

Identification of Novel β -Tubulin Inhibitors Using a Combined *In Silico/In Vitro* Approach

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Supporting Information

1. Actives dataset generation

Table S1. List of ligands active against tubulin used to generate the actives dataset for pharmacophore model training.

ID	Name	Tubulin activity	SMILES	Reference
AS1	Thiabendazole	81.6 ± 1% inhibition at 20 μ M	C1=CC=C2C(=C1)NC(=N2)C3=CSC=N3	¹
AS2	Albendazole	IC ₅₀ = 6.9 μ M	CCCSC1=CC2=C(C=C1)N=C(N2)NC(=O)OC	²

AS3	Fenbendazole	$IC_{50} = 5.4 \mu M$	<chem>COC(=O)Nc1nc2ccc(\$c3cccc3)cc2[nH]1</chem>	2
AS7	Mebendazole	$IC_{50} = 6.1 \mu M$	<chem>COC(=O)NC1=NC2=C(N1)C=C(C=C2)C(=O)C3=CC=CC=C3</chem>	2
AS8	Flubendazol	$IC_{50} = 3.5 \mu M$	<chem>COC(=O)Nc1nc2cc(C(=O)c3ccc(F)cc3)ccc2[nH]1</chem>	2
AS9	Parbendazol	$IC_{50} = 3.1 \mu M$	<chem>CCCCc1ccc2[nH]c(NC(=O)OC)nc2c1</chem>	2
AS10	Oxfendazole (Mebendazole inhibitory binding)	$IB_{50} = 1.3 \mu M$	<chem>COC(=O)NC1=NC2=C(N1)C=C(C=C2)S(=O)C3=CC=CC=C3</chem>	3
AS11	Oxibendazole	$IC_{50} = 2.4 \mu M$	<chem>CCCOc1ccc2[nH]c(NC(=O)OC)nc2c1</chem>	2
AS5	Nocodazole	$IC_{50} = 1.82 \pm 0.06 \mu M$	<chem>COC(=O)NC1=NC2=C(N1)C=C(C=C2)C(=O)C3=CC=CS3</chem>	4
AS12	CHEMBL459773	$IC_{50} = 4.23 \mu M$	<chem>COC1=C(OC)C(OC)=CC(C(OC)C2=CSC(C3=CC=CC=C3)=N2)=O=C1</chem>	5
AS13	CHEMBL3426932	$IC_{50} = 2.36 \mu M$	<chem>COC1=C(OC)C(OC)=CC(C2=C(C3=CC=C(OCC)C=C3)N=C(NS2)=C1)C</chem>	6
AS14	CHEMBL273081	$IC_{50} = 16.75 \mu M$	<chem>C1C=C(/N=C2S/C(N/2)=O)=C/C3=CC=C(N(C)C)C=C3)C=CC=C1</chem>	7
AS15	24045-22-5	$IC_{50} = 4.0 \pm 0.6 \mu M$	<chem>O=C(N(1)/C(SC1=N\C2=CC=C(O)C=CC2)=C\3=CC(OC)=C(O)C=C3</chem>	8
AS16	1596344-07-8	$IC_{50} = 5.6 \pm 0.3 \mu M$	<chem>O=C(N(1)/C(SC1=N\C2=CC=C(O)C=CC2)=C\4=C(O)C=CC=C4)</chem>	8
AS17	463979-31-9	$IC_{50} = 6.28 \mu M$	<chem>O=C(N(1)/C(SC1=N\C2=CC=C(O)C=CC2)=C\3=CC(OC)=CC=C3</chem>	9
AS4	356542-94-4	$IC_{50} = 1.5 \mu M$	<chem>O=C(C1=CC(OC)=C(O)C=C1)/C=C/C2=C(OC)C=C(OC)C=C2OC</chem>	10, 11
AS18	CHEMBL1163178	$IC_{50} = 6.8 \pm 0.6 \mu M$	<chem>O=C(C1=CC(Cl)=C(Cl)C=C1)C(S(C(N(CC)CC)=S)C2=CC=CC=C2</chem>	12
AS19	CHEMBL220818	$IC_{50} = 2.2 \mu M$	<chem>O=C(C1=CC(OC)=C(OC)C(OC)=C1)/C=C/C2=CC=C(C=C=C3)C3=C2</chem>	13
AS20	CHEMBL285901	$IC_{50} = 2.8 \mu M$	<chem>O=C(C1=CC(OC)=C(OC)C(OC)=C1)/C=C/C2=CC=C(N(C)C)C=C2</chem>	13
AS21	1585234-76-9	$IC_{50} = 1.6 \pm 0.32 \mu M$	<chem>COC1=C(C(OC)=CC(OC)=C1)/C=C/C(C2=CC(N)=CC=C2)=N\O</chem>	14
AS22	CHEMBL3393090	$IC_{50} = 1.3 \pm 0.0 \mu M$	<chem>OC1=CC(/C=C/C(C2=CC=CC=C2)=O)=C=C1OC</chem>	15
AS23	CHEMBL3393091	$IC_{50} = 3.4 \pm 0.2 \mu M$	<chem>FC(C=C1)=CC=C1C2=CC=CC=C2C(/C=C/C3=CC=C(O)C(B(O)O)=C3)=O</chem>	15
AS24	CHEMBL3393092	$IC_{50} = 3.6 \pm 0.0 \mu M$	<chem>FC(C=C1)=CC=C1C2=CC=CC=C2C(/C=C/C3=CC=C(N(C)C)C=C3)=O</chem>	15
AS25	CHEMBL3393099	$IC_{50} = 0.49 \pm 0.27 \mu M$	<chem>FC1=CC=CC(C2=CC=CC=C2C(/C=C/C3=CC=C(O)=C3)OC)=O=C1</chem>	15
AS26	CHEMBL3818930	$IC_{50} = 1.99 \pm 0.03 \mu M$	<chem>O=C(/C(CC1=C2OC)=C/C3=C4=CNC4=CC=C3)C1=CC(OC)=C2OC</chem>	16
AS27	CHEMBL3818850	$IC_{50} = 2.68 \pm 0.15 \mu M$	<chem>O=C(C1=CC(OC)=C(OC)C(OC)=C1)/C(C)=C/C2=CN C3=C2C=CC(OC)=C3</chem>	17

