

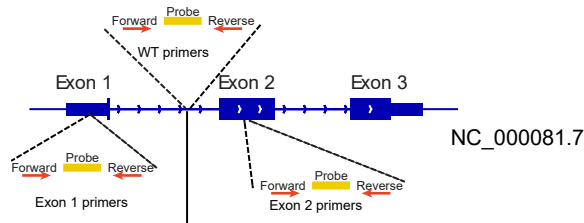
**Supplemental information**

**Premature aging and reduced cancer incidence  
associated with near-complete  
body-wide *Myc* inactivation**

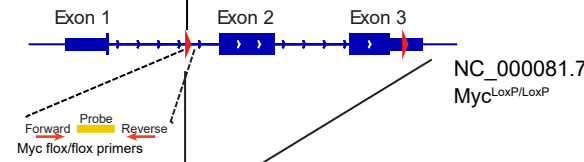
**Huabo Wang, Jie Lu, Taylor Stevens, Alexander Roberts, Jordan Mandel, Raghunandan Avula, Bingwei Ma, Yijen Wu, Jinglin Wang, Clinton Van't Land, Toren Finkel, Jerry E. Vockley, Merlin Airik, Rannar Airik, Radhika Muzumdar, Zhenwei Gong, Michel S. Torbenson, and Edward V. Prochownik**

Fig. S1

A



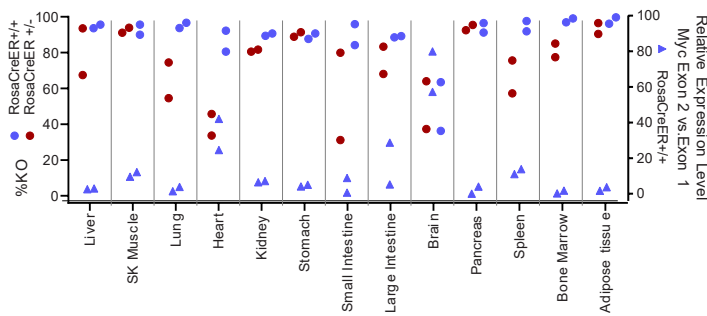
B



C



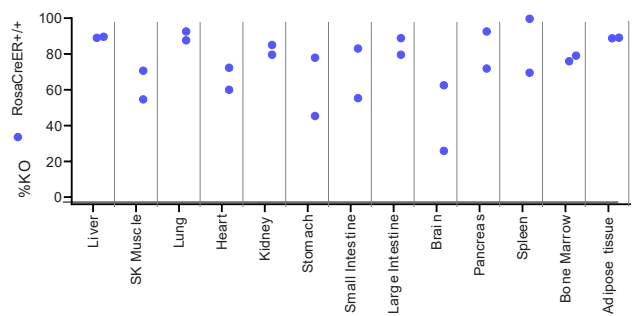
E



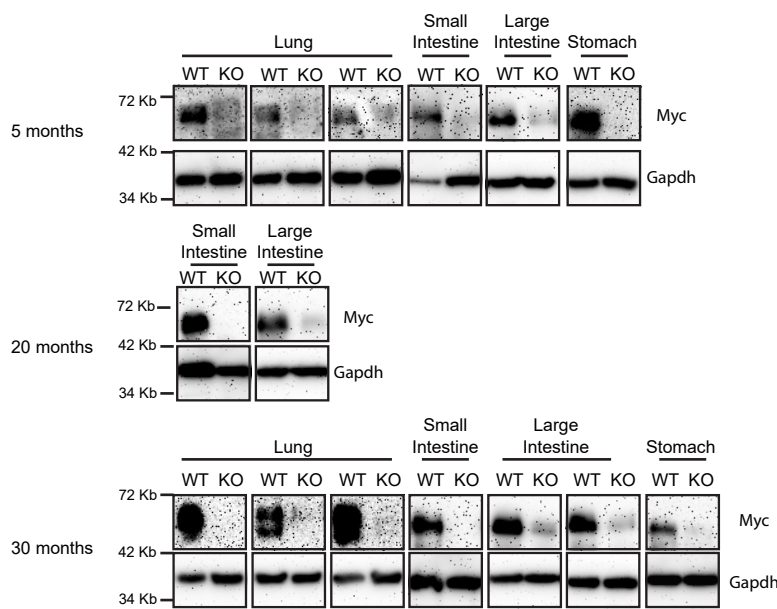
D

		Start (bp)	End (bp)	Sequence (5' -3')
WT (NC_000081.7)	Forward	61858492	61858513	GGGAATCCTCACATTCTACTT
	Reverse	61858643	61858626	GATTCAGCACTGGGTGCA
	Probe	61858549	61858572	/56-FAM/TAGGAAGACTGCGGTGAGTCGTGA/3IABkFQ/
Myc <sup>loxP/loxP</sup> (NC_000081.7)	Forward	61858570	61858588	TGATCTGAGCGGTCCGTA
	Reverse	61858697	61858675	CACCTCCCTTCTACACTCTAAAC
	Probe	Loxp	Loxp	/56-FAM/CCTGCACGA/ZEN/TCCGGAACCCCTTAAT/3IABkFQ/
Myc <sup>-/-</sup> (KO) (NC_000081.7)	Forward	61858570	61858588	TGATCTGAGCGGTCCGTA
	Reverse	61858697	61858675	TAAAGTCCCAAAGACTCCAG
	Probe	Loxp	Loxp	/56-FAM/CCTGCACGA/ZEN/TCCGGAACCCCTTAAT/3IABkFQ/
Exon 1 (NC_000081.7)	Forward	61857404	61857425	GCTGTAGTAATCCAGCGAGAG
	Reverse	61857507	61857488	ACTCCAGAGTGCCCTCTTA
	Probe	61857447	61857468	/56-FAM/TTGGAAGAG/ZEN/CCGTGTGTCCAGA/3IABkFQ/
Exon 2 (NC_000081.7)	Forward	61859421	61859442	CTCCGTACAGCCCTATTTTCATC
	Reverse	61859542	61859524	TGGGAAGCAGCTCGAATTT
	Probe	61859464	61859480	/56-FAM/TATCACCAG/ZEN/CAACAGCA
Cre (OM228709.1)	Forward	3366	3344	GCGGTCTGGCAGTAAAACTATC
	Reverse	3265	3287	GTGAAACAGCATTGCTGTCACTT
	Probe	3323	3300	/56-FAM/AAACATGCT/ZEN/TCATCGTCCGG/3IABkFQ/
Rosa WT (NC_000072.7)	Forward	113053072	113053056	CTGGCTTCTGAGGACCG
	Reverse	113052875	113052894	CCGAAAATCTGTGGGAAGTC
	Probe	113052960	113052983	/56-FAM/TTTAAGCT/ZEN/GCCCAAGACTCCC/3IABkFQ/

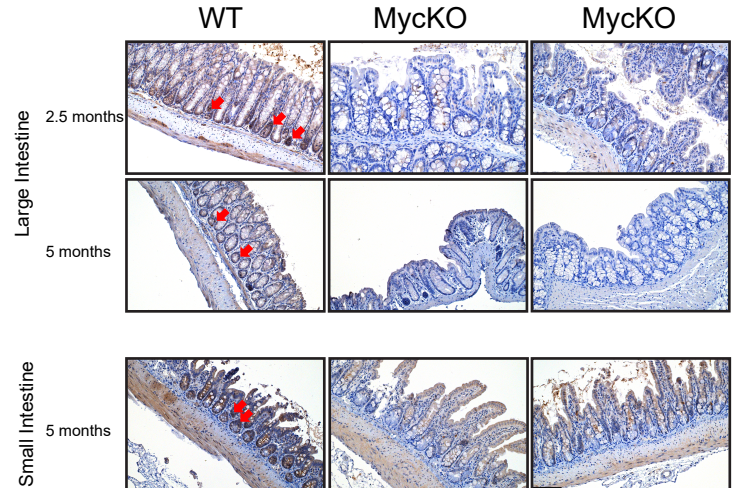
F



G



H





**Figure S1. Strategy for qPCR-based Taq-Man-based assay for identifying *Myc* alleles, related to STAR Methods.**

(A, B). The *Myc* loci of WT and *myc<sup>loxP/loxP</sup>* (B6.129S6-*Myc<sup>tm2Fwa</sup>*/Mmjax) mice. LoxP sites are indicated by red triangles. In A, the red arrows indicate the primer sets used to amplify the WT allele. The reverse primer in intron 1 overlaps the inserted LoxP site in the (*Myc<sup>loxP/loxP</sup>*) allele shown in B. Amplification with this primer set is thus possible only in WT mice and generates a PCR fragment detected with the unique TaqMan probe. In B, red arrows indicate the primer set flanking the 5' LoxP site that specifically amplifies this region and is detected with TaqMan probe.

(C). The excised and recombined *Myc* locus following CreER activation.

(D). Sequences and genomic locations for primers and probes shown in A-C as well as the qRT-PCR primers and probes used for qRT-PCR. Annotations for primers and probes are based on the numbering of the murine genomic sequence: <https://www.ncbi.nlm.nih.gov/gene/17869>.

