

# Supporting Information for

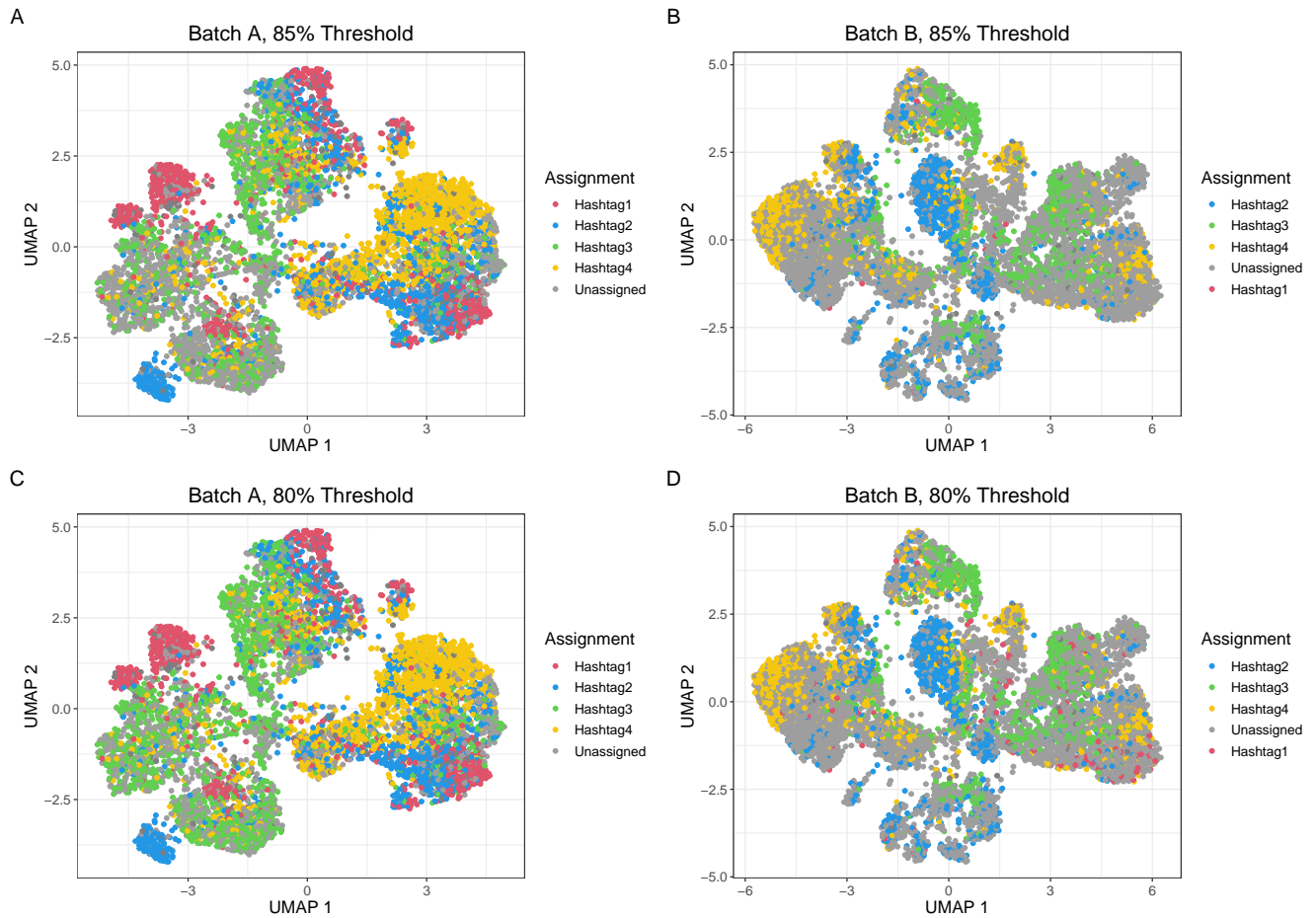
## Performance of computational algorithms to deconvolve heterogeneous bulk tumor tissue depends on experimental factors

Hippen, Omran, Weber, Jung, Drapkin, Doherty, Hicks, and Greene

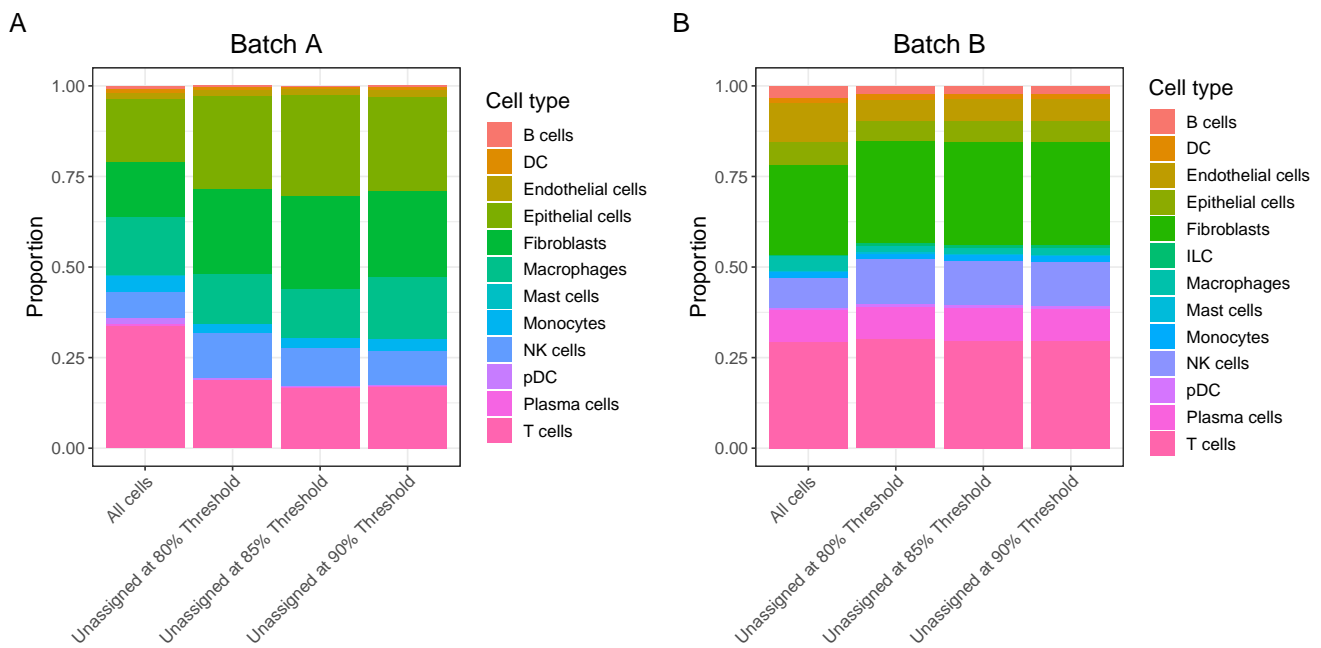
Casey S. Greene, e-mail: [casey.s.greene@cuanschutz.edu](mailto:casey.s.greene@cuanschutz.edu)

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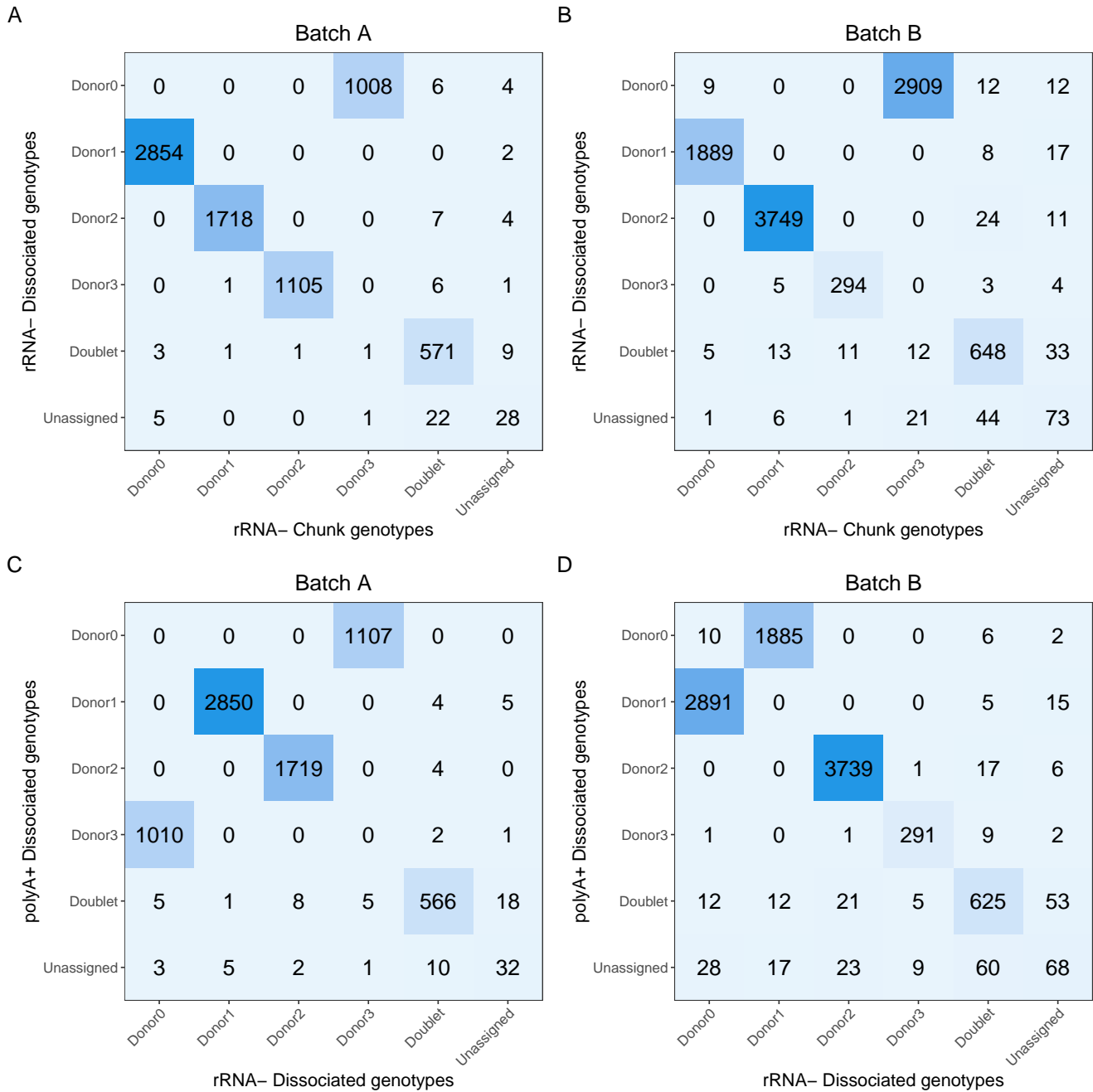
Figs S1 to S7



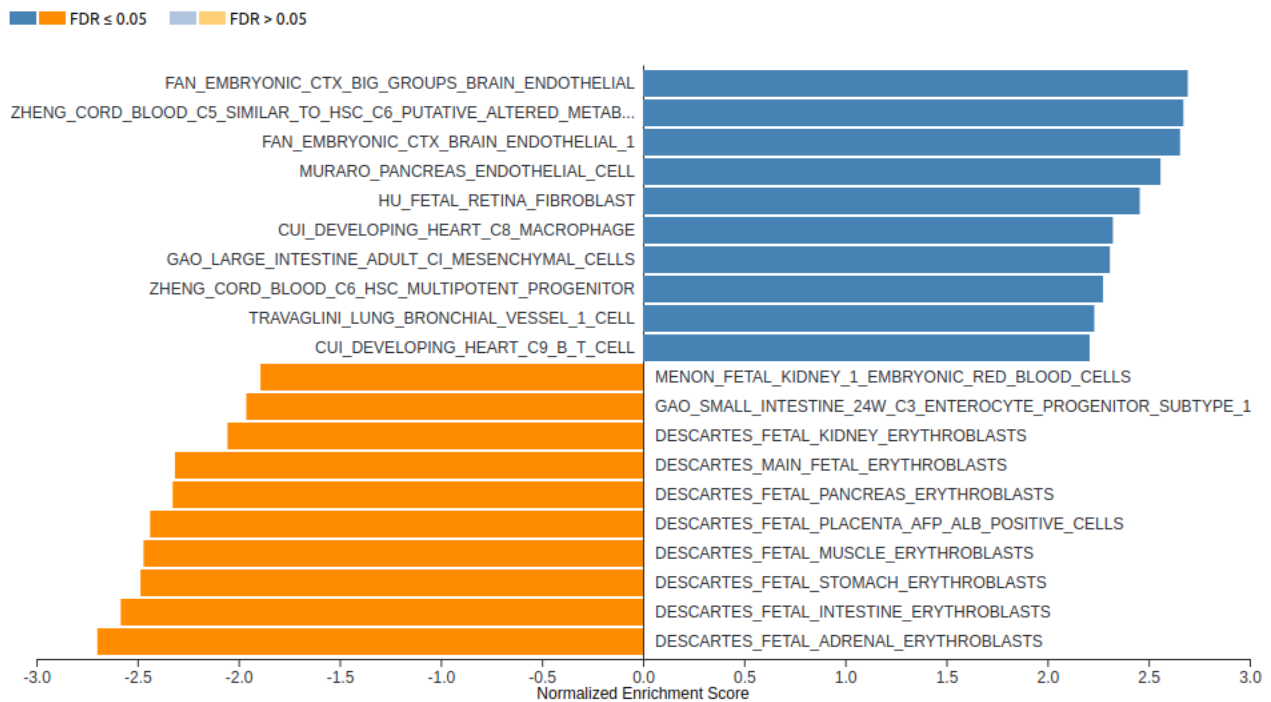