AS28	CHEMBL595808	$IC_{50} = 12 \mu M$	<chem>COC1=C(OC)C(OC)=CC(C(/C=C/C2=C(OC)C=C(OC)C=C2OC)=O)=C1</chem>	¹⁸
AS29	CHEMBL9859	$IC_{50} = 0.46 \mu M$	<chem>COC1=C(OC)C(OC)=CC(C(/C=C/C2=CC=C(OC)C(O)=C2)=O)=C1</chem>	¹⁹
AS30	CHEMBL107	$IC_{50} = 6.7 \pm 0.05 \mu M$	<chem>CC(=O)N[C@H]1CCCC2=C(C(=C(C(=C2C3=CC=C(C(=O)C=C13)OC)OC)OC)OC)OC</chem>	²⁰
AS31	CHEMBL208189	$IC_{50} = 8.3 \mu M$	<chem>CC(=O)CN[C@H]1CCCC2=CC(=C(C(=C2C3=CC=C(C(=O)C=C13)OC)OC)OC)OC</chem>	²¹
AS32	CHEMBL206877	$IC_{50} = 4.6 \pm 0.17 \mu M$	<chem>COC1=CC=C2C(=CC1=O)[C@H](CCC3=CC(=C(C(=C32)OC)OC)NC4=C(C=C(C=C4)[N+](=O)[O-])[N+](=O)[O-]</chem>	²¹
AS33	CHEMBL2180999	$IC_{50} = 1.62 \mu M$	<chem>COC1=CC=C2C(=CC1=O)[C@H](CCC3=CC(=C(C(=C32)OC)OC)NCC4=C(C(=CC=C4)F)F</chem>	²²
AS34	CHEMBL4553296	$IC_{50} = 0.05 \mu M$	<chem>O(C)C1=C2C=3C([C@@H](NC(=O)CCC2=CC(OC)=C1OC)=CC(=O)C(NC4=CC(F)(F)F)=C(Cl)C=C4)=CC3</chem>	²³
AS35	CHEMBL1946973	$IC_{50} = 1.1 \pm 0.1 \mu M$	<chem>COC1=C(OC)C(OC)=CC(N2C(C3=CC=C(OCC)C=C3)=NN=N2)=C1</chem>	²⁴
AS36	CHEMBL1823147	$IC_{50} = 3.0 \pm 0.6 \mu M$	<chem>COC1=C(OC)C(OC)=CC(C(2=C(C(OC)C3=C(O)C(OC)=CC=C23)=O)=C1</chem>	²⁵
AS37	CHEMBL3580699	$IC_{50} = 14 \pm 2.2 \mu M$	<chem>COC1=C(OC)C(OC)=CC(C2=C(NC3=CC=C(OC)C=C3)N(C=CC=C4)C4=N2)=C1</chem>	²⁶
AS38	7-Methoxy-4-(3,4,5-trimethoxyphenyl)quinolin-2(1H)-one	$IC_{50} = 2.56 \pm 0.15 \mu M$	<chem>COC1=C(OC)C(OC)=CC(C(2=CC=C(CC)C=C2N3)=CC3=O)=C1</chem>	²⁷
AS39	CHEMBL221765	$IC_{50} = 2.0 \mu M$	<chem>COC1=CC(SC2=C(C(OC)=O)NC3=CC=C(OC)C=C23)=CC(OC)=C1OC</chem>	²⁸
AS40	CHEMBL380871	$IC_{50} = 4.5 \pm 0.1 \mu M$	<chem>COC1=CC(SC2=C(C(OC)=O)NC3=CC=C([N+]([O-])=O)C=C23)=CC(OC)=C1OC</chem>	²⁹
AS41	1179587-98-4	$IC_{50} = 1.3 \pm 0.1 \mu M$	<chem>COC1=CC(SC2=C(C(OC)=O)NC3=CC=C(Br)C=C23)=CC(OC)=C1OC</chem>	³⁰
AS42	1179588-09-0	$IC_{50} = 0.67 \pm 0.02 \mu M$	<chem>COC1=CC(C(2=C(C(OC)=O)NC3=CC=C(OC)C=C23)=O)=CC(OC)=C1OC</chem>	³⁰
AS43	1839121-23-1	$IC_{50} = 8.3 \mu M$	<chem>O=C(C(NC1=CC=C(OC)N=C1)O)C2=CN(CC3OCC3)C4=C2C=CC=C4</chem>	³¹
AS44	CHEMBL3747333	$IC_{50} = 6.6 \mu M$	<chem>O=C(C(NC1=CC=C(OC)C(OC)=O)C2=CN(CC3OCC3)C3=C2C=CC=C3</chem>	³¹
AS45	1582297-75-3	$IC_{50} = 1.65 \mu M$	<chem>CC1=C([Se](C2=CC(OC)=C(OC)C(OC)=C2)O)C3=C(C(=N1)C=CC=C3</chem>	³²
AS46	CHEMBL4092940	$IC_{50} = 11.2 \pm 1.13 \mu M$	<chem>C1C=CC(NC(C(2=CC=C(S2)=O)=C3NC4=C([N+]([O-])=O)C=NC(N5CCN(C)CC5)=N4)=C3C=C1</chem>	³³
AS47	CHEMBL1940254	$IC_{50} = 1.1 \pm 0.5 \mu M$	<chem>COC1=CC(SC2=C(C(3=CC=CN3)NC4=CC=CC=C4)=CC(OC)=C1OC</chem>	³⁴

AS48	CHEMBL1940258	$IC_{50} = 0.74 \pm 0.05 \mu M$	<chem>COC1=CC(SC2=C(C3=CC=CS3)NC4=CC=CC=C24)=CC(OC)=C1OC</chem>	34
AS49	CHEMBL2313790	$IC_{50} = 1.3 \pm 0.06 \mu M$	<chem>COC1=CC(SC2=C(N3C=CN=CC4=CC=CC=C24)=CC(OC)=C1OC</chem>	35
AS50	CHEMBL3597212	$IC_{50} = 1.1 \pm 0.1 \mu M$	<chem>COC1=CC(SC2=C(C3=CC=CC3)NC4=CC(OC)=CC=C24)=CC(OC)=C1OC</chem>	36
AS51	CHEMBL3597223	$IC_{50} = 1.2 \pm 0.1 \mu M$	<chem>COC1=CC(=CC(=C1OC)O)C2=NC3=C2C=CC(=C3Cl)Cl)C4=CC=CC=C4</chem>	36
AS52	CHEMBL89341	$IC_{50} = 1.1 \pm 0.4 \mu M$	<chem>COC1=CC(C(=C2=C(C3=CC=O)=C(OC)C=C3)NC4=CC(OC)=CC=C24)=O)=CC(OC)=C1OC</chem>	37
AS53	CHEMBL2442345	$IC_{50} = 1.1 \pm 0.4 \mu M$	<chem>COC1=CC(C(=C2=C(C3=CC=O)=C(OC)C=C3)NC4=CC(OC)=CC=C24)=O)=CC(OC)=C1OC</chem>	37
AS54	CHEMBL20684	$IC_{50} = 2.2 \mu M$	<chem>COC1=CC=C(S(NC2=C3C=CN=C2N3)CC=C(O)C=C3)(=O)C=C1</chem>	38
AS55	CHEMBL281995	$IC_{50} = 2.1 \mu M$	<chem>COC1=CC=C(S(NC2=C3C=C=C(Cl)N3)CC=C2)(=O)=O)C=C1</chem>	38
AS56	CHEMBL20289	$IC_{50} = 9.5 \mu M$	<chem>COC1=CC=C(S(NC2=C3C=C(Cl)N3)CC=C2)(=O)=O)C=C1</chem>	38
AS57	CHEMBL20296	$IC_{50} = 88.5 \mu M$	<chem>CIC1=CC=C(S(NC2=C3C=C=C(Cl)N3)CC=C2)(=O)=O)C=C1</chem>	39
AS58	CHEMBL20800	$IC_{50} = 2.9 \mu M$	<chem>CC1=CC=C(C=C1S(=O)(=O)NC2=CC=CC3=C2NC(=C3)O</chem>	39
AS59	SCHEMBL3072090	$IC_{50} = 2.5 \mu M$	<chem>CC1=CC=C(S(NC2=C3C=C(N3)=O)CC=C2)(=O)=O)C=C1</chem>	40
AS60	CHEMBL216379	$IC_{50} = 1.1 \pm 0.1 \mu M$	<chem>O=C(NC1=C2C(CCNS(=O)(=O)C=C3)(=O)=O)CC=C1)C4=CC=NC=C4</chem>	41
AS61	CHEMBL216327	$IC_{50} = 1.2 \pm 0.1 \mu M$	<chem>O=C(NC1=C2C(CCNS(=O)(=O)C=C3)(=O)=O)CC=C1)C4=CC=CO4</chem>	41
AS62	CHEMBL230296	$IC_{50} = 0.6 \mu M$	<chem>COC1=NC(OC)=C(NS(C2=CC3=C(N(C)C)C4=C3C=CC=C4)C=C2)(=O)=O)C=C1</chem>	42
AS63	CHEMBL79280 (Betabulin)	$IC_{50} = 2 \mu M$	<chem>COC1=C(F)C=C(NS(C2=C(F)C(F)=C2F)(=O)=O)C=C1</chem>	43
AS64	CHEMBL2397992	$IC_{50} = 3.68 \mu M$	<chem>COC1=CC(OC)=CC(OC)=C1/C=C/S(NC2=CC=C(OC)C(N)=C2)(=O)=O</chem>	44
AS65	CHEMBL141689	$IC_{50} = 3.6 \mu M$	<chem>COC1=CC=C(CNC2=NC(NCC3=CC=C(OC)C=C3)=C4C(N(C(C)C)=N4)=N2)C=C1</chem>	45
AS66	CHEMBL2001473	Tubulin polymerization MIC = 20 μM	<chem>COC1=CC=C(CNC2=NC(NCC3=CC=C(OC)C=C3)=N C(NC4CCCC4)=N2)C=C1</chem>	46
AS67	CHEMBL1968484	Tubulin polymerization MIC = 10 μM	<chem>COC1=CC=C(CNC2=NC(NCC3=CC=C(OC)C=C3)=N C(NC(C)C)=N2)C=C1</chem>	46
AS68	CHEMBL1976996	Tubulin polymerization MIC = 5 μM	<chem>COC1=CC(OC)C=C(CNC2=NC(NCC3=CC=C(OC)C=C3)=N C(NC4CCCC4)=N2)C=C1</chem>	46

