathology Department of Lyon-Sud University Hospital	2006–2019	NS	NS	IC	NS	1645	8.4
Keane 2019 ¹²⁰	Australia & New Zealand	Princess Alexandra Hospital, Canberra Hospital, Royal North Shore Hospital, Australasian Leukaemia and Lymphoma Group Discovery Centre	2003–2014	40.0	NS (18–90)	IC	NS	433	6.9
Tracy 2018 ¹²¹	Iowa & Minnesota	University of Iowa or Mayo Clinic Rochester	2002–2012	59.4	63 (20–89)	HIV-	≥30	362	4.4
Petrella 2017 ¹²²	Belgium, France, Switzerland	Lymphoma Study Association trial LNHO3-6B	2003–2012	68.4	70 (60–80)	HIV-	NS	285	1.1
Naeini 2016 ⁸⁰	California	Clariant Pathology Services/ Neogenomics	2008–2015	57.0	67 (11–96)	NS	≥10	567	5.8
Tisi 2016 ¹²³	Italy	Catholic University of the Sacred Heart, Rome	2006–2013	NS	NS	HIV-	NS	52	17.3
Ziarkiewicz 2016 ¹²⁴	Poland	Medical University of Warsaw	1994–2011	50.0	63.5 (23–86)	IC	>5	74	12.2
Morton 2014 ¹²⁵	California	Los Angeles Residual Tissue Repository	1977–2003	48.3	NS	HIV-	"All or nearly all"	111	1.8
Ok 2014 ¹²⁶	"Western countries"	International DLBCL Rituximab-CHOP Consortium Program Study	NS	57.5	63 (16–95)	HIV-	≥10	703	4.0
Slack 2014 ¹²⁷	Canada	British Columbia Cancer Agency	1999–2006	63.3	64 (16–92)	IC	"Majority of tumor cells"	385	2.9
Hofscheier 2011 ¹²⁸	Germany	Institute of Pathology, Tubingen	2000–2009	NS	72 (51–92)	IC	"Majority of tumor cells"	169	3.6
Gibson 2009 ¹²⁹	Ohio	Department of Clinical Pathology, Cleveland Clinic Pathology at the University Hospitals of Basel, Switzerland; Bologna, Italy; Innsbruck, Austria; & the Triemli Hospital, Zurich, Switzerland	2002–2007	NS	NS (60–NS)	IC	NS	95	5.3
Hoeller 2009 ¹³⁰	Austria, Italy & Switzerland	Department of Clinical Pathology, Cleveland Clinic Pathology at the University Hospitals of Basel, Switzerland; Bologna, Italy; Innsbruck, Austria; & the Triemli Hospital, Zurich, Switzerland	NS	52.5	NS (50–93)	HIV-	≥10	188	4.3
D'Amore 1996 ¹³¹	Denmark	Danish Lymphoma Study Group (LYFO Registry)	1983–NS	NS	NS	IC	NS	95	4.2
Studies conducted among people with HIV									
Ramos 2020 ¹³²	US	Many study sites (Randomized controlled trial)	2012–2017	NS	NS	HIV+	NS	61	26.2
Morton 2014 ¹³	California	Los Angeles Residual Tissue Repository	1977–2003	100.0	NS	HIV+	"All or nearly all"	47	63.8
Chao 2012 ¹³³	California	Kaiser Permanente Southern and Northern California Health Plans	1996–2007	91.4	NS	HIV+	≥75	70	31.4
Chadburn 2009 ¹³⁴	California, Florida, Illinois, Massachusetts, New Jersey, New York, Ohio	Clinical trials AMC010 (45 pts) & AMC034 (36 pts)	NS	86.5	41	HIV+	"Majority of neoplastic cells"	78	29.5
Vaghefi 2006 ¹³⁵	France	NS	1984–2002	NS	NS	HIV+	NS	8	87.5

DLBCL = diffuse large B-cell lymphoma, EBV = Epstein-Barr virus, HIV = human immunodeficiency virus, IC = immunocompetent, NS = not specified, Pos = positive, pts = patients, US = United States, yrs = years

^{a.} Inclusion criteria: tissue specimens from 10 of more cases tested for EBV, EBER ISH detection, conducted in Canada, Europe or the US, cases aged 15 and older.

^{b.} In addition to excluding HIV+ cases, some studies (reported as IC) made additional exclusions based on immune status (e.g., excluding organ transplant recipients).

eFigure 9. Forest Plot of EBV Prevalence (%) in DLBCL Tumor Tissues



CI = confidence interval, DLBCL = diffuse large B-cell lymphoma, EBV = Epstein-Barr virus, ES = effect size, PWH = people with human immunodeficiency virus, I^2 = index of consistency

eAppendix 20. Gastric Carcinoma

The association between EBV and GC was first reported in a case of lymphoepithelial-like gastric carcinoma,¹³⁶ and afterwards, the association was observed in gastric adenocarcinoma.¹³⁷ Since then, several meta-analyses have addressed the prevalence of EBV in GC.¹³⁸⁻¹⁴² The most recent systematic review by Tavakoli and colleagues, including studies from 26 countries, estimated a pooled prevalence of EBV infection (via EBER ISH detection) among GC patients of 8.77% (CI: 7.73–9.92%).¹⁴³

We identified seven studies conducted in the US (**eTable 16**), where the pooled prevalence of EBV was 13.6% for males and 1.9% for females (**eFig. 10**). After combining these calculated PAFs with those for *H. pylori*, the final PAFs were 12.8% (8.3–17.8%) for males and 1.8% (0.3–4.1%) for females.

eTable 16. Characteristics of Studies Reporting on EBV Prevalence in GC Cases

Study ^a	Region	Source of cases	Diagnosis dates	Mean/median age in yrs	Males		Females	
					Tested N	Pos %	Tested N	Pos %
Kim 2019 ¹⁴⁴	New York	Memorial Sloan Kettering Cancer Center	2006–2016	68.0	24	20.8	19	5.3
Ma 2016 ¹⁴⁵	Pennsylvania	University of Pittsburgh Medical Center	2004–2015	73.0	25	24.0	19	5.3
Truong 2009 ¹⁴⁶	Texas	University of Texas M. D. Anderson Cancer Center	1987–2006	EBV+: 60.0 EBV-: 67.0	147	7.5	88	1.1
Grogg 2003 ¹⁴⁷	Minnesota	Mayo Clinic	1990–1998	68.4	69 ^b	5.8	38	0.0
Vo 2002 ¹⁴⁸	Texas, Louisiana, Minnesota	Touro Infirmary, St Luke's Baptist Hospital, Audie Murphy Memorial Veterans Administration Hospital	NS	EBV+: 66.5 EBV-: 68.3	78	14.1	30	0.0
Shibata 1993 ¹⁴⁹	Hawaii	Japan-Hawaii Cancer Study	1965–NS	EBV+: 69.5 EBV-: 69.1	99	14.1	88	5.7
Shibata 1992 ¹³⁷	Los Angeles	LAC+USC Medical Center, Hospital of the Good Samaritan ^c	NS	NS	99	21.2	39	2.6

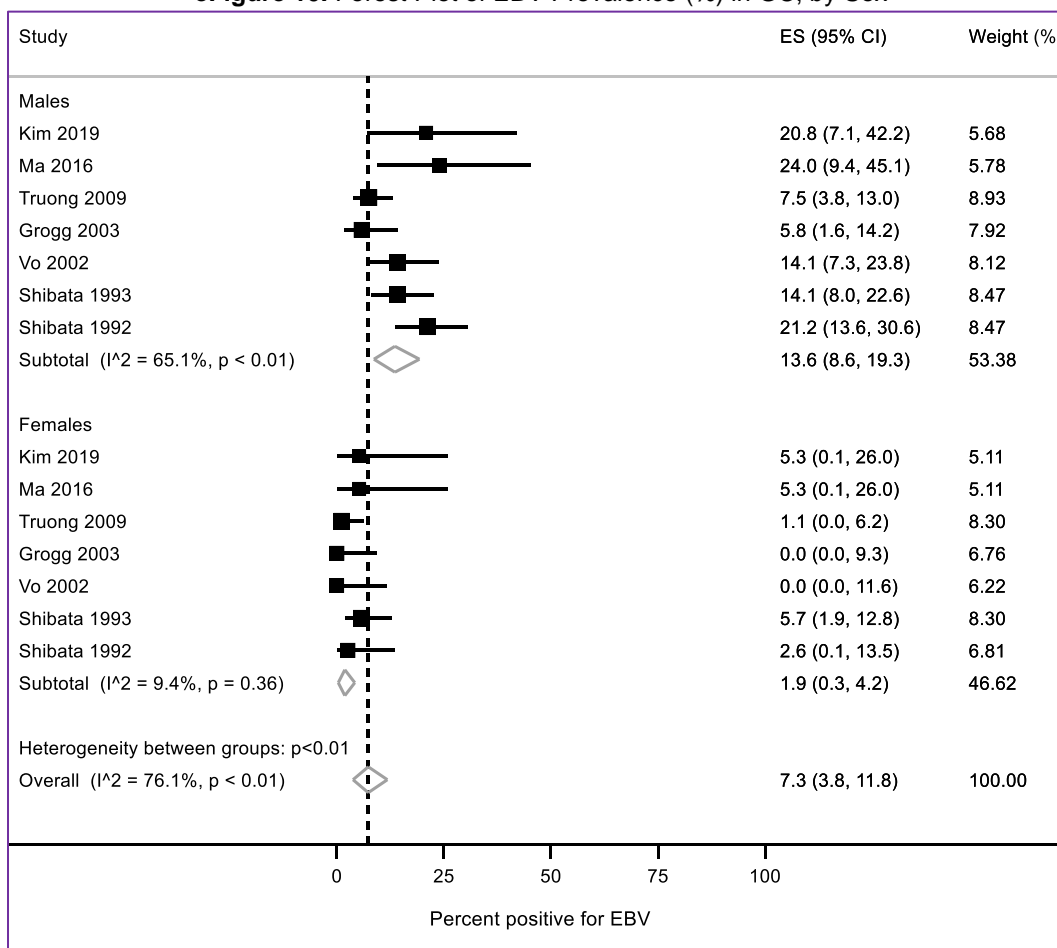
EBER ISH = Epstein-Barr encoding region *in situ* hybridization, EBV = Epstein-Barr virus, GC = gastric carcinoma, NS = not specified, Pos = positive, US = United States, yrs = years

a. Inclusion criteria: tissue specimens from 10 or more cases tested for EBV, EBER ISH detection, conducted in the US, cases aged 15 and older, EBV prevalence reported by sex.

b. Removed three cases of known EBV-positive gastric carcinoma who were added to the series from the consultation files.

c. Cases positive for EBV sequences via PCR were then studied by ISH.

eFigure 10. Forest Plot of EBV Prevalence (%) in GC, by Sex



CI = confidence interval, EBV = Epstein-Barr virus, ES = effect size, GC = gastric carcinoma, I^2 = index of consistency

eAppendix 21. Human Papillomavirus

The most recent monograph (volume 100B) classified HPV types 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58 and 59 as Group 1 carcinogens.⁴ HPV68 is considered ‘probably’ carcinogenic (Group 2A), and several HPV types (26, 30, 34, 53, 66, 67, 69, 70, 73, 82, 85, 97) as ‘possibly’ carcinogenic (Group 2B).⁴ Persistent HPV infection is the strongest risk factor for anal, penile, vaginal and vulvar cancers, with virtually all cervical cancers being caused by HPV infection.¹⁵⁰

eAppendix 22. Anal SCC

After combining five studies (**eTable S17**) that met our inclusion criteria, the pooled prevalence of HR-HPV in anal SCCs was 90.2% for males and 96.3% for females (**eFig. 11**). We found that 100% of anal SCCs among PWH were attributable to HR-HPV. This finding is supported by studies conducted in Europe (not shown); Kreuter and colleagues (2010) found HR-HPV in all nine HIV+ males diagnosed with anal SCCs from 2003 to 2009 in Germany;¹⁵¹ Arana (2015) et al. reported that among 14 HIV+ males and five HIV+ females diagnosed with anal SCC in France from 2007 to 2009, all were HR-HPV+.¹⁵² It has been estimated that 32.5% of anal SCCs in males and 3.0% in females were diagnosed in PWH in the US from 2001 to 2015.¹⁵³ Only two of the five included studies reported HPV results by HIV status; among these two studies, 31.9% of cases were PWH. We assumed that proportion of cases that are PWH in studies where the HIV status of cases was not reported (Herfs 2017,¹⁵⁴ Alemany 2015,¹⁵⁵ Steinau 2013¹⁵⁶) would be similar to that among the two studies (Zhu 2021¹⁵⁷ and Meyer 2013¹⁵⁸) where HIV status was reported. For this reason, we combined all studies/cases to get PAFs for each males and females.

