

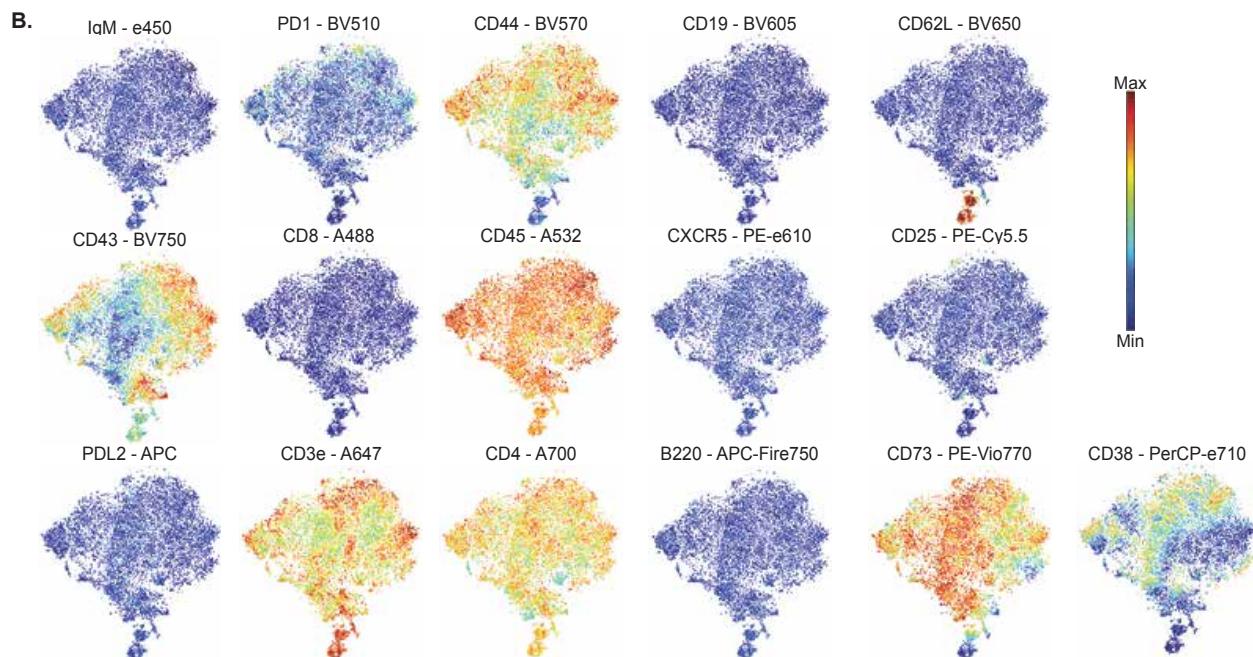
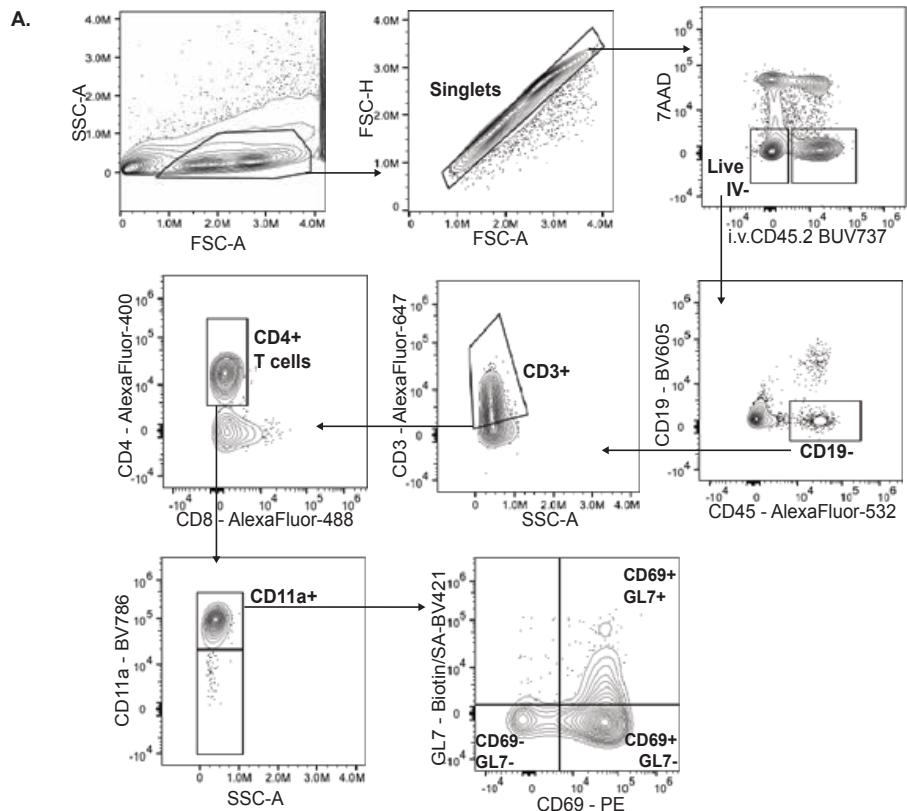
## **SUPPLEMENTAL MATERIAL**

