

Supplementary Information

Lactate dehydrogenase D is a general dehydrogenase for D-2-hydroxyacids and is associated with D-lactic acidosis

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mmLDHD

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1      10      20
mmLDHD .....MALLRVATQRLSPWRSFCSRGSQGG.....
hsLDHD .....MARLLRSATWELFPWRGYCSQKAKE.....
btLDHD .....MATLLRPASWGLFSWRGYCSRGTQGR.....
xLLDHD .....MSLQAAARIKRALTYCVNSWTCRTYC.....
dRLDHD .....MTLFRHLVRIITSPRLPFCGSSRRFSAKTA.....
scDL1D1 MLWKRTCTRLIKPIAQPRGRLLVRRSCYRYASTGTGSTDSSQWLKYSVLIASSATLFGYLFAKNLYSRETAKEDLIEKLEMVKKIDP

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mmLDHD

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          α1          α2          β1          α3
          30         40         50         60         70         80
mmLDHD .....LSQDFVEALKAVVGGSPHVS...TASAVREQHGHDESMHR...CQFPED...AVVWPQNVQVSRVALSLC
hsLDHD .....LCRDFVEALKAVVGGSSHVS...TAAVREQHGRDESMHR...CEFPED...AVVWPONVEQVSRVALSLC
btLDHD .....LSAGFVEALKAVVGGSPHVS...TAAVREQHGHDESMHR...CEFPED...AVVWPONVEQVSRVALSLC
xLLDHD .....MKKEFVDELRTVVGESNTS...TAMAVREQHGRDESMHR...CRFPED...VVVWPONVDQVSKMAALCMC
dRLDHD .....AVERVVSSFRVVGDEGVS...VGSVAVREQHGRDESMHR...CRFPED...VVVWPONVEQVSRVALSLC
scDL1D1 VNSTLKLSSLDSPDYLHDFVKIDKVVLEKQVLEGNKPEYSDAKSDDLDAHSDTYFNTHHSPEQRPRITLLFPHITTEVSKILKHL

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mmLDHD

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          β2          TT          β3          η1          β4          α4          α5
          90         100        110        120        130        140        150        160        170
mmLDHD YNQGVPVPIFFGCTGVEGGVCAVGGVGVINLTH...MDRIELNTEDFSVVVEFGVTRKALNTHLSDSLGLWFPVDPGADASLCLGM
hsLDHD YRQGVPIIIPFGCTGLEGGVCAVGGVGVINLTH...MDRIELNTEDFSVVVEFGVTRKALNTHLSDSLGLWFPVDPGADASLCLGM
btLDHD YRQGVPIIIPFGCTGLEGGVCAVGGVGVINLTH...MDRIELNTEDFSVVVEFGVTRKALNTHLSDSLGLWFPVDPGADASLCLGM
xLLDHD YRNSVPIIIPFGCTGLEGGVCAVGGVGVINLTH...MDRIELNTEDFSVVVEFGVTRKALNTHLSDSLGLWFPVDPGADASLCLGM
dRLDHD HHYVRLPIIIPFGCTGLEGGVCAVGGVGVINLTH...MDRIELNTEDFSVVVEFGVTRKALNTHLSDSLGLWFPVDPGADASLCLGM
scDL1D1 HDNMPVVPVPSGCTSLGCHFLPRLTIGDITVVDLSKFMNNVVKVDFDKLDELITVQAGLPWEDLNDYLSHGLMFGCDPGLPAQIGC

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mmLDHD

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          η2          η3          β5          β6          η4
          170        180        190        200        210        220
mmLDHD AATGASGTNAVRYGTMRENVINLEVLVLPDGRLLHTAGRG...RHY...RKSAGYNLTLGLFVGS
hsLDHD AATGASGTNAVRYGTMRENVINLEVLVLPDGRLLHTAGRG...RHF...RFGFWPEIPHHTAWYSPCVSLGF...RKSAGYNLTLGLFVGS
btLDHD AATGASGTNAVRYGTMRENVINLEVLVLPDGRLLHTAGRG...RRFR...RKSAGYNLTLGLFVGS
xLLDHD AATGASGTNAVRYGTMRENVINLEVLVLPDGRILHTAGRG...R...RKTAGYNLTLGLFVGS
dRLDHD AATGASGTNAVRYGTMRENVINLEVLVLPDGTILHTAGRG...R...RKTAGYNLTLGLFVGS
scDL1D1 IANSCSCTNAVRYGTMKENIINMTIVLPDGTIVKTKKRE...R...RKSAGYNLTLGLFVGS

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mmLDHD

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          β7          β8          α6          β9          α7          β10
          230        240        250        260        270        280        290        300
mmLDHD ECTLGLITITSLTLRLHPAPEATVAATCAFPSVQAAVDSTVQILOAAVPVARIIEFLDDVMMDDACNRHSKLN...CPVAPTLFLFEFHG
hsLDHD ECTLGLITITATTLRLHPAPEATVAATCAFPSVQAAVDSTVHILQAAVPVARIIEFLDEVMDDACNRHSKLN...CLVAPTLEFLFEFHG
btLDHD ECTLGLITITAA TLRLHPAPEATVAATCAFPTVQAAVDSTVHILQAAVPVARIIEFLDEVMDDACNRHSKLN...CCVAPTLEFLFEFHG
xLLDHD ECTLGLITITKASLRLHGPPEAMVAATCAFPSVQAAVDSTVQILOCGVPIARIEFLDDVMTGACNRFNGLS...YVPTLPTLEFLFEFHG
dRLDHD ECTLGLITITKALRLYGVPEAMVSAATCAFPSVQAAVDSTVQILOAGVPIARIEFLDDVMTNACNRFNGLS...YVPTLPTLEFLFEFHG
scDL1D1 ECTLGLITITPEATVVKCHVKPKAETVAIVSFDITKDAACASNLITSGIHLNAMEELLDENMKLINASESITRCDWVETKPTMFEKIGC