eTable 17. Characteristics of Studies Reporting on HR-HPV Prevalence in Invasive Anal SCCs, by Sex and HIV Status

7.5	Region(s)	Source of cases	Diagnosis dates	Histology	Detection methods HR-HPV types tested for ^b	Specimen	HIV status	Males		Females	
								Tested N	Pos %	Tested N	Pos %
Zhu 2021 ¹⁵⁷	Massachusetts	Pathology archives	2000–2020	SCC	PCR, MGP, HPV GP5/GP6, L1 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68	FFPE	HIV- HIV+	34 12	64.7 100.0	70 0	88.6 NA
Herfs 2017 ¹⁵⁴	Little Rock (Arkansas), Boston (Massachusetts)	Pathology archives	2001–2015	SCC	PCR-RT 16, 18, 31, 33, 34, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68	FFPE	Unknown (14/154 HIV+)	23	91.3	27	88.9
Alemaný 2015 ¹⁵⁵	Multiple	Pathology archives	1999–2009	SCC	SPF-10 PCR, DEIA, LIPA ₂₅ 16, 18, 31, 33, 34, 35, 39, 45, 51, 52, 53, 56, 58, 59, 66, 68, 70, 73	FFPE	Unknown	35	88.6	57	100.0
Meyer 2013 ¹⁵⁸	New York	Surgical pathology files	1997–2009	SCC	SPF-10 PCR, DEIA, LIPA ₂₅ 16, 18, 31, 33, 35, 39, 45, 51, 56, 58, 59, 66, 68, 73	NS	HIV- HIV+	13 10	100.0 100.0	17 2	100.0 100.0
Steinau 2013 ¹⁵⁶	Florida, Hawaii, Iowa, Kentucky, Louisiana, Michigan, California	Cancer registries, tissue repositories	1995–2005	133 SCC, 2 other ^c	PCR, LA, INNO-LiPA 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68	FFPE	Unknown	48	91.7	87	96.6

FFPE = formalin-fixed paraffin-embedded, HIV = human immunodeficiency virus, HPV = human papillomavirus, HR = high-risk, LA = Linear Array, MGP = modified general primer, NA = not applicable, NS = not specified,

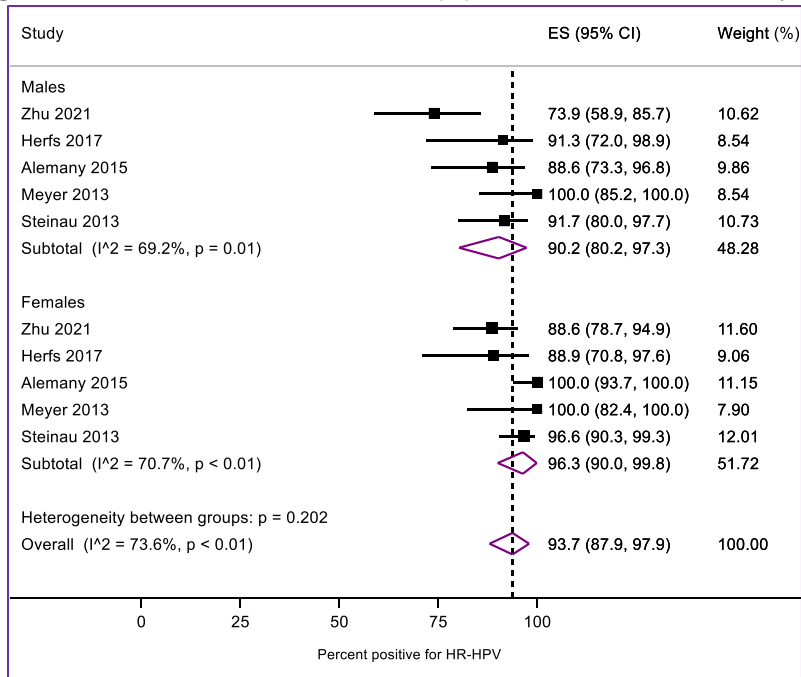
PCR = polymerase chain reaction, Pos = positive, RT = "RealTime", SCC = squamous cell carcinoma, US = United States

^{a.} Inclusion criteria: invasive anal SCC tissue specimens, PCR detection, 10 or more cases, US-based study population, published after 1995, data stratified by sex or available upon request.

^{b.} HR-HPV types included: 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 68, 26, 30, 34, 53, 66, 67, 69, 70, 73, 82, 85, 97.

^{c.} We excluded 11 adenocarcinomas by removing five cases from males and six from females; 2/11 adenocarcinomas were HPV+ and one positive case was removed from each sex.

eFigure 11. Forest Plot of the Prevalence (%) of HR-HPV in Anal SCC, by Sex^a



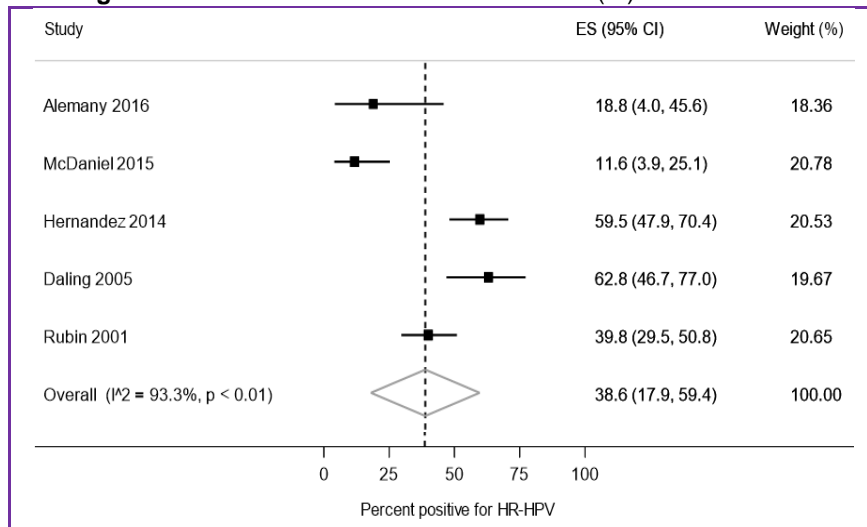
CI = confidence interval, HIV = human immunodeficiency virus, HR-HPV = high-risk human papillomavirus, I^2 = index of consistency, SCC = squamous cell carcinoma

^a The figures include those positive for HIV (e.g., Zhu 2021 included 34 HIV negative and 12 HIV positive males).

eAppendix 23. Penile Cancer

Pooling five studies (eTable S18 and eFig. 12) meeting the inclusion criteria provided a prevalence in cases of 38.6% (CI: 17.9–59.4%).

eFigure 12. Forest Plot of HR-HPV Prevalence (%) in Penile Cancer



CI = confidence interval, ES = effect size, HPV = human papillomavirus, HR = high-risk, I^2 = index of consistency

eTable 18. Characteristics of Studies Reporting on HR-HPV Prevalence in Penile Cancers

Study ^a	Region(s)	Source of cases	Diagnosis dates	Histology	Detection methods HR-HPV types genotyped ^b	Specimen	HIV status	Tested N	Pos %
Alemaný 2016 ¹⁵⁹	Hawaii ^c , Iowa	Pathology archives	1994–2004	NS	SPF-10, DEIA, LiPA ₂₅ , 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59	FFPE	Unknown	16	18.8
McDaniel 2015 ¹⁶⁰	Michigan	Pathology archives	2005–2013	SCC	GP5+/GP6+, MY09/MY11, CP, 16, 33	FFPE	Unknown	43	11.6
Hernandez 2014 ¹⁶¹	California, Florida, Hawaii ^c , Iowa, Kentucky, Louisiana, Michigan	Population-based cancer registries, residual tissue repositories	1998–2005	NS (majority SCC)	PCR, LA, INNO-LiPA ₂₅ , 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68	FFPE	Unknown	79	59.5
Daling 2005 ¹⁶²	Washington	Population-based cancer registry	1979–1998	NS	PCR-MY09/MY11, L1, 16, 18, 31, 33, 35, 45	PE	Unknown	43	62.8
Rubin 2001 ¹⁶³	Connecticut, Michigan, New York, Texas	Pathology archives	NS	SCC	PCR SPF-10, LiPA ₂₅ , 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68, 70	FFPE	Unknown	88	39.8

CP = consensus primers, FFPE = formalin-fixed paraffin-embedded, HIV = human immunodeficiency virus, HPV = human papillomavirus, HR = high-risk, LA = linear array, NS = not specified, PCR = polymerase chain reaction, PE = paraffin-embedded, Pos = positive, SCC = squamous cell carcinoma

^a. Inclusion criteria: invasive penile cancers tissue specimens, PCR detection, 10 or more cases, North American study population, published after 1995.

^b. HR-HPV types included: 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 68, 26, 30, 34, 53, 66, 67, 69, 70, 73, 82, 85, 97.

^a. Three cases overlapped.

eAppendix 24. Vaginal Cancer

Two studies met our inclusion criteria (**eTable S19**). The pooled prevalence of HR-HPV types in invasive vaginal cancers was 72.2% (CI: 62.8–81.7%), where the Sinno 2014 study received a weight of 74.7% and the Daling 2002 study a weight of 25.3% (forest plot not shown).

eTable 19. Characteristics of Studies Reporting on HR-HPV^a Prevalence in Vaginal Cancers

Study ^a	Region(s)	Source of cases	Diagnosis dates	Histology	HIV status	Detection methods HR-HPV types tested ^b	Specimen	Tested N	Pos % (95% CI)
Sinno 2014 ¹⁶⁴	California, Florida, Hawaii, Kentucky, Louisiana, Iowa, Michigan	Population-based cancer registries, residual tissue repositories	1994–2005	NS (86% SCC)	Unknown	LA, INNO-LiPA 16, 18, 31, 33, 35, 39, 45, 51, 52, 53, 56, 58, 59, 66, 67, 68, 70, 73, 82	FFPE	60	75.0 (62.1–85.3)
Daling 2002 ¹⁶⁵	Washington	Population-based cancer registry	1981–1998	SCC	Unknown	PCR-L1, MY09/MY11 16, 18/45, 31	PE	25	64.0 (42.5–82.0)

FFPE = formalin-fixed paraffin-embedded, HIV = human immunodeficiency virus, HPV = human papillomavirus, HR = high-risk, LA = linear array, NS = not specified, PCR = polymerase chain reaction, PE = paraffin-embedded, Pos = positive, SCC = squamous cell carcinoma, US = United States

^a. Inclusion criteria: invasive vaginal cancer tissue specimens, PCR detection, 10 or more cases, North American study population, published after 1995.

^b. HR-HPV types included: 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 68, 26, 30, 34, 53, 66, 67, 69, 70, 73, 82, 85, 97.

eAppendix 25. Vulvar Cancer

Since HPV is more prevalent in vulvar cancers diagnosed among younger women, and vulvar cancer incidence is higher among older women,¹⁶⁶ HR-HPV prevalence was analyzed by age group (**eTable S20**). The pooled prevalence of HR-HPV in cases was 74.4% for women aged <50 years and 45.7% for women aged \geq 50 years old (**eFig. 13**).

eTable 20. Characteristics of Studies Reporting on the Prevalence of HR- HPV in Vulvar Cancer Cases, by Age Group

