# Supporting information for

# Assessment of Four Engineered PET Degrading Enzymes Considering Large-Scale Industrial Applications

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## **KEYWORDS**

polyethylene terephthalate (PET), polyethylene terephthalate hydrolases, industrial enzymatic

PET recycling, enzyme engineering, PET hydrolysis reaction conditions

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**Supplementary Figure 1.** SDS-PAGE analysis of 1- LCC<sup>ICCG</sup>, 2- FAST-PETase, 3- HotPETase, 4- PES-H1<sup>L92F/Q94Y</sup>. M for Protein ladder Precision Plus.



**Supplementary Figure 2.** DSF thermal denaturation curves for FAST-PETase (yellow), PES-H1<sup>L92F/Q94Y</sup> (purple) and LCC<sup>ICCG</sup> (blue) in potassium phosphate buffer 100 mM, pH 8.0, and for HotPETase (red) in glycine-OH buffer 100 mM, pH 9.2.



**Supplementary Figure 3. 16.5% (w/w) PcW-PET depolymerization performed in a reactor using PES-H1<sup>L92F/Q94Y</sup> over 48 h of hydrolysis.** Reaction performed at 60 °C, pH 8.0 using 16.5% (w/w) of PcW-PET and PES-H1<sup>L92F/Q94Y</sup> at a concentration of 1 mg<sub>enzyme</sub> g<sub>PET</sub>-1. Purple dots represent the PET conversion in % measured by the NaOH consumption considering an exclusive production of TPA and MEG (2 mol of NaOH is consumed to titrate 1 mol of the diacid TPA). Purple crosses represent the percentage of PET conversion adjusted by considering the TPA/MHET ratio measured by UHPLC analysis at different time (1 mol of NaOH is consumed to titrate 1 mol of the monoacid MHET).



Supplementary Figure 4. Comparison of 20% (w/w) PcW-PET depolymerizations performed by the four enzymes at bioreactor scale. Enzyme-based PET depolymerizations were performed using FAST-PETase at 50 °C, pH 8.0 (orange), HotPETase at 60 °C, pH 9.2 (red), PES-H1<sup>L92F/Q94Y</sup> at 60 °C, pH 8.0 (purple) and LCC<sup>ICCG</sup> at 68 °C, pH 8.0 (blue) of a 200 g<sub>PET</sub> kg<sup>-1</sup> solution with 1 mg<sub>enzyme</sub> g<sub>PET</sub><sup>-1</sup>. Dots represent the PET conversion in % measured by the NaOH consumption considering an exclusive production of TPA and EG (2 mol of NaOH is consumed to titrate 1 mol of the diacid TPA). Crosses represent the percentage of PET conversion adjusted by considering the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET ratio (1 mol of NaOH is consumed to titrate 1 mol of the TPA/MHET).



**Supplementary Figure 5. 20% (w/w) PcW-PET depolymerization performed in a reactor using PES-H1<sup>L92F/Q94Y</sup> over 48 h of hydrolysis.** Reaction performed at 60 °C, pH 8.0 using 20% (w/w) of PcW-PET and PES-H1<sup>L92F/Q94Y</sup> at a concentration of 1 mg<sub>enzyme</sub> g<sub>PET</sub>-1. Purple dots represent the PET conversion in % measured by the NaOH consumption considering an exclusive production of TPA and MEG (2 mol of NaOH is consumed to titrate 1 mol of the diacid TPA). Purple crosses represent the percentage of PET conversion adjusted by considering the TPA/MHET ratio measured by UHPLC analysis at different time (1 mol of NaOH is consumed to titrate 1 mol of the monoacid MHET).

**Supplementary Table 1.**  $T_m$  assessments of FAST-PETase, HotPETase, PES-H1<sup>L92F/Q94Y</sup> and LCC<sup>ICCG</sup> in 100 mM potassium phosphate buffer pH 8.0, and of HotPETase in 100 mM glycine-OH buffer pH 9.2 performed by DSF and comparison with the previously published  $T_m$ .

Enzyme	T <sub>m</sub> assessed	T <sub>m</sub> reported	
	(°C)	(°C)	
FAST-PETase	63.3 ± 0.0	67.1 <sup>1</sup>	
		DPBS pH 7.0	
HotPETase	80.7 ± 0.0	82.5 <sup>2</sup>	
		50 mM glycine-OH, pH 9.2	
PES-H1 <sup>L92F/Q94Y</sup>	77.6 ± 0.2	78.2 <sup>3</sup>	
		50 mM potassium phosphate pH 8.0	
LCCICCG	91.7 ± 0.2	$94.0 \pm 0.2^4$	
		20 mM tris-HCl, 300 mM NaCl, pH 8.0	

**Supplementary Table 2.** Performances measured at different temperatures during the hydrolysis of Gf-PET for FAST-PETase, HotPETase, PES-H1<sup>L92F/Q94Y</sup> and LCC<sup>ICCG</sup> in 100 mM potassium phosphate buffer pH 8.0, and for HotPETase in 100 mM glycine-OH buffer pH 9.2. Red hashes indicate the value of specific activity that was calculated over 2 h of reaction instead of 4 h and might be underestimated due to poor stability of the enzyme at specified temperature.