(E). qPCR and qRT-PCR-based quantification of *Myc* locus excision and expression in ~6-7 wk old mice that had been treated for 5 days with tamoxifen at the time of weaning (~4 wks and weights >15 g). 2 mice with 2 copies of CreER and 2 with a single copy of CreER were used to determine how *Myc* locus excision efficiency was influenced by CreER copy number. After confirming the number of CreER alleles using the TaqMan-based approach described above, the WT:KO *Myc* allele ratio was determined from a standard curve of known amounts of each DNA (Wang et al., 2018; Wang et al., 2022b)(Wang et al., 2018; Wang et al., 2022b). qRT PCR reactions for *Myc* transcripts (<https://www.ncbi.nlm.nih.gov/gene/17869>) were performed on RNAs extracted from tissues of mice with two copies of ROSA-CreER as previously described

<sup>1,2</sup>

(F). Persistence of *Myc* locus deletion. qPCR performed as described in E on the indicated

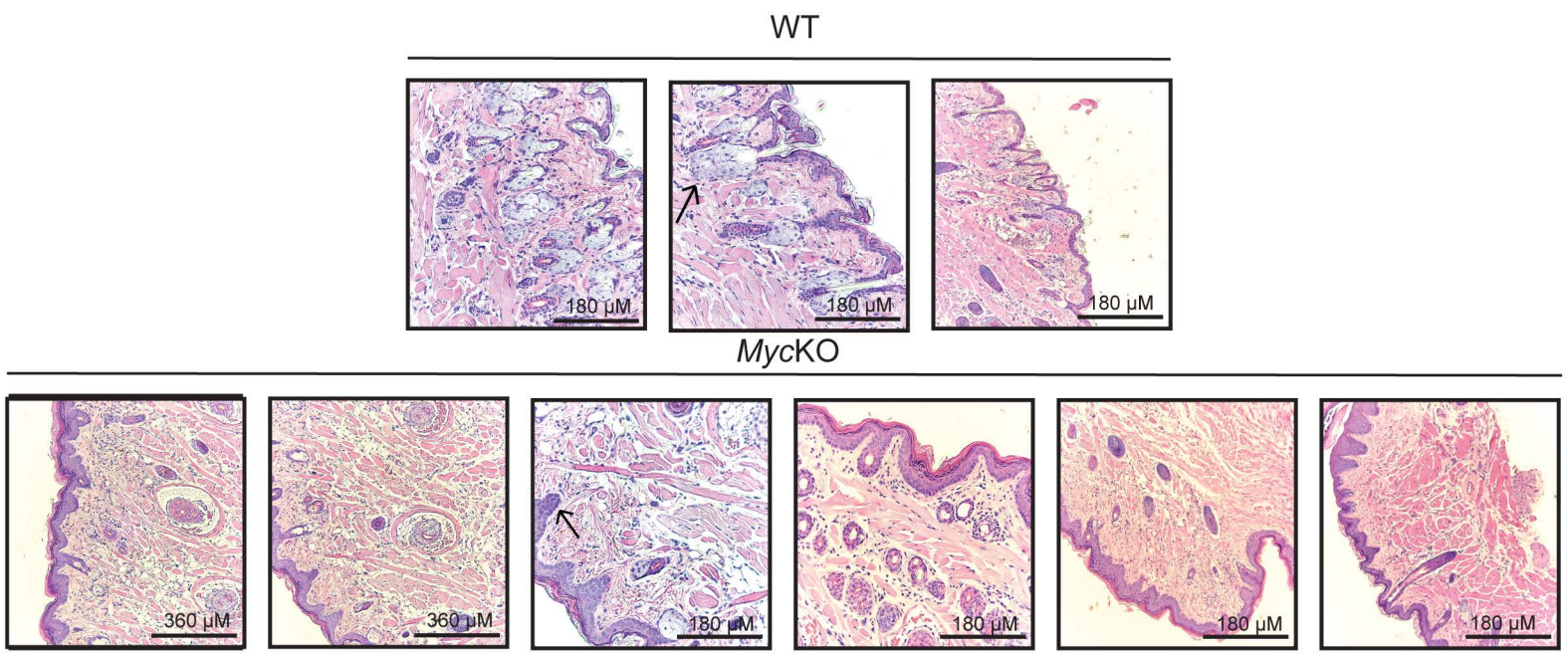
tissues from 20-22 month-old *Myc*KO mice. See Table S1 for a summary of qPCR and qRT-PCR results performed on various tissues of additional mice.

(G). Immuno-blotting for *Myc* protein in the indicated tissues from 5-6 months, 20-22 months and 30-33 months old WT and *Myc*KO mice.

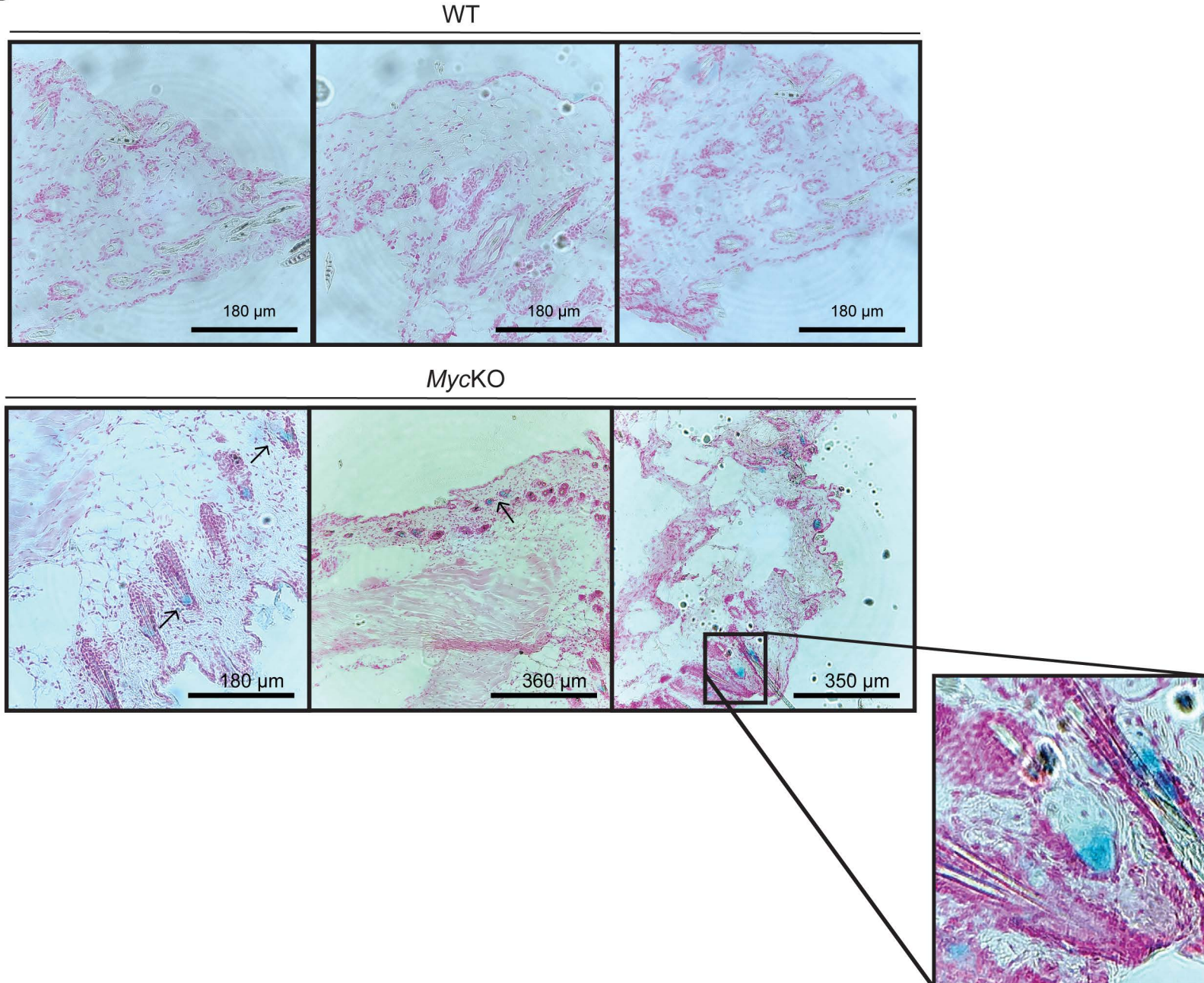
(H). Immunohistochemical staining for *Myc* protein in the small and large intestines of WT and *Myc*KO mice. Note, in both tissues, the absence of detectable *Myc* in the crypts which is where the highest levels of expression are confined. <sup>3</sup> Scale bar = 100  $\mu$ m.