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mmLDHD

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          α8          β11          α9          β12          α10
          310        320        330        340        350        360        370        380
mmLDHD SQQITLAEQLQRTEAITQDNGGSHFSWAKEAEKRNELWAARHNAWYAALALSPG...SKAYSTDVCPVSRRLPEILVETKEDLEK
hsLDHD SQQALEEQLQRTEIVQNGGSHFSWAKEAEERSRLWAARHNAWYAALALTRPG...CKGYSTDVCPVSRRLPEILVOTKEDLN
btLDHD SEQALAEQVQRTEIIRHNGGSHFSWAKEAEERSRLWAARHNAWYAALALTRPG...CKGYSTDVCPVSRRLPEILVOTKEDLE
xLLDHD ITENGLKKEQVQRTEITQNGGSHFTWAKDEETRSRLWAARHNAWYAALALTRPG...CKGYSTDVCPVSRRLPEILVETKADLI
dRLDHD ITENGLKKEQVQRTEITQNGGSHFAWAKDEETRSRLWAARHNAWYAALALTRPG...CKAYSTDVCPVSRRLPEILVETKADLI
scDL1D1 RFPNIVNALVDVKAQAQLNHCNSFQEAQDDDKLELWPAKVAALWSVLDADKSKDKSAKIWIITDVAVPVSQFDRVIVHETKEDLEK

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mmLDHD

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          β13          η5          β14          α11          α12          α13
          390        400        410        420        430        440        450        460        470
mmLDHD ASKLTGATVGHVGDGNFHCILLVDPDDAEQRVKAFANLGRRALALHGTCTGEHGCIGKRRQILQEEVGPVGVETMRQLKNTL
hsLDHD ASGLTGSIVGHVGDGNFHCILLVNPDDAEELGRVKAFANLGRRALALHGTCTGEHGCIGKRRQILQEEVGA VGVETMRQLKAVL
btLDHD AWRLTGTIVGHVGDGNFHCILLVDPEDPEELLRVQAFANLGRRALALHGTCTGEHGCIGKRRQILPEEVGA VGMETMRQIKATL
xLLDHD ESRLTGTIVGHVGDGNFHCIMVNLADKDEEVRVQKDFNRLARALAMNNGTCTGEHGCIGKRRQILPEEVGELVGTATMQLKNTL
dRLDHD SNNTITGIVGHVGDGNFHCILLVDPNDTDEEVRVHSTFTERLARALAMNNGTCTGEHGCIGKRRQILPEEVGLAIEVMKGLKASL
scDL1D1 ASKRLINATVGHAGDGNFHFIVYR...TPEHETCSQLVDMVGRALANADGTCTGEHGCIGKRRQILPEEVGELVGTATMQLKAVL

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mmLDHD

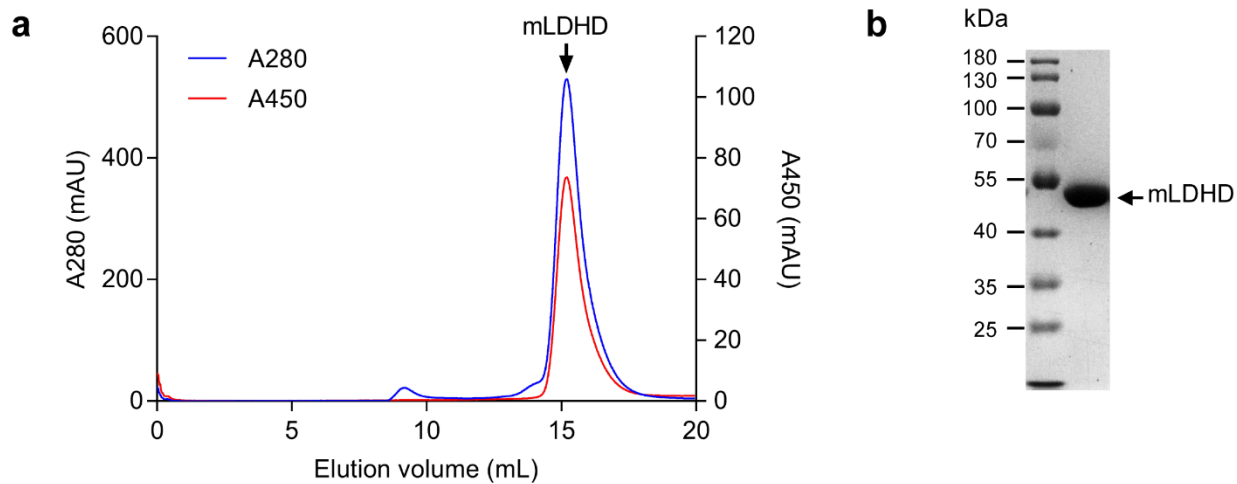
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          TT
          480
mmLDHD DFRGLMNPQKVL...
hsLDHD DPQGLMNPQKVL...
btLDHD DPQGLMNPQKPEEGTVLRLQEEGHFRGRSLSQLLCP...
xLLDHD DPKNLMNPQKVV...
dRLDHD DFRNLMNPQKVLLELTQTNTEQ...
scDL1D1 DPKRIMNPKIKTFRKTDNPEPANDYR...

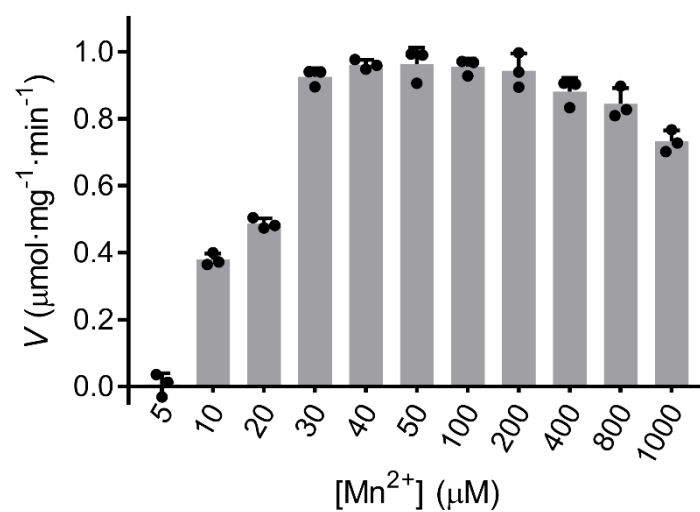
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- ▲ FAD-binding site
- ▲ Substrate-binding site (subsite A)
- ▲ Substrate-binding site (subsite B)
- ▲ Disease-associated mutation sites