**Fig. S1. Relaxed probability thresholds for hash demultiplexing increase number of assigned cells.** A) Assignments for Batch A where any cell with greater than 85% probability of originating from a sample is assigned to that sample. B) Assignments for Batch B at the 85% probability threshold. C) Assignments for Batch A at a threshold of greater than 80% probability of originating from a sample. D) Assignments for Batch B at the 80% probability threshold.



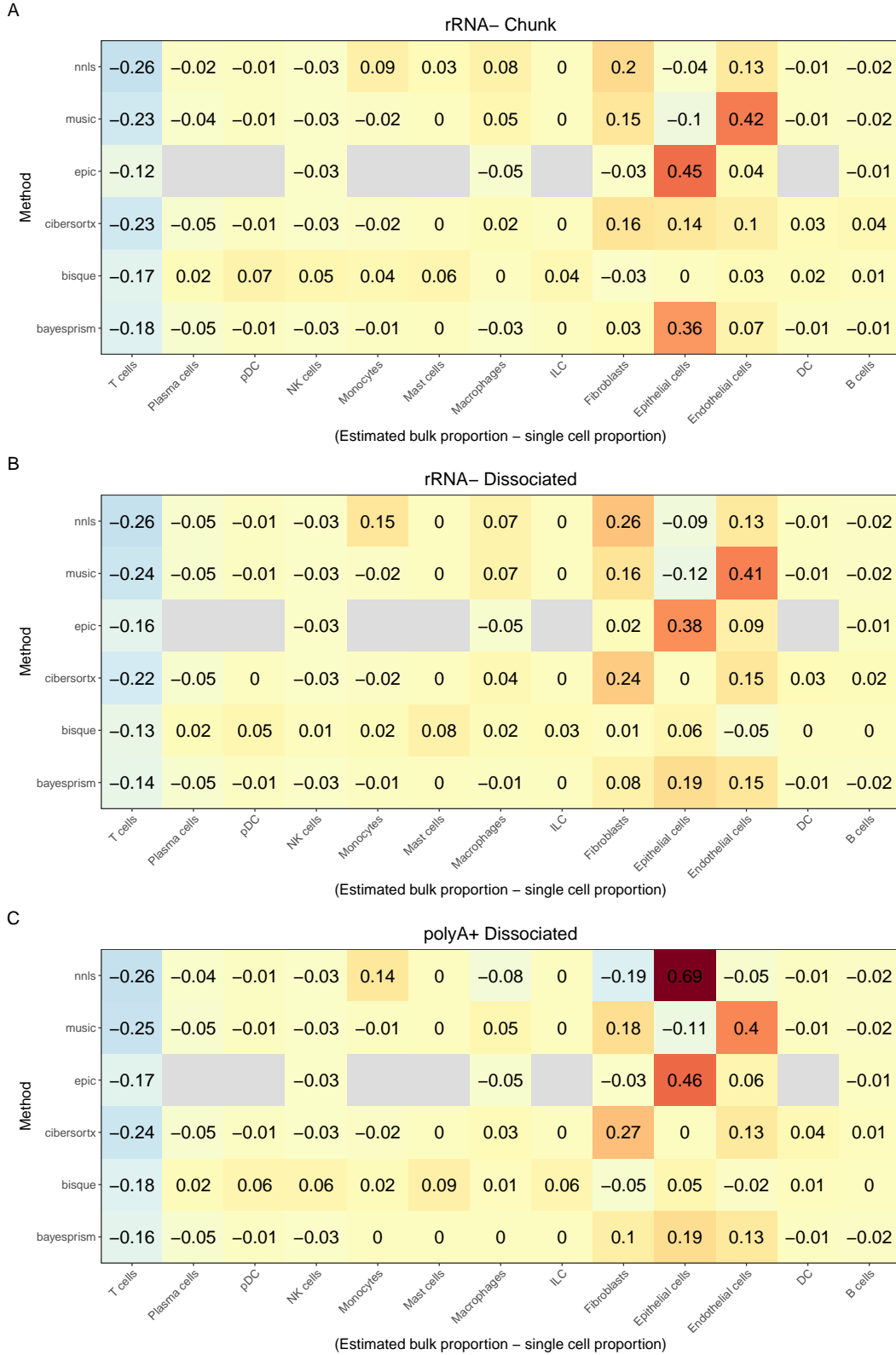
**Fig. S2. Hash demultiplexing demonstrates cell type bias.** A) Proportion of cell types in Batch A across all cells and in unassigned cells at various probability