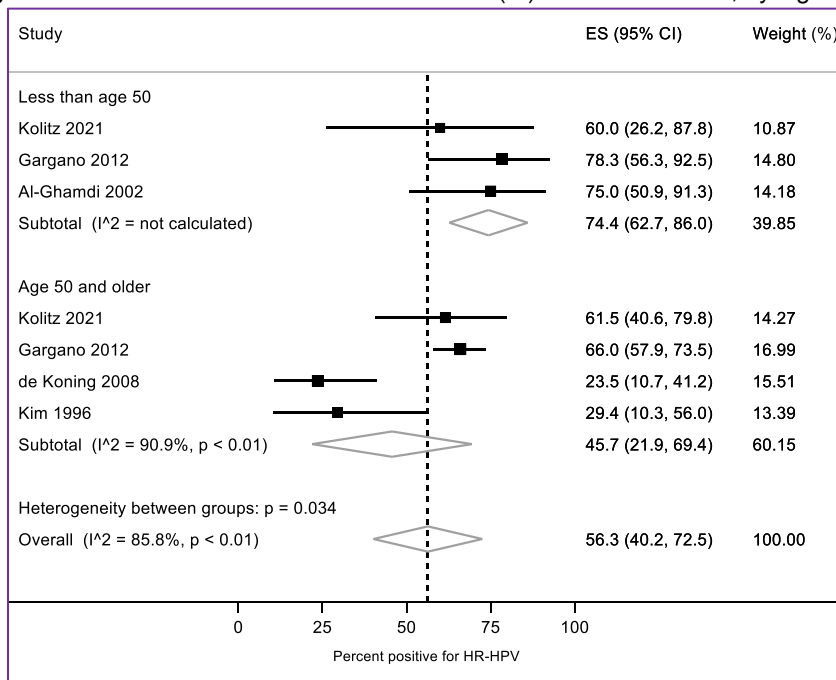
Study ^a	Region(s)	Source of cases	Diagnosis dates	Histology	HIV	Detection methods HR-HPV ^b types tested	Specimen	Age <50 yrs		Age ≥50 yrs	
								Tested N	Pos %	Tested N	Pos %
Kolitz 2021 ¹⁶⁷	Texas	Pathology archives	2010–2020	SCC	None	Consensus PCR-L1, NS	FFPE	10	60.0	26	61.5
Gargano 2012 ¹⁶⁸	California, Florida, Hawaii, Iowa Kentucky, Louisiana, Michigan	Population-based cancer registries, residual tissue repositories	1995–2005	NS	Unknown	LA, INNO-LiPA 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68	FFPE	23	78.3	153	66.0
de Koning 2008 ¹⁶⁹	New York	Pathology department	1990–2005	SCC	Unknown	SPF-10, LiPA ₂₅ 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68, 70	PE	--	--	34	23.5
Al-Ghamdi 2002 ¹⁷⁰	British Columbia, Yukon, Canada	Population-based cancer registry	1970–1998	SCC	One HIV+	PCR-MY09/MY11, PCR-GP5/GP6, TS 16, 18	FFPE	20	75.0	--	--
Kim 1996 ¹⁷¹	Maryland, Florida	Pathology archives	1989–1994	SCC	Unknown	PCR-MY09/MY11, PCR-L1, TS, Sequencing 16, 18	Fresh	--	--	17	29.4

FFPE = formalin-fixed paraffin-embedded, HIV = human immunodeficiency virus, HPV = human papillomavirus, HR = high-risk, LA = linear array, NS = not specified, PE = paraffin-embedded, PCR = polymerase chain reaction, Pos = positive, SCC = squamous cell carcinoma, TS = type-specific, US = United States, yrs = years

^a. Inclusion criteria: invasive vulvar cancers tissue specimens, PCR detection, 10 or more cases, North American study population, published after 1995, data stratified by age or available upon request.

^b. HR-HPV types included: 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 68, 26, 30, 34, 53, 66, 67, 69, 70, 73, 82, 85, 97.

eFigure 13. Forest Plot for HR-HPV Prevalence (%) in Vulvar Cancer, by Age Group



CI = confidence interval, HR-HPV = high-risk human papillomaviruses, I² = index of consistency

eAppendix 26. Head and Neck Cancers

When attributing HNCs to HPV detection of the oncoproteins E6 and E7 is recognized as the gold standard,^{172,173} because they are produced by HR-HPVs and must be present for viral replication to occur. We only considered the prevalence of HPV16 as the association between HNCs and HPV is most established for this type. Twenty-one studies met the inclusion criteria (**eTable S21**). The PAFs were 60.3% for the oropharynx (**eFig. 14**), 7.9% for the oral cavity (**eFig. 15**) and 12.7% for the larynx (**eFig. 16**).

eTable 21. Characteristics of Studies Reporting on HPV16 Prevalence Detected via E6 and/or E7 in HNCs

Study ^a	Region ^b	Diagnosis dates	Detection method(s)	Specimen	Anatomical site					
					Oropharynx		Oral cavity		Larynx	
					Tested N	Pos ^c %	Tested N	Pos ^c %	Tested N	Pos ^c %
Lewis 2021 ¹⁷⁴	Tennessee	2000–2018	qRT-PCR E6/E7	FFPE	259	81.9	--	--	--	--
Mazul 2016 ¹⁷⁵	North Carolina	2002–2006	TS-PCR E7	FFPE	238	63.4	--	--	--	--
Hooper 2015 ¹⁷⁶	Oregon	--	PCR-E6, E7	FF	44	68.2	24	8.3	19	0.0
Zandberg 2015 ¹⁷⁷	Maryland	1992–2007	PCR-E6	FFPE	194	34.5	--	--	--	--
Isayeva 2014 ¹⁷⁸	Alabama	2004–2012	RT-PCR E6/E7	PE	102	48.0	--	--	--	--
Lingen 2013 ¹⁷⁹ ^d	California, Illinois, Ohio, Ontario (CA)	2005–2011	qRT-PCR E6 or 7	FFPE	--	--	409	3.7	--	--
Walline 2013 ¹⁸⁰	Michigan	2001-2011	PCR-E6	FFPE	208	78.8	104	4.8	--	--
Jordan 2012 ¹⁸¹	California, Illinois, Ohio, Ontario (CA)	2000–2009	qPCR E6	FFPE	235	62.1	--	--	--	--
Stephen 2012 ¹⁸²	Michigan	1999–2005	qRT-PCR E6	FFPE	--	--	--	--	77	27.3
Chaturvedi 2011 ¹⁸³	Hawaii, Iowa, Los Angeles, California	1984–2004	qRT-PCR E6	FFPE	216	35.2	--	--	--	--
Schlecht 2011 ¹⁸⁴	New York	NS	TS-PCR E6/E7	FF, PE	23	52.2	29	27.6	27	18.5
Agoston 2010 ¹⁸⁵	Massachusetts	NS	PCR-E7	FFPE	126	58.7	--	--	--	--
Kingma 2010 ¹⁸⁶	Oklahoma & Montana	2005–2007	RT-PCR-E6	FFPE	61	49.2	--	--	--	--
Jo 2009 ¹⁸⁷	California	2000–2003	PCR-E7	FF, FFPE	14	92.9	--	--	--	--
Settle 2009 ¹⁸⁸	Maryland	1995–2006	PCR-E6	PE	--	--	28	10.7	55	7.3
Tezal 2009 ¹⁸⁹	New York	1999–2005	TS-PCR E6	PE	30	70.0	--	--	--	--
Cohen 2008 ¹⁹⁰	Pennsylvania	1996–2001	TS-PCR E7	PE	35	68.6	--	--	--	--
Liang 2008 ¹⁹¹	Minnesota	2004–2006	TS-PCR E6	FF	--	--	51	2.0	--	--
Worden 2008 ¹⁹²	Michigan	NS	RT-PCR E6	NS	42	64.3	--	--	--	--
Zhao 2005 ¹⁹³	Maryland	1984–2002	RT-PCR E6/E7	Frozen	26	57.7	38	15.8	16	18.8
Strome 2002 ¹⁹⁴	Minnesota	1987–1995	TS-PCR E6	PE	52	40.4	--	--	--	--

CA = Canada, FF = fresh-frozen, FFPE = formalin-fixed paraffin embedded, HNCs = head and neck cancers, HPV = human papillomavirus, NS = not specified, PE = paraffin embedded, PCR = polymerase chain reaction, Pos = positive, qRT-PCR = real-time quantitative reverse transcription, RT = real-time, TS = type-specific, US = United states

-- Indicates the cancer was not included in the original study or that it overlapped with another included study.

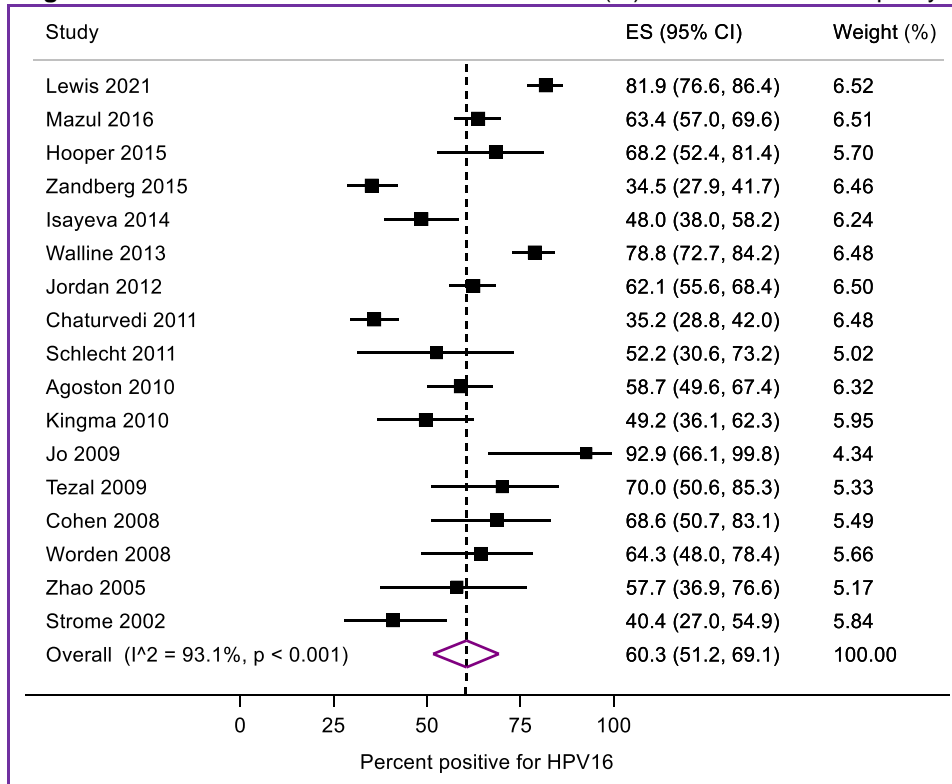
^{a.} Inclusion criteria: site specific results (e.g., base versus oral tongue), detection in cancer tissue, invasive and untreated cancer, detection with E6 and/or E7 for HPV16, did not test specimens for E6/7 based on previous HPV results, North American study population, and published in 2000 or later.

^{b.} Only cases from Chaturvedi et al.'s 2011 study originated from population-based cancer registries, the remaining studies cases came from clinics, hospitals, and pathology departments.

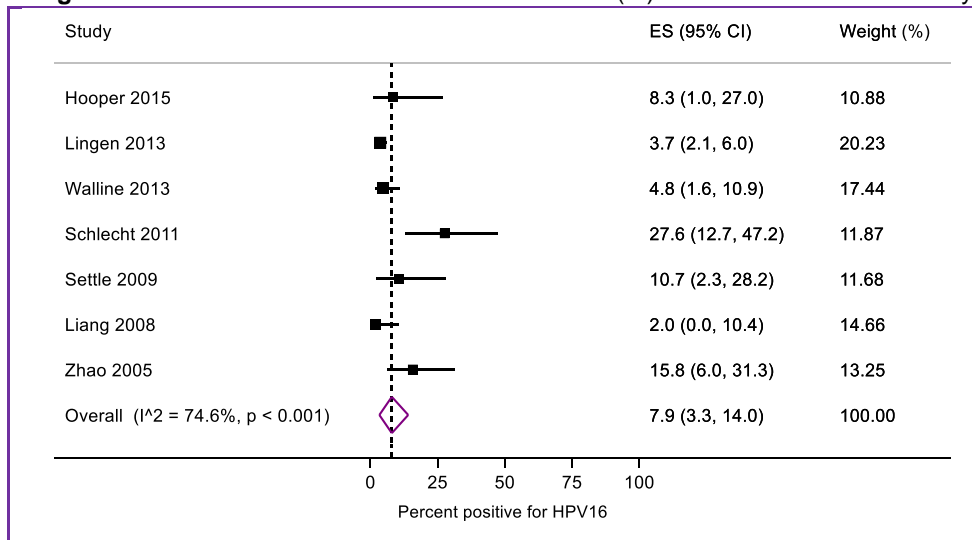
^{c.} Tested positive for E6 and/or E7.

^{d.} Lingen 2013 included four *in situ* cases.