### **GL7 ligand expression defines a novel subset of CD4<sup>+</sup> T<sub>RM</sub> cells in lungs recovered from pneumococcus.**

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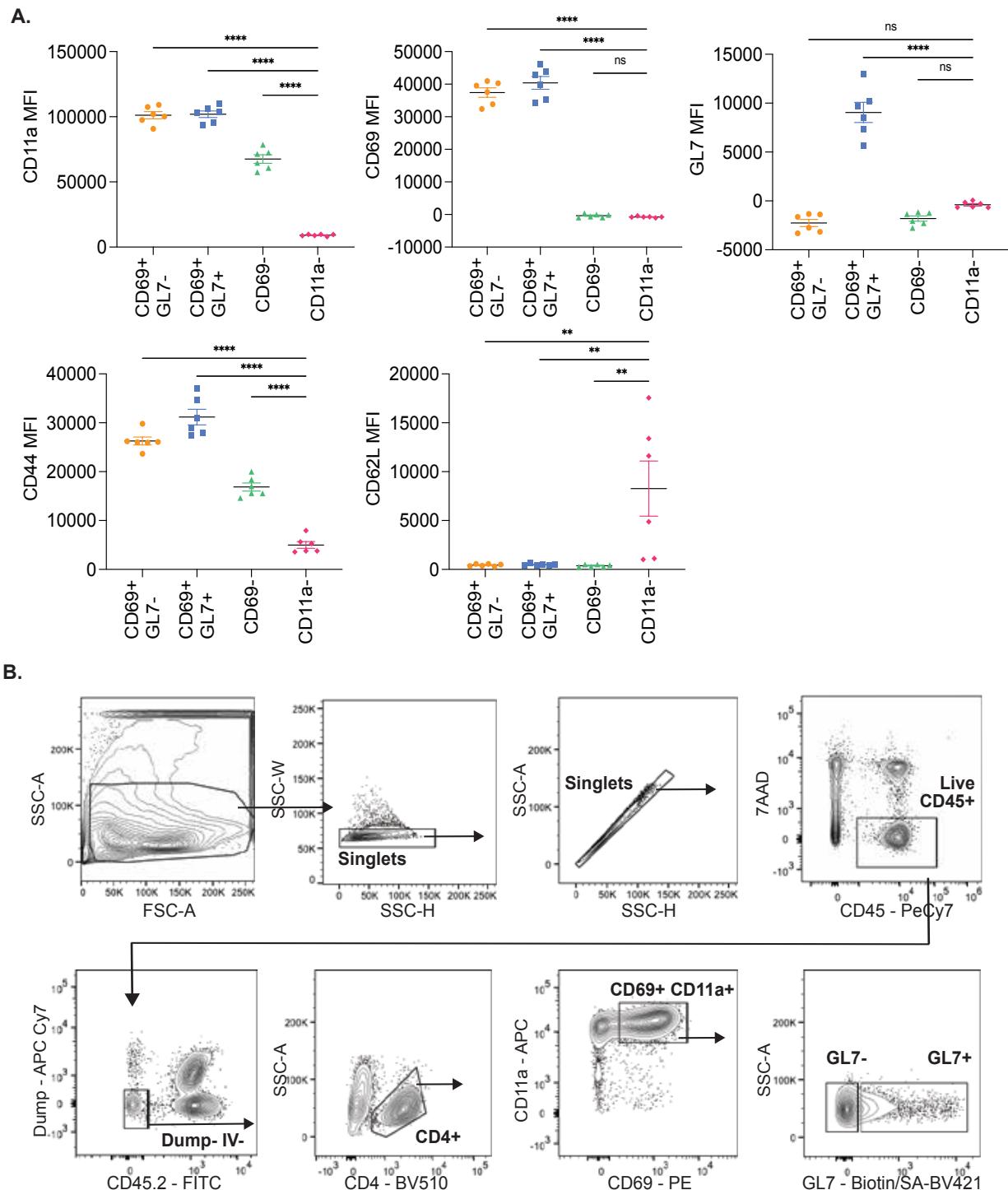
## SUPPLEMENTAL FIGURES

**Supplemental Figure 1**



**Supplemental Figure 1. Identification of lung CD4<sup>+</sup> T<sub>RM</sub> subsets.** **A)** Manual gating strategy to define live lung (i.v. CD45.2<sup>-</sup>) CD4<sup>+</sup> T cell subsets in experienced mouse lungs after 3 min of i.v. injection of anti-CD45.2 antibody. **B)** Heat maps depiction of expression level of surface markers overlayed on opt-SNE plots of lung (i.v. CD45.2<sup>-</sup>) CD4<sup>+</sup>T cell from n=6 experienced mice.