thresholds. Epithelial cells and fibroblasts are proportionally greater and T cells proportionally lesser in unassigned cells than in all cells. B) Proportion of cell types in Batch B.



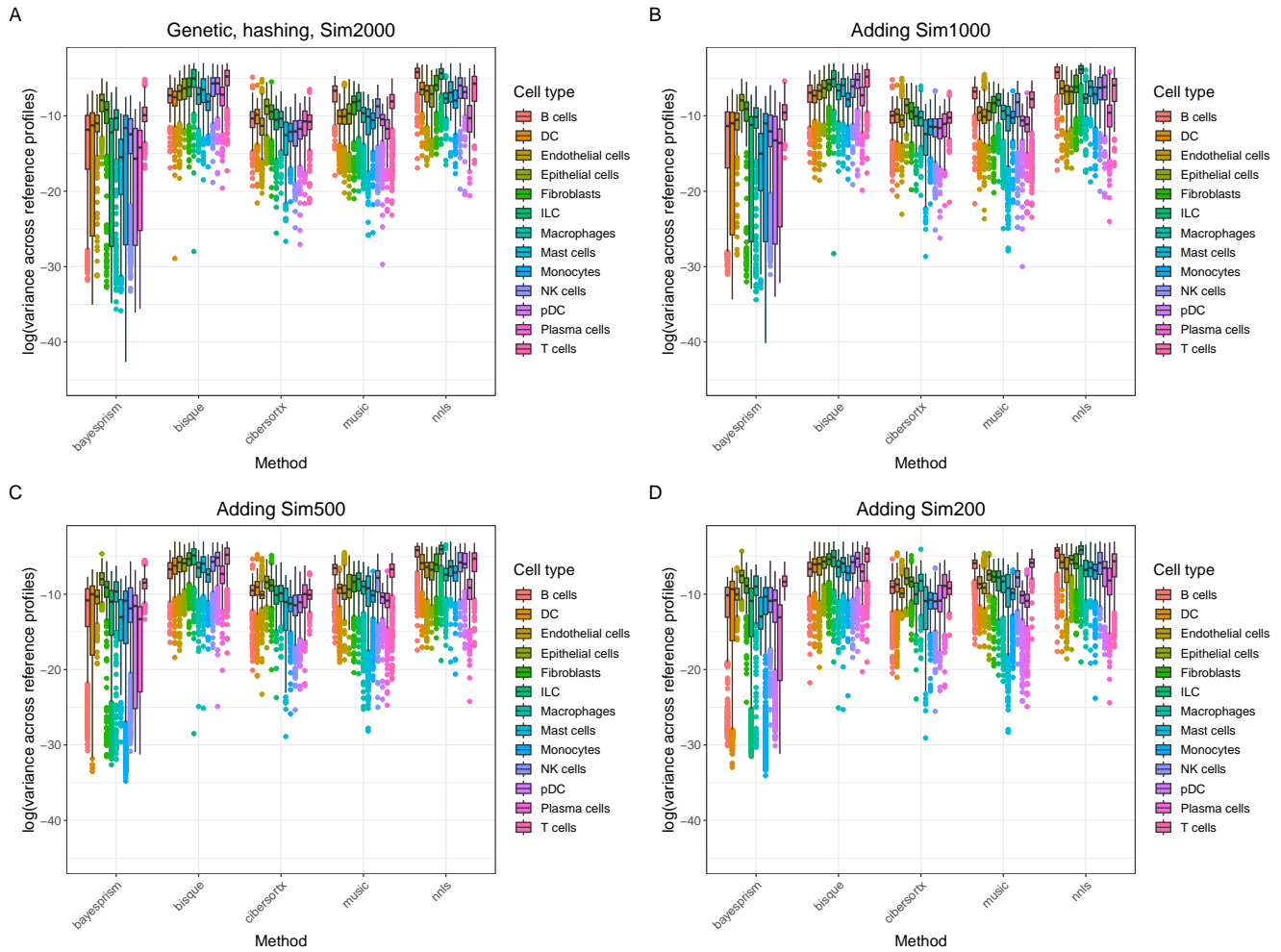
**Fig. S3. Genetic demultiplexing is concordant across source of bulk reference genotypes.** A) Confusion matrix of genetic demultiplexing assignments for Batch A when using reference genotypes from rRNA<sup>-</sup> Chunk samples vs rRNA<sup>-</sup> Dissociated samples. B) Genetic demultiplexing assignments for Batch B using reference genotypes from rRNA<sup>-</sup> Chunk samples vs