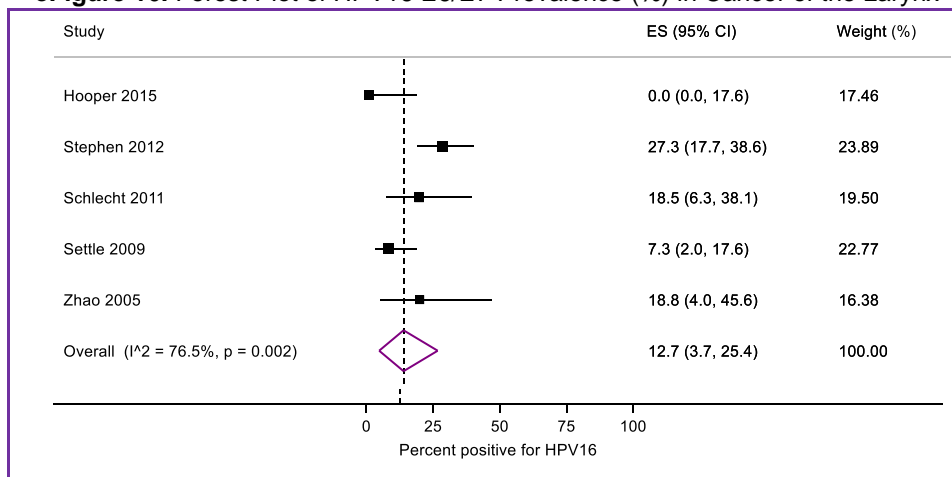
eFigure 14. Forest Plot of HPV16 E6/E7 Prevalence (%) in Cancer of the Oropharynx



eFigure 15. Forest Plot of HPV16 E6/E7 Prevalence (%) in Cancer of the Oral Cavity



eFigure 16. Forest Plot of HPV16 E6/E7 Prevalence (%) in Cancer of the Larynx



CI = confidence interval, ES = effect size, HPV = human papillomavirus, I^2 = index of consistency

eAppendix 27. Merkel Cell Polyomavirus

MCPyV a small double-stranded DNA virus, is named after skin cells called Merkel cells because these skin cells are believed to be the primary target for this virus. The discovery of the MCPyV genome in Merkel cell carcinoma biopsies in 2008,¹⁹⁵ prompted IARC to evaluate MCPyV in February 2012.¹⁹⁶ MCPyV is prevalent but remains dormant for the vast majority of people infected.

eAppendix 28. Merkel Cell Carcinoma of the Skin

This cancer is a rare and aggressive neuroendocrine tumor of the skin. Merkel cell carcinoma is marked by the uncontrolled growth of Merkel cells (cells responsible for sensation). Most often, Merkel cell carcinoma develops in sun-exposed areas of the body. Risk factors for Merkel cell carcinoma include older age, fair skin, history of intensive sun exposure, and a weaker immune system. The eleven studies meeting the inclusion criteria reported MCPyV prevalence in Merkel cell carcinoma ranging from 25.0–100.0% (**eTable S22**). The pooled prevalence of MCPyV in Merkel cell carcinoma was 70.3% (**eFig. 17**).

eTable 22. Characteristics of Studies Reporting on MCPyV Prevalence in Merkel Cell Carcinoma of the Skin

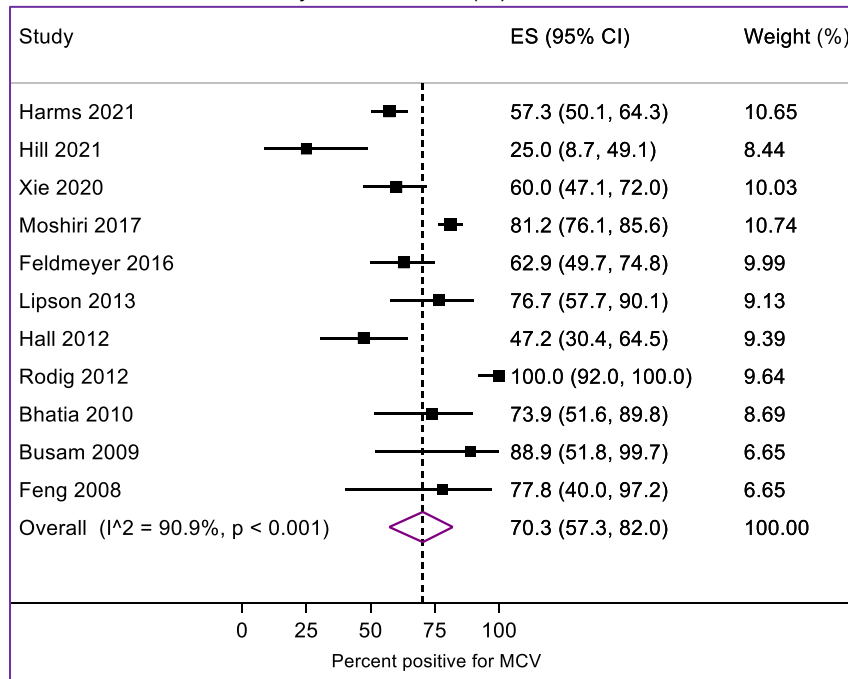
Study ^a	Source of cases	Diagnosis dates	Male %	Median age (in yrs)	Specimen	Detection method(s)	Tested N	Pos %
Harms 2021 ¹⁹⁷	Michigan Medicine Cutaneous Oncology Database; Cutaneous Oncology Program database, and Pathology laboratory information systems	1999–2019	65.8	74	FFPE	IHC (CM2B4), ISH (T, ST, LT), RT qPCR (LT2)	199	57.3
Hill 2021 ¹⁹⁸	University of Pennsylvania and Marshfield clinic	1996–2012	56.3	73	FFPE	Nested qPCR; OneStep RT-PCR system	20	25.0
Xie 2020 ¹⁹⁹	Mayo Clinic	NS	67.7	73	FFPE	IHC (MCPyV large T-antigen)	65	60.0
Moshiri 2017 ²⁰⁰	University of Washington	1980–2015	62.8	71	FFPE	IHC (CM2B4 and Ab3), RT qPCR (Large T, Small T, VP 2)	282 ^b	81.2
Feldmeyer 2016 ²⁰¹	University of Texas MD Anderson Cancer Center	2002–2015	72.6	71	FFPE	IHC (MCPyV T-antigen)	62	62.9
Lipson 2013 ²⁰²	Johns Hopkins Hospital surgical pathology archives	2000–2011	65.3	65	FFPE	RT qPCR (VP1, LT3)	30	76.7
Hall 2012 ²⁰³	University of California, San Francisco Dermatopathology Service	1996–2010	41.7	77	NS	IHC (CM2B4)	36	47.2
Rodig 2012 ²⁰⁴	Referral specialty clinic	NS	63.6	74	FFPE	RT qPCR (LT2, Set 6, 7, 9, LT3)	44	100.0
Bhatia 2010 ²⁰⁵	Ohio State University Medical Center	1994–2007	73.9	77	FFPE	qPCR (MCPyV, EU375804)	23	73.9
Busam 2009 ²⁰⁶	Memorial Sloan-Kettering Cancer Center	NS	66.7	80	Frozen	RT qPCR (T primer sets)	9	88.9
Feng 2008 ¹⁹⁵	Cooperative Human Tissue Network (US)	NS	88.9	58	Frozen	PCR (VP1), nested PCR (VP1-2), PCR-Southern hybridization	9	77.8

FFPE = formalin-fixed paraffin embedded, HPV = human papillomavirus, IHC = immunohistochemistry, ISH = *in situ* hybridization, MCPyV = Merkel cell polyomavirus, NS = not specified, PCR = polymerase chain reaction, Pos = positive, qRT-PCR = real-time quantitative reverse transcription, RT = real-time, US = United states, yrs = years

^{a.} Inclusion criteria: Merkel cell carcinoma tissues (adjacent tissues excluded) arising from ≥8 cancer cases, results presented per person not tissue (if multiple specimens collected), detection via PCR, IHC and/or ISH, US study population.

^{b.} Included 40 recurrent cases.

eFigure 17. Forest Plot of MCPyV Prevalence (%) in Merkel Cell Carcinoma of the Skin



CI = confidence interval, ES = effect size, I^2 = index of consistency, MCPyV = Merkel cell polyomavirus

eReferences

1. De Silva AP, De Livera AM, Lee KJ, Moreno-Betancur M, Simpson JA. Multiple imputation methods for handling missing values in longitudinal studies with sampling weights: Comparison of methods implemented in Stata. *Biom J.* 2021;63(2):354-371. doi:10.1002/bimj.201900360
2. National Center for Health Statistics (NCHS). Module 4: Variance Estimation. 2021. <https://wwwn.cdc.gov/nchs/nhanes/tutorials/module4.aspx>
3. Johnson CL, Paulose-Ram R, Ogden CL, et al. National health and nutrition examination survey: analytic guidelines, 1999-2010. *Vital Health Stat 2.* 2013;(161):1-24.
4. International Agency for Research on Cancer. *Biological Agents.* Vol. 100 B. 2012. *IARC Monographs on the Evaluation of Carcinogenic Risks to Humans.*
5. Zhao LH, Liu X, Yan HX, et al. Genomic and oncogenic preference of HBV integration in hepatocellular carcinoma. *Nat Commun.* 2016;7:12992. doi:10.1038/ncomms12992
6. Hassan MM, Spitz MR, Thomas MB, et al. The association of family history of liver cancer with hepatocellular carcinoma: a case-control study in the United States. *J Hepatol.* 2009;50(2):334-41. doi:10.1016/j.jhep.2008.08.016
7. Ognjanovic S, Yuan JM, Chaptman AK, Fan Y, Yu MC. Genetic polymorphisms in the cytokine genes and risk of hepatocellular carcinoma in low-risk non-Asians of USA. *Carcinogenesis.* 2009;30(5):758-62. doi:10.1093/carcin/bgn286
8. Davila JA, Morgan RO, Shaib Y, McGlynn KA, El-Serag HB. Diabetes increases the risk of hepatocellular carcinoma in the United States: a population based case control study. *Gut.* 2005;54(4):533-9. doi:10.1136/gut.2004.052167
9. Hassan MM, Hwang LY, Hatten CJ, et al. Risk factors for hepatocellular carcinoma: synergism of alcohol with viral hepatitis and diabetes mellitus. *Hepatology.* 2002;36(5):1206-13. doi:10.1053/jhep.2002.36780
10. Nomura A, Stemmermann GN, Chyou PH, Tabor E. Hepatitis B and C virus serologies among Japanese Americans with hepatocellular carcinoma. *J Infect Dis.* 1996;173(6):1474-6. doi:10.1093/infdis/173.6.1474
11. Di Bisceglie AM, Order SE, Klein JL, et al. The role of chronic viral hepatitis in hepatocellular carcinoma in the United States. *Am J Gastroenterol.* 1991;86(3):335-8.
12. Miettinen OS. Proportion of disease caused or prevented by a given exposure, trait or intervention. *Am J Epidemiol.* 1974;99(5):325-32.
13. Morton LM, Slager SL, Cerhan JR, et al. Etiologic heterogeneity among non-Hodgkin lymphoma subtypes: the InterLymph Non-Hodgkin Lymphoma Subtypes Project. *J Natl Cancer Inst Monogr.* 2014;2014(48):130-44. doi:10.1093/jncimonographs/lgu013
14. Anderson LA, Pfeiffer R, Warren JL, et al. Hematopoietic malignancies associated with viral and alcoholic hepatitis. *Cancer Epidemiol Biomarkers Prev.* 2008;17(11):3069-75. doi:10.1158/1055-9965.EPI-08-0408
15. Dal Maso L, Franceschi S. Hepatitis C virus and risk of lymphoma and other lymphoid neoplasms: a meta-analysis of epidemiologic studies. *Cancer Epidemiol Biomarkers Prev.* 2006;15(11):2078-85. doi:10.1158/1055-9965.EPI-06-0308
16. Morton LM, Sampson JN, Cerhan JR, et al. Rationale and Design of the International Lymphoma Epidemiology Consortium (InterLymph) Non-Hodgkin Lymphoma Subtypes Project. *J Natl Cancer Inst Monogr.* 2014;2014(48):1-14. doi:10.1093/jncimonographs/lgu005
17. Mbulaiteye SM, Morton LM, Sampson JN, et al. Medical history, lifestyle, family history, and occupational risk factors for sporadic Burkitt lymphoma/leukemia: the Interlymph Non-Hodgkin Lymphoma Subtypes Project. *J Natl Cancer Inst Monogr.* 2014;2014(48):106-14. doi:10.1093/jncimonographs/lgu003