AS69	CHEMBL141477	$IC_{50} = 2 \mu M$	<chem>COC1=CC=C(CNC2=NC(NCC3=CC=C(OC)C=C3)=C4C(N(C5CCCC5)C=N4)=N2)C=C1</chem>	47
AS70	SCHEMBL2905972	$IC_{50} = 16 \pm 2 \mu M$	<chem>COC1=CC=C(CNC2=NC(NCC3=CC=C(OC)C=C3)=C4C(C(C(C)C)=NN4)=N2)C=C1</chem>	48
AS71	CHEMBL513813	$IC_{50} = 3.4 \mu M$	<chem>COC1=CC=C(CNC2=NC3=CC=NN3C(NCC4=CC=C(OC)C=C4)=N2)C=C1</chem>	45
AS72	CHEMBL466716	$IC_{50} = 3.6 \mu M$	<chem>COC1=CC=C(CNC2=NC3=CC=C(C(C(C)C)=NN3C(NCC4=CC=C(OC)C=C4)=N2)C=C1</chem>	45
AS6	CHEMBL61	$IC_{50} = 1.3 \pm 0.06 \mu M$	<chem>CO[C@H]1C2[C@H](CO[C@H]1O)[C@@H](O)C3=CC4=C(OCO4)C=C13)C=C5OC</chem>	49
AS73	CHEMBL2165487	$IC_{50} = 7.7 \pm 1.1 \mu M$	<chem>COc1cc([C@H]2c3cc4c(cc3[C@H](O)(C3O)CCC3)[C@H]3COC(=O)[C@H]23)OCO4)cc(O)c1OC</chem>	50
AS74	CHEMBL2165463	21.1 % inhibition at 1 μM	<chem>COc1cc([C@H]2c3cc4c(cc3[C@H](O)(C3O)CC3)[C@H]3COC(=O)[C@H]23)OCO4)cc(O)c1OC</chem>	50
AS75	CHEMBL2165468	84.5% inhibition at 1 μM ($IC_{50} = 0.45 \pm 0.05 \mu M$)	<chem>O=C1[C@H]2([C@H](C=C3C([C@H]([C@H](C=C3)OCO4)C5=CC(OC)=C(OC)C(OC)=C5)[H])=C4=C(C3)OCO4)C5=CC(OC)=C(OC)C(OC)=C5[H]</chem>	50
AS76	266685-31-8	$IC_{50} = 0.17 \mu M$	<chem>CO[C@H]1C2=C(COC2=O)NC3=CC4=C(OCO4)C=C13)C=C5OC</chem>	51
AS77	1260403-30-2	$IC_{50} = 0.27 \mu M$	<chem>O=C1OCC2=C1C(C3=CC4=C(C=C3N2)OCO4)/C=C/C5=CC(OC)=C(OC)C(OC)=C5</chem>	51
AS78	CHEMBL114642	$IC_{50} = 2.3 \mu M$	<chem>O=C1C2=C(C=CC=C2)C(C3=CC=C(OC)C(O)=C3)C4=CC=CC=C41</chem>	52
AS79	CHEMBL392633	$IC_{50} = 0.56 \mu M$	<chem>CO[C@H]1C2=C(C=C1S(=O)(=O)OC2=C3C=CC=C3=CC4=CC=CC=C42)</chem>	53
AS80	CHEMBL455138	$IC_{50} = 0.52 \mu M$	<chem>O=C1C2=C(C=CC=C2)/C(C3=CC=CC=C31)=C\CC4=CC=C(OC)C(OC)=C4</chem>	54
AS81	CHEMBL1210677	$IC_{50} = 1.5 \mu M$	<chem>O=C1C2=C(C=CC=C2)/C(C3=CC=CC=C31)=N\CC4=CC=C(OC)C(OC)=C4</chem>	55
AS82	CHEMBL1814612	$IC_{50} = 0.44 \mu M$	<chem>O=C1C2=C(C=CC=C2)/C(C3=CC=CC=C31)=N\CC4=CC=C(OC)C(OC)=C4</chem>	56
AS83	CHEMBL1814615	$IC_{50} = 1.11 \mu M$	<chem>O=C1C2=C(C=CC=C2Cl)/C(C3=CC=CC=C31)=N\CC4=CC=C(OC)C(OC)=C4</chem>	56
AS84	CHEMBL4585656	51.0 \pm 11.2 % inhibition at 10 μM	<chem>OC1=C(C)C=C2C([C@H](C)CC/C=C(C)\C2)=C1</chem>	57
AS85	CHEMBL4551469	56.3 \pm 7.5 % inhibition at 10 μM	<chem>C[C@H]1C2=CC(OC(C3=CC=C3)=O)=C(C)C=C2C/C(C)=C\CC1</chem>	57
AS86	2301070-52-8	58.1 \pm 25.3 % inhibition at 10 μM	<chem>OC1=C(C)C=C2C([C@H](C)CCC/C(C)=C\2)=C1</chem>	57
AS87	CHEMBL4590152	38.7 \pm 25.4 % inhibition at 10 μM	<chem>C[C@H]1C2=CC(OC(C3=CC=C3)=O)=C(C)C=C2/C=C(C)\CCC1</chem>	57

AS88	CHEMBL4542864	$46.9 \pm 5.3\%$ inhibition at 10 μM	<chem>OC1=C(C)C=C2C([C@H](C)CC[C@H]3[C@@](C)(O3)C2)=C1</chem>	57
AS89	CHEMBL4550701	$47.9 \pm 8.1\%$ inhibition at 10 μM	<chem>C[C@H]1C2=CC(OC(C3=CC=CC=C3)=O)=C(C)C=C2C[C@@](C)(O4)[C@@@H]4CC1</chem>	57
AS90	CHEMBL4553976	$30.3 \pm 5.4\%$ inhibition at 10 μM	<chem>OC1=C(C)C=C2C([C@H](C)CCC[C@@]3(C)[C@@H]2O3)=C1</chem>	57
AS91	CHEMBL4538683	$97.9 \pm 15.3\%$ inhibition at 10 μM	<chem>C[C@H]1C2=CC(OC(C3=CC=CC=C3)=O)=C(C)C=C2[C@@H](O4)[C@@]4(C)CCC1</chem>	57
AS92	CHEMBL4567416	$43.7 \pm 27.5\%$ inhibition at 10 μM	<chem>OC1=C(C)C=C2C([C@](O3)CC[C@H]3[C@H](C)C2)=C1</chem>	57
AS93	CHEMBL4582836	$26.1 \pm 15.8\%$ inhibition at 10 μM	<chem>C[C@]1(O2)C3=CC(OC(C4=CC=CC=C4)=O)=C(C)C=C3C[C@@H](C)[C@@H]2CC1</chem>	57
AS94	CHEMBL4578873	$38.7 \pm 2.5\%$ inhibition at 10 μM	<chem>OC1=C(C)C=C2C([C@@]3(C)CCC[C@H](C)[C@@H]2O3)=C1</chem>	57
AS95	CHEMBL4593906	$9.2 \pm 4.1\%$ inhibition at 10 μM	<chem>C[C@]12C3=CC(OC(C4=CC=CC=C4)=O)=C(C)C=C3[C@@H](O2)[C@@H](C)CCC1</chem>	57