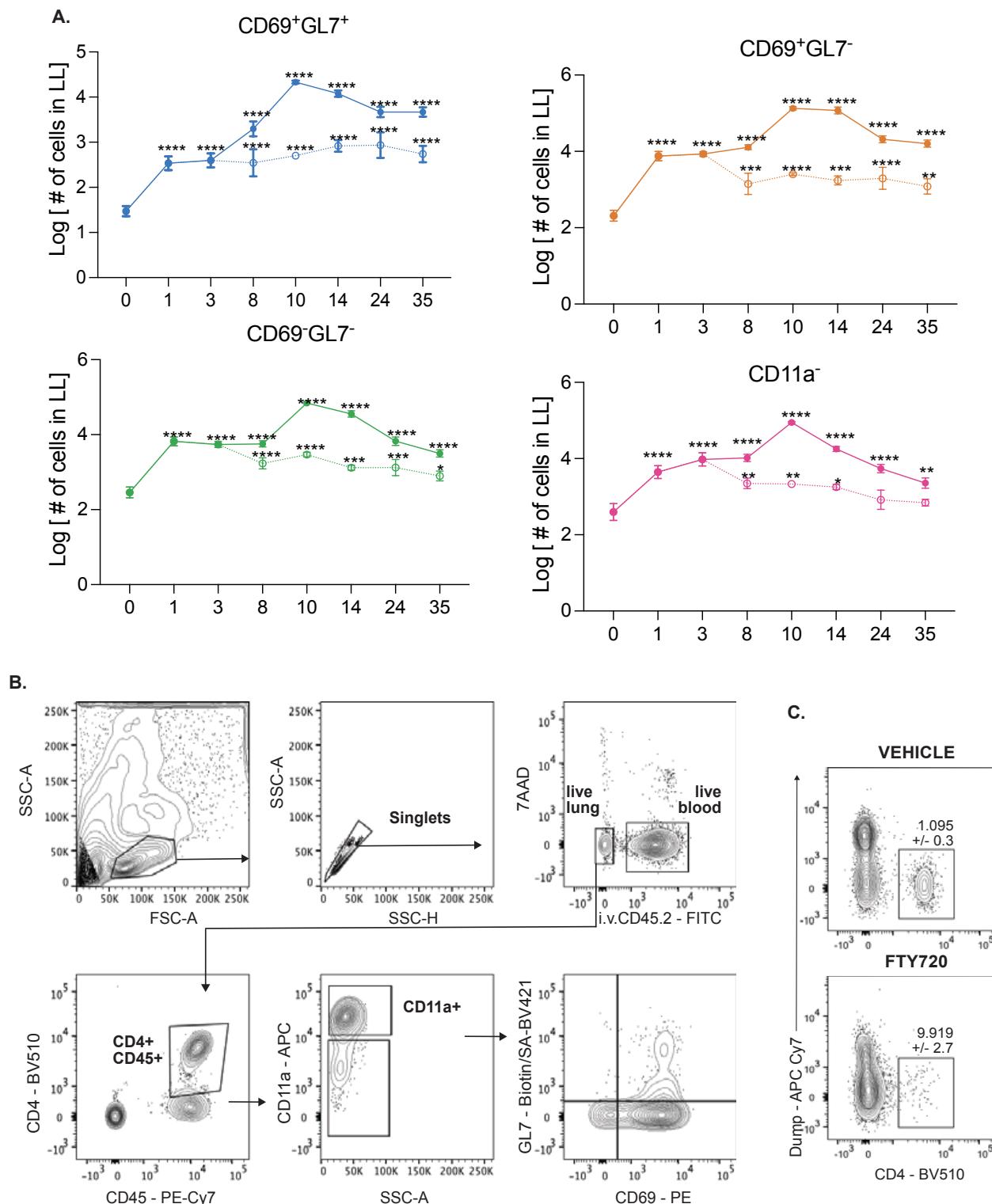
## Supplemental Figure 2



**Supplemental Figure 2. Characterization of lung CD4<sup>+</sup> T<sub>RM</sub> subsets. A)** Mean fluorescence intensity (MFI) of expression in all subsets of lung (i.v. CD45.2<sup>-</sup>) CD4<sup>+</sup> T cells. One-way ANOVA with Dunnett's multiple comparisons test against CD11alo subset, \*\*p=0.002, \*\*\*\*p<0.0001. n=6, two independent experiments. **B)** Sorting gating

strategy to separate GL7+ and GL7- subsets of lung (i.v. CD45.2-) CD4<sup>+</sup> CD69<sup>+</sup>CD11a<sup>+</sup> cells. Sorted cells from 6 mice pooled during sort. Anti-CD8 and anti-CD11c were included in dump color cocktail.