18. Slager SL, Benavente Y, Blair A, et al. Medical history, lifestyle, family history, and occupational risk factors for chronic lymphocytic leukemia/small lymphocytic lymphoma: the InterLymph Non-Hodgkin Lymphoma Subtypes Project. *J Natl Cancer Inst Monogr.* 2014;2014(48):41-51. doi:10.1093/jncimonographs/lgu001
19. Cerhan JR, Krickler A, Paltiel O, et al. Medical history, lifestyle, family history, and occupational risk factors for diffuse large B-cell lymphoma: the InterLymph Non-Hodgkin Lymphoma Subtypes Project. *J Natl Cancer Inst Monogr.* 2014;2014(48):15-25. doi:10.1093/jncimonographs/lgu010
20. Vajdic CM, Landgren O, McMaster ML, et al. Medical history, lifestyle, family history, and occupational risk factors for lymphoplasmacytic lymphoma/Waldenstrom's macroglobulinemia: the InterLymph Non-Hodgkin Lymphoma Subtypes Project. *J Natl Cancer Inst Monogr.* 2014;2014(48):87-97. doi:10.1093/jncimonographs/lgu002
21. Bracci PM, Benavente Y, Turner JJ, et al. Medical history, lifestyle, family history, and occupational risk factors for marginal zone lymphoma: the InterLymph Non-Hodgkin Lymphoma Subtypes Project. *J Natl Cancer Inst Monogr.* 2014;2014(48):52-65. doi:10.1093/jncimonographs/lgu011
22. Petrick JL, Yang B, Altekruse SF, et al. Risk factors for intrahepatic and extrahepatic cholangiocarcinoma in the United States: A population-based study in SEER-Medicare. *PLoS One.* 2017;12(10):e0186643. doi:10.1371/journal.pone.0186643
23. Choi J, Ghooz HM, Peeraphatdit T, et al. Aspirin use and the risk of cholangiocarcinoma. *Hepatology.* 2016;64(3):785-96. doi:10.1002/hep.28529
24. Shaib YH, El-Serag HB, Nooka AK, et al. Risk factors for intrahepatic and extrahepatic cholangiocarcinoma: a hospital-based case-control study. *Am J Gastroenterol.* 2007;102(5):1016-21. doi:10.1111/j.1572-0241.2007.01104.x
25. Shaib YH, El-Serag HB, Davila JA, Morgan R, McGlynn KA. Risk factors of intrahepatic cholangiocarcinoma in the United States: a case-control study. *Gastroenterology.* 2005;128(3):620-6. doi:10.1053/j.gastro.2004.12.048
26. Hooi JKY, Lai WY, Ng WK, et al. Global Prevalence of *Helicobacter pylori* Infection: Systematic Review and Meta-analysis. *Gastroenterology.* 2017;doi:10.1053/j.gastro.2017.04.022
27. Peleteiro B, Bastos A, Ferro A, Lunet N. Prevalence of *Helicobacter pylori* infection worldwide: a systematic review of studies with national coverage. *Dig Dis Sci.* 2014;59(8):1698-709. doi:10.1007/s10620-014-3063-0
28. Yang L, Kartsonaki C, Yao P, et al. The relative and attributable risks of cardia and non-cardia gastric cancer associated with *Helicobacter pylori* infection in China: a case-cohort study. *Lancet Public Health.* 2021;6(12):e888-e896. doi:10.1016/S2468-2667(21)00164-X
29. Colquhoun A, Arnold M, Ferlay J, Goodman KJ, Forman D, Soerjomataram I. Global patterns of cardia and non-cardia gastric cancer incidence in 2012. *Gut.* 2015;64(12):1881-8. doi:10.1136/gutjnl-2014-308915
30. Cavaleiro-Pinto M, Peleteiro B, Lunet N, Barros H. *Helicobacter pylori* infection and gastric cardia cancer: systematic review and meta-analysis. *Cancer Causes Control.* 2011;22(3):375-87. doi:10.1007/s10552-010-9707-2
31. Hansen S, Vollset SE, Derakhshan MH, et al. Two distinct aetiologies of cardia cancer; evidence from premorbid serological markers of gastric atrophy and *Helicobacter pylori* status. *Gut.* 2007;56(7):918-25. doi:10.1136/gut.2006.114504
32. Lochhead P, El-Omar EM. *Helicobacter pylori* infection and gastric cancer. *Best Pract Res Clin Gastroenterol.* 2007;21(2):281-97. doi:10.1016/j.bpg.2007.02.002
33. Plummer M, Franceschi S, Vignat J, Forman D, de Martel C. Global burden of gastric cancer attributable to *Helicobacter pylori*. *Int J Cancer.* 2015;136(2):487-90. doi:10.1002/ijc.28999

34. Gonzalez CA, Megraud F, Buissonniere A, et al. Helicobacter pylori infection assessed by ELISA and by immunoblot and noncardia gastric cancer risk in a prospective study: the Eurgast-EPIC project. *Ann Oncol*. 2012;23(5):1320-4. doi:10.1093/annonc/mdr384
35. Mitchell H, English DR, Elliott F, et al. Immunoblotting using multiple antigens is essential to demonstrate the true risk of Helicobacter pylori infection for gastric cancer. *Aliment Pharmacol Ther*. 2008;28(7):903-10. doi:10.1111/j.1365-2036.2008.03792.x
36. Franco EL. Measurement errors in epidemiological studies of human papillomavirus and cervical cancer. In: Muñoz NB, F.X.; Meheus, A., ed. *The Epidemiology of Human Papillomavirus and Cervical Cancer*. International Agency for Research on Cancer; 1992.
37. Persson C, Jia Y, Pettersson H, Dillner J, Nyrén O, W Y. H. pylori Seropositivity before Age 40 and Subsequent Risk of Stomach Cancer: A Glimpse of the True Relationship? *PLoS One*. 2011;6(3):e17404.
38. Knekt P, Teppo L, Aromaa A, Rissanen H, Kosunen TU. Helicobacter pylori IgA and IgG antibodies, serum pepsinogen I and the risk of gastric cancer: changes in the risk with extended follow-up period. *Int J Cancer*. 2006;119(3):702-5. doi:10.1002/ijc.21884
39. Nomura AM, Lee J, Stemmermann GN, Nomura RY, Perez-Perez GI, Blaser MJ. Helicobacter pylori CagA seropositivity and gastric carcinoma risk in a Japanese American population. *J Infect Dis*. 2002;186(8):1138-44. doi:10.1086/343808
40. Parsonnet J, Samloff IM, Nelson LM, Orentreich N, Vogelmann JH, Friedman GD. Helicobacter pylori, pepsinogen, and risk for gastric adenocarcinoma. *Cancer Epidemiol Biomarkers Prev*. 1993;2(5):461-6.
41. Siman JH, Engstrand L, Berglund G, Forsgren A, Floren CH. Helicobacter pylori and CagA seropositivity and its association with gastric and oesophageal carcinoma. *Scand J Gastroenterol*. 2007;42(8):933-40. doi:10.1080/00365520601173863
42. Canadian Cancer Society. MALT lymphoma. Accessed April 4 2023, <http://www.cancer.ca/en/cancer-information/cancer-type/non-hodgkin-lymphoma/non-hodgkin-lymphoma/malt-lymphoma/?region=bc>
43. Zullo A, Hassan C, Cristofari F, et al. Effects of Helicobacter pylori eradication on early stage gastric mucosa-associated lymphoid tissue lymphoma. *Clin Gastroenterol Hepatol*. 2010;8(2):105-10. doi:10.1016/j.cgh.2009.07.017
44. Zucca E, Arcaini L, Buske C, et al. Marginal zone lymphomas: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2020;31(1):17-29. doi:10.1016/j.annonc.2019.10.010
45. Ladetto M, Dreyling M. EHA Endorsement of ESMO Clinical Practice Guidelines for Marginal Zone Lymphomas. *Hemasphere*. 2020;4(2):e351. doi:10.1097/HS9.0000000000000351
46. Jung K, Kim DH, Seo HI, Gong EJ, Bang CS. Efficacy of eradication therapy in Helicobacter pylori-negative gastric mucosa-associated lymphoid tissue lymphoma: A meta-analysis. *Helicobacter*. 2021;26(2):e12774. doi:10.1111/hel.12774
47. Parsonnet J, Hansen S, Rodriguez L, et al. Helicobacter pylori infection and gastric lymphoma. *N Engl J Med*. 1994;330(18):1267-71. doi:10.1056/NEJM199405053301803
48. Zhang Y. Epidemiology of esophageal cancer. *World J Gastroenterol*. 2013;19(34):5598-606. doi:10.3748/wjg.v19.i34.5598
49. Patel N, Benipal B. Incidence of Esophageal Cancer in the United States from 2001-2015: A United States Cancer Statistics Analysis of 50 States. *Cureus*. 2018;10(12):e3709. doi:10.7759/cureus.3709
50. Whiteman DC, Parmar P, Fahey P, et al. Association of Helicobacter pylori infection with reduced risk for esophageal cancer is independent of environmental and genetic modifiers. *Gastroenterology*. 2010;139(1):73-83; quiz e11-2. doi:10.1053/j.gastro.2010.04.009

51. Gao H, Li L, Zhang C, et al. Systematic Review with Meta-analysis: Association of *Helicobacter pylori* Infection with Esophageal Cancer. *Gastroenterol Res Pract*. 2019;2019:1953497. doi:10.1155/2019/1953497
52. Rokkas T, Sechopoulos P, Pistiolas D, Kothonas F, Margantinis G, Koukoulis G. The relationship of *Helicobacter pylori* infection and colon neoplasia, on the basis of meta-analysis. *Eur J Gastroenterol Hepatol*. 2013;25(11):1286-94. doi:10.1097/MEG.0b013e328363d3cd
53. Islami F, Kamangar F. *Helicobacter pylori* and esophageal cancer risk: a meta-analysis. *Cancer Prev Res (Phila)*. 2008;1(5):329-38. doi:10.1158/1940-6207.CAPR-08-0109
54. Zhuo X, Zhang Y, Wang Y, Zhuo W, Zhu Y, Zhang X. *Helicobacter pylori* infection and oesophageal cancer risk: association studies via evidence-based meta-analyses. *Clin Oncol (R Coll Radiol)*. 2008;20(10):757-62. doi:10.1016/j.clon.2008.07.005
55. Xie FJ, Zhang YP, Zheng QQ, et al. *Helicobacter pylori* infection and esophageal cancer risk: an updated meta-analysis. *World J Gastroenterol*. 2013;19(36):6098-107. doi:10.3748/wjg.v19.i36.6098
56. Nie S, Chen T, Yang X, Huai P, Lu M. Association of *Helicobacter pylori* infection with esophageal adenocarcinoma and squamous cell carcinoma: a meta-analysis. *Dis Esophagus*. 2014;27(7):645-53. doi:10.1111/dote.12194
57. de Martel C, Llosa AE, Farr SM, et al. *Helicobacter pylori* infection and the risk of development of esophageal adenocarcinoma. *J Infect Dis*. 2005;191(5):761-7. doi:10.1086/427659
58. El-Omar EM, Oien K, El-Nujumi A, et al. *Helicobacter pylori* infection and chronic gastric acid hyposecretion. *Gastroenterology*. 1997;113(1):15-24. doi:10.1016/s0016-5085(97)70075-1
59. Polyzos SA, Zeglinas C, Artemaki F, et al. *Helicobacter pylori* infection and esophageal adenocarcinoma: a review and a personal view. *Ann Gastroenterol*. 2018;31(1):8-13. doi:10.20524/aog.2017.0213
60. Gall A, Fero J, McCoy C, et al. Bacterial Composition of the Human Upper Gastrointestinal Tract Microbiome Is Dynamic and Associated with Genomic Instability in a Barrett's Esophagus Cohort. *PLoS One*. 2015;10(6):e0129055. doi:10.1371/journal.pone.0129055
61. Holleczer B, Schotker B, Brenner H. *Helicobacter pylori* infection, chronic atrophic gastritis and risk of stomach and esophagus cancer: Results from the prospective population-based ESTHER cohort study. *Int J Cancer*. 2020;146(10):2773-2783. doi:10.1002/ijc.32610
62. Fruh M, Zhou W, Zhai R, et al. Polymorphisms of inflammatory and metalloproteinase genes, *Helicobacter pylori* infection and the risk of oesophageal adenocarcinoma. *Br J Cancer*. 2008;98(4):689-92. doi:10.1038/sj.bjc.6604234
63. Vicari JJ, Peek RM, Falk GW, et al. The seroprevalence of cagA-positive *Helicobacter pylori* strains in the spectrum of gastroesophageal reflux disease. *Gastroenterology*. 1998;115(1):50-7. doi:10.1016/s0016-5085(98)70364-6
64. Chow WH, Blaser MJ, Blot WJ, et al. An inverse relation between cagA+ strains of *Helicobacter pylori* infection and risk of esophageal and gastric cardia adenocarcinoma. *Cancer Res*. 1998;58(4):588-90.
65. Weston AP, Badr AS, Topalovski M, Cherian R, Dixon A, Hassanein RS. Prospective evaluation of the prevalence of gastric *Helicobacter pylori* infection in patients with GERD, Barrett's esophagus, Barrett's dysplasia, and Barrett's adenocarcinoma. *Am J Gastroenterol*. 2000;95(2):387-94. doi:10.1111/j.1572-0241.2000.01758.x
66. El-Omar EM, Rabkin CS, Gammon MD, et al. Increased risk of noncardia gastric cancer associated with proinflammatory cytokine gene polymorphisms. *Gastroenterology*. 2003;124(5):1193-201. doi:10.1016/s0016-5085(03)00157-4