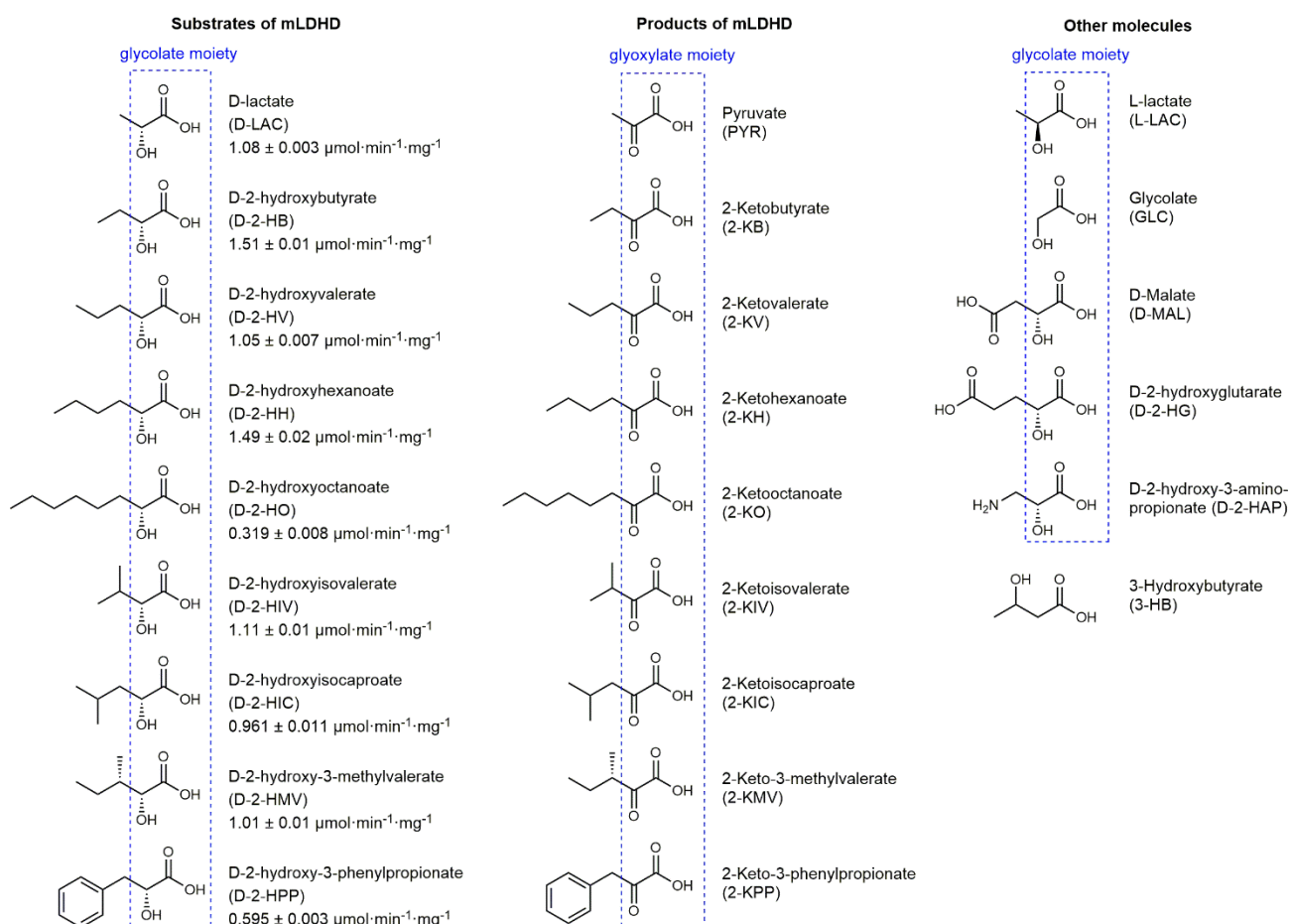
Supplementary Fig 1. Sequence alignment of LDHD from six representative eukaryotes. Amino acid sequences of FAD-dependent D-lactate dehydrogenases from *H. sapiens* (hsLDHD, Uniprot: Q86WU2), *M. musculus* (mmLDHD, Uniprot: Q7TNG8), *B. taurus* (btLDHD, Uniprot: Q148K4), *X. laevis* (xlLDHD, Uniprot: A0A1L8GLK1), *D. rerio* (drLDHD, Uniprot: F1QXM5), and *S. cerevisiae* (scDLD1, Uniprot: P32891) are aligned. The 23-residue insertion of hsLDHD is indicated by green box. The secondary structures of mmLDHD are placed on the top of the sequence alignment. Residues of mmLDHD involved in the FAD binding are indicated by red triangles. Residues of mmLDHD composing the substrate-binding subsites A and B are indicated by blue and green triangles, respectively. The disease-associated mutations of human LDHD are indicated by black triangles.



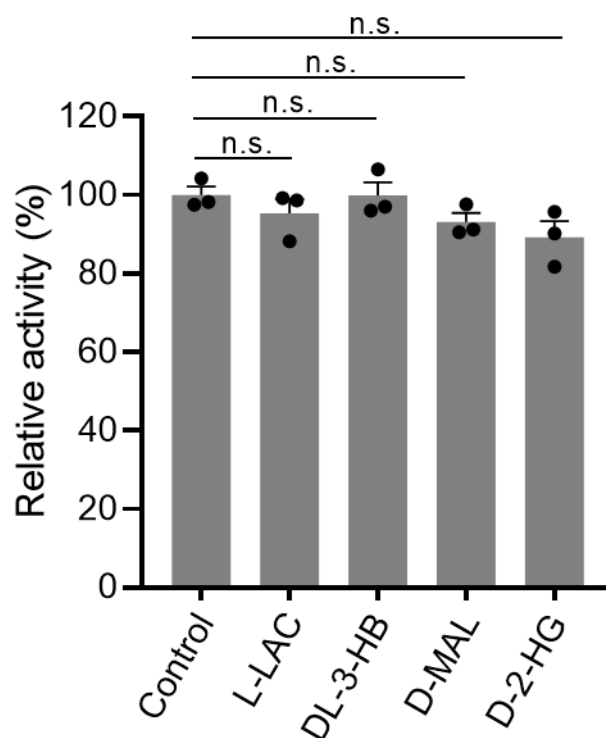
Supplementary Fig 2. Biochemical characterization of mLDHD. **a.** Size-exclusion chromatography analysis of mLDHD. The absorbances at 280 nm and 450 nm are monitored. **b.** SDS-PAGE analysis of mLDHD. The purified mLDHD protein has high purity and homogeneity and exists as a monomer in solution. Source data are provided as a Source Data file.