2. Description and optimisation of pharmacophore models

Model SB1

Pharmacophore SB1 was derived from Nocodazole bound to chain B of porcine tubulin (PDB: 5CA1).⁵⁸ Based on the automatically generated pharmacophore model, removal and tolerance adjusting of features and Xvols coat modifications, led to the model consisting of two HC features and one HBD (interaction with GLU198B). LB pharmacophore features from active compounds **AS2**, **AS7**, **AS8**, **AS10**, and **AS11** were merged, and the feature coordinates were extracted and manually added to model SB1, leading to the final model consisting of an additional HBD, two HBAs, two AI features and eleven Xvols.

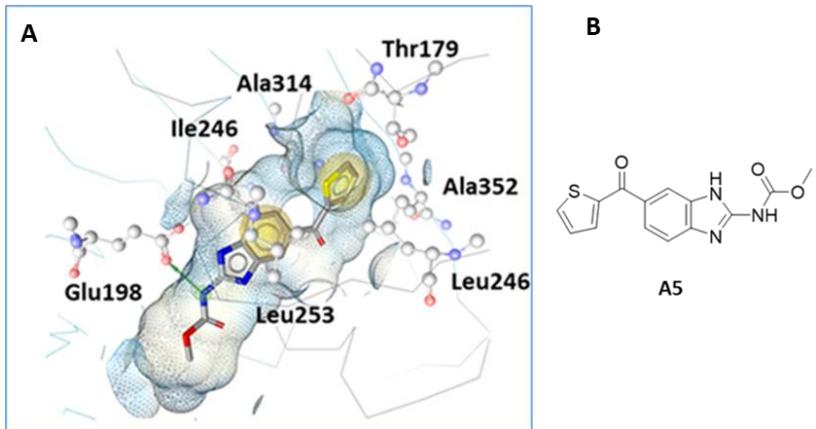


Figure S1. A) Automatically generated pharmacophore model of Tubulin CBS 3D crystal structure complex with nocodazole **A5** based on (PDB 5CA1)⁵⁸, B) Structure of the ligand nocodazole **A5**.

Model SB2

Pharmacophore model SB2 was derived from [2-(1H-indol-4-yl)-1H-imidazol-4-yl](3,4,5-trimethoxyphenyl)methanone bound to chain B of bovine tubulin (PDB: 6O5M).⁵⁹ Based on the automatically generated pharmacophore model, removal and tolerance adjusting of features and Xvols coat modifications, led to the model consisting of one HC feature, three HBAs towards HOH67B and Cys239B, two HBDs (interactions with Thr179A, ASN347B), and 18 Xvols.

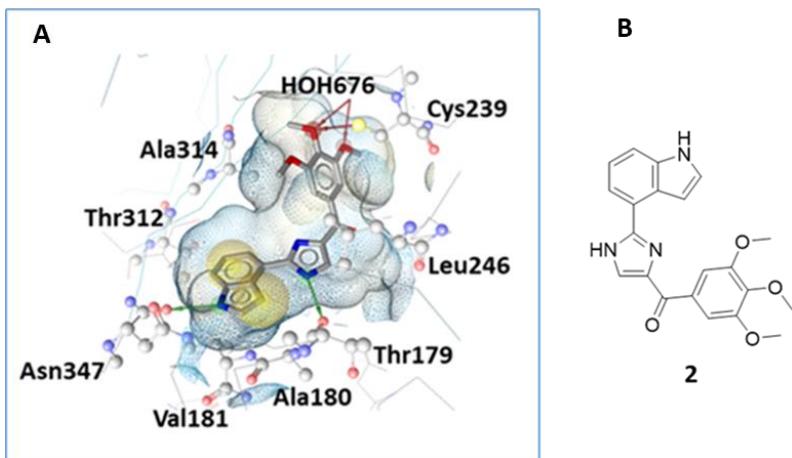


Figure S2. A) Automatically generated pharmacophore model of Tubulin CBS 3D crystal structure complex with the indol-imidazol-trimethoxyphenyl **2** based on (PDB 6O5M)⁵⁹, B) Structure of the ligand **2**.

Model LB3

Pharmacophore model LB3 was developed by automatic clustering of **AS55**, **AS56**, **AS57**, **AS63**, **AS85**, **AS86**, **AS87**, **AS88**, **AS89**, **AS90**, **AS91**, **AS92**, **AS93**, **AS94**, and **AS95** and creating a merged feature pharmacophore. After automatic pharmacophore model generation, removal and tolerance adjusting of features and Xvols coat modifications were optimised to find as many actives as possible while excluding inactives. The final model contained one AI feature, three HC features, two HBAs and 36 Xvols.

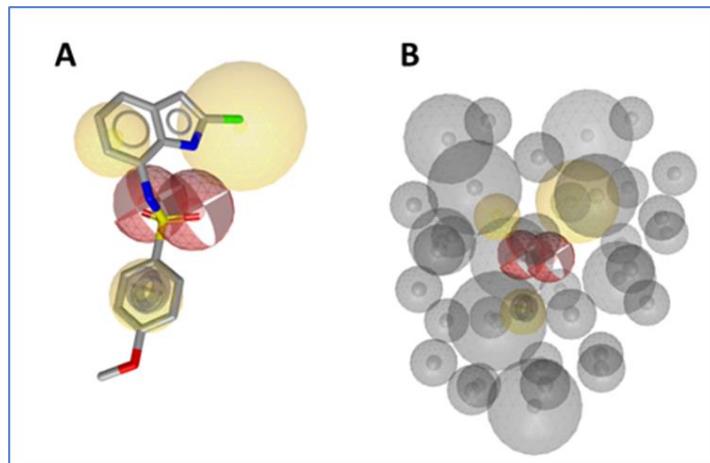


Figure S3. A) Optimised LB pharmacophore model LB3 aligned with base ligand **AS55** and B) the model LB3 with Xvols spheres. (HC, yellow spheres), (AI, purple sphere) Hydrogen bond (HBA; red arrows/spheres), and (Xvols; grey sphere).

Model LB4

Pharmacophore model LB4 was developed by automatic clustering and aligning the myosverin derivatives **AS66**, **AS67**, **AS68**, **AS69**, **AS70**, **AS71**, and **AS72** and creating a merged feature pharmacophore. After automatic pharmacophore model generation, removal and tolerance adjusting of features and Xvols coat modifications were optimised to find as many actives as possible while excluding inactives. The final model contained two HC features, five HBAs, two HBDs and 54 Xvols.