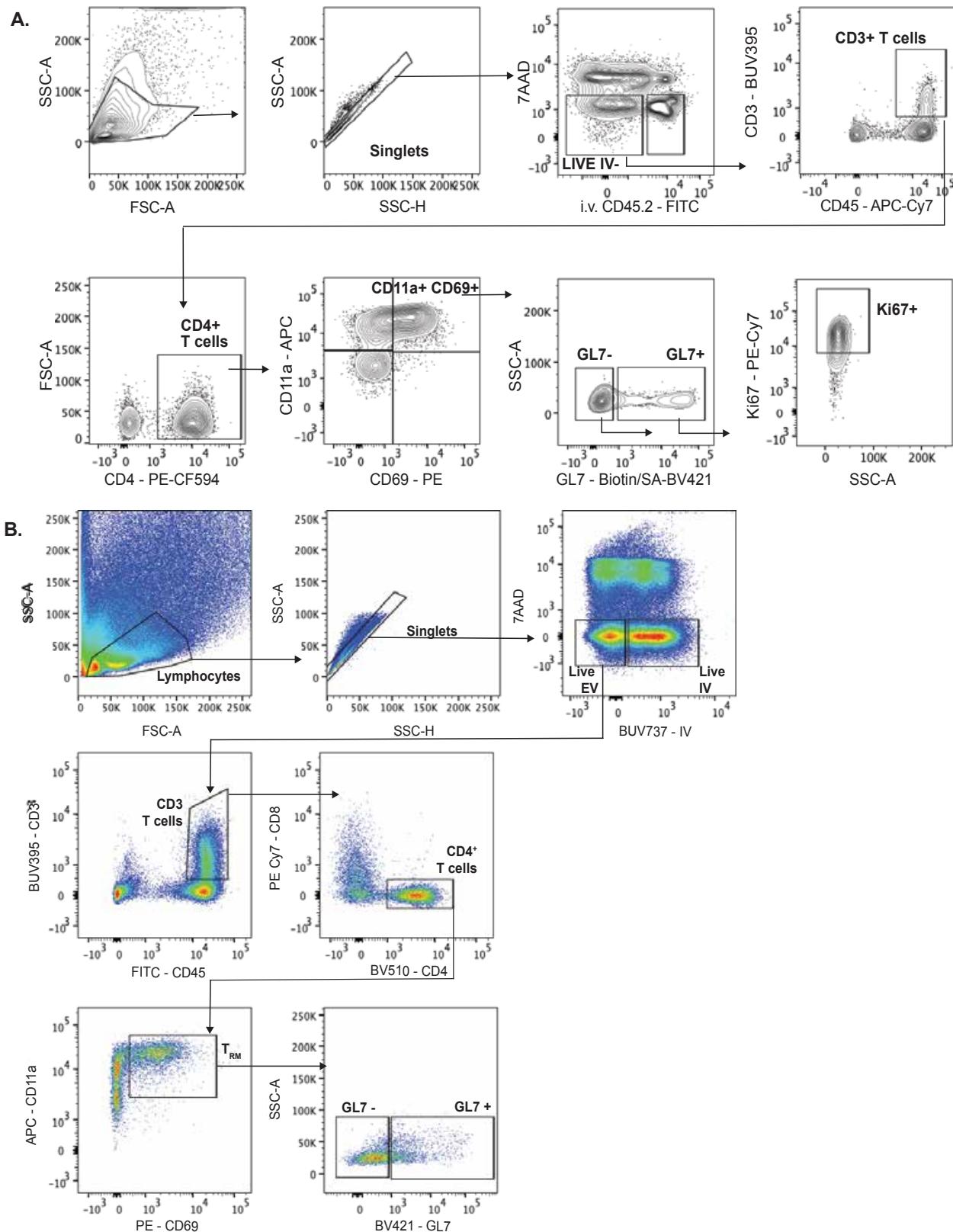
### Supplemental Figure 3



**Supplemental Figure 3. Dynamics of lung CD4<sup>+</sup> T cell subsets. A)** Cell counts of each subset of lung (i.v.CD45.2<sup>-</sup>) CD4<sup>+</sup> T cells across heterotypic immune timeline after