Fig. S2

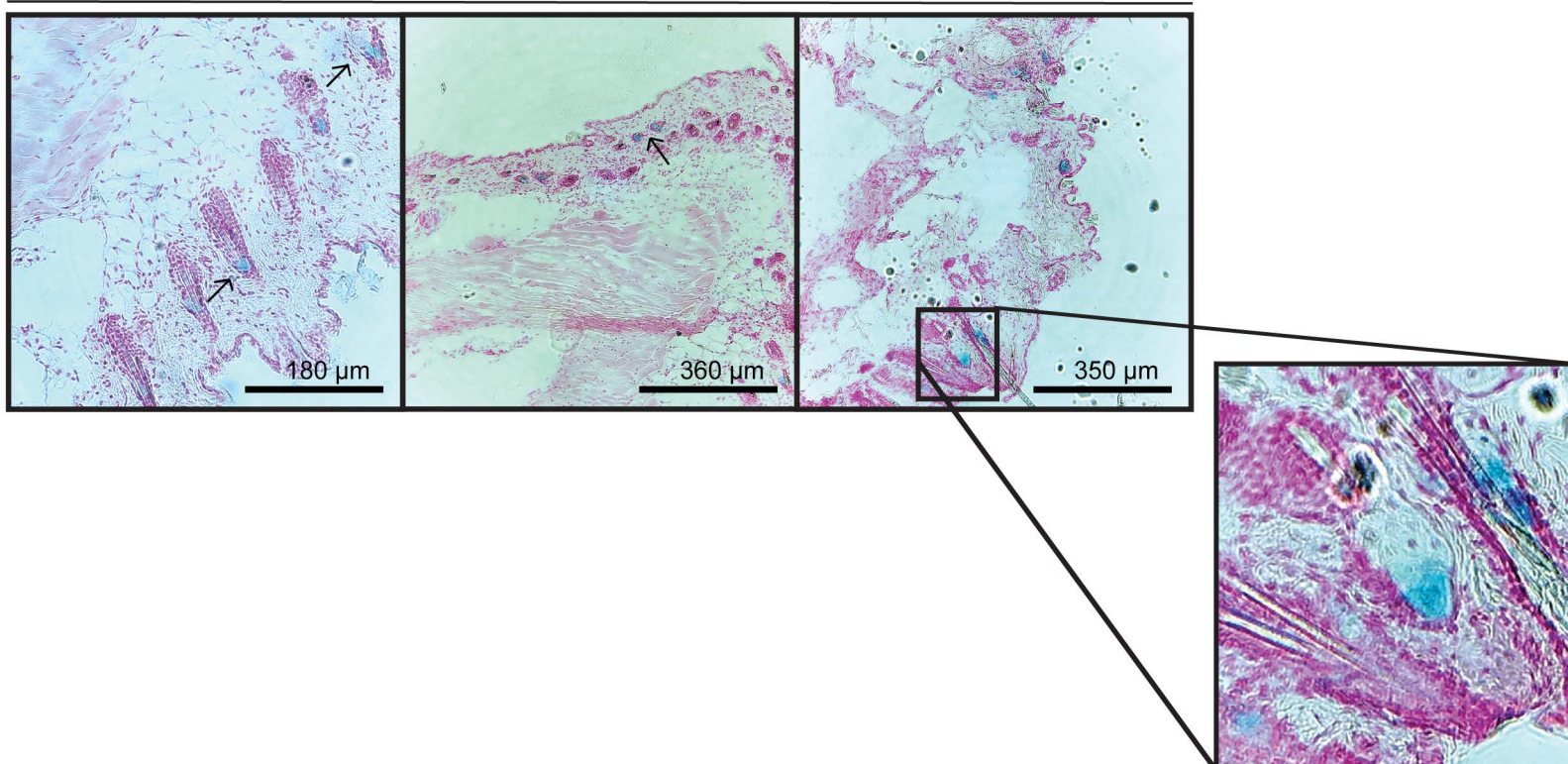
A



B



*MyckKO*

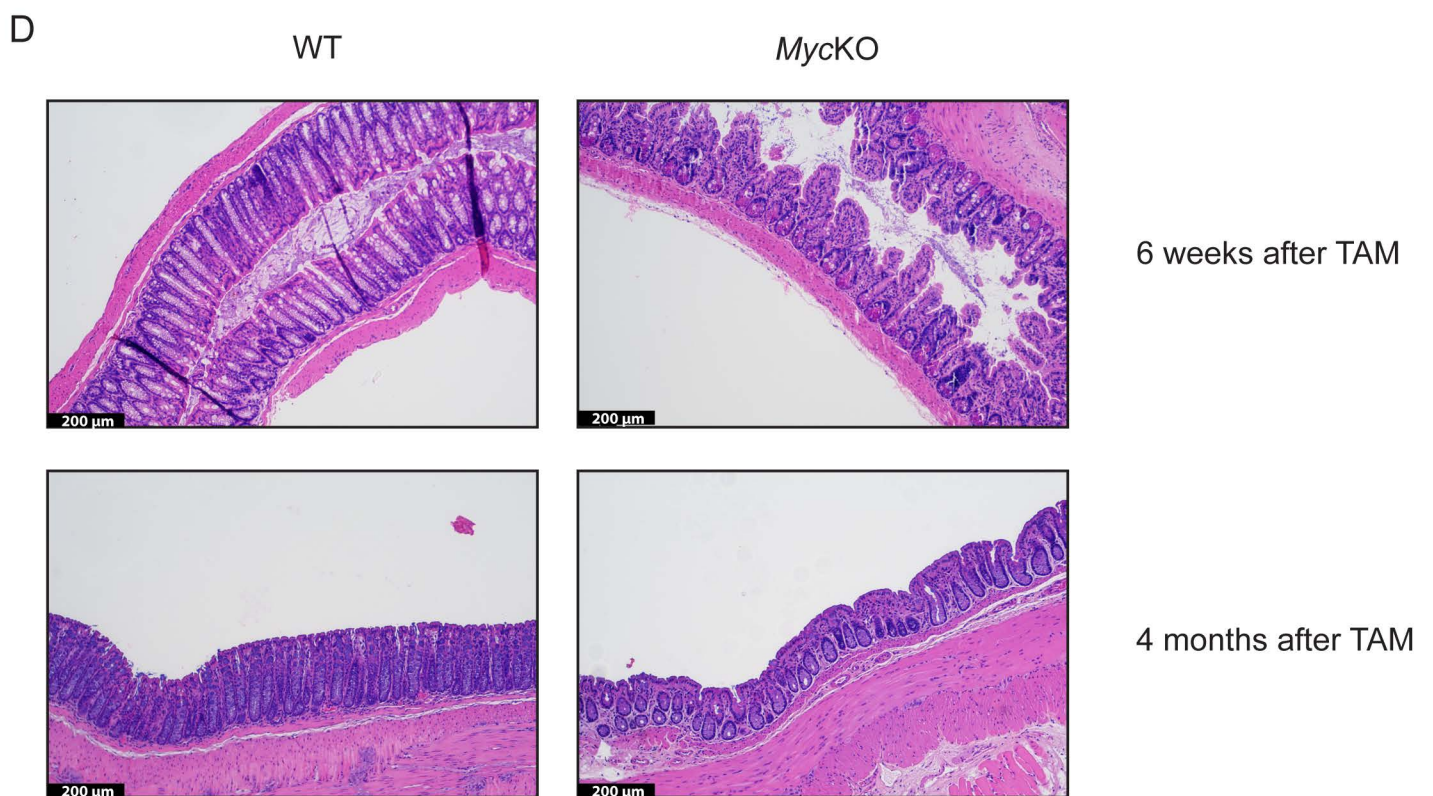
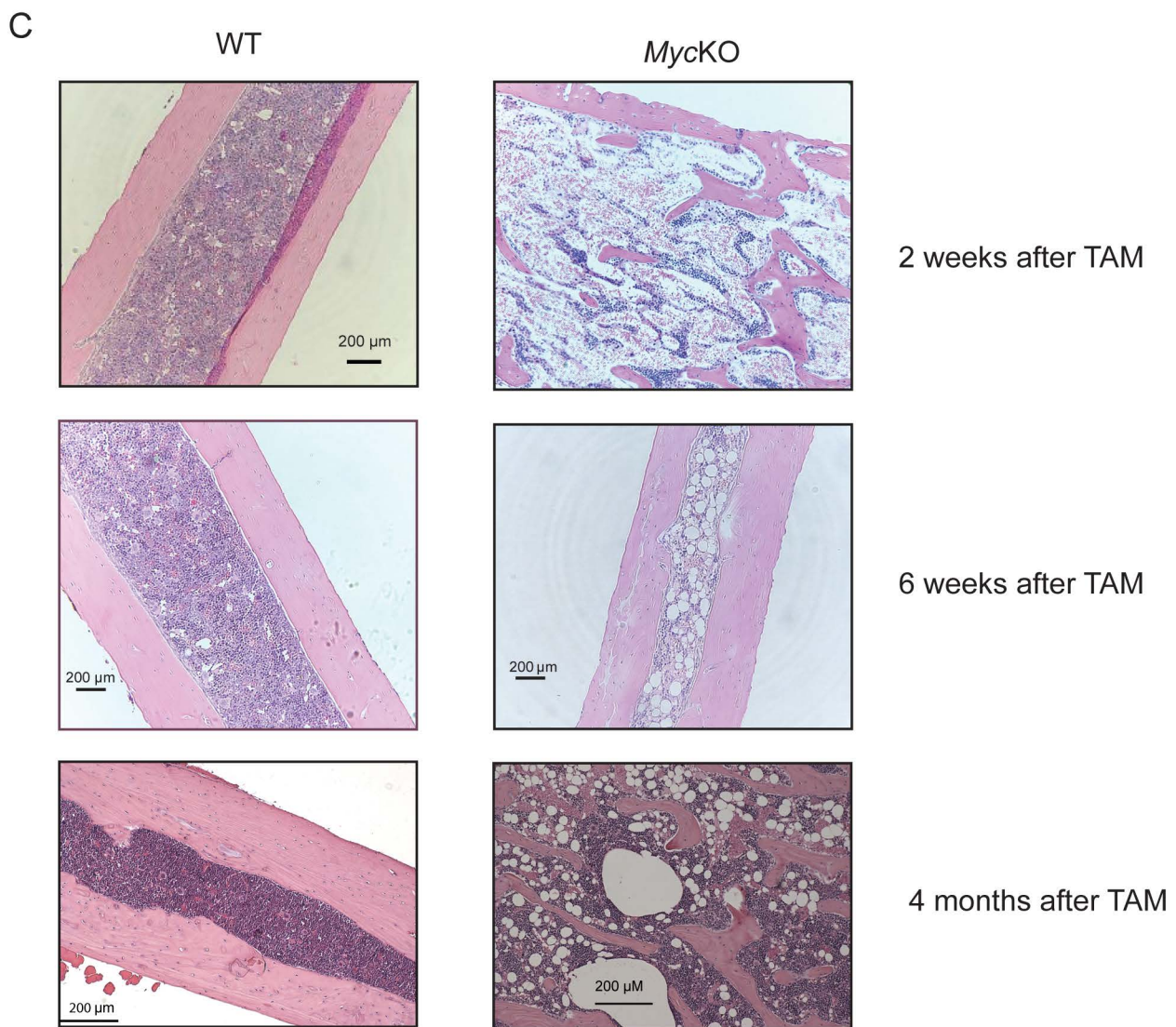
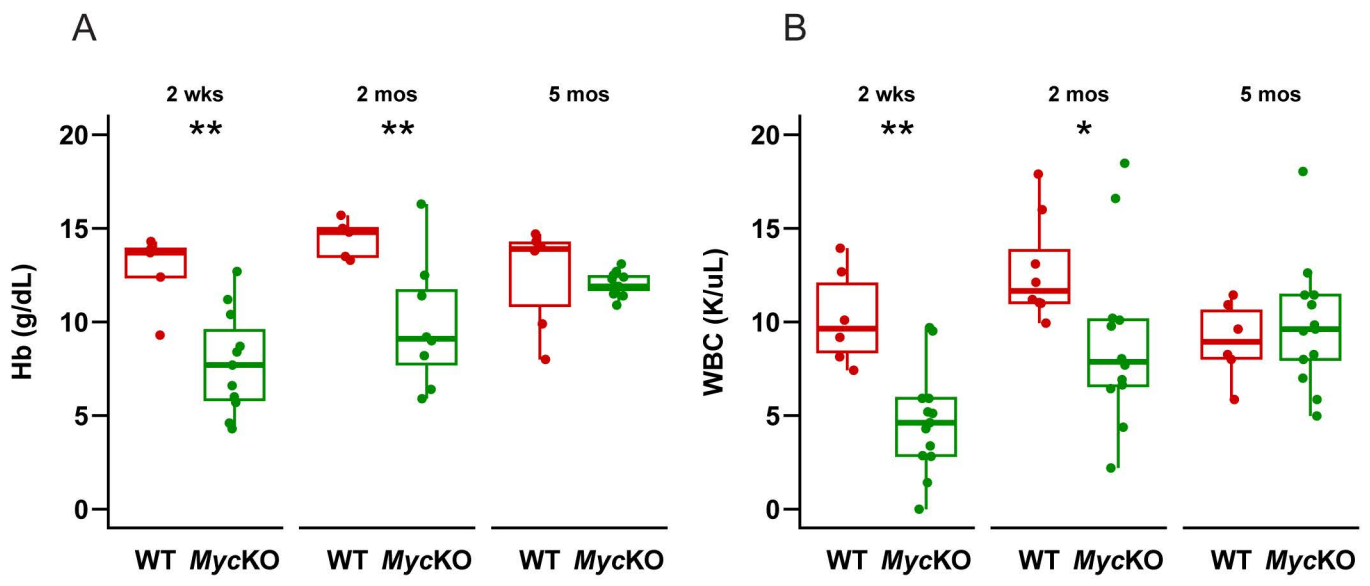