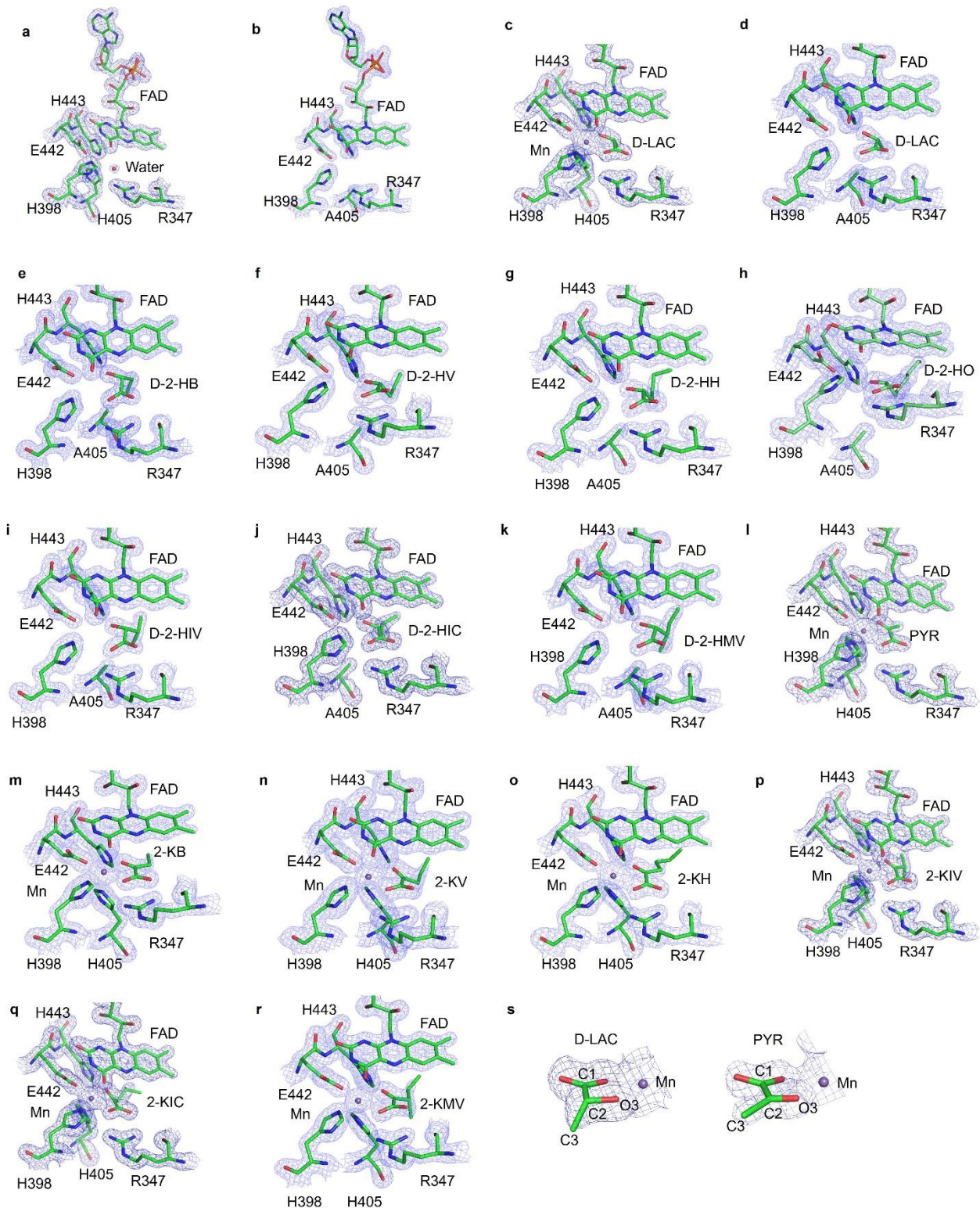
Supplementary Fig 3. Specific activity of mLDHD towards D-LAC with varied Mn²⁺ concentrations at the standard reaction conditions. The error bars show the standard errors of the mean (SEM) of three independent measurements. Source data are provided as a Source Data file.



Supplementary Fig 4. Chemical structures of organic acids used in this study. The common glycolate moiety of 2-hydroxyacids and the common glyoxylate moiety of 2-ketoacids are indicated with dashed boxes. The specific activity of mLDHD towards different substrates (1 mM) is listed beside the substrate structure (n = 3 independent experiments).