Temperature	Performance criteria	FAST-PETase	HotPETase	PES-H1 <sup>L92F/Q94Y</sup>	LCC <sup>ICCG</sup>
	Specific activity (µmol h <sup>-1</sup> mg <sub>enzyme</sub> <sup>-1</sup> )	245.5 ± 12.5	210.9 ± 13.5	$67.2 \pm 0.6$	55.8 ± 15.3
45 °C	PET depolymerization at 24h (%)	8.7 ± 2.3	$10.4 \pm 0.7$	$4.9 \pm 0.4$	$5.2 \pm 0.6$
	PET final depolymerization (%)	$9.0 \pm 2.3$	$12.2 \pm 0.9$	$6.6 \pm 0.6$	$8.0 \pm 0.6$
	Specific activity (µmol h <sup>-1</sup> mg <sub>enzyme</sub> <sup>-1</sup> )	347.5 ± 46.1	429.7 ± 56.1	151.0 ± 22.4	176.7 ± 7.4
50 °C	PET depolymerization at 24h (%)	9.9 ± 3.3	15.3 ± 3.2	8.8 ± 3.1	12.8 ± 0.6
	PET final depolymerization (%)	9.6 ± 3.2	$16.2 \pm 3.7$	11.9 ± 5.1	20.8 ± 1.7
	Specific activity (µmol h <sup>-1</sup> mg <sub>enzyme</sub> <sup>-1</sup> )	148.4 ± 55.6 #	1059.5 ± 35.5	$369.4 \pm 33.4$	575.7 ± 45.8
60 °C	PET depolymerization at 24h (%)	$1.5 \pm 0.4$	25.4 ± 3.7	10.9 ± 1.7	23.1 ± 3.4
	PET final depolymerization (%)	$1.6 \pm 0.4$	25.0 ± 3.8	10.7 ± 1.8	28.0 ± 4.9
	Specific activity (µmol h <sup>-1</sup> mg <sub>enzyme</sub> <sup>-1</sup> )	20.8 ± 11.0 #	1350.8 ± 135.8 #	481.3 ± 44.7 #	752.8 ± 64.5
65 °C	PET depolymerization at 24h (%)	$0.3 \pm 0.4$	12.7 ± 1.7	7.2 ± 1.2	31.3 ± 9.8
	PET final depolymerization (%)	$0.3 \pm 0.4$	12.3 ± 1.5	7.0 ± 1.2	34.4 ± 10.5
	Specific activity (µmol h <sup>-1</sup> mg <sub>enzyme</sub> <sup>-1</sup> )	7.6 ± 3.9 #	1038.5 ± 137.7 #	404.3 ± 39.9 <b>#</b>	962.8 ± 28.7
68 °C	PET depolymerization at 24h (%)	$0.08 \pm 0.02$	9.1 ± 0.8	$3.9 \pm 0.6$	41.3 ± 5.8
	PET final depolymerization (%)	$0.12 \pm 0.02$	8.3 ± 0.8	$3.9 \pm 0.6$	45.8 ± 5.0

**Supplementary Table 3.** Performances of FAST-PETase at pH 8.0, 50 °C, HotPETase at pH 9.2, 60 °C, PES-H1<sup>L92F/Q94Y</sup> at pH 8.0, 60 °C and LCC<sup>ICCG</sup> at pH 8.0, 68 °C during the hydrolysis of 16.5% (w/w) PcW-PET in reactors at 1 mg<sub>enzyme</sub> g<sub>PET<sup>-1</sup></sub>. n.a. - not applicable; n.d. - not determined.