rRNA<sup>-</sup> Dissociated samples. C) Confusion matrix of genetic demultiplexing assignments for Batch A when using reference genotypes from rRNA<sup>-</sup> Dissociated samples vs polyA<sup>+</sup> Dissociated samples. D) Genetic demultiplexing assignments for Batch B using reference genotypes from rRNA<sup>-</sup> Dissociated samples vs polyA<sup>+</sup> Dissociated samples.



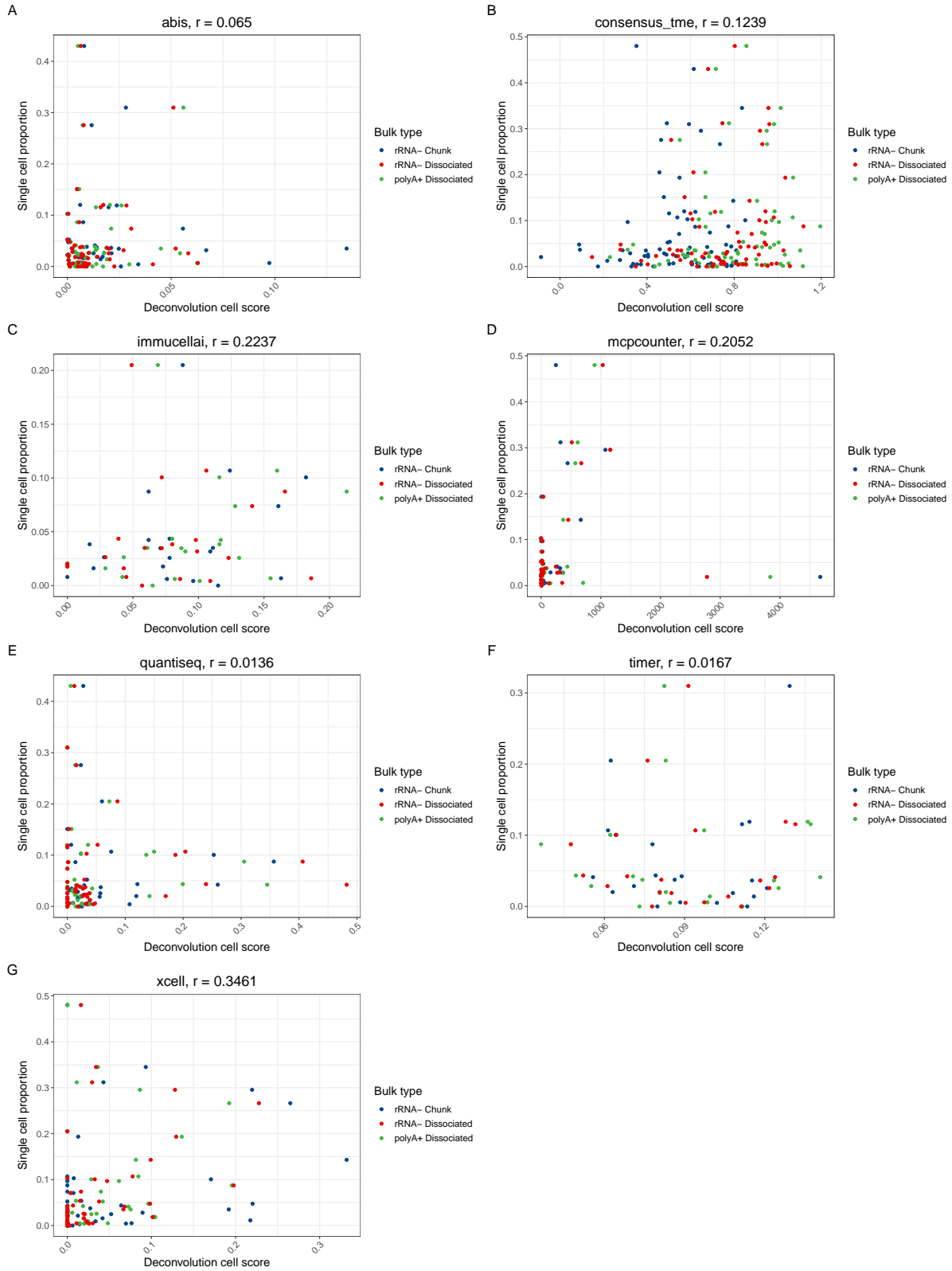
**Fig. S4. Stromal cell types are more abundant in dissociated bulk samples.** Results from Gene Set Enrichment Analysis of rRNA<sup>+</sup> Chunk samples vs rRNA<sup>-</sup> Dissociated