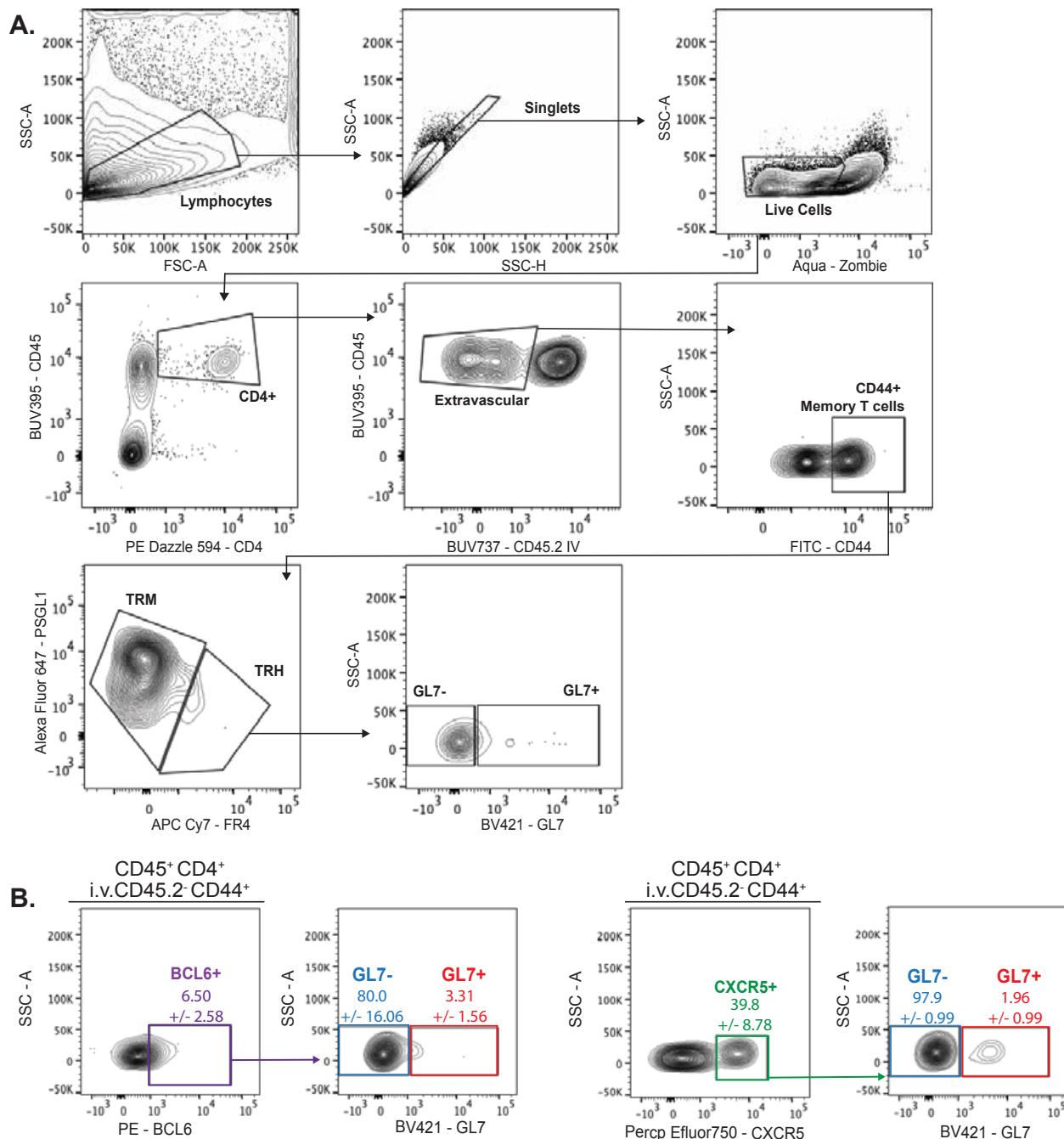
receiving 1x or 2x i.t. infections with Sp19F at days 0 and 7. Data shown as log transformed mean and SEM. n=6 per group, two independent experiments. 2-way ANOVA with Dunnett's multiple comparison tests against Odpi, \*p=0.017 - 0.049, \*\*p=0.0017 - 0.0047, \*\*\*p=0.0001 – 0.0008, \*\*\*\*p<0.0001. FTY720 or vehicle i.p. to experienced mice at 1mg/kg every other day for 12 days. **B)** Gating strategy to identify lung (i.v.CD45.2<sup>-</sup>) and blood (i.v.CD45.2<sup>+</sup>) CD45<sup>+</sup>CD4<sup>+</sup> T cells in FTY720 or vehicle treated mice. **C)** Representative flow cytometry plots and mean ± standard deviation of frequency of live blood (i.v.CD45.2<sup>+</sup>) CD45<sup>+</sup>CD4<sup>+</sup> from LL, n=10 per group, two independent experiments.