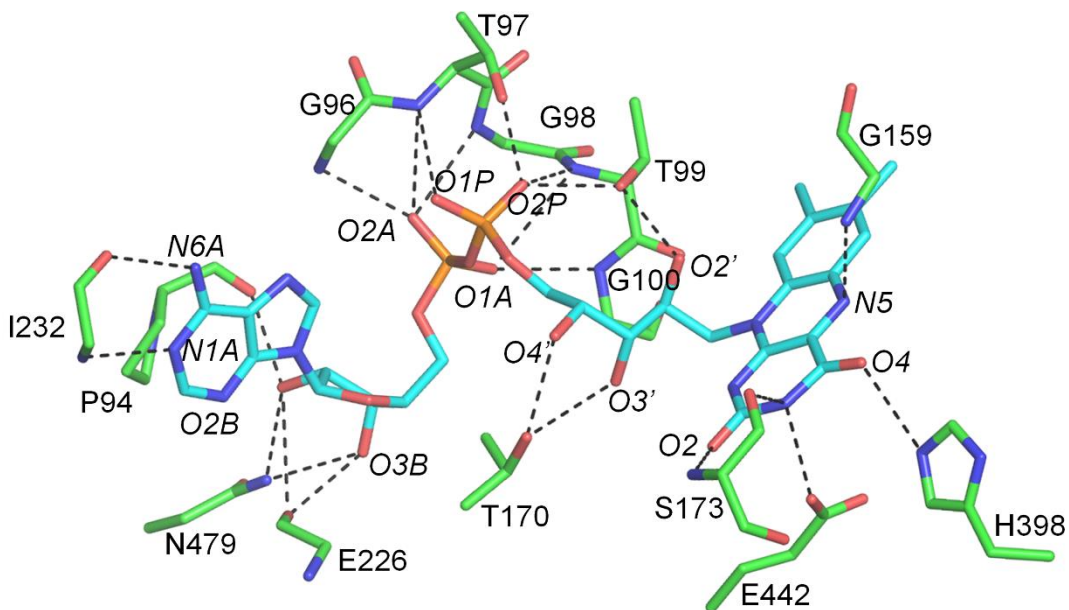


Supplementary Fig 5. Specific activity of mLDHD towards D-LAC in the absence and presence of several substrate analogs. The specific activity of mLDHD towards D-LAC (100 μ M) was measured in the absence (control) and presence of indicated substrate analogs (1 mM) including L-lactate (L-LAC), DL-hydroxybutyrate (DL-3-HB), D-malate (D-MAL), and D-2-hydroxyglutarate (D-2-HG). The data are presented as percentage of the specific activity in the presence of substrate analog relative to that in the absence of substrate analog (control). The error bars represent the standard errors of the mean (SEM) of three independent experiments ($n = 3$). The p values were calculated with two-sided Student's t -test (0.33 for L-LAC, 0.98 for DL-3-HB, 0.09 for D-MAL and 0.08 for D-2-HG). n.s., not significant ($p > 0.05$). Source data are provided as a Source Data file.

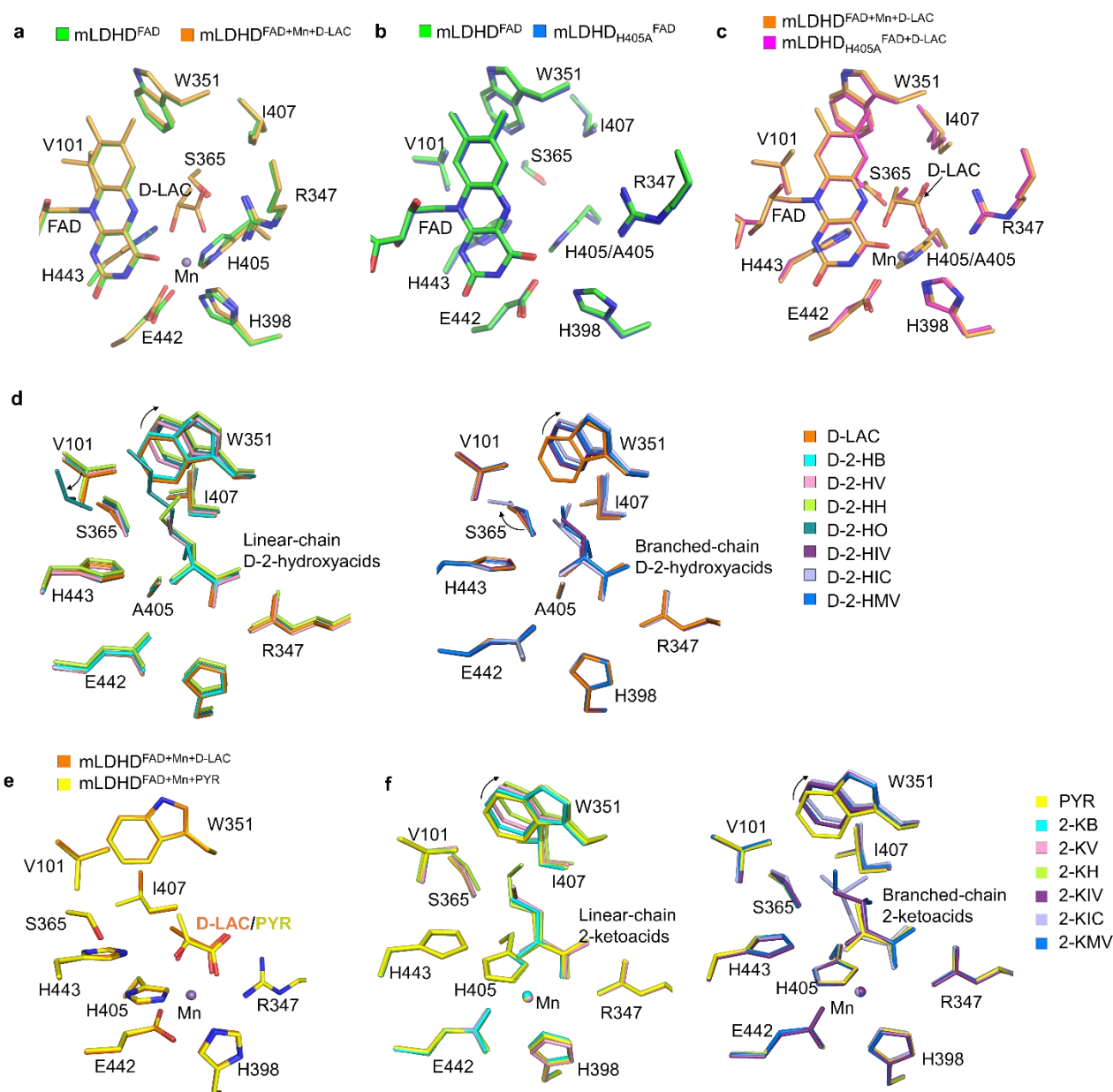


Supplementary Fig 6. Representative simulated annealing composite $2F_o-F_c$ omit maps (1.5σ contour level) for the active sites in different mLDHD structures. The FAD, substrate or product, and key residues at the active site are shown with stick models, and the metal ion and water molecule are shown as cyan sphere and red sphere, respectively. **a.** The structure of the FAD-bound WT mLDHD. **b.** The structure of the FAD-bound mLDHD_{H405A}. **c.** The structure of WT mLDHD in complex with FAD, Mn²⁺ and D-lactate (D-LAC). **d.** The structure of mLDHD_{H405A} in complex with

FAD and D-LAC. **e-k.** The structures of mLDHD_{H405A} in complexes with FAD and different substrates: D-2-hydroxybutyrate (D-2-HB, **e**), D-2-hydroxyvalerate (D-2-HV, **f**), D-2-hydroxyhexanoate (D-2-HH, **g**), D-2-hydroxyoctanoate (D-2-HO, **h**), D-2-hydroxyisovalerate (D-2-HIV, **i**), D-2-hydroxyisocaproate (D-2-HIC, **j**), and D-2-hydroxy-3-methylvalerate (D-2-HMV, **k**). **l-r.** The structures of WT mLDHD in complexes with FAD, Mn²⁺ and different products: pyruvate (PYR, **l**), 2-ketobutyrate (2-KB, **m**), 2-ketovalerate (2-KV, **n**), 2-ketohexanoate (2-KH, **o**), 2-ketoisovalerate (2-KIV, **p**), 2-ketoisocaproate (2-KIC, **q**), and 2-keto-3-methylvalerate (2-KMV, **r**). **s.** Comparison of the electron density maps for D-LAC in the mLDHD^{FAD+Mn+D-LAC} structure and PYR in the mLDHD^{FAD+Mn+PYR} structure showing the co-planarity and non-planarity of the C2 atom with the C1, C3 and O3 atoms, respectively.