**Figure S2. Histopathology of skin from alopecic regions of *MyckKO* mice and premature onset of senescence, related to Figure 1B&C.**

(A). H&E-stained sections of peri-orbital skin from a 5 mo *MyckKO* mouse and normal skin from the corresponding region of a WT control mouse. In WT panels, the arrow shows normal hair follicles and their adjacent sebaceous glands. In *MyckKO* panels, the arrows show thickening and hyperkeratinization of the epidermis, loss of surface invaginations and a generalized paucity of hair follicles and sebaceous glands. (B). SA- $\beta$ -gal staining of the same areas from the WT and *MyckKO* mice shown in A. Arrows indicate SA- $\beta$ -gal-positive cells in the latter samples adjacent to rare hair follicles. Most of SA- $\beta$ -gal-positive cells detected resided within the inner and outer root sheath and base of the follicle (<http://eulep.pdn.cam.ac.uk/~skinbase/>). A higher power magnification of one of the fields containing SA- $\beta$ -gal-positive cells is shown to the right.



Fig. S3



**Figure S3. *Myck*KO mice develop transient mild-moderate, anemia, leukopenia and bone marrow hypoplasia, related to Figure 1&3.**

(A). Hemoglobin levels in WT and age-matched *Myck*KO mice at the indicated ages, which are expressed relative to the time of starting 4-hydroxytamoxifen therapy. N = 6-14 at each age, Unpaired t test, \*= $p < 0.05$ , \*\*= $p < 0.01$ .

(B). Peripheral white blood cell counts performed at the times shown in A.

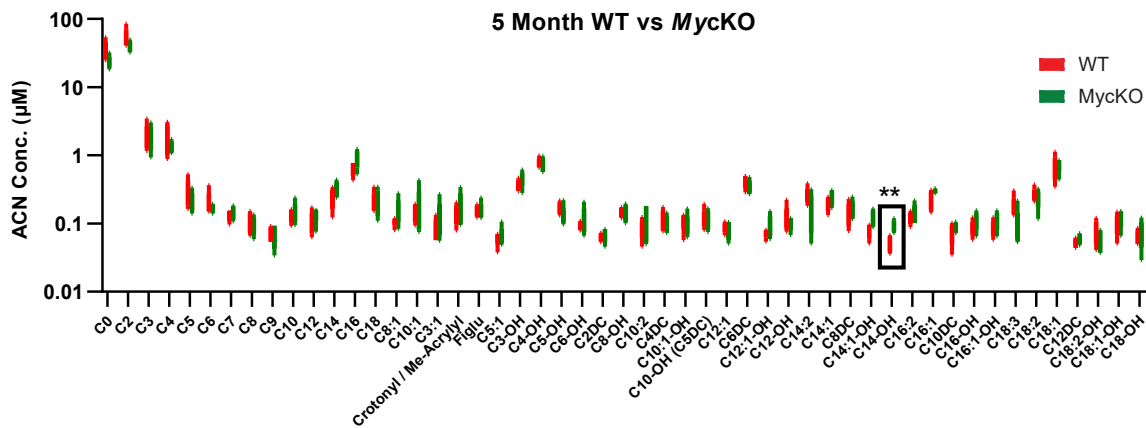
(C). H&E-stained bone marrows of WT and *Myck*KO mice of the indicated ages.

(D). H&E-stained colonic tissues from WT and *Myck*KO mice of the indicated ages.