## Supplemental Figure 4



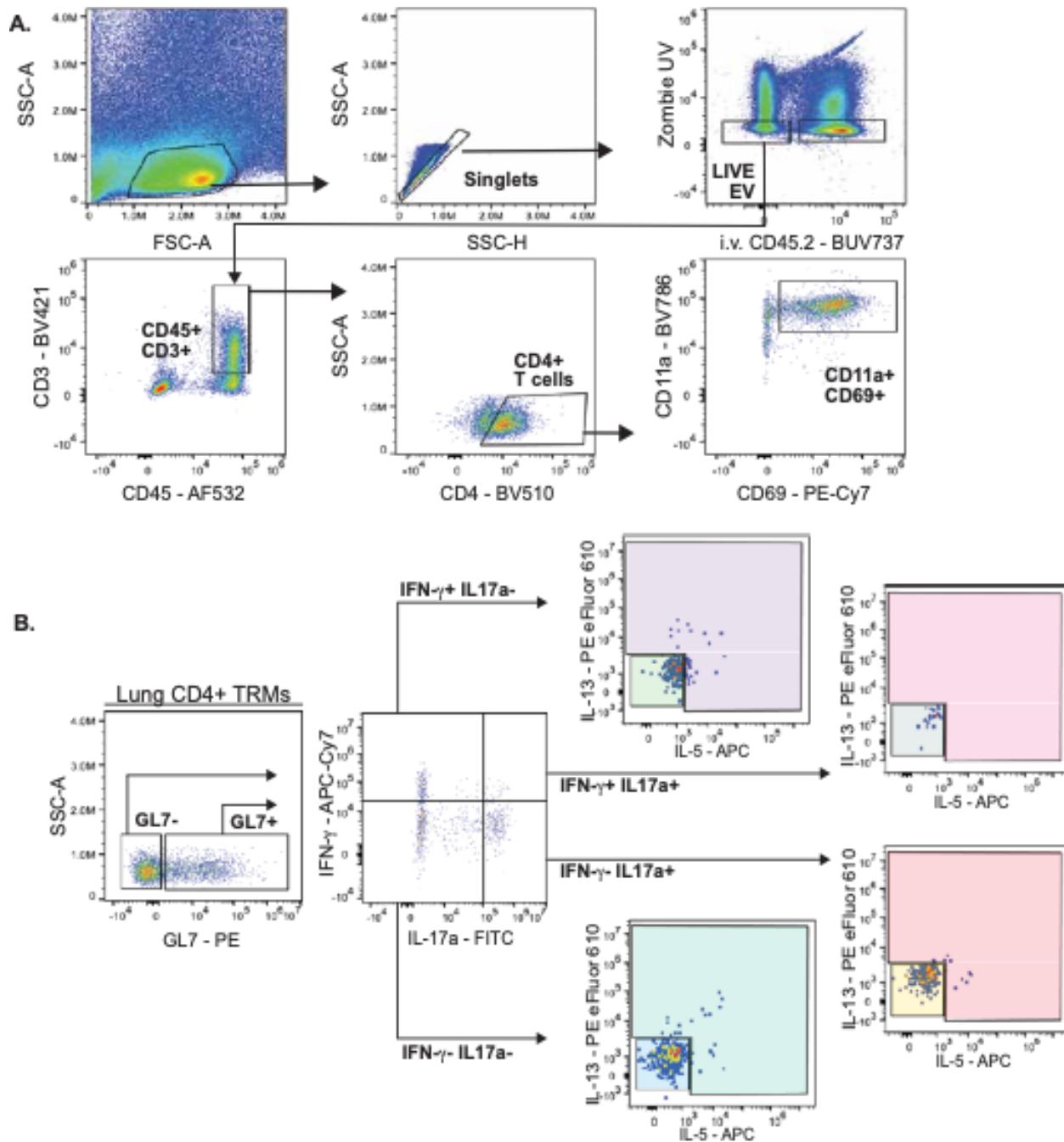
**Supplemental Figure 4. Ki67 Characterization of GL7<sup>+</sup> T<sub>RM</sub> subsets.** **A)** Gating strategy to evaluate Ki67 expression in GL7<sup>+</sup> and GL7<sup>-</sup> subsets of lung (i.v.CD45.2<sup>-</sup>) CD4<sup>+</sup> CD69<sup>+</sup>CD11a<sup>+</sup> T cells. **B)** Gating strategy to evaluate percent of GL7<sup>+</sup> and GL7<sup>-</sup> cells of lung (i.v.CD45.2<sup>-</sup>) CD4<sup>+</sup> CD69<sup>+</sup>CD11a<sup>+</sup> T cells in cells infected with Sp19F or killed KPN or IAV.

## Supplemental Figure 5



**Supplemental Figure 5. Gating strategy to evaluate TRM vs TRH phenotypes in memory T cells. A)** Gating strategy used to evaluate expression of GL7 in PSGL1 and FR4 subsets of lung (i.v.CD45.2-) CD4<sup>+</sup>CD44<sup>+</sup> memory T cells in mouse lungs processed with manual digestion method. **B)** Representative flow cytometry plots and mean +/- standard error of mean of GL7 subsets in BCL6+ and CXCR5+ populations in lung (i.v.CD45.2-) CD4<sup>+</sup>CD44<sup>+</sup> memory T cells in mouse lungs processed with manual digestion method, n=6, two independent experiments.