					Calculated PET conversion	
Enzymo	Timo	TPA molar	- MHET molar	NaOH	after TPA/MHET molar	Residual solid
Enzyme	Time	TFA IIIQiai		consumption	ratio adjustment	weight
	(h)	(%)	(%)	(%)	(%)	(%)
	3.0	42	58	10.3	14.5	n.a.
FAST-PETase	6.0	48	52	12.1	16.3	n.a.
	24.0	64	36	14.9	18.1	18.6
	3.0	79	21	12.5	14.0	n.a.
HotPETase	6.0	98	2	17.6	17.9	n.a.
	24.0	100	0	26.4	26.4	28.7
	3.0	43	57	26.8	37.4	n.a.
	6.0	51	49	41.2	54.4	n.a.
PES-H1L92F/Q94Y	24.0	78	22	80.0	90.1	n.a.
	30.0	85	15	86.5	93.5	n.a.
	48.0	99	1	95.7	96.2	98.6
	3.0	57	43	45.3	57.6	n.a.
LCCICCG	6.0	71	29	67.3	78.6	n.a.
	24.0	100	0	97.3	97.3	98.1

**Supplementary Table 4.** Performances of FAST-PETase at pH 8.0, 50 °C, HotPETase at pH 9.2, 60 °C, PES-H1<sup>L92F/Q94Y</sup> at pH 8.0, 60 °C and LCC<sup>ICCG</sup> at pH 8.0, 68 °C during the hydrolysis of 20% (w/w) PcW-PET in reactors at 1 mg<sub>enzyme</sub> g<sub>PET<sup>-1</sup></sub>. n.a. - not applicable; n.d. - not determined.

					Calculated PET conversion	
Enzyme	Time	TPA molar	MHET molar	NaOH consumption	after TPA/MHET molar ratio adjustment	Residual solid weight
	(h)	(%)	(%)	(%)	(%)	(%)
	3.5	45	55	10.5	14.5	n.a.
FAST-PETase	7.8	54	46	12.4	16.1	n.a.
	24.0	83	17	14.9	16.3	16.3
HotPETase	3.0	82	18	12.0	13.2	n.a.
	6.0	98	2	16.3	16.5	n.a.
	24.0	100	0	24.3	24.3	24.1
	3.2	35	64	25.8	38.6	n.a.
	7.1	43	56	38.4	54.2	n.a.
PES-H1 <sup>L92F/Q94Y</sup>	24.0	n.d.	n.d.	61.1	n.d.	n.a.
	27.5	68	32	64.0	75.9	n.a.
	47.2	85	14	75.6	81.9	81.1
LCC <sup>icce</sup>	1.5	57	43	28.3	35.9	n.a.
	3.7	61	39	47.8	59.5	n.a.
	8.0	73	27	72.5	83.7	n.a.
	24.0	100	0	97.2	97.2	98.1

Maximum productivity	Average productivity		
[g <sub>TPAeq</sub> L <sup>-1</sup> h <sup>-1</sup> ]	[g <sub>TPAeq</sub> L <sup>-1</sup> h <sup>-1</sup> ]		
15.9	1.1ª		
17.0	1.8ª		
17.3	4.4 <sup>a</sup> ; 2.7 <sup>b</sup>		
34.5	7.0 <sup>a</sup>		
	Махітит productivity [g <sub>трАеq</sub> L <sup>-1</sup> h <sup>-1</sup> ] 15.9 17.0 17.3 34.5		

**Supplementary Table 5.** Productivities of the four different PET hydrolases using 20% (w/w) postconsumer colored-flake PET waste powder (PcW-PET) as substrate.

 $^{\rm a}$  at 24 h.  $^{\rm b}$  at 48 h.

Nucleotide and expressed amino sequences for production of LCC<sup>ICCG</sup>, FAST-PETase, HotPETase and PES-H1<sup>L92F/Q94Y</sup>. All nucleotide sequences were codon-optimized for expression in *E. coli* and were inserted between *NdeI* and *XhoI* restriction enzyme sites of pET-26b(+) bacterial expression plasmid. Codons mutated from the wild type enzyme are indicated in red.

#### LCC<sup>ICCG</sup> nucleotide sequence:

ATGAGCAACCCGTACCAGCGTGGCCCGAATCCGACCCGCAGCGCACTGACCGCAGATGGCCCGTTTAGCGTGGCA ACCTACACCGTCTCACGCCTGTCAGTCTCGGGTTTTGGCGGGGCGTGATTTATTACCCGACCGGCACGTCTCTG ACGTTCGGTGGCATCGCGATGAGTCCGGGTTATACCGCAGATGCTAGCTCTCTGGCATGGCTGGGTCGTCGCCTG GCTTCCCATGGCTTTGTGGTTCTGGTGATTAACACGAATTCACGTTTCGATGGCCCGGACAGCCGCGCCTCTCAG CTGAGTGCCGCCCTGAACTACCTGCGTACCAGTTCCCCGAGCGCCGTTCGCGCACGTCTGGATGCAAATCGTCTG GCGGTTGCCGGTCATTCTATGGGTGGCGGTGGCACCCTGCGTATTGCAGAACAAAACCCGAGCCTGAAAGCGGCT GTCCCGCTGGCCCGTGGCCACCCGATAAAACGTTTAATACCAGTGTCCCGGTGCTGATTGTTGGCGCAGAAGCT GACACCGTGGCCCGGTTTCGCAGCATGCCATCCCGTTTTATCAAAACCTGCCGAGCACCACGCCGAAAGTTTAC GTCGAACTGTGCAACGCATCGCACTGCCGATAGCAACAATGCGGCCATTTCCGTTTATACGATCTCATGG ATGAAACTGTGGGTCGATAATGACACCGGTTACCGCCAGTTCCTGTGTAATGTGAACGACCCGGCTCTGTGCGCAC TTCCGCACCAATAATCGCCACTGCCAACTCGAGCACCACCACCACCACCACCACTGA