Scale bar = 200  $\mu$ m

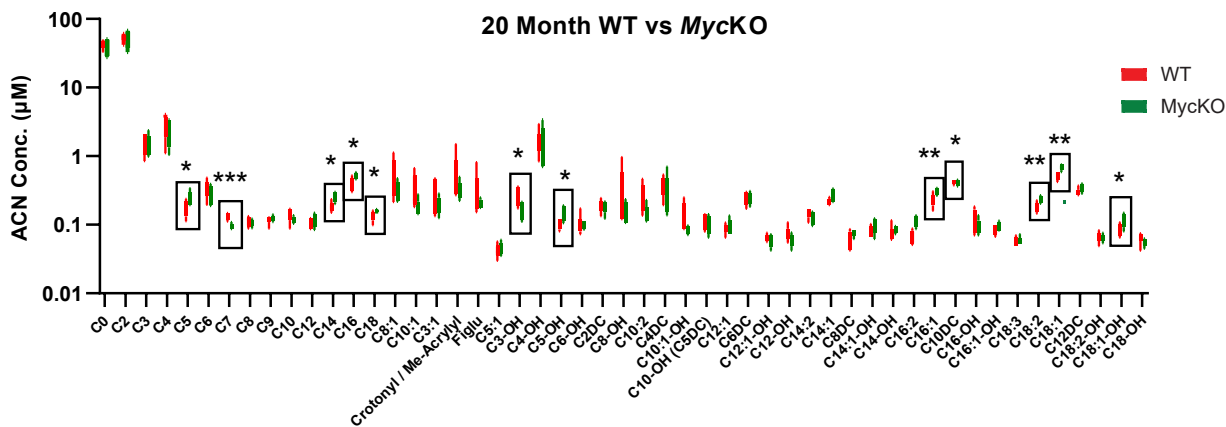
Fig. S4

A



	Below threshold?	P value		Below threshold?	P value		Below threshold?	P value	
C0	No	0.082805	Crotonyl / Me-Acrylyl	No	0.341981	C14:2	No	0.110939	
C2	No	0.107371	Figlu	No	0.931718	C14:1	No	0.174621	
C3	No	0.891553	C5:1	No	0.726059	C8DC	No	0.640394	
C4	No	0.310665	C3-OH	No	0.206181	C14:1-OH	No	0.052795	
C5	No	0.243173	C4-OH	No	0.735646	C14-OH	Yes	**	0.003299
C6	No	0.395319	C5-OH	No	0.158956	C16:2	No	0.426658	
C7	No	0.342432	C6-OH	No	0.359565	C16:1	No	0.066959	
C8	No	0.408316	C2DC	No	0.437708	C10DC	No	0.267349	
C9	No	0.932381	C8-OH	No	0.815773	C16-OH	No	0.179856	
C10	No	0.639306	C10:2	No	0.357354	C16:1-OH	No	0.179856	
C12	No	0.252238	C4DC	No	0.519504	C18:3	No	0.093283	
C14	No	0.123141	C10:1-OH	No	0.495535	C18:2	No	0.605520	
C16	No	0.112089	C10-OH (C5DC)	No	0.804210	C18:1	No	0.788717	
C18	No	0.812097	C12:1	No	0.551986	C12DC	No	0.268356	
C8:1	No	0.179578	C6DC	No	0.184811	C18:2-OH	No	0.243451	
C10:1	No	0.757984	C12:1-OH	No	0.258790	C18:1-OH	No	0.368712	
C3:1	No	0.365815	C12-OH	No	0.468176	C18-OH	No	0.548166	

B



	Below threshold?	P value		Below threshold?	P value		Below threshold?	P value		
C0	No	0.301834	Crotonyl / Me-Acrylyl	No	0.264035	C14:2	No	0.197887		
C2	No	0.611193	Figlu	No	0.333312	C14:1	No	0.073848		
C3	No	0.856345	C5:1	No	0.498076	C8DC	No	0.096298		
C4	No	0.485919	C3-OH	Yes	*	0.016120	C14:1-OH	No	0.177224	
C5	Yes	*	0.035634	C4-OH	No	0.768323	C14-OH	No	0.335738	
C6	No	0.260523	C5-OH	Yes	*	0.012020	C16:2	Yes	**	0.003900
C7	Yes	***	0.000783	C6-OH	No	0.996976	C16:1	Yes	*	0.014118
C8	No	0.497159	C2DC	No	0.720109	C10DC	No	0.686387		
C9	No	0.496558	C8-OH	No	0.294558	C16-OH	No	0.293995		
C10	No	0.222842	C10:2	No	0.137534	C16:1-OH	No	0.150042		
C12	No	0.359745	C4DC	No	0.413080	C18:3	No	0.969488		
C14	Yes	*	0.013240	C10:1-OH	No	0.125231	C18:2	Yes	**	0.008329
C16	Yes	*	0.044914	C10-OH (C5DC)	No	0.590178	C18:1	Yes	**	0.003805
C18	Yes	*	0.037790	C12:1	No	0.708297	C12DC	No	0.117119	
C8:1	No	0.319791	C6DC	No	0.770883	C18:2-OH	No	0.595252		
C10:1	No	0.124682	C12:1-OH	No	0.590031	C18:1-OH	Yes	*	0.043729	
C3:1	No	0.442019	C12-OH	No	0.130382	C18-OH	No	0.225025		

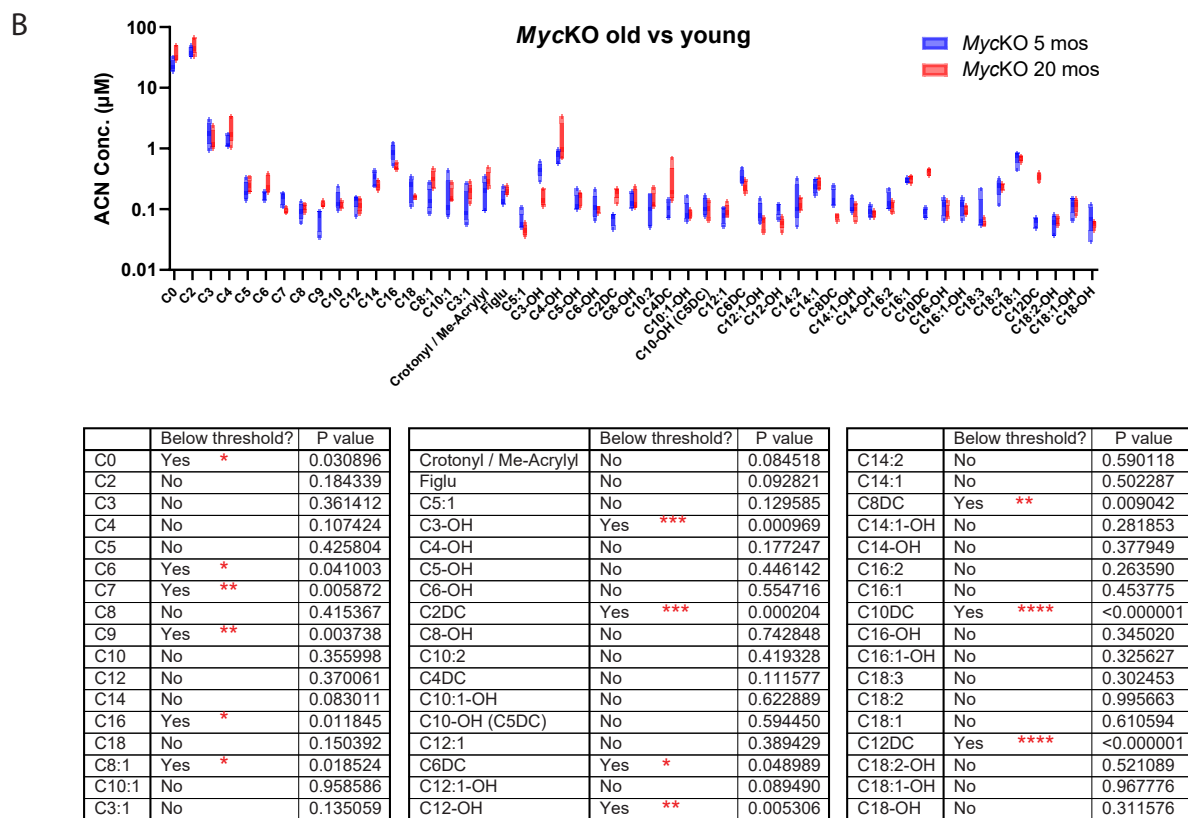
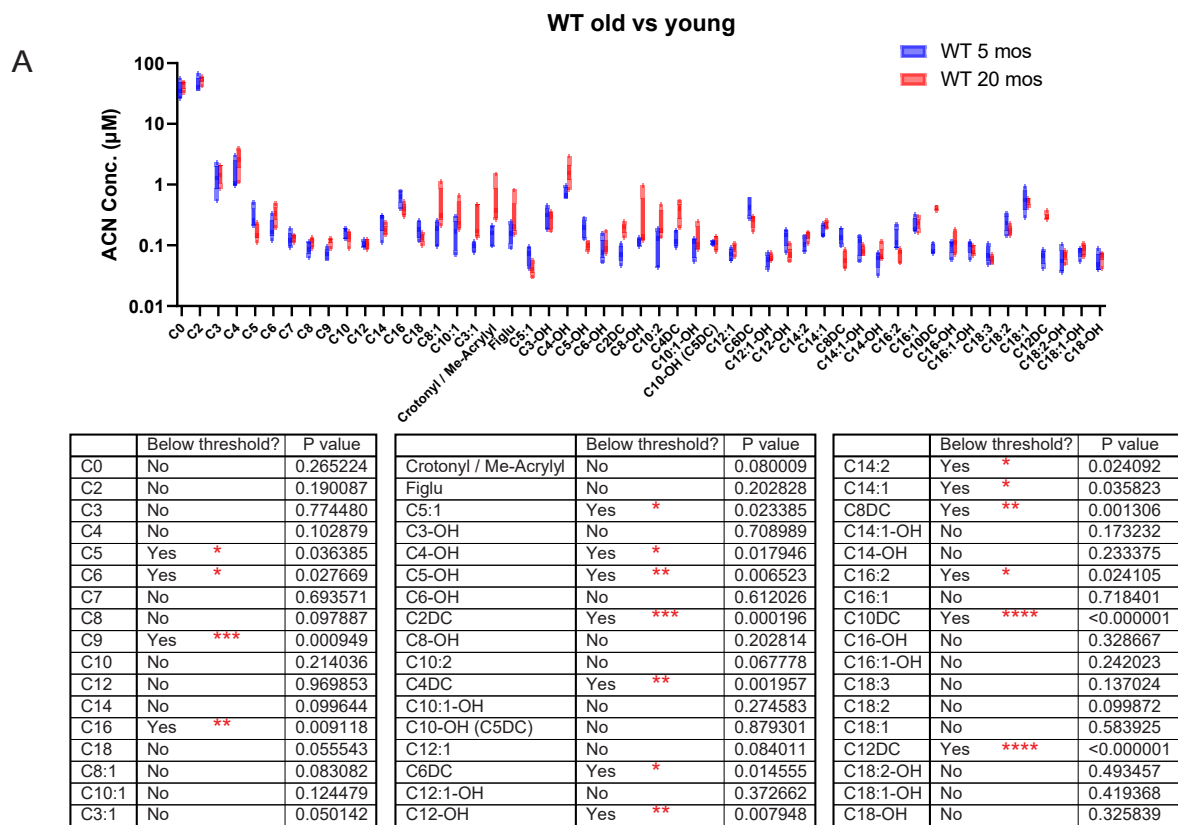
**Figure S4. Young and old *MyckO* mice show evidence of Complex I defects and more generalized mitochondrial dysfunction, related to Figure 4F&G**