67. Anandasabapathy S, Jhamb J, Davila M, Wei C, Morris J, Bresalier R. Clinical and endoscopic factors predict higher pathologic grades of Barrett dysplasia. *Cancer*. 2007;109(4):668-74. doi:10.1002/cncr.22451
68. Wu AH, Crabtree JE, Bernstein L, et al. Role of *Helicobacter pylori* CagA+ strains and risk of adenocarcinoma of the stomach and esophagus. *Int J Cancer*. 2003;103(6):815-21. doi:10.1002/ijc.10887
69. Oberg S, Peters JH, Nigro JJ, et al. *Helicobacter pylori* is not associated with the manifestations of gastroesophageal reflux disease. *Arch Surg*. 1999;134(7):722-6. doi:10.1001/archsurg.134.7.722
70. Peek RM, Jr., Vaezi MF, Falk GW, et al. Role of *Helicobacter pylori* cagA(+) strains and specific host immune responses on the development of premalignant and malignant lesions in the gastric cardia. *Int J Cancer*. 1999;82(4):520-4. doi:10.1002/(sici)1097-0215(19990812)82:4<520::aid-ijc9>3.0.co;2-7
71. Khan G, Hashim MJ. Global burden of deaths from Epstein-Barr virus attributable malignancies 1990-2010. *Infect Agent Cancer*. 2014;9(1):38. doi:10.1186/1750-9378-9-38
72. Gulley ML, Tang W. Laboratory assays for Epstein-Barr virus-related disease. *J Mol Diagn*. 2008;10(4):279-92. doi:10.2353/jmoldx.2008.080023
73. Richter J, John K, Staiger AM, et al. Epstein-Barr virus status of sporadic Burkitt lymphoma is associated with patient age and mutational features. *Br J Haematol*. 2021;196(3):681-689. doi:10.1111/bjh.17874
74. Dupont Harwood C, Eriksen PRG, Clasen-Linde E, et al. Clinicopathologic characteristics of Burkitt lymphoma of the head and neck in a non-endemic region-a Danish nationwide study. *Acta Oto Laryngologica*. 2021;
75. Mbulaiteye SM, Pullarkat ST, Nathwani BN, et al. Epstein-Barr virus patterns in US Burkitt lymphoma tumors from the SEER residual tissue repository during 1979-2009. *APMIS*. 2014;122(1):5-15. doi:10.1111/apm.12078
76. Kasprzak A, Spachacz R, Wachowiak J, Stefanska K, Zabel M. Epstein-Barr virus (EBV) infection in B-cell non-Hodgkin's lymphomas in children: Virus latency and its correlation with CD21 and CD23 molecules. 2007;45(3):169-179.
77. Karajannis MA, Hummel M, Oschlies I, et al. Epstein-Barr virus infection and viral gene expression in pediatric non-Hodgkin lymphomas. In: Stathopoulou FT, ed. *Genome and Proteome in Oncology*. Nova Biomedical; 2005:297-325:chap 16.
78. Teitell MA, Lones MA, Perkins SL, Sanger WG, Cairo MS, Said JW. TCL1 expression and Epstein-Barr virus status in pediatric Burkitt lymphoma. 124(4):569-575.
79. Haralambieva E, Schuurung E, Rosati S, et al. Interphase Fluorescence In Situ Hybridization for Detection of 8q24/MYC Breakpoints on Routine Histologic Sections: Validation in Burkitt Lymphomas from Three Geographic Regions. *Genes Chromosomes and Cancer*. 2004;40(1):10-18.
80. Naeini YB, Wu A, O'Malley DP. Aggressive B-cell lymphomas: frequency, immunophenotype, and genetics in a reference laboratory population. *Ann Diagn Pathol*. 2016;25:7-14. doi:10.1016/j.anndiagpath.2016.07.008
81. Lenze D, Leoncini L, Hummel M, et al. The different epidemiologic subtypes of Burkitt lymphoma share a homogenous micro RNA profile distinct from diffuse large B-cell lymphoma. 25(12):1869-1876.
82. Hansen PB, Penkowa M, Kirk O, et al. Human immunodeficiency virus-associated malignant lymphoma in eastern Denmark diagnosed from 1990-1996: clinical features, histopathology, and association with Epstein-Barr virus and human herpesvirus-8. *Eur J Haematol*. 2000;64(6):368-75. doi:10.1034/j.1600-0609.2000.90126.x

83. Davi F, Delecluse HJ, Guet P, et al. Burkitt-like lymphomas in AIDS patients: characterization within a series of 103 human immunodeficiency virus-associated non-Hodgkin's lymphomas. Burkitt's Lymphoma Study Group. *J Clin Oncol.* 1998;16(12):3788-95. doi:10.1200/JCO.1998.16.12.3788
84. Spina M, Tirelli U, Zagonel V, et al. Burkitt's lymphoma in adults with and without human immunodeficiency virus infection: a single-institution clinicopathologic study of 75 patients. *Cancer.* 1998;82(4):766-74.
85. Shiels MS, Pfeiffer RM, Hall HI, et al. Proportions of Kaposi sarcoma, selected non-Hodgkin lymphomas, and cervical cancer in the United States occurring in persons with AIDS, 1980-2007. *JAMA.* 2011;305(14):1450-9. doi:10.1001/jama.2011.396
86. Glaser SL, Clarke CA, Gulley ML, et al. Population-based patterns of human immunodeficiency virus-related Hodgkin lymphoma in the Greater San Francisco Bay Area, 1988-1998. *Cancer.* 2003;98(2):300-9. doi:10.1002/cncr.11459
87. Thompson LD, Fisher SI, Chu WS, Nelson A, Abbondanzo SL. HIV-associated Hodgkin lymphoma: a clinicopathologic and immunophenotypic study of 45 cases. *Am J Clin Pathol.* 2004;121(5):727-38. doi:10.1309/PNVQ-0PQG-XHVY-6L7G
88. Besson C, Lancar R, Prevot S, et al. High Risk Features Contrast With Favorable Outcomes in HIV-associated Hodgkin Lymphoma in the Modern cART Era, ANRS CO16 LYMPHOVIR Cohort. *Clin Infect Dis.* 2015;61(9):1469-75. doi:10.1093/cid/civ627
89. Hentrich M, Berger M, Wyen C, et al. Stage-adapted treatment of HIV-associated Hodgkin lymphoma: results of a prospective multicenter study. *J Clin Oncol.* 2012;30(33):4117-23. doi:10.1200/JCO.2012.41.8137
90. Carbone A, Gloghini A, Larocca LM, et al. Human immunodeficiency virus-associated Hodgkin's disease derives from post-germinal center B cells. *Blood.* 1999;93(7):2319-26.
91. Tirelli U, Errante D, Dolcetti R, et al. Hodgkin's disease and human immunodeficiency virus infection: clinicopathologic and virologic features of 114 patients from the Italian Cooperative Group on AIDS and Tumors. *J Clin Oncol.* 1995;13(7):1758-67. doi:10.1200/JCO.1995.13.7.1758
92. Shiels MS, Koritzinsky EH, Clarke CA, Suneja G, Morton LM, Engels EA. Prevalence of HIV Infection among U.S. Hodgkin lymphoma cases. *Cancer Epidemiol Biomarkers Prev.* 2014;23(2):274-81. doi:10.1158/1055-9965.EPI-13-0865
93. Linabery AM, Erhardt EB, Richardson MR, et al. Family history of cancer and risk of pediatric and adolescent Hodgkin lymphoma: A Children's Oncology Group study. 2015;137(9):2163-2174.
94. Siddon A, Lozovatsky L, Mohamed A, Hudnall SD. Human herpesvirus 6 positive Reed-Sternberg cells in nodular sclerosis Hodgkin lymphoma. *British Journal of Haematology.* 2012;158(5):635-643.
95. Glaser SL, Gulley ML, Clarke CA, et al. Racial/ethnic variation in EBV-positive classical Hodgkin lymphoma in California populations. 123(7):1499-1507.
96. Heller KN, Arrey F, Steinherz P, et al. Patients with Epstein Barr virus-positive lymphomas have decreased CD4(+) T-cell responses to the viral nuclear antigen 1. 2008;123(12):2824-31.
97. Chang ET, Zheng T, Lennette ET, et al. Heterogeneity of risk factors and antibody profiles in Epstein-Barr virus genome-positive and -negative Hodgkin lymphoma. 2004;189(12):2271-81.
98. Vasef MA, Ubaidat MA, Khalidi HS, Almasri NM, Al-Abbadi M, Annab HZ. Association between Epstein-Barr virus and classic Hodgkin lymphoma in Jordan: a comparative study with Epstein-Barr virus-associated Hodgkin lymphoma in North America. *South Med J.* 2004;97(3):273-7. doi:10.1097/O1.SMJ.0000090035.09019.8D
99. Andriko JW, Aguilera NS, edkar MA, Abbondanzo SL. Childhood Hodgkin's disease in the United States: An analysis of histologic subtypes and association with Epstein-Barr virus. 1997;10(4):366-371.

100. Razzouk BI, Gan YJ, Mendonca C, et al. Epstein-Barr virus in pediatric Hodgkin disease: Age and histiotype are more predictive than geographic region. *Medical and Pediatric Oncology*. 1997;28(4):248-254.
101. Elenitoba-Johnson KS, Medeiros LJ, Khorsand J, King TC. P53 expression in Reed-Sternberg cells does not correlate with gene mutations in Hodgkin's disease. *Am J Clin Pathol*. 1996;106(6):728-38.
102. Lin AY, Kingma DW, Lennette ET, et al. Epstein-Barr virus and familial Hodgkin's disease. *Blood*. 1996;88(8):3160-5.
103. Shah KM, Young LS. Epstein-Barr virus and carcinogenesis: beyond Burkitt's lymphoma. *Clin Microbiol Infect*. 2009;15(11):982-8. doi:10.1111/j.1469-0691.2009.03033.x
104. Mertens R, Granzen B, Lassay L, Gademann G, Hess CF, Heimann G. Nasopharyngeal carcinoma in childhood and adolescence: concept and preliminary results of the cooperative GPOH study NPC-91. Gesellschaft fur Padiatrische Onkologie und Hamatologie. *Cancer*. 1997;80(5):951-9.
105. Polychronopoulou S, Kostaridou S, Panagiotou JP, et al. Nasopharyngeal carcinoma in childhood and adolescence: A single institution's experience with treatment modalities during the last 15 years. *Cancer*. 21(5):393-402.
106. *OpenEpi: Open Source Epidemiologic Statistics for Public Health*. 2013. www.OpenEpi.com
107. Verma N, Patel S, Osborn V, et al. Prognostic significance of human papillomavirus and Epstein-Bar virus in nasopharyngeal carcinoma. *Head Neck*. 2020;42(9):2364-2374. doi:10.1002/hed.26245
108. Jiang W, Chamberlain PD, Garden AS, et al. Prognostic value of p16 expression in Epstein-Barr virus-positive nasopharyngeal carcinomas. *Head Neck*. 2016;38 Suppl 1:E1459-66. doi:10.1002/hed.24258
109. Dogan S, Hedberg ML, Ferris RL, Rath TJ, Assaad AM, Chiosea SI. Human papillomavirus and Epstein-Barr virus in nasopharyngeal carcinoma in a low-incidence population. *Head Neck*. 2014;36(4):511-6. doi:10.1002/hed.23318
110. Lin Z, Khong B, Kwok S, et al. Human papillomavirus 16 detected in nasopharyngeal carcinomas in white Americans but not in endemic Southern Chinese patients. *Head Neck*. 2014;36(5):709-14. doi:10.1002/hed.23362
111. Stenmark MH, McHugh JB, Schipper M, et al. Nonendemic HPV-positive nasopharyngeal carcinoma: association with poor prognosis. *Int J Radiat Oncol Biol Phys*. 2014;88(3):580-8. doi:10.1016/j.ijrobp.2013.11.246
112. Wilson DD, Crandley EF, Sim A, et al. Prognostic significance of p16 and its relationship with human papillomavirus in pharyngeal squamous cell carcinomas. *JAMA Otolaryngol Head Neck Surg*. 2014;140(7):647-53. doi:10.1001/jamaoto.2014.821
113. Singhi AD, Califano J, Westra WH. High-risk human papillomavirus in nasopharyngeal carcinoma. *Head Neck*. 2012;34(2):213-8. doi:10.1002/hed.21714
114. International Agency for Research on Agency. IARC Monographs. World Health Organization. <http://monographs.iarc.fr/>
115. Haverkos BM, Pan Z, Gru AA, et al. Extranodal NK/T Cell Lymphoma, Nasal Type (ENKTL-NT): An Update on Epidemiology, Clinical Presentation, and Natural History in North American and European Cases. *Curr Hematol Malig Rep*. 2016;11(6):514-527. doi:10.1007/s11899-016-0355-9
116. Li S, Feng X, Li T, et al. Extranodal NK/T-cell lymphoma, nasal type: a report of 73 cases at MD Anderson Cancer Center. *Am J Surg Pathol*. 2013;37(1):14-23. doi:10.1097/PAS.0b013e31826731b5
117. Chan JKC, Quintanilla-Martinez L, Ferry JA, Peh SC. Extranodal NK/T-cell lymphoma, nasal type. In: Swerdlow SH, Campo E, Harris NK, et al, eds. *World Health Organization Classification of Tumours of Haematopoietic and Lymphoid Tissues*. IARC Press; 2008:285-288.