samples. Gene signatures associated with endothelial cells, fibroblasts, macrophages, and other immune cells (blue) are more abundant in rRNA<sup>-</sup> Dissociated samples, whereas red blood cell gene signatures (orange) are more abundant in rRNA<sup>+</sup> Chunk samples.



**Fig. S5. Deconvolution estimates vary based on input bulk type.** A) The average difference between estimated cell type proportion in rRNA<sup>-</sup> Chunk samples minus the corresponding cell type proportion in scRNA-seq Individual samples. Gray boxes represent cell types not estimated by a given method. B) The difference in cell type proportion based on rRNA<sup>-</sup> Dissociated sample deconvolution estimates and scRNA-seq Individual samples. C) The difference in cell type proportion based on polyA<sup>+</sup> Dissociated sample deconvolution estimates and scRNA-seq Individual samples.



**Fig. S6. Robustness to very small reference profiles** A) The variance of deconvolution proportion estimates, stratified by cell type and method, when using our default reference profile (genetic), the reference profile of cells assigned to a sample by hash demultiplexing (hashing), and a simulated sample of approximately 2000 cells (Sim2000). B) Variance of proportion estimates across the same reference profiles as in A but also including results from Sim1000. C) Same as B but also including results from Sim500. D) Same as C but also including results from Sim200.



**Fig. S7. Alternate deconvolution methods that return cell type scores do not match single cell proportions.** A-G) Correlation between the cell type score returned by the deconvolution method and the corresponding proportion of cells in the scRNA-seq Individual sample. The name of the deconvolution method and the Pearson correlation ( $r$  value) is shown at the top of each panel.