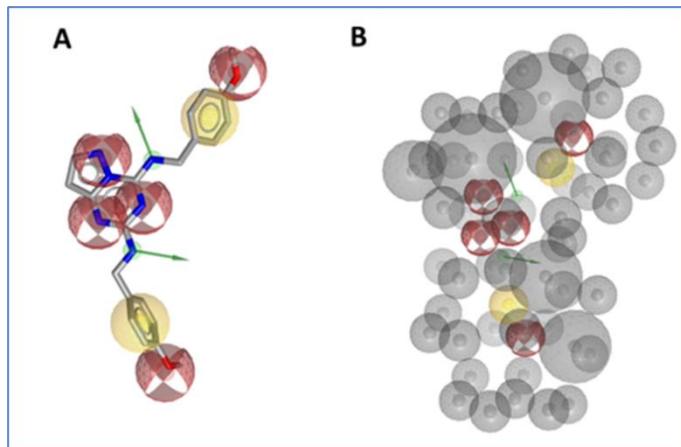


Figure S4. A) Optimised LB pharmacophore model LB4 aligned with base ligand **AS71** and B) the model LB4 with Xvol spheres. (HC, yellow spheres), (AI, purple sphere), (HBD; green arrows/spheres), Hydrogen bond acceptors (HBA; red arrows/spheres), and (Xvols; grey sphere).

Model LB5

Pharmacophore model LB5 was developed by automatic clustering and aligning the sesquiterpenoids **AS85**, **AS86**, **AS87**, **AS88**, **AS89**, **AS90**, **AS91**, **AS92**, **AS93**, and **AS94** and creating a merged features pharmacophore. After automatic pharmacophore model generation, removal and tolerance adjusting of features and Xvols coat modifications were optimised to find as many actives as possible while excluding inactives. The final model contained one AI feature, three HC features, one HBA, and 49 Xvols.

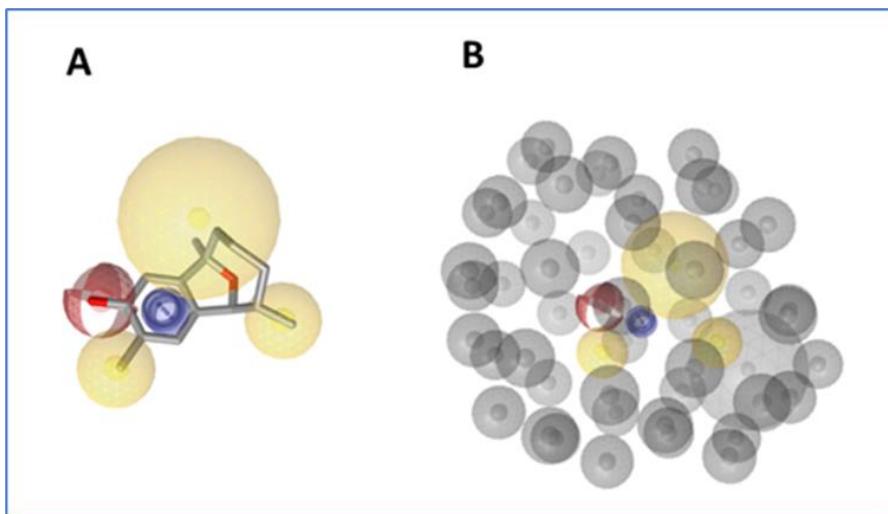


Figure S5. A) Optimised LB pharmacophore model LB5 aligned with base ligand **AS94** and B) the model LB5 with Xvols spheres. (HC, yellow spheres), (AI, purple sphere), (HBA; red arrows/spheres), and (Xvols; grey sphere).

Model LB6

Pharmacophore model LB6 was developed by manually selecting and aligning the indole-sulfonamide derivatives **AS58**, **AS59**, **AS60**, and **AS61**, creating a shared features pharmacophore. After automatic pharmacophore model generation, removal and tolerance adjusting of features and Xvols coat modifications were optimised to find as many actives as possible while excluding inactives. The final model contained two AI features, two HC features, two HBAs, one HBD and 35 Xvols.

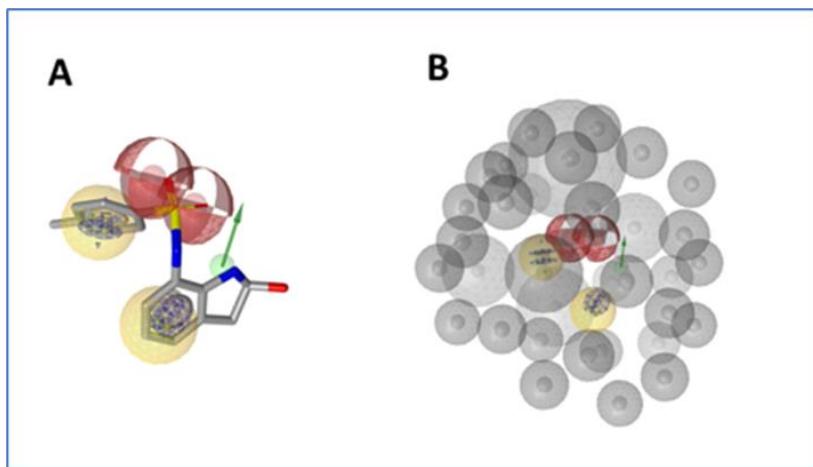


Figure S6. A) Optimised LB pharmacophore model LB6 aligned with base ligand **AS59** and B) the model LB6 with Xvol spheres. (HC, yellow spheres), (AI, purple sphere), (HBD; green arrows/spheres), (HBA; red arrows/spheres), and (Xvols; grey sphere).

Model LB7

Pharmacophore model LB7 was developed by manually selecting and aligning **AS30**, **AS33**, **AS34**, **AS75**, and **AS77** and creating a merged features pharmacophore. After automatic pharmacophore model generation, removal and tolerance adjusting of features and Xvols coat modifications were optimised to find as many actives as possible while excluding inactives. The final model contained one AI feature, one HC feature, four HBAs, one HBD and 68 Xvols.

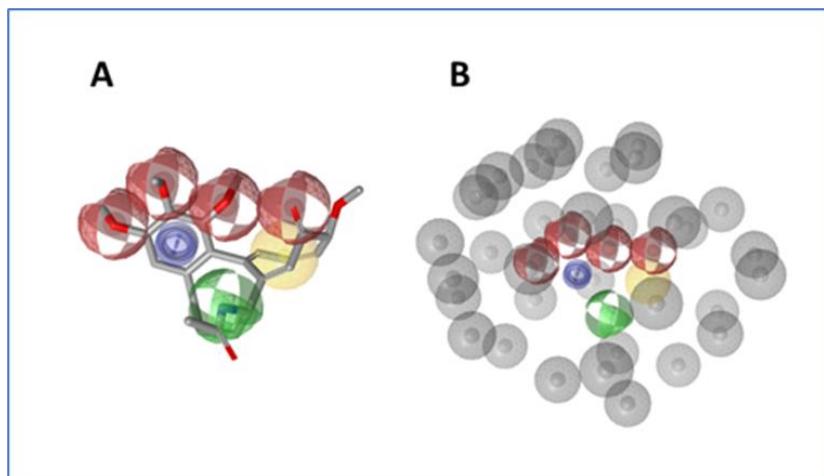


Figure S7. A) Optimised LB pharmacophore model LB7 aligned with base ligand **AS30** and B) the model LB7 with Xvols spheres. (HC, yellow spheres), (AI, purple sphere), (HBD; green arrows/spheres), (HBA; red arrows/spheres), and (Xvols; grey sphere).

Model LB8

Pharmacophore model LB8 was developed by manually selecting and aligning **AS1**, **AS3**, **AS8**, **AS12**, **AS13**, **AS14**, **AS15**, **AS16**, and **AS17** and creating a merged features pharmacophore. After automatic pharmacophore model generation, removal and tolerance adjusting of features and Xvols coat modifications were optimised to find as many actives as possible while excluding inactives. The final model contained one AI feature, two HC features, two HBAs and 30 Xvols.