118. American Cancer Society. Types of B-cell Lymphoma. Accessed April 4, 2023, <https://www.cancer.org/cancer/non-hodgkin-lymphoma/about/b-cell-lymphoma.html>
119. Bourbon E, Maucourt-Boulch D, Fontaine J, et al. Clinicopathological features and survival in EBV-positive diffuse large B-cell lymphoma not otherwise specified. *Blood Adv*. 2021;5(16):3227-3239. doi:10.1182/bloodadvances.2021004515
120. Keane C, Tobin J, Gunawardana J, et al. The tumour microenvironment is immuno-tolerogenic and a principal determinant of patient outcome in EBV-positive diffuse large B-cell lymphoma. *Eur J Haematol*. 2019;103(3):200-207. doi:10.1111/ejh.13274
121. Tracy SI, Habermann TM, Feldman AL, et al. Outcomes among North American patients with diffuse large B-cell lymphoma are independent of tumor Epstein-Barr virus positivity or immunosuppression. *Haematologica*. 2018;103(2):297-303. doi:10.3324/haematol.2017.176511
122. Petrella T, Copie-Bergman C, Briere J, et al. BCL2 expression but not MYC and BCL2 coexpression predicts survival in elderly patients with diffuse large B-cell lymphoma independently of cell of origin in the phase 3 LNH03-6B trial. *Ann Oncol*. 2017;28(5):1042-1049. doi:10.1093/annonc/mdx022
123. Tisi MC, Cupelli E, Santangelo R, et al. Whole blood EBV-DNA predicts outcome in diffuse large B-cell lymphoma. *Leuk Lymphoma*. 2016;57(3):628-34. doi:10.3109/10428194.2015.1072766
124. Ziarkiewicz M, Wolosz D, Dzieciatkowski T, et al. Epstein-Barr Virus-Positive Diffuse Large B cell Lymphoma in the Experience of a Tertiary Medical Center in Poland. *Arch Immunol Ther Exp (Warsz)*. 2016;64(2):159-69. doi:10.1007/s00005-015-0341-2
125. Morton LM, Kim CJ, Weiss LM, et al. Molecular characteristics of diffuse large B-cell lymphoma in human immunodeficiency virus-infected and -uninfected patients in the pre-highly active antiretroviral therapy and pre-rituximab era. *Leuk Lymphoma*. 2014;55(3):551-7. doi:10.3109/10428194.2013.813499
126. Ok CY, Li L, Xu-Monette ZY, et al. Prevalence and clinical implications of Epstein-Barr virus infection in de novo diffuse large B-cell lymphoma in Western countries. *Clin Cancer Res*. 2014;20(9):2338-49. doi:10.1158/1078-0432.CCR-13-3157
127. Slack GW, Steidl C, Sehn LH, Gascoyne RD. CD30 expression in de novo diffuse large B-cell lymphoma: a population-based study from British Columbia. *Br J Haematol*. 2014;167(5):608-17. doi:10.1111/bjh.13085
128. Hofschneider A, Ponciano A, Bonzheim I, et al. Geographic variation in the prevalence of Epstein-Barr virus-positive diffuse large B-cell lymphoma of the elderly: a comparative analysis of a Mexican and a German population. *Mod Pathol*. 2011;24(8):1046-54. doi:10.1038/modpathol.2011.62
129. Gibson SE, Hsi ED. Epstein-Barr virus-positive B-cell lymphoma of the elderly at a United States tertiary medical center: an uncommon aggressive lymphoma with a nongerminal center B-cell phenotype. *Hum Pathol*. 2009;40(5):653-61. doi:10.1016/j.humpath.2008.10.007
130. Hoeller S, Tzankov A, Pileri SA, Went P, Dirnhofer S. Epstein-Barr virus-positive diffuse large B-cell lymphoma in elderly patients is rare in Western populations. *Human Pathology*. 2010;41(3):352-357. doi:<https://doi.org/10.1016/j.humpath.2009.07.024>
131. d'Amore F, Johansen P, Houmand A, Weisenburger DD, Mortensen LS. Epstein-Barr virus genome in non-Hodgkin's lymphomas occurring in immunocompetent patients: highest prevalence in nonlymphoblastic T-cell lymphoma and correlation with a poor prognosis. Danish Lymphoma Study Group, LYFO. *Blood*. 1996;87(3):1045-55.
132. Ramos JC, Sparano JA, Chadburn A, et al. Impact of Myc in HIV-associated non-Hodgkin lymphomas treated with EPOCH and outcomes with vorinostat (AMC-075 trial). *Blood*. 2020;136(11):1284-1297. doi:10.1182/blood.2019003959

133. Chao C, Silverberg MJ, Martinez-Maza O, et al. Epstein-Barr virus infection and expression of B-cell oncogenic markers in HIV-related diffuse large B-cell Lymphoma. *Clin Cancer Res.* 2012;18(17):4702-12. doi:10.1158/1078-0432.CCR-11-3169
134. Chadburn A, Chiu A, Lee JY, et al. Immunophenotypic analysis of AIDS-related diffuse large B-cell lymphoma and clinical implications in patients from AIDS Malignancies Consortium clinical trials 010 and 034. *J Clin Oncol.* 2009;27(30):5039-48. doi:10.1200/JCO.2008.20.5450
135. Vaghefi P, Martin A, Prevot S, et al. Genomic imbalances in AIDS-related lymphomas: relation with tumoral Epstein-Barr virus status. *AIDS.* 2006;20(18):2285-91. doi:10.1097/QAD.0b013e328010ac5b
136. Burke AP, Yen TS, Shekitka KM, Sobin LH. Lymphoepithelial carcinoma of the stomach with Epstein-Barr virus demonstrated by polymerase chain reaction. *Mod Pathol.* 1990;3(3):377-80.
137. Shibata D, Weiss LM. Epstein-Barr virus-associated gastric adenocarcinoma. *Am J Pathol.* 1992;140(4):769-74.
138. Murphy G, Pfeiffer R, Camargo MC, Rabkin CS. Meta-analysis shows that prevalence of Epstein-Barr virus-positive gastric cancer differs based on sex and anatomic location. *Gastroenterology.* 2009;137(3):824-33. doi:10.1053/j.gastro.2009.05.001
139. Li S, Du H, Wang Z, Zhou L, Zhao X, Zeng Y. Meta-analysis of the relationship between Epstein-Barr virus infection and clinicopathological features of patients with gastric carcinoma. *Sci China Life Sci.* 2010;53(4):524-30. doi:10.1007/s11427-010-0082-8
140. Sousa H, Pinto-Correia AL, Medeiros R, Dinis-Ribeiro M. Epstein-Barr virus is associated with gastric carcinoma: the question is what is the significance? *World J Gastroenterol.* 2008;14(27):4347-51.
141. Camargo MC, Murphy G, Koriyama C, et al. Determinants of Epstein-Barr virus-positive gastric cancer: an international pooled analysis. *Br J Cancer.* 2011;105(1):38-43. doi:10.1038/bjc.2011.215
142. Lee JH, Kim SH, Han SH, An JS, Lee ES, Kim YS. Clinicopathological and molecular characteristics of Epstein-Barr virus-associated gastric carcinoma: a meta-analysis. *J Gastroenterol Hepatol.* 2009;24(3):354-65. doi:10.1111/j.1440-1746.2009.05775.x
143. Tavakoli A, Monavari SH, Solaymani Mohammadi F, Kiani SJ, Armat S, Farahmand M. Association between Epstein-Barr virus infection and gastric cancer: a systematic review and meta-analysis. *BMC Cancer.* 2020;20(1):493. doi:10.1186/s12885-020-07013-x
144. Kim TS, da Silva E, Coit DG, Tang LH. Intratumoral Immune Response to Gastric Cancer Varies by Molecular and Histologic Subtype. *Am J Surg Pathol.* 2019;43(6):851-860. doi:10.1097/PAS.0000000000001253
145. Ma C, Patel K, Singhi AD, et al. Programmed Death-Ligand 1 Expression Is Common in Gastric Cancer Associated With Epstein-Barr Virus or Microsatellite Instability. *Am J Surg Pathol.* 2016;40(11):1496-1506. doi:10.1097/PAS.0000000000000698
146. Truong CD, Feng W, Li W, et al. Characteristics of Epstein-Barr virus-associated gastric cancer: a study of 235 cases at a comprehensive cancer center in U.S.A. *J Exp Clin Cancer Res.* 2009;28:14. doi:10.1186/1756-9966-28-14
147. Grogg KL, Lohse CM, Pankratz VS, Halling KC, Smyrk TC. Lymphocyte-rich gastric cancer: associations with Epstein-Barr virus, microsatellite instability, histology, and survival. *Mod Pathol.* 2003;16(7):641-51. doi:10.1097/01.MP.0000076980.73826.C0
148. Vo QN, Geradts J, Gulley ML, Boudreau DA, Bravo JC, Schneider BG. Epstein-Barr virus in gastric adenocarcinomas: association with ethnicity and CDKN2A promoter methylation. *J Clin Pathol.* 2002;55(9):669-75. doi:10.1136/jcp.55.9.669

149. Shibata D, Hawes D, Stemmermann GN, Weiss LM. Epstein-Barr virus-associated gastric adenocarcinoma among Japanese Americans in Hawaii. *Cancer Epidemiol Biomarkers Prev.* 1993;2(3):213-7.
150. Forman D, de Martel C, Lacey CJ, et al. Global burden of human papillomavirus and related diseases. *Vaccine.* 2012;30 Suppl 5:F12-23. doi:10.1016/j.vaccine.2012.07.055
151. Kreuter A, Potthoff A, Brockmeyer NH, et al. Anal carcinoma in human immunodeficiency virus-positive men: results of a prospective study from Germany. *Br J Dermatol.* 2010;162(6):1269-77. doi:10.1111/j.1365-2133.2010.09712.x
152. Arana R, Flejou JF, Si-Mohamed A, Bauer P, Etienney I. Clinicopathological and virological characteristics of superficially invasive squamous-cell carcinoma of the anus. *Colorectal Dis.* 2015;17(11):965-72. doi:10.1111/codi.12951
153. Zhang ER, Pfeiffer RM, Austin A, et al. Impact of HIV on Anal Squamous Cell Carcinoma Rates in the United States, 2001-2015. *J Natl Cancer Inst.* 2022;114(9):1246-1252. doi:10.1093/jnci/djac103
154. Herfs M, Longuespee R, Quick CM, et al. Proteomic signatures reveal a dualistic and clinically relevant classification of anal canal carcinoma. *J Pathol.* 2017;241(4):522-533. doi:10.1002/path.4858
155. Alemany L, Saunier M, Alvarado-Cabrero I, et al. Human papillomavirus DNA prevalence and type distribution in anal carcinomas worldwide. *Int J Cancer.* 2015;136(1):98-107. doi:10.1002/ijc.28963
156. Steinau M, Unger ER, Hernandez BY, et al. Human papillomavirus prevalence in invasive anal cancers in the United States before vaccine introduction. *J Low Genit Tract Dis.* 2013;17(4):397-403. doi:10.1097/LGT.0b013e31827ed372
157. Zhu X, Jamshed S, Zou J, et al. Molecular and immunophenotypic characterization of anal squamous cell carcinoma reveals distinct clinicopathologic groups associated with HPV and TP53 mutation status. *Mod Pathol.* 2021;34(5):1017-1030. doi:10.1038/s41379-020-00729-y
158. Meyer JE, Panico VJ, Marconato HM, Sherr DL, Christos P, Pirog EC. HIV positivity but not HPV/p16 status is associated with higher recurrence rate in anal cancer. *J Gastrointest Cancer.* 2013;44(4):450-5. doi:10.1007/s12029-013-9543-1
159. Alemany L, Cubilla A, Halec G, et al. Role of Human Papillomavirus in Penile Carcinomas Worldwide. *Eur Urol.* 2016;69(5):953-61. doi:10.1016/j.eururo.2015.12.007
160. McDaniel AS, Hovelson DH, Cani AK, et al. Genomic Profiling of Penile Squamous Cell Carcinoma Reveals New Opportunities for Targeted Therapy. *Cancer Res.* 2015;75(24):5219-27. doi:10.1158/0008-5472.CAN-15-1004
161. Hernandez BY, Goodman MT, Unger ER, et al. Human papillomavirus genotype prevalence in invasive penile cancers from a registry-based United States population. *Front Oncol.* 2014;4:9. doi:10.3389/fonc.2014.00009
162. Daling JR, Madeleine MM, Johnson LG, et al. Penile cancer: importance of circumcision, human papillomavirus and smoking in in situ and invasive disease. *Int J Cancer.* 2005;116(4):606-16. doi:10.1002/ijc.21009
163. Rubin MA, Kleter B, Zhou M, et al. Detection and typing of human papillomavirus DNA in penile carcinoma: evidence for multiple independent pathways of penile carcinogenesis. *Am J Pathol.* 2001;159(4):1211-8. doi:10.1016/S0002-9440(10)62506-0
164. Sinno AK, Saraiya M, Thompson TD, et al. Human papillomavirus genotype prevalence in invasive vaginal cancer from a registry-based population. *Obstet Gynecol.* 2014;123(4):817-21. doi:10.1097/AOG.000000000000171
165. Daling JR, Madeleine MM, Schwartz SM, et al. A population-based study of squamous cell vaginal cancer: HPV and cofactors. *Gynecol Oncol.* 2002;84(2):263-70. doi:10.1006/gyno.2001.6502