(A). 5 month-old mice. The results from Figure 4F are again shown along with the actual values obtained for each serum acyl carnitine. Unpaired t test, \*\*= $p < 0.01$

(B). 20 month-old mice. The results from Figure 4G are again shown along with the actual values obtained for each serum acyl carnitine. Unpaired t test, \*= $p < 0.05$ , \*\*= $p < 0.01$ , \*\*\*= $p < 0.001$ .



Fig. S5



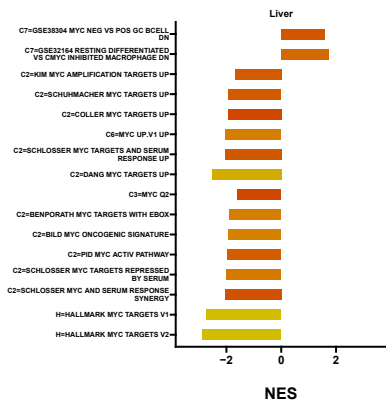
**Figure S5. Both young and old WT and *MycKO* mice show evidence of age-related differences in serum acyl carnitine levels, related to Figure 4F&G**

(A). 5 month old and 20 month-old WT mice. Unpaired t test,  $*=p < 0.05$ ,  $**=p < 0.01$ ,  $***=p < 0.001$ ,  $****=p < 0.0001$ .

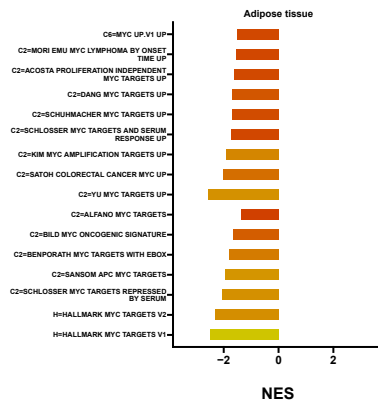
(B). 5 month old and 20 month-old *MycKO* mice. Unpaired t test,  $*=p < 0.05$ ,  $**=p < 0.01$ ,  $***=p < 0.001$ ,  $****=p < 0.0001$ .

Fig. S6

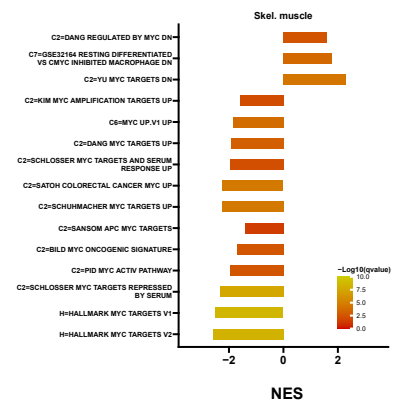
A



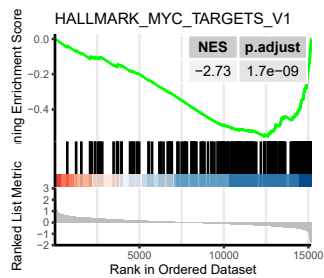
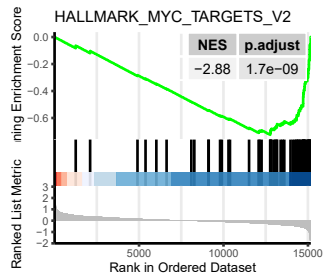
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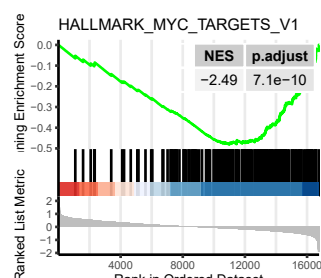
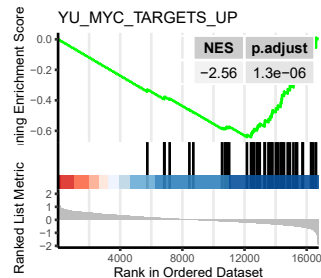
C



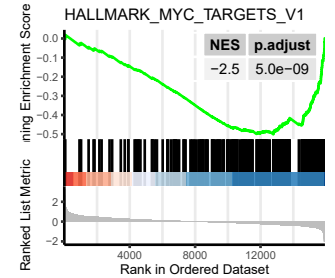
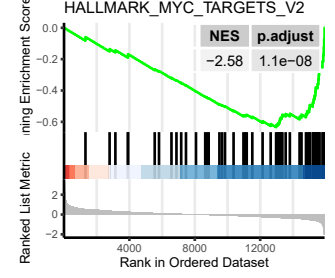
D