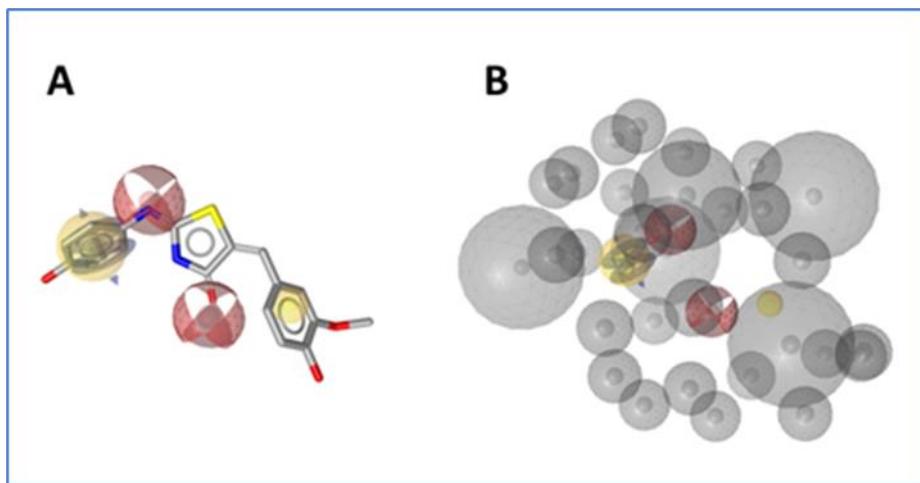


Figure S8. A) Optimised LB pharmacophore model LB8 aligned with base ligand **AS15** and B) the model LB8 with Xvol spheres. (HC, yellow spheres), (AI, purple sphere), (HBA; red arrows/spheres), and (Xvols; grey sphere).

Model LB9

Pharmacophore model LB9 was developed by manually selecting and aligning **AS3**, **AS8**, **AS12**, and **AS13** and creating a merged features pharmacophore. After automatic pharmacophore model generation, removal and tolerance adjusting of features and Xvols coat modifications were optimised to find as many actives as possible while excluding inactives. The final model contained two AI features, one HC feature, three HBAs and 39 Xvols.

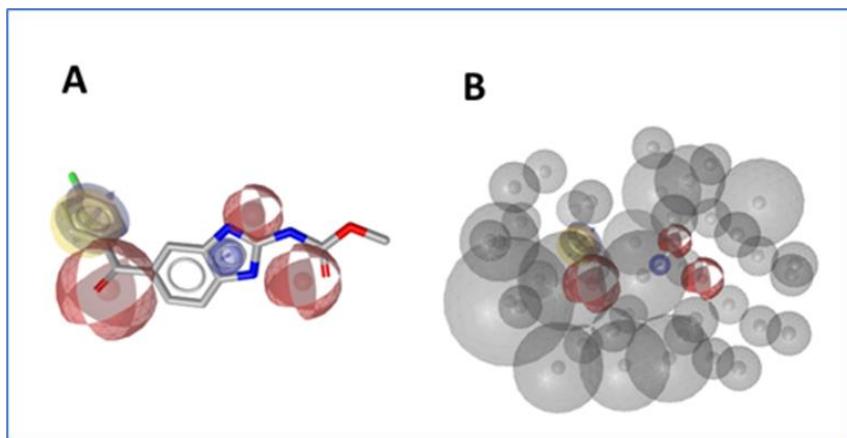


Figure S9. A) Optimised LB pharmacophore model LB9 aligned with base ligand **AS8** and B) the model LB9 with Xvols spheres. (HC, yellow spheres), (AI, purple sphere), (HBA; red arrows/spheres), and (Xvols; grey sphere).

Model LB10

Pharmacophore model LB10 was developed by automatic clustering and aligning **AS79**, **AS80**, **AS81**, and **AS82** creating a merged features pharmacophore. After automatic pharmacophore model generation, removal and tolerance adjusting of features and Xvols coat modifications were optimised to find as many actives as possible while excluding inactives. The final model contained two AI features, two HC features, two HBAs and 45 Xvols.

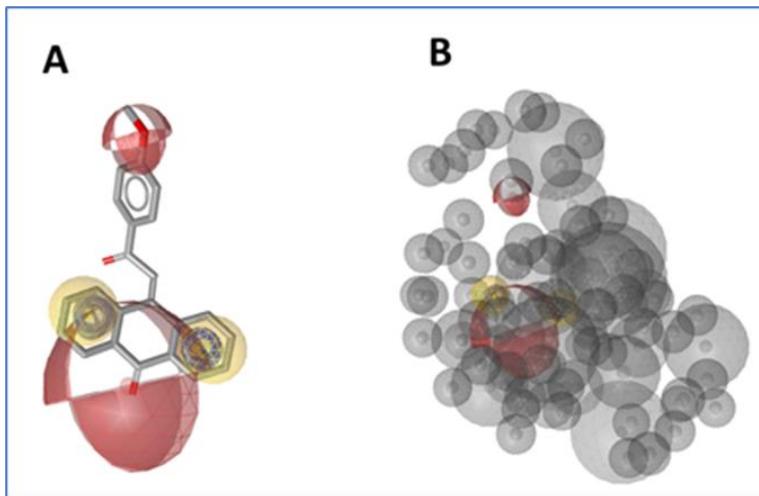


Figure S10. A) Optimised LB pharmacophore model LB10 aligned with base ligand **AS80** and B) the model LB10 with Xvol spheres. (HC, yellow spheres), (Al, purple sphere), (HBA; red arrows/spheres), and (Xvols; grey sphere).

Model LB11

Pharmacophore model LB11 was developed by manually selecting and aligning **AS13**, **AS27**, **AS43**, and **AS72** and creating a merged features pharmacophore. After automatic pharmacophore model generation, removal and tolerance adjusting of features and Xvols coat modifications were optimised to find as many actives as possible while excluding inactives. The final model contained one HC feature, four HBAs and 39 Xvols.

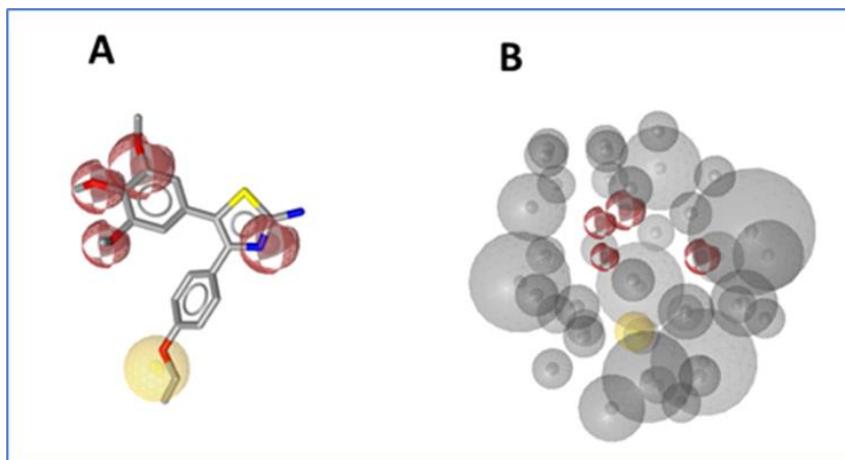


Figure S11. A) Optimised LB pharmacophore model LB11 aligned with base ligand **AS13** and B) the model LB11 with Xvol spheres. (HC, yellow spheres), (AI, purple sphere), (HBA; red arrows/spheres), and (Xvols; grey sphere).

Model DS2

The model DS2 was based on **AS64**, a diphenyl sulfonamide. It contained three HC features, two AI features that are mapped on the two phenyl rings, two HBD features and 26 Xvols.