#### LCC<sup>ICCG</sup> expressed amino acid sequence (amino acid numbering starts at 35):

M<sup>35</sup>SNPYQRGPNPTRSALTADGPFSVATYTVSRLSVSGFGGGVIYYPTGTSLTFGGIAMSPGYTADASSLAWLGRR LASHGFVVLVINTNSRFDGPDSRASQLSAALNYLRTSSPSAVRARLDANRLAVAGHSMGGGGTLRIAEQNPSLKA AVPLTPWHTDKTFNTSVPVLIVGAEADTVAPVSQHAIPFYQNLPSTTPKVYVELCNASHIAPNSNNAAISVYTIS WMKLWVDNDTRYRQFLCNVNDPALCDFRTNNRHCQLEHHHHHH

# FAST-PETase nucleotide sequence with a pelB leader sequence (lowercase) upfront the nucleotide sequence encoding for the mature protein (uppercase):

 CCGCAGGCACCGTGGCATAGCAGTACCAACTTTAGTAGCGTTACGGTTCCGACCCTGATTTTTGCTTGTGAAAAT GATAGCATTGCACCGGTTAATAGCAGCGCACTGCCGATTTATGATTCAATGAGCCAGAATGCAAAACAGTTTCTG GAAATTAAAGGCGGTAGCCATTCTTGTGCCAATAGTGGTAATAGCAATCAGGCACTGATTGGTAAAAAGGGTGTT GCCTGGATGAAACGTTTTATGGATAACGATACCGTTATAGCACCTTTGCATGTGAAAATCCGAATAGTACCGCC GTTAGTGATTTTCGCACCGCCAAATTGCAGTCTCGAGCACCACCACCACCACCACTGA

#### FAST-PETase expressed amino acid sequence (amino acid numbering starts at 28):

Q<sup>28</sup>TNPYARGPNPTAASLEASAGPFTVRSFTVSRPSGYGAGTVYYPTNAGGTVGAIAIVPGYTARQSSIKWWGPRL ASHGFVVITIDTNSTLDQPESRSSQQMAALRQVASLNGTSSSPIYGKVDTARMGVMGWSMGGGGSLISAANNPSL KAAAPQAPWHSSTNFSSVTVPTLIFACENDSIAPVNSSALPIYDSMSQNAKQFLEIKGGSHSCANSGNSNQALIG KKGVAWMKRFMDNDTRYSTFACENPNSTAVSDFRTANCSLEHHHHHH

#### HotPETase nucleotide sequence:

#### HotPETase expressed amino acid sequence (amino acid numbering starts at 27):

M<sup>27</sup>QTNPYARGPNPTAASLEASAGPFTVRSFTVARPVGYGAGTVYYPTNAGGTVGAIAIVPGYTATQSSINWWGPR LASHGFVVITIDTNSTLDKPESRSSQQMAALRQVASLNGTSSSPIYGKVDTARGGVMGWSMGGGGSLISAANNPS LKAAAVMAPWHSSTNFSSVTVPTLIFACENDRIAPVKEYALPIYDSMSLNAKQFLEICGGSHSCACSGNSNQALI GMKGVAWMKRFMDNDTRYSQFACENPNSTAVCDFRTANCSLEHHHHHH

### PES-H1<sup>L92F/Q94Y</sup> nucleotide sequence:

## PES-H1<sup>L92F/Q94Y</sup> expressed amino acid sequence (amino acid numbering starts at 0):

M<sup>0</sup>ANPYERGPDPTESSIEAVRGPFAVAQTTVSRLQADGFGGGTIYYPTDTSQGTFGAVAISPGFTAGQESIAWLG PRIASQGFVVITIDTITRFDYPDSRGRQLQAALDHLRTNSVVRNRIDPNRMAVMGHSMGGGGALSAAANNTSLEA AIPLQGWHTRKNWSSVRTPTLVVGAQLDTIAPVSSHSEAFYNSLPSDLDKAYMELRGASHLVSNTPDTTTAKYSI AWLKRFVDDDLRYEQFLCPAPDDFAISEYRSTCPFLEHHHHHH

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