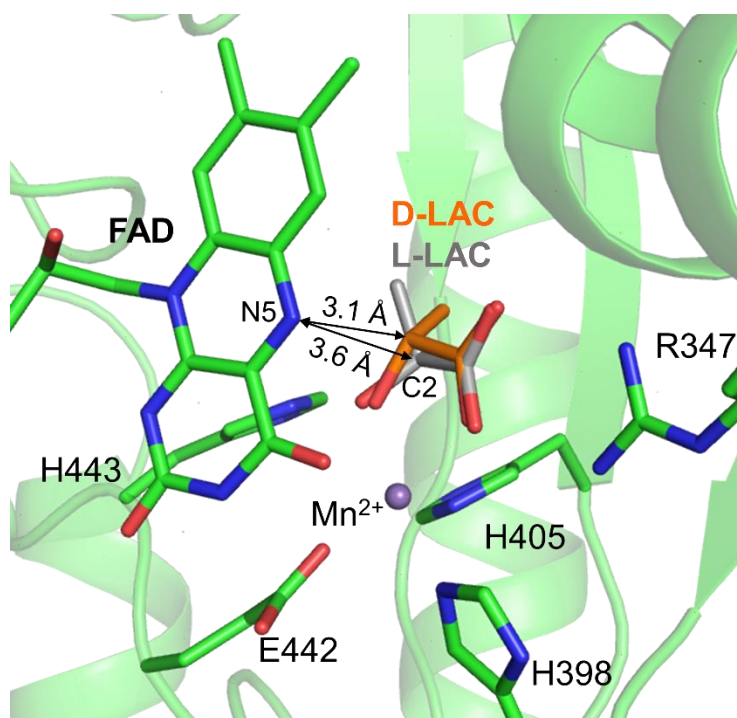


Supplementary Fig 7. Hydrogen-bonding interactions between FAD and the surrounding residues in the FAD-bound WT mLDHD structure. FAD is shown as cyan stick and the protein residues in green stick. The hydrogen bonds are indicated by dashed lines.