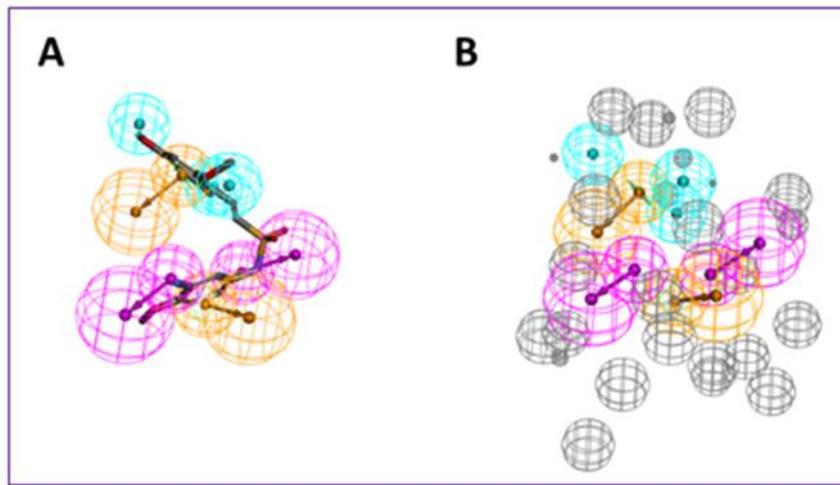


Figure S12. A) Optimised LB pharmacophore model DS2 aligned with base ligand **AS64**. B) the model DS2 with Xvol spheres. (HC, blue spheres), (AI, brown spheres), (HBD; purple), and (Xvols; grey sphere).

Model DS5

Model DS5 was based on **AS75**, a podophyllotoxin. It contains four HC features, two AI features, five HBA features and 34 Xvols.

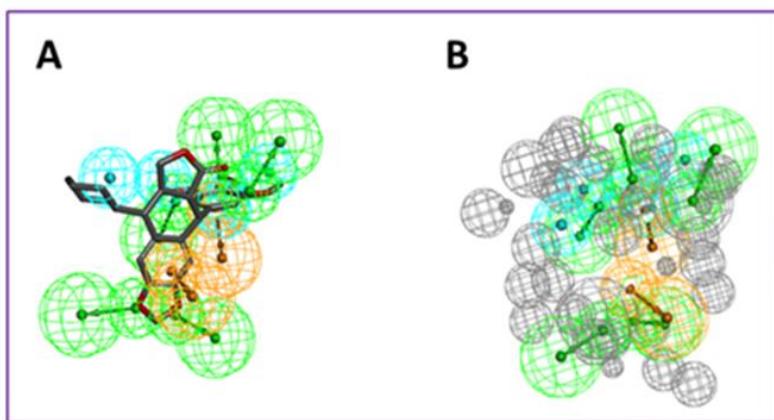


Figure S13. A) Optimised LB pharmacophore model DS5 aligned with base ligand **AS75**. B) the model DS5 with Xvol spheres. (HC, blue spheres), (AI, brown spheres), (HBA; green sphere), and exclusion volumes (Xvols; grey sphere).

Model DS8

The model DS8 was calculated for **AS77**, a modified podophyllotoxin scaffold with an amine functionality inserted in the central ring and an elongated linker to the trimethoxyphenylring.

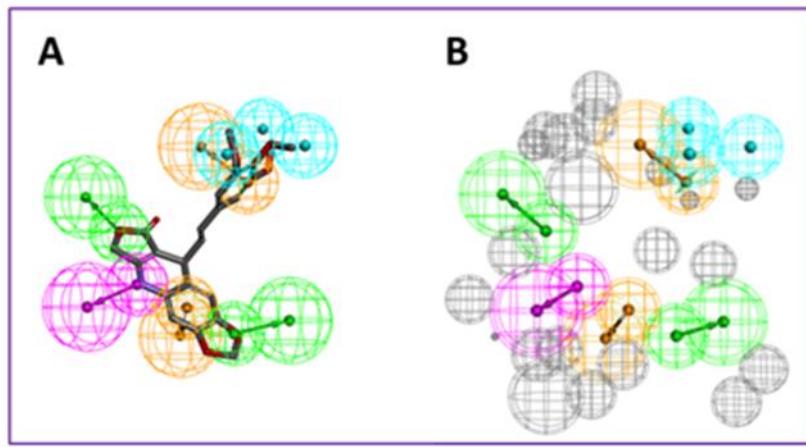


Figure S14. A) Optimised LB pharmacophore model DS8 aligned with base ligand **AS77**. B) the model DS8 with Xvol spheres. (HC, blue spheres), (AI, brown spheres), (HBA; green sphere) (HBD; purple sphere), and (Xvols; grey sphere).

Model DS9

Model DS9 was derived from **AS79**, an anthracene sulfonate. It consists of a HC feature, three AI features, two HBA features and 60 Xvols.

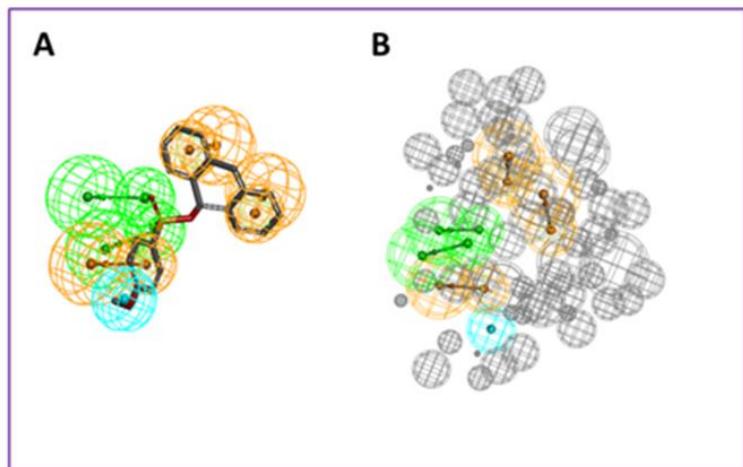


Figure S15. A) Optimised LB pharmacophore model DS9 aligned with base ligand **AS79**. B) the model DS9 with Xvol spheres. (HC, blue spheres), (AI, brown spheres), (HBA; green sphere), and (Xvols; grey sphere).

Model DS12

The model DS12 was based on **AS29**, a chalcone. The final model contains two HC, two AI, two HBA features and 118 Xvols.

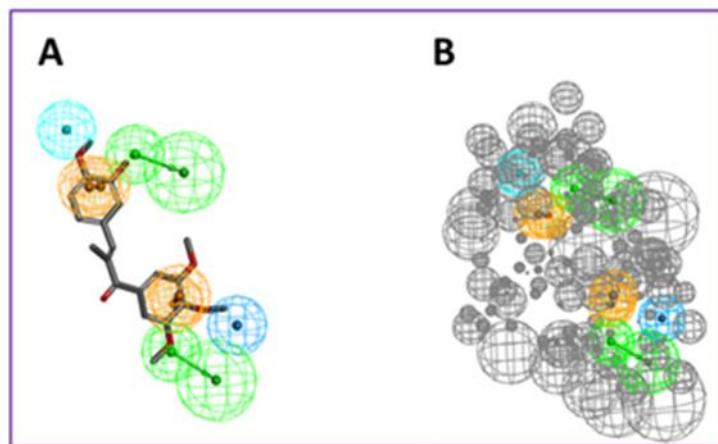


Figure S16. A) Optimised LB pharmacophore model DS12 aligned with base ligand **AS29**. B) the model DS12 with Xvol spheres. (HC, blue spheres), (AI, brown spheres), (HBA; green sphere), and (Xvols; grey sphere).