E



F

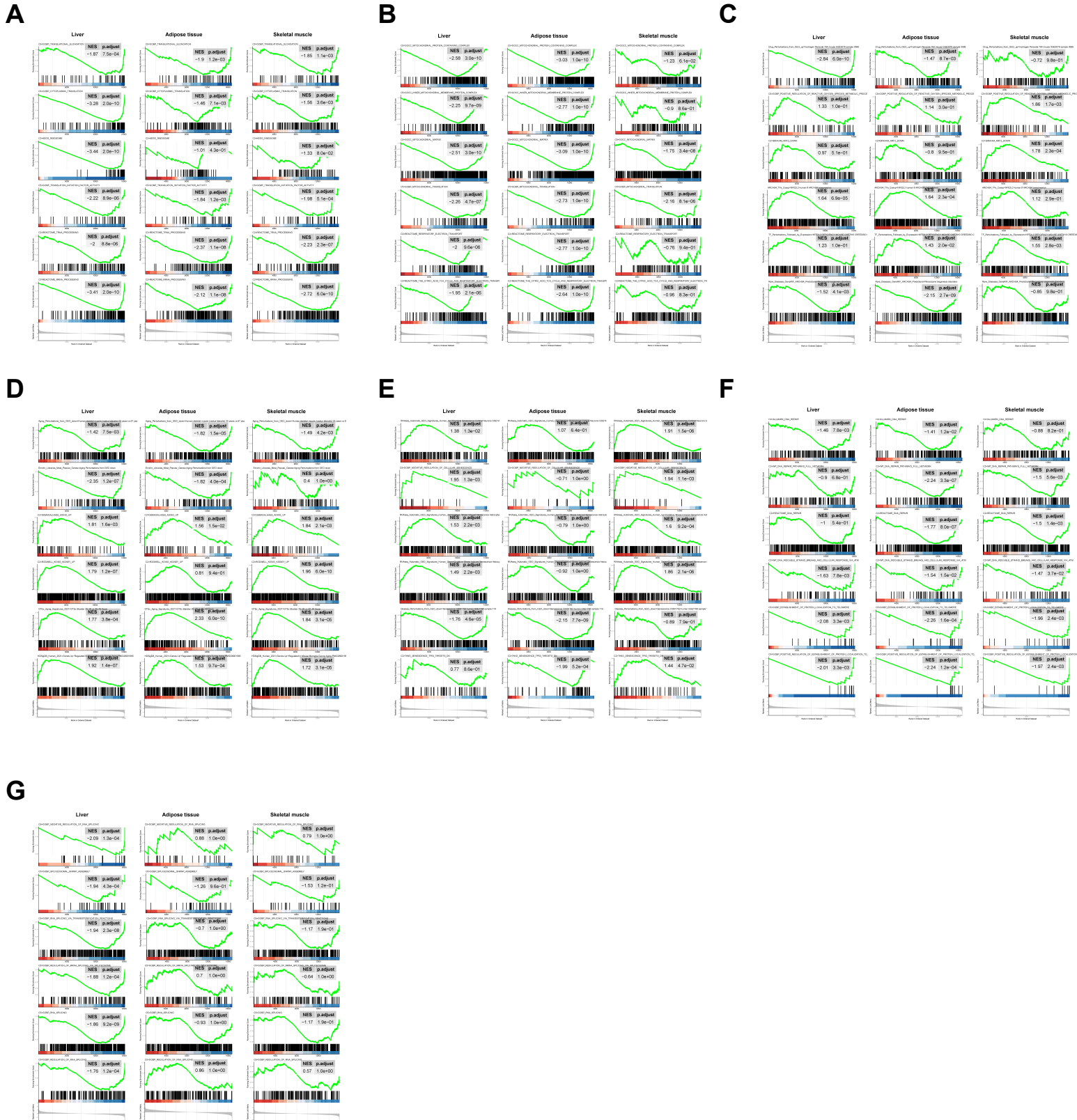


**Figure S6. Dysregulation of Myc target gene sets in 5 mo MycKO tissues, related to Figure 1&6.**

(A-C). GSEA summaries of positively-regulated direct Myc targets in liver, adipose tissue, and skeletal muscle, respectively from 5 month-old MycKO mice. All Myc targets from the MSigDB Collection (<http://www.gsea-msigdb.org/gsea/msigdb/collections.jsp>) were analyzed by GSEA to determine the overall directionality of expression of each set's component transcripts.

(D-F). Select examples of individual GSEA profiles from A-C.

Fig. S7



**Figure S7. GSEA profiles in liver, adipose tissue and skeletal muscle from 5 month-old WT and MycKO mice, related to Figure 6A.** Normalized enrichment scores and q values are indicated in the upper right corner of each profile. Data used to generate the ridge plots shown in Figure 6A are re-graphed and included here.

(A) GSEA profiles for “Translation/ribosomal structure and function”.

(B) GSEA profiles for “Mitochondrial structure and function”.

(C) GSEA profiles for “Oxidative stress response”.

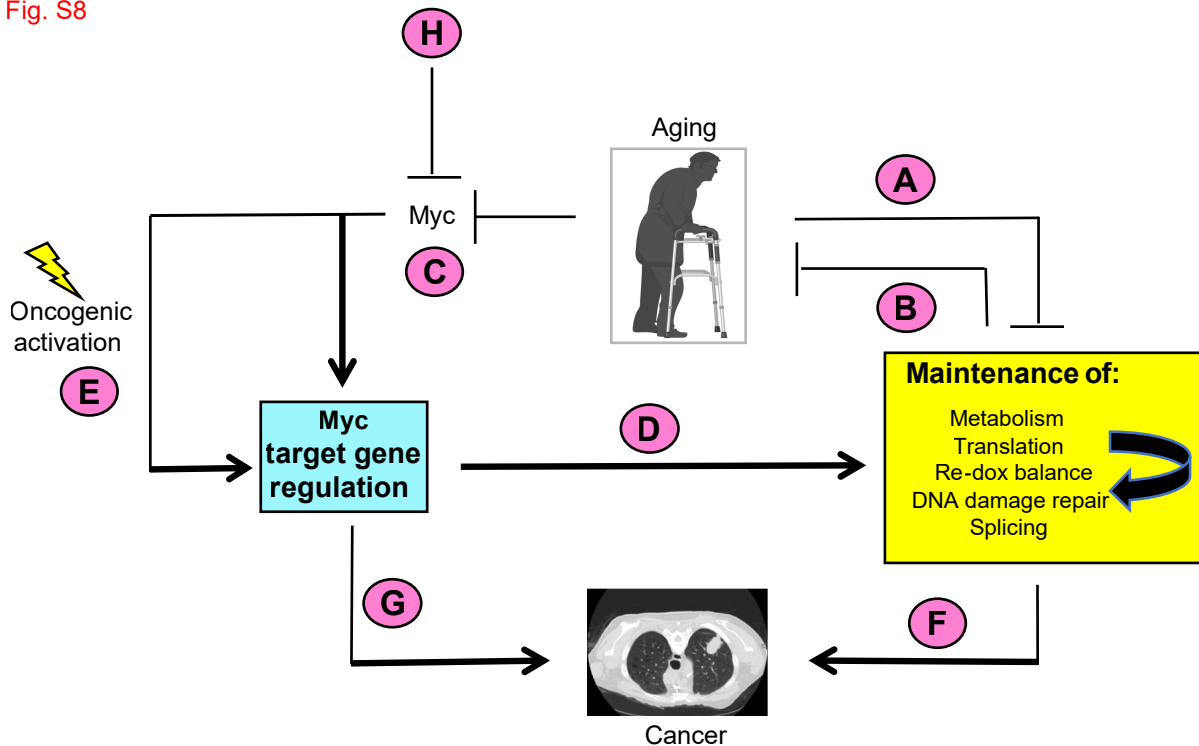
(D) GSEA profiles for “Aging”.

(E) GSEA profiles for “Senescence”.

(F) GSEA profiles for “DNA damage response and repair”.

(G) GSEA profiles for “mRNA splicing”.

Fig. S8





**Figure S8. Model depicting the cooperation between normal aging and Myc regulation, related to Figures 1-7 .**

(A). Normal aging and senescence are associated with the accumulation of Myc-independent defects involving the maintenance of mitochondrial and ribosomal structure and function, redox balance, DNA damage recognition and repair and splicing (yellow box) (Figure 7A).<sup>4-17</sup> Individual functional defects therein can negatively impact other functions (yellow box: curved arrow). For example, defective mitochondrial function can generate ROS, which in turn can cause oxidative DNA damage and impair translation.<sup>18-22</sup>

(B). Defects in the above functions and pathways conversely accelerate aging.<sup>5,9,11,13,23-26</sup>

(C). Normal aging is also associated with declines in Myc levels, causing dysregulation of Myc target gene expression (blue box) (Figure 7F-I).

(D). Normal levels of Myc and its target genes are needed to maintain the baseline cellular functions depicted in the yellow box.<sup>2,27,28</sup> These cooperate with the age-dependent and Myc-independent pathways shown in A.

(E). Oncogenic activation of Myc can dysregulate Myc target genes, thus altering the functions depicted in the yellow box in ways that sustain maximal rates of tumor growth.<sup>2,27-30</sup>

(F). Dysregulation of the functions described for E and D support the initiation, maintenance and/or evolution of cancer.<sup>27-30</sup>

(G). Other Myc target genes not shown in the yellow box, such as those pertaining to cell cycle and survival, can independently contribute to the development of cancer when they are dysregulated as a result of Myc over-expression.<sup>29,31-33</sup> Some of these genes may be so-called “pathological targets” that contain low-affinity Myc binding sites and are only activated in response to the excessive levels of Myc associated with certain tumors.<sup>29,34</sup>

(H). *Myc*KO mice no longer regulate their target genes. As a result, they lose the ability to maintain the functions depicted in the yellow box, thereby hastening the onset of aging via the Myc-dependent pathway that support these functions (B, C and D). Reductions in Myc also eliminate a major age-dependent oncogenic pathway (E), thus leading to a reduced lifetime incidence of cancer (F and G) that contributes to the increased longevity of these mice.

**Table S3. Antibodies used for the current studies, related to STAR Methods.**

<b>Name of antigen target</b>	<b>Type of antibody</b>	<b>Vendor</b>	<b>Catalog number</b>	<b>dilution</b>	<b>Purpose</b>
<b>Glut 1</b>	Rabbit mAb	Abcam	Ab115730	1:2,000	WB
<b>Glut 2</b>	Rabbit pAb*	Proteintech	20436-1-AP	1:1,000	WB
<b>Glut 4</b>	Mouse mAb	Cell signaling	2213	1:1,000	WB
<b>GAPDH</b>	Mouse mAb	Sigma	G8795	1:10,000	WB
<b>γH2A.X</b>	Rabbit mAb	Abcam	ab81299	1:200	IHF
<b>PDH</b>	Rabbit mAb	Cell signaling	3205	1:1,000	WB
<b>c-Myc</b>	Rabbit mAb	Cell signaling	13987	1:1,000 1:400	WB
<b>c-Myc(N-262)</b>	Rabbit pAb	Santa Cruze	sc-764	1:250	IHC
<b>p-PDH</b>	Rabbit pAb	Cal biochem	AP1062	1:300	WB
<b>PFK-L</b>	Rabbit pAb	Avivva sys bio	ARP45774_T100	1:400	WB
<b>PFK-M</b>	Mouse mAb	R+D systems	MAB7687	1:3,000	WB
<b>IgG</b>	HRP-Goat-anti-rabbit	Cell signaling	7074	1:5,000 1:2,000	Secondary for WB
<b>IgG</b>	HRP-Goat-anti-mouse	Cell signaling	7076	1:10,000	Secondary for WB