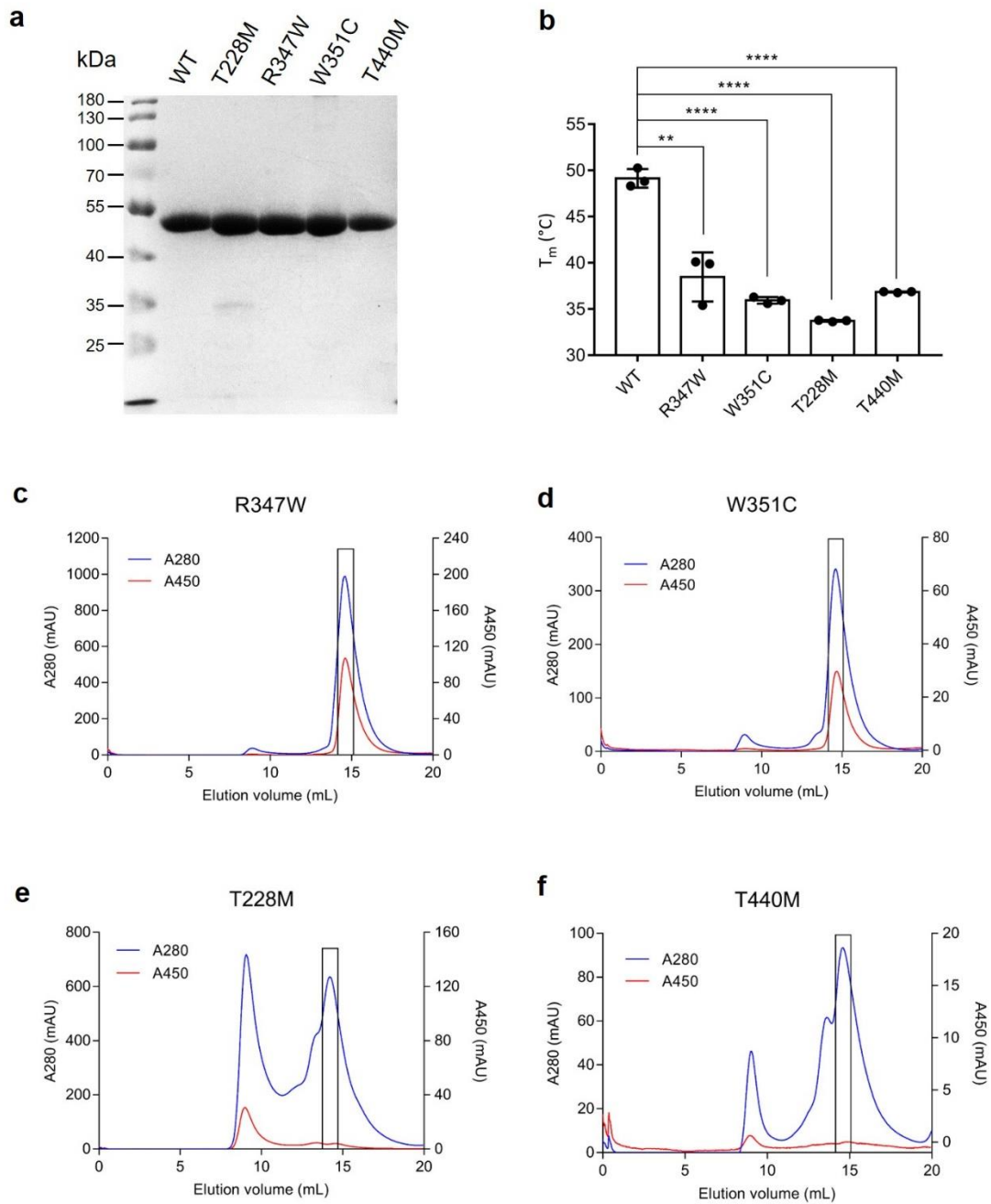


Supplementary Fig 8. Comparisons of the active sites in different mLDHD structures. a. Comparison of the active sites in the WT mLDHD^{FAD} and mLDHD^{FAD+Mn+D-LAC} structures. **b.** Comparison of the active sites in the WT mLDHD^{FAD} and mutant mLDHD_{H405A}^{FAD} structures. **c.** Comparison of the active sites in the WT mLDHD^{FAD+Mn+D-LAC} and mutant mLDHD_{H405A}^{FAD+D-LAC} structures. **d.** Comparison of the active sites in different substrate-bound mLDHD_{H405A} structures showing the conformational changes of the key residues forming the substrate-binding subsite B to varied extents along with the increase of the size of the hydrophobic moiety attached to the C2 atom of substrate. Left: comparison of the active sites in structures bound with D-LAC and other D-2-hydroxyacid substrates with linear aliphatic moieties including D-2-hydroxybutyrate (D-2-HB), D-2-hydroxyvalerate (D-2-HV), D-2-hydroxyhexanoate (D-2-HH), and D-2-hydroxyoctanoate (D-2-HO). Right: comparison of the active sites in structures bound with D-LAC and other D-2-hydroxyacid substrates with branched aliphatic moieties including D-2-hydroxyisovalerate (D-2-HIV), D-2-hydroxyisocaproate (D-2-HIC), and D-2-hydroxy-3-methylvalerate (D-2-HMV). **e.** Comparison of

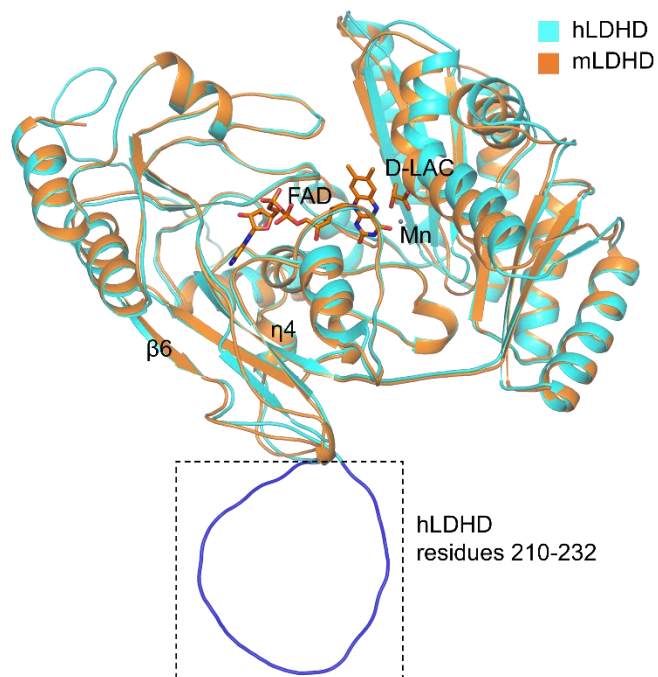
the active sites in the WT mLDHD^{FAD+Mn+D-LAC} and mLDHD^{FAD+Mn+PYR} structures. **f.** Comparison of the active sites in different product-bound mLDHD structures showing the conformational changes of the key residues forming the substrate-binding subsite B to varied extents along with the increase of the size of the hydrophobic moiety attached to the C2 atom of product. Left: comparison of the active sites in structures bound with PYR and other 2-ketoacid products with linear aliphatic moieties including 2-ketobutyrate (2-KB), 2-ketovalerate (2-KV), and 2-ketohexanoate (2-KH). Right: comparison of the active sites in structures bound with PYR and other 2-ketoacid products with branched aliphatic moieties including 2-ketoisovalerate (2-KIV), 2-ketoisocaproate (2-KIC), and 2-keto-3-methylvalerate (2-KMV).



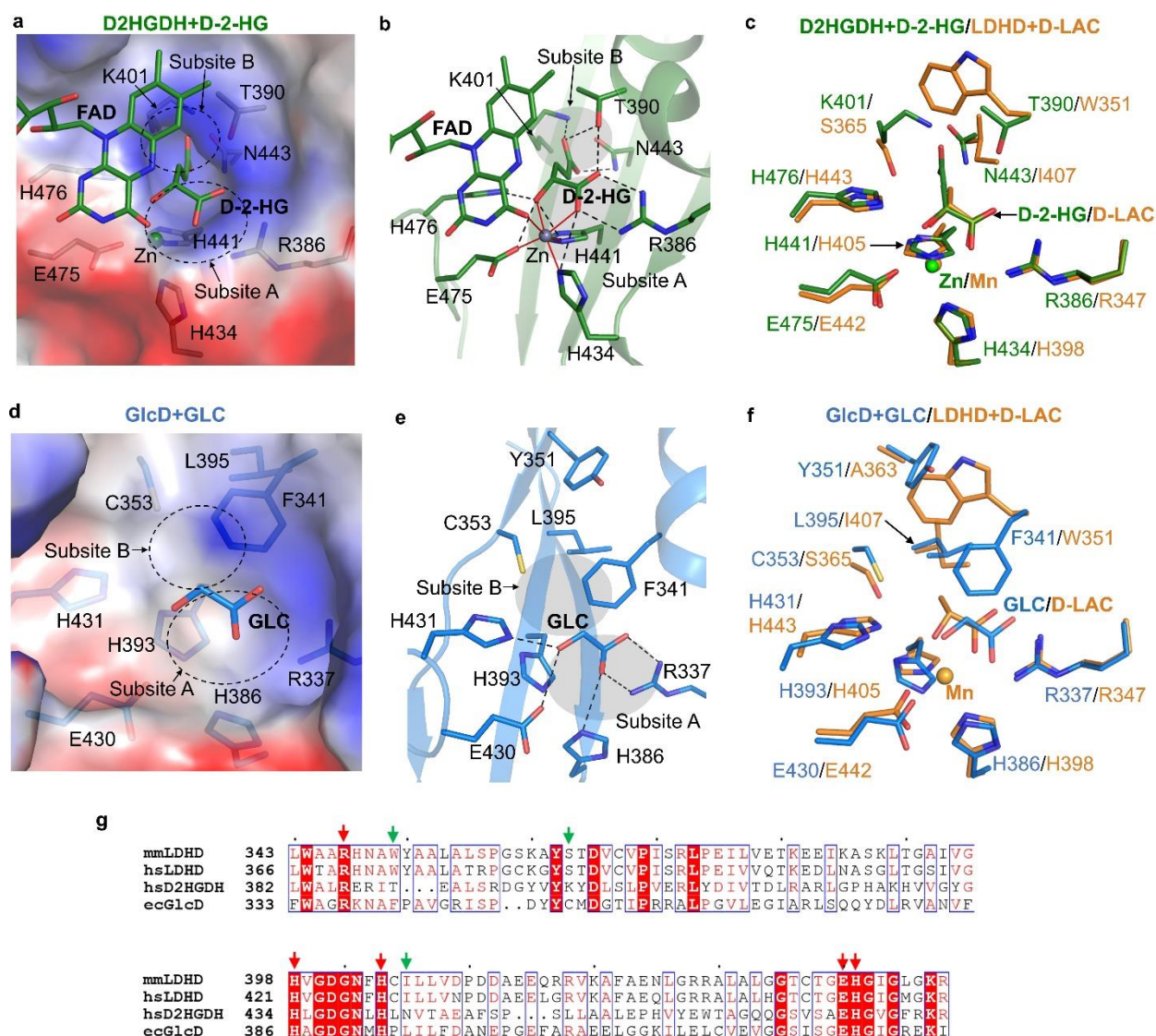
Supplementary Fig 9. Docking of L-LAC into the active site of the mLDHD^{FAD+Mn+D-LAC} structure. The docked L-lactate (L-LAC, grey) and the bound D-lactate (D-LAC, orange) are superimposed.