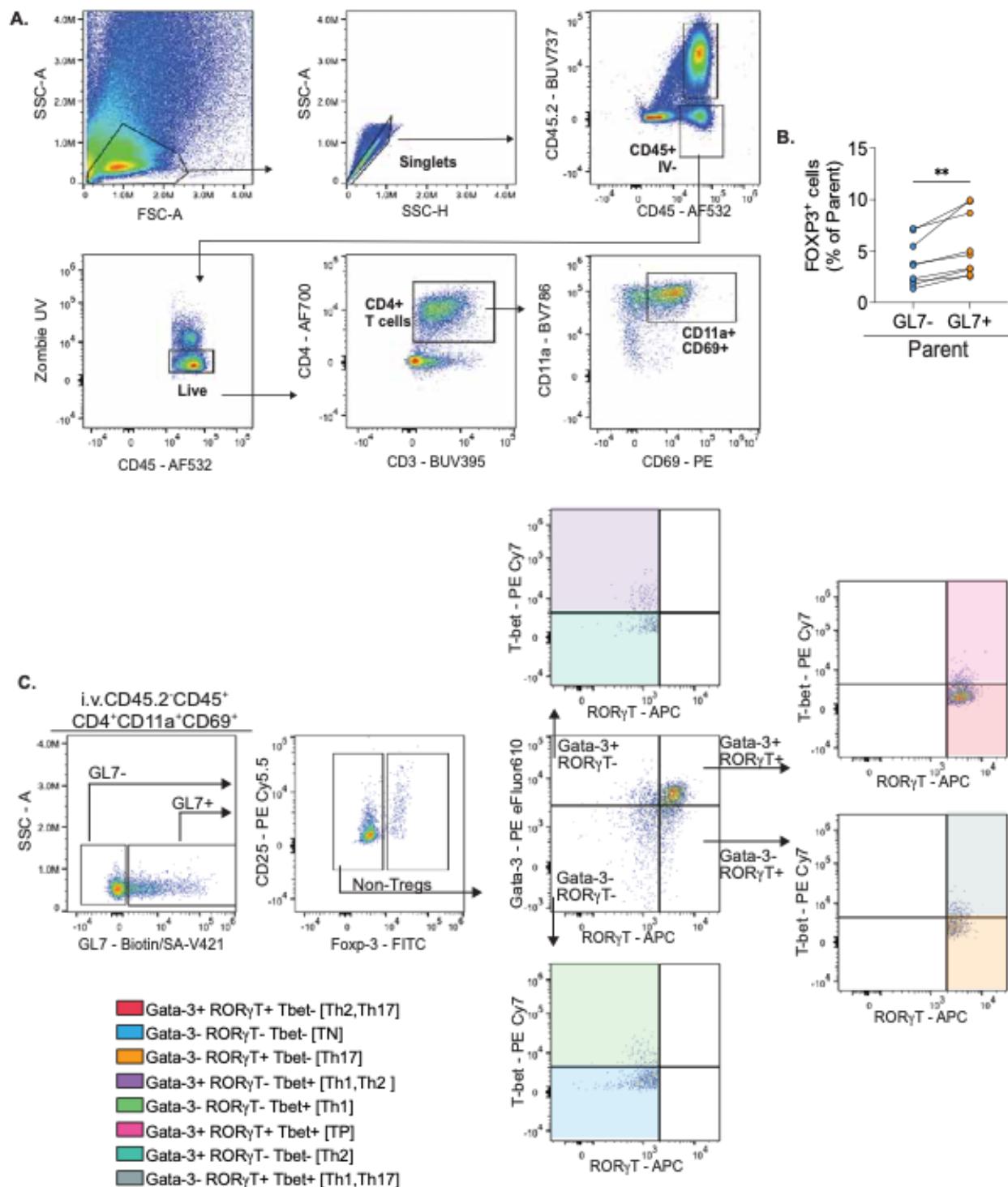
## Supplemental Figure 6



**Supplemental Figure 6. Gating strategy to evaluate intracellular cytokine staining profiles of GL7<sup>+</sup> T<sub>RM</sub> subsets.**

**A)** Intracellular cytokine staining (ICS) gating strategy used to evaluate cytokine secretion in GL7<sup>+</sup> and GL7<sup>-</sup> subsets of lung (i.v.CD45.2<sup>-</sup>) CD4<sup>+</sup> T<sub>RM</sub> cells. **B)** Nested gating strategy to identify expression of one, two, three, or four cytokines in GL7<sup>+</sup> and GL7<sup>-</sup> subsets of lung (i.v.CD45.2<sup>-</sup>) CD4<sup>+</sup> T<sub>RM</sub> cells.

## Supplemental Figure 7



**Supplemental Figure 7. Gating strategy for functional evaluation of GL7<sup>+</sup> T<sub>RM</sub> subsets.**

**A)** Manual gating strategy to identify expression of transcription factors in subsets of lung CD4<sup>+</sup> T<sub>RM</sub> cells. **B)** Quantification of percent of FOXP3+ cells in parent population (GL7<sup>+</sup> or GL7<sup>-</sup> T<sub>RM</sub>), n=9, two independent experiments, paired T-test, \*\*p=0.0032.