\*pAb=Polyclonal antibody

## References

1. Wang, H., Dolezal, J.M., Kulkarni, S., Lu, J., Mandel, J., Jackson, L.E., Alencastro, F., Duncan, A.W., and Prochownik, E.V. (2018). Myc and ChREBP transcription factors cooperatively regulate normal and neoplastic hepatocyte proliferation in mice. *J Biol Chem* 293, 14740-14757. 10.1074/jbc.RA118.004099.
2. Wang, H., Stevens, T., Lu, J., Airik, M., Airik, R., and Prochownik, E.V. (2022). Disruption of Multiple Overlapping Functions Following Stepwise Inactivation of the Extended Myc Network. *Cells* 11. 10.3390/cells11244087.
3. Muncan, V., Sansom, O.J., Tertoolen, L., Pheesse, T.J., Begthel, H., Sancho, E., Cole, A.M., Gregorieff, A., de Alboran, I.M., Clevers, H., and Clarke, A.R. (2006). Rapid loss of intestinal crypts upon conditional deletion of the Wnt/Tcf-4 target gene c-Myc. *Mol Cell Biol* 26, 8418-8426. 10.1128/MCB.00821-06.
4. Anisimova, A.S., Alexandrov, A.I., Makarova, N.E., Gladyshev, V.N., and Dmitriev, S.E. (2018). Protein synthesis and quality control in aging. *Aging (Albany NY)* 10, 4269-4288. 10.18632/aging.101721.
5. Balaban, R.S., Nemoto, S., and Finkel, T. (2005). Mitochondria, oxidants, and aging. *Cell* 120, 483-495. 10.1016/j.cell.2005.02.001.
6. Bhadra, M., Howell, P., Dutta, S., Heintz, C., and Mair, W.B. (2020). Alternative splicing in aging and longevity. *Hum Genet* 139, 357-369. 10.1007/s00439-019-02094-6.
7. Bratic, A., and Larsson, N.G. (2013). The role of mitochondria in aging. *J Clin Invest* 123, 951-957. 10.1172/JCI64125.
8. Deschenes, M., and Chabot, B. (2017). The emerging role of alternative splicing in senescence and aging. *Aging Cell* 16, 918-933. 10.1111/ace1.12646.
9. Edgar, D., Shabalina, I., Camara, Y., Wredenberg, A., Calvaruso, M.A., Nijtmans, L., Nedergaard, J., Cannon, B., Larsson, N.G., and Trifunovic, A. (2009). Random point mutations with major effects on protein-coding genes are the driving force behind premature aging in mtDNA mutator mice. *Cell Metab* 10, 131-138. 10.1016/j.cmet.2009.06.010.
10. Gonskikh, Y., and Polacek, N. (2017). Alterations of the translation apparatus during aging and stress response. *Mech Ageing Dev* 168, 30-36. 10.1016/j.mad.2017.04.003.
11. Jang, J.Y., Blum, A., Liu, J., and Finkel, T. (2018). The role of mitochondria in aging. *J Clin Invest* 128, 3662-3670. 10.1172/JCI120842.
12. Meshorer, E., and Soreq, H. (2002). Pre-mRNA splicing modulations in senescence. *Aging Cell* 1, 10-16. 10.1046/j.1474-9728.2002.00005.x.
13. Opresko, P.L., and Shay, J.W. (2017). Telomere-associated aging disorders. *Ageing Res Rev* 33, 52-66. 10.1016/j.arr.2016.05.009.
14. Park, Y., and Gerson, S.L. (2005). DNA repair defects in stem cell function and aging. *Annu Rev Med* 56, 495-508. 10.1146/annurev.med.56.082103.104546.
15. Short, K.R., Bigelow, M.L., Kahl, J., Singh, R., Coenen-Schimke, J., Raghavakaimal, S., and Nair, K.S. (2005). Decline in skeletal muscle mitochondrial function with aging in humans. *Proc Natl Acad Sci U S A* 102, 5618-5623. 10.1073/pnas.0501559102.
16. Srivastava, S. (2017). The Mitochondrial Basis of Aging and Age-Related Disorders. *Genes (Basel)* 8. 10.3390/genes8120398.
17. Turi, Z., Lacey, M., Mistrik, M., and Moudry, P. (2019). Impaired ribosome biogenesis: mechanisms and relevance to cancer and aging. *Aging (Albany NY)* 11, 2512-2540. 10.18632/aging.101922.
18. Ghosh, A., and Shcherbik, N. (2020). Effects of Oxidative Stress on Protein Translation: Implications for Cardiovascular Diseases. *Int J Mol Sci* 21. 10.3390/ijms21082661.

19. Kirkinezos, I.G., and Moraes, C.T. (2001). Reactive oxygen species and mitochondrial diseases. *Semin Cell Dev Biol* *12*, 449-457. 10.1006/scdb.2001.0282.
20. Molenaars, M., Janssens, G.E., Williams, E.G., Jongejan, A., Lan, J., Rabot, S., Joly, F., Moerland, P.D., Schomakers, B.V., Lezzerini, M., et al. (2020). A Conserved Mito-Cytosolic Translational Balance Links Two Longevity Pathways. *Cell Metab* *31*, 549-563 e547. 10.1016/j.cmet.2020.01.011.
21. Rosca, M.G., Vazquez, E.J., Chen, Q., Kerner, J., Kern, T.S., and Hoppel, C.L. (2012). Oxidation of fatty acids is the source of increased mitochondrial reactive oxygen species production in kidney cortical tubules in early diabetes. *Diabetes* *61*, 2074-2083. 10.2337/db11-1437.
22. Vafa, O., Wade, M., Kern, S., Beeche, M., Pandita, T.K., Hampton, G.M., and Wahl, G.M. (2002). c-Myc can induce DNA damage, increase reactive oxygen species, and mitigate p53 function: a mechanism for oncogene-induced genetic instability. *Mol Cell* *9*, 1031-1044. 10.1016/s1097-2765(02)00520-8.
23. Kauppila, T.E.S., Kauppila, J.H.K., and Larsson, N.G. (2017). Mammalian Mitochondria and Aging: An Update. *Cell Metab* *25*, 57-71. 10.1016/j.cmet.2016.09.017.
24. Lenaz, G., Bovina, C., Castelluccio, C., Fato, R., Formiggini, G., Genova, M.L., Marchetti, M., Pich, M.M., Pallotti, F., Parenti Castelli, G., and Biagini, G. (1997). Mitochondrial complex I defects in aging. *Mol Cell Biochem* *174*, 329-333.
25. Stefanatos, R., and Sanz, A. (2011). Mitochondrial complex I: a central regulator of the aging process. *Cell Cycle* *10*, 1528-1532. 10.4161/cc.10.10.15496.
26. Trifunovic, A., and Larsson, N.G. (2008). Mitochondrial dysfunction as a cause of ageing. *J Intern Med* *263*, 167-178. 10.1111/j.1365-2796.2007.01905.x.
27. Dang, C.V., O'Donnell, K.A., Zeller, K.I., Nguyen, T., Osthus, R.C., and Li, F. (2006). The c-Myc target gene network. *Semin Cancer Biol* *16*, 253-264. 10.1016/j.semcancer.2006.07.014.
28. Dolezal, J.M., Wang, H., Kulkarni, S., Jackson, L., Lu, J., Ranganathan, S., Goetzman, E.S., Bharathi, S.S., Beezhold, K., Byersdorfer, C.A., and Prochownik, E.V. (2017). Sequential adaptive changes in a c-Myc-driven model of hepatocellular carcinoma. *J Biol Chem* *292*, 10068-10086. 10.1074/jbc.M117.782052.
29. Prochownik, E.V. (2022). Regulation of Normal and Neoplastic Proliferation and Metabolism by the Extended Myc Network. *Cells* *11*. 10.3390/cells11243974.
30. Shachaf, C.M., Kopelman, A.M., Arvanitis, C., Karlsson, A., Beer, S., Mandl, S., Bachmann, M.H., Borowsky, A.D., Ruebner, B., Cardiff, R.D., et al. (2004). MYC inactivation uncovers pluripotent differentiation and tumour dormancy in hepatocellular cancer. *Nature* *431*, 1112-1117. 10.1038/nature03043.
31. Dang, C.V. (2011). Therapeutic targeting of Myc-reprogrammed cancer cell metabolism. *Cold Spring Harb Symp Quant Biol* *76*, 369-374. 10.1101/sqb.2011.76.011296.
32. Dang, C.V. (2012). MYC on the path to cancer. *Cell* *149*, 22-35. 10.1016/j.cell.2012.03.003.
33. Gabay, M., Li, Y., and Felsher, D.W. (2014). MYC activation is a hallmark of cancer initiation and maintenance. *Cold Spring Harb Perspect Med* *4*. 10.1101/cshperspect.a014241.
34. Prochownik, E.V., and Wang, H. (2022). Normal and Neoplastic Growth Suppression by the Extended Myc Network. *Cells* *11*. 10.3390/cells11040747.