Supplementary Fig 10. Biochemical analysis of the mLDHD mutants containing mutations corresponding to the disease-associated mutations of hLDHD. **a.** SDS-PAGE analysis of WT mLDHD and the T228M, R347C, W351C, and T440M mLDHD mutants. **b.** Thermostability analysis of WT mLDHD and the T228M, R347C, W351C, and T440M mLDHD mutants ($n = 3$ independent experiments). The p values were calculated with two-sided Student's t -test. **, $p < 0.01$ (2.9×10^{-3} for R347W); ****, $p < 0.0001$ (2.8×10^{-5} for W351C, 1.2×10^{-5} for T228M and 3.0×10^{-5} for T440M). **c-f.** Elution curves of the R347C (**c**), W351C (**d**), T228M (**e**), and T440M (**f**) mLDHD mutants from the size-exclusion chromatography analysis. The absorbances at 280 nm and 450 nm were monitored. The peaks corresponding to the target protein are indicated by boxes. Source data are provided as a Source Data file.



Supplementary Fig 11. Comparison of the Alphafold2 predicted hLDHD structure (Q86WU2) and the mLDHD^{FAD+Mn+D-LAC} structure. The Alphafold2 predicted hLDHD structure is colored in cyan with the insertion in blue. The mLDHD^{FAD+Mn+D-LAC} structure is colored in orange.



Supplementary Fig 12. Comparison of the substrate-binding sites of LDHD, D2HGDH and GlcD. **a.** Electrostatic surface of the substrate-binding site of human D2HGDH (hsD2HGDH) bound with D-2-HG and Zn^{2+} (PDB 6LPP). **b.** Interactions between D-2-HG, Zn^{2+} and the surrounding residues. The hydrogen bonds are shown in black dashed lines and the coordinate bonds are shown in red solid lines. **c.** Superposition of the key residues composing the substrate-binding sites of mouse LDHD and human D2HGDH bound with D-LAC and D-2-HG, respectively. **d.** Electrostatic surface of the substrate-binding site of the predicted model of *E. coli* GlcD (ecGlcD) bound with substrate glycolate (GLC). **e.** Interactions between GLC and the surrounding residues. The hydrogen bonds are shown in black dashed lines. **f.** Superposition of the key residues composing the substrate-binding sites of mouse LDHD and *E. coli* GlcD with bound substrates D-LAC and GLC, respectively. **g.** Sequence alignment of mmLDHD, hsLDHD, hsD2HGDH, and ecGlcD showing the key residues forming the substrate-binding site. Residues located at the substrate-binding subsite A are indicated by red arrows, and those at the substrate-binding subsite B are indicated by green arrows.

Supplementary Table 1. Functional roles of LDHD mutations in the catalytic reaction and pathogenesis.

mLDHD	hLDHD	Specific activity ($\mu\text{m}\cdot\text{min}^{-1}\cdot\text{mg}^{-1}$) ^a	Residual activity compared to WT	Structure location	Functional role
WT	-	1.083 \pm 0.003	-	-	-
Active-site mutations					
R347A	R370A	N.D. ^b	N.D.	α 9	Impair substrate binding
H398A	H421A	N.D.	N.D.	β 13- η 5 loop	Impair metal ion binding
H405A	H428A	N.D.	N.D.	β 14	Impair metal ion binding
E442A	E465A	N.D.	N.D.	α 11- α 12 loop	Impair metal ion binding
H443A	H466A	N.D.	N.D.	α 11- α 12 loop	Impair substrate binding
Disease-associated mutations					
T228M	T251M	0.024 \pm 0.02	2.2%	η 4- β 7 loop	Impair FAD binding
R347W	R370W	0.045 \pm 0.027	4.2%	α 9	Impair substrate binding
W351C	W374C	0.127 \pm 0.014	11.8%	α 9	Impair substrate binding
T440M	T463M	0.014 \pm 0.005	1.3%	α 11- α 12 loop	Impair FAD binding

^a The specific activities of WT and mutant mLDHD enzymes were measured at the standard conditions: 50 mM Tris-HCl (pH 7.4), 2 μ g enzyme, 50 μ M MnCl₂, 200 μ M PMS, 100 μ M DCIP, and 1 mM D-lactate. The molar ratio of FAD:protein was determined to be 0.73 \pm 0.04 for WT mLDHD, 0.44 \pm 0.02 for the W351C mutant, and 0.48 \pm 0.008 for the R347W mutant, respectively. The specific activities of WT and mutant mLDHD enzymes were corrected according to the concentration of active enzyme in the reaction solution calculated based on the FAD occupancy. The specific activity values are presented as the mean \pm SEM (n = 3 independent experiments).

^b N.D., not detectable.