**C)** Nested gating strategy to identify expression of one, two or more transcription factors in both GL7<sup>+</sup> and GL7<sup>-</sup> subsets of lung (i.v.CD45.2<sup>-</sup>) CD4<sup>+</sup> T<sub>RM</sub> cells. Lung CD4<sup>+</sup> T<sub>RM</sub> cells gated as shown in **A**. Subsets labeled based on the lineage-defining transcription factors (LDTF) they express.

## SUPPLEMENTAL TABLES

Supplemental Table 1: List of antibodies used in this study				
Marker	Conjugate	Clone	Vendor	Catalogue #
B220	APC-Fire750	RA3-6B2	Biolegend	103259
BCL6	AF488	K112-91	BD Biosciences	561524
BCL6	PE	K112-91	BD Biosciences	561522
CD103	BV711	2E7-	Biolegend	121435
CD11a	BV786	M17	BD Biosciences	740866
CD11a	APC	M17/4	invitrogen	17-011-82
CD19	BV605	6D5	Biolegend	115539
CD25	PE-Cy5.5	PC61.5	Thermo Fisher	35-0251-80
CD3	AF647	145-2C11	Biolegend	100322
CD3	BV421	145-2c11	Biolegend	100335
CD3	BUV395	145-2c11	BD Biosciences	563565
CD38	PerCP-e710	90	Thermo Fisher	46-0381-80
CD4	AF700	RM4-4	Biolegend	116021
CD4	BV510	GK1.5	Biolegend	100449
CD4	PE-CF594	RM4-5	BD Biosciences	562314
CD4	PE-Dazzle594	GK1.5	Biolegend	100456
CD43	BV750	S7	BD Biosciences	747277
CD44	BV570	IM7	Biolegend	103037
CD44	FITC	IM7	Biolegend	103006
CD45	AF532	30-F11	Thermo Fisher	58-0451-82
CD45	APC Cy7	30-F11	Biolegend	103115
CD45	BUV395	HI30	BD Biosciences	563791
CD45.2	BUV737	104	BD Biosciences	612778
CD62L	BV650	MEL-14.	BD Biosciences	564108
CD69	PE	H1.2F3	Biolegend	104508
CD69	PECY7	H1.2F3	Biolegend	104512
CD73	PE-Vio770	REA778	Miltenyi	130-111-519
CD8	AF488	53-6.7	Biolegend	100723
CXCR5	PE-e610	SPRCL5	Thermo Fisher	61-7185-82
CXCR5	PerCP-eFluor750	SPRCL5	Thermo Fisher	46-7185-82
FOXP3	FITC	FJK-16s	Invitrogen	2126755
FR4	APC Fire-750	12A5	Biolegend	125013
FR4	PerCP Cy5.5	12A5	Biolegend	125018
Gata-3	PE-e610	TWAJ	Invitrogen	2187622
GL7	Biotin	GL7	Biolegend	144616
GL7	PE	GL7	Biolegend	144608
ICOS	BV650	C398.4	BD Biosciences	568041
IFN- $\gamma$	APC Cy7	XMG1.2	Biolegend	505850
IgD	AF488	11-26c.2a	Biolegend	405717
IgM	e450	eB121-15F9	Thermo Fisher	48-5890-82
IL-13	PE E610	eBIO13A	Thermo Fisher	61-7133-82

IL-17a	FITC	TC11-18H10.1	Biolegend	506908
IL-5	APC	TRFK5	Biolegend	504306
Ki67	PeCy7	16A8	Biolegend	652426
PD1	BV510	29F.1A12	Biolegend	135241
PD1	PE-Cy7	29F.1A12	Biolegend	135216
PDL2	APC	TY25	Biolegend	107210
PSGL1	Alexa Fluor 647	4RA10	Thermo Fisher	16-6763-84
ROR $\gamma$ T	APC	B2D	Invitrogen	17-6981-80
Streptavidin	BV421	n/a	Biolegend	405225
Streptavidin	APC-Fire750	n/a	Biolegend	405260
T-bet	PE-Cy7	4B10	Biolegend	644823
ZOMBIE	AQUA	n/a	Biolegend	423101
ZOMBIE	UV	n/a	Biolegend	423107
	7AAD	n/a	BD Biosciences	559925

**Supplemental Table 2: Parameters for duplex fluorescent IHC assay performed in this study**

Sequence Order	Antigen Target	Species Origin	Clone	Manufacturer	Catalog	Primary probe or antibody Dilution	Antigen Retrieval (Ventana)	Incubation Temperature	Incubation Period	Fluorophore
1	CD4	Rabbit	D7D2Z	CST	25229	1:50	CC1	RT	24 Hrs.	Opal 570 1:100
2	GL7	Rat	GL7	BioLegend	14460 2	1:100	CC1	RT	2 Hrs.	Opal 520 1:100

CC1-cell conditioning 1-Tris based antigen retrieval buffer; CST-cell signaling technology