166. De Vuyst H, Clifford GM, Nascimento MC, Madeleine MM, Franceschi S. Prevalence and type distribution of human papillomavirus in carcinoma and intraepithelial neoplasia of the vulva, vagina and anus: a meta-analysis. *Int J Cancer*. 2009;124(7):1626-36. doi:10.1002/ijc.24116
167. Koltz E, Lucas E, Hosler GA, et al. Human Papillomavirus Positive and Negative Vulvar Squamous Cell Carcinoma Are Biologically but Not Clinically Distinct. *J Invest Dermatol*. 2021;doi:10.1016/j.jid.2021.10.009
168. Gargano JW, Wilkinson EJ, Unger ER, et al. Prevalence of human papillomavirus types in invasive vulvar cancers and vulvar intraepithelial neoplasia 3 in the United States before vaccine introduction. *J Low Genit Tract Dis*. 2012;16(4):471-9. doi:10.1097/LGT.0b013e3182472947
169. de Koning MN, Quint WG, Pirog EC. Prevalence of mucosal and cutaneous human papillomaviruses in different histologic subtypes of vulvar carcinoma. *Mod Pathol*. 2008;21(3):334-44. doi:10.1038/modpathol.3801009
170. Al-Ghamdi A, Freedman D, Miller D, et al. Vulvar squamous cell carcinoma in young women: a clinicopathologic study of 21 cases. *Gynecol Oncol*. 2002;84(1):94-101. doi:10.1006/gyno.2001.6466
171. Kim YT, Thomas NF, Kesis TD, Wilkinson EJ, Hedrick L, Cho KR. p53 mutations and clonality in vulvar carcinomas and squamous hyperplasias: evidence suggesting that squamous hyperplasias do not serve as direct precursors of human papillomavirus-negative vulvar carcinomas. *Hum Pathol*. 1996;27(4):389-95.
172. Bishop JA, Ma XJ, Wang H, et al. Detection of transcriptionally active high-risk HPV in patients with head and neck squamous cell carcinoma as visualized by a novel E6/E7 mRNA in situ hybridization method. *Am J Surg Pathol*. 2012;36(12):1874-82. doi:10.1097/PAS.0b013e318265fb2b
173. Rietbergen MM, Leemans CR, Bloemena E, et al. Increasing prevalence rates of HPV attributable oropharyngeal squamous cell carcinomas in the Netherlands as assessed by a validated test algorithm. *Int J Cancer*. 2013;132(7):1565-71. doi:10.1002/ijc.27821
174. Lewis JS, Jr., Mirabello L, Liu P, et al. Oropharyngeal Squamous Cell Carcinoma Morphology and Subtypes by Human Papillomavirus Type and by 16 Lineages and Sublineages. *Head Neck Pathol*. 2021;15(4):1089-1098. doi:10.1007/s12105-021-01318-4
175. Mazul AL, Rodriguez-Ormaza N, Taylor JM, et al. Prognostic significance of non-HPV16 genotypes in oropharyngeal squamous cell carcinoma. *Oral Oncol*. 2016;61:98-103. doi:10.1016/j.oraloncology.2016.08.019
176. Hooper JE, Hebert JF, Schilling A, et al. Hybrid Capture 2 is as effective as PCR testing for high-risk human papillomavirus in head and neck cancers. *Appl Immunohistochem Mol Morphol*. 2015;23(4):266-72. doi:10.1097/PDM.0000000000000036
177. Zandberg DP, Liu S, Goloubeva OG, Schumaker LM, Cullen KJ. Emergence of HPV16-positive oropharyngeal cancer in Black patients over time: University of Maryland 1992-2007. *Cancer Prev Res (Phila)*. 2015;8(1):12-9. doi:10.1158/1940-6207.CAPR-14-0089-T
178. Isayeva T, Xu J, Dai Q, et al. African Americans with oropharyngeal carcinoma have significantly poorer outcomes despite similar rates of human papillomavirus-mediated carcinogenesis. *Hum Pathol*. 2014;45(2):310-9. doi:10.1016/j.humpath.2013.09.006
179. Lingen MW, Xiao W, Schmitt A, et al. Low etiologic fraction for high-risk human papillomavirus in oral cavity squamous cell carcinomas. *Oral Oncol*. 2013;49(1):1-8. doi:10.1016/j.oraloncology.2012.07.002
180. Walline HM, Komarck C, McHugh JB, et al. High-risk human papillomavirus detection in oropharyngeal, nasopharyngeal, and oral cavity cancers: comparison of multiple methods. *JAMA Otolaryngol Head Neck Surg*. 2013;139(12):1320-7. doi:10.1001/jamaoto.2013.5460

181. Jordan RC, Lingen MW, Perez-Ordóñez B, et al. Validation of methods for oropharyngeal cancer HPV status determination in US cooperative group trials. *Am J Surg Pathol*. 2012;36(7):945-54. doi:10.1097/PAS.0b013e318253a2d1
182. Stephen JK, Chen KM, Shah V, et al. Human papillomavirus outcomes in an access-to-care laryngeal cancer cohort. *Otolaryngol Head Neck Surg*. 2012;146(5):730-8. doi:10.1177/0194599811434482
183. Chaturvedi AK, Engels EA, Pfeiffer RM, et al. Human papillomavirus and rising oropharyngeal cancer incidence in the United States. *J Clin Oncol*. 2011;29(32):4294-301. doi:10.1200/JCO.2011.36.4596
184. Schlecht NF, Brandwein-Gensler M, Nuovo GJ, et al. A comparison of clinically utilized human papillomavirus detection methods in head and neck cancer. *Mod Pathol*. 2011;24(10):1295-305. doi:10.1038/modpathol.2011.91
185. Agoston ES, Robinson SJ, Mehra KK, et al. Polymerase chain reaction detection of HPV in squamous carcinoma of the oropharynx. *Am J Clin Pathol*. 2010;134(1):36-41. doi:10.1309/AJCP1AAWXE5JJCLZ
186. Kingma DW, Allen RA, Moore W, et al. HPV genotype distribution in oral and oropharyngeal squamous cell carcinoma using seven in vitro amplification assays. *Anticancer Res*. 2010;30(12):5099-104.
187. Jo S, Juhasz A, Zhang K, et al. Human papillomavirus infection as a prognostic factor in oropharyngeal squamous cell carcinomas treated in a prospective phase II clinical trial. *Anticancer Res*. 2009;29(5):1467-74.
188. Settle K, Posner MR, Schumaker LM, et al. Racial survival disparity in head and neck cancer results from low prevalence of human papillomavirus infection in black oropharyngeal cancer patients. *Cancer Prev Res (Phila)*. 2009;2(9):776-81. doi:10.1158/1940-6207.CAPR-09-0149
189. Tezal M, Sullivan Nasca M, Stoler DL, et al. Chronic periodontitis-human papillomavirus synergy in base of tongue cancers. *Arch Otolaryngol Head Neck Surg*. 2009;135(4):391-6. doi:10.1001/archoto.2009.6
190. Cohen MA, Basha SR, Reichenbach DK, Robertson E, Sewell DA. Increased viral load correlates with improved survival in HPV-16-associated tonsil carcinoma patients. *Acta Otolaryngol*. 2008;128(5):583-9. doi:10.1080/00016480701558880
191. Liang XH, Lewis J, Foote R, Smith D, Kademani D. Prevalence and significance of human papillomavirus in oral tongue cancer: the Mayo Clinic experience. *J Oral Maxillofac Surg*. 2008;66(9):1875-80. doi:10.1016/j.joms.2008.04.009
192. Worden FP, Kumar B, Lee JS, et al. Chemoselection as a strategy for organ preservation in advanced oropharynx cancer: response and survival positively associated with HPV16 copy number. *J Clin Oncol*. 2008;26(19):3138-46. doi:10.1200/JCO.2007.12.7597
193. Zhao M, Rosenbaum E, Carvalho AL, et al. Feasibility of quantitative PCR-based saliva rinse screening of HPV for head and neck cancer. *Int J Cancer*. 2005;117(4):605-10. doi:10.1002/ijc.21216
194. Strome SE, Savva A, Brissett AE, et al. Squamous cell carcinoma of the tonsils: a molecular analysis of HPV associations. *Clin Cancer Res*. 2002;8(4):1093-100.
195. Feng H, Shuda M, Chang Y, Moore PS. Clonal integration of a polyomavirus in human Merkel cell carcinoma. *Science*. 2008;319(5866):1096-100. doi:10.1126/science.1152586
196. International Agency for Research on Agency. *Malaria and Some Polyomaviruses (SV40, BK, JC, and Merkel Cell Viruses)*. Vol. 104. 2014. *IARC Monographs on the Evaluation of Carcinogenic Risk to Humans*.

197. Harms KL, Zhao L, Johnson B, et al. Virus-positive Merkel Cell Carcinoma Is an Independent Prognostic Group with Distinct Predictive Biomarkers. *Clin Cancer Res*. 2021;27(9):2494-2504. doi:10.1158/1078-0432.CCR-20-0864
198. Hill NT, Kim D, Busam KJ, Chu EY, Green C, Brownell I. Distinct Signatures of Genomic Copy Number Variants Define Subgroups of Merkel Cell Carcinoma Tumors. *Cancers (Basel)*. 2021;13(5)doi:10.3390/cancers13051134
199. Xie H, Kaye FJ, Isse K, et al. Delta-Like Protein 3 Expression and Targeting in Merkel Cell Carcinoma. *Oncologist*. 2020;25(9):810-817. doi:10.1634/theoncologist.2019-0877
200. Moshiri AS, Doumani R, Yelistratova L, et al. Polyomavirus-Negative Merkel Cell Carcinoma: A More Aggressive Subtype Based on Analysis of 282 Cases Using Multimodal Tumor Virus Detection. *J Invest Dermatol*. 2017;137(4):819-827. doi:10.1016/j.jid.2016.10.028
201. Feldmeyer L, Hudgens CW, Ray-Lyons G, et al. Density, Distribution, and Composition of Immune Infiltrates Correlate with Survival in Merkel Cell Carcinoma. *Clin Cancer Res*. 2016;22(22):5553-5563. doi:10.1158/1078-0432.CCR-16-0392
202. Lipson EJ, Vincent JG, Loyo M, et al. PD-L1 expression in the Merkel cell carcinoma microenvironment: association with inflammation, Merkel cell polyomavirus and overall survival. *Cancer Immunol Res*. 2013;1(1):54-63. doi:10.1158/2326-6066.CIR-13-0034
203. Hall BJ, Pincus LB, Yu SS, Oh DH, Wilson AR, McCalmont TH. Immunohistochemical prognostication of Merkel cell carcinoma: p63 expression but not polyomavirus status correlates with outcome. *J Cutan Pathol*. 2012;39(10):911-7. doi:10.1111/j.1600-0560.2012.01964.x
204. Rodig SJ, Cheng J, Wardzala J, et al. Improved detection suggests all Merkel cell carcinomas harbor Merkel polyomavirus. *J Clin Invest*. 2012;122(12):4645-53. doi:10.1172/JCI64116
205. Bhatia K, Goedert JJ, Modali R, Preiss L, Ayers LW. Merkel cell carcinoma subgroups by Merkel cell polyomavirus DNA relative abundance and oncogene expression. *Int J Cancer*. 2010;126(9):2240-6. doi:10.1002/ijc.24676
206. Busam KJ, Jungbluth AA, Rekhman N, et al. Merkel cell polyomavirus expression in merkel cell carcinomas and its absence in combined tumors and pulmonary neuroendocrine carcinomas. *Am J Surg Pathol*. 2009;33(9):1378-85. doi:10.1097/PAS.0b013e3181aa30a5