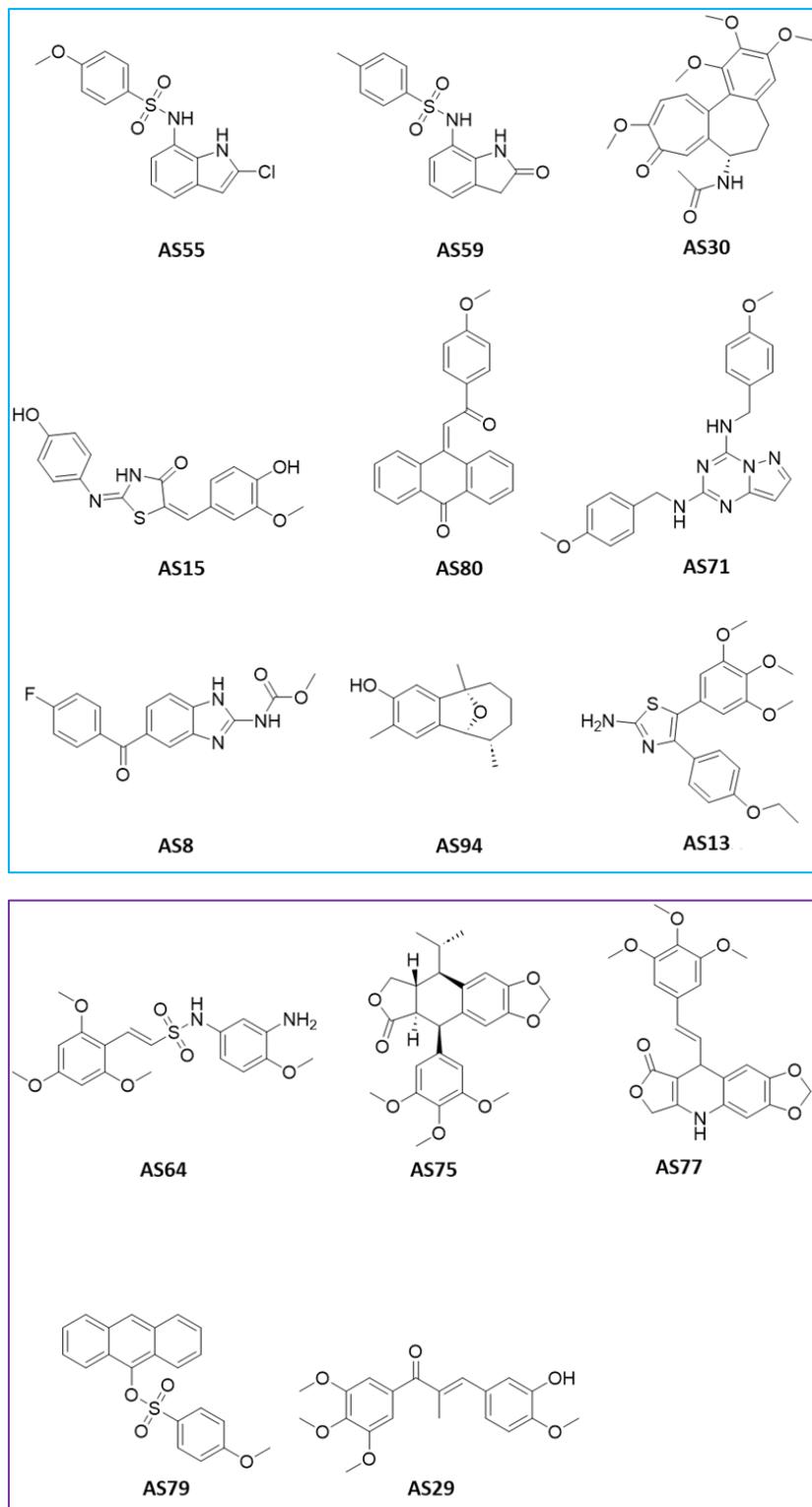


Figure S17. Chemical structures of the main active scaffolds used to generate the LB pharmacophore models. Compounds in the blue box were used for the LS models and compounds in the purple box were used for the DS models.

3. Virtual screening of pharmacophore models with consensus hits

Table S2. Virtual screening results with consensus hits obtained by the respective pharmacophore models.

Models	Consensus hit (n=2)		
	Specs (Synth)	Specs (NP)	PhytoChem (PC)
DS2/DS8	0	0	1
DS5/DS8	0	3	0
DS8/DS9	4	1	0
DS5/DS9	9	0	0
DS2/DS9	13	0	0
DS12/DS9	5	0	0
DS12/DS2	3	0	0
LB8/DS9	3	0	0
LB8/DS2	1	0	0
LB8/DS12	2	0	0
LB7/DS9	2	0	0
LB6/DS9	1	0	0
LB5/DS9	19	0	0
LB5/DS2	10	0	0
LB5/DS12	3	0	0
LB5/DS8	4	0	0
LB5/DS6	3	0	0
LB3/DS9	46	0	0
LB3/DS5	1	0	0
LB3/DS2	5	0	0
LB3/DS12	3	0	0
LB3/LB9	1	0	0
LB3/LB8	4	0	0
LB3/LB6	16	0	0
LB3/LB5	167	0	0
SB2/LB5	4	0	0
LB11/DS9	6	0	0
LB11/DS2	1	0	0
LB11/DS12	1	0	0
LB11/LB9	1	0	0
LB11/LB8	1	0	0
LB11/LB7	1	0	0

LB11/LB6	6	0	0
LB11/LB5	5	0	0
LB11/LB3	6	0	0
LB11/SB2	3	0	0
LB10/DS9	6	0	0
LB10/DS5	5	0	0
LB10/LB3	2	0	0
LB10/LB11	3	0	0
SB1/LB5	4	0	0
SB1/LB3	1	0	0
Consensus hit (n=3)			
Models	Specs (Synth)	Specs (NP)	PhytoChem (PC)
DS5/DS8/DS9	0	3	0
LB10 / LB5 / DS9	1	0	0
LB10 / DS9/DS12	2	0	0
LB11 /LB6/DS2	1	0	0
LB3/LB5/LB8	1	0	0
LB3/LB5/DS9	2	0	0
LB5/DS2/DS8	1	0	0
DS2/DS5/DS9	1	0	0
LB3/LB5/DS2	1	0	0

4. Test compound selection of hits obtained from virtual screening of the databases.

Table S3. Virtual screening hits selected for experimental validation. Red box indicates insoluble compounds or fluorescent interference and thus excluded compounds.

Specs	Virtual Hit	SB1	SB2	LB3	LB4	LB5	LB6	LB7	LB8	LB9	LB10	LB11	DS2	DS5	DS8	DS9	DS12
AA-504/21163091	SC1													X			
			X														
AB-323/13887441	SC2		X												X		
AC-776/41252593	SC3									X				X			
AF-399/14738025	SC4									X							
AG-205/13184005	SC5			X													
AG-205/21054014	SC6														X		
AG-205/36265044	SC7	X															
AG-389/15452424	SC8									X	X						X
AG-401/30827061	SC9								X	X							X
AG-690/09788017	SC10											X					
AJ-292/21122017	SC11														X		
AJ-916/12583009	SC12							X		X							
AK-087/42718249	SC13							X									
AK-087/42718376	SC14															X	
AK-693/40962733	SC15							X									

AK-968/41922685	SC16	X																						
AM-807/13614425	SC17		X																					
AN-329/43211079	SC18																							
AN-465/14334022	SC19																							
AN-465/41674183	SC20																							
AN-648/42098983	SC21		X																					
AN-652/12103469	SC22																							
AO-022/43514129	SC23		X																					
AO-313/21215019	SC24																							
AO-365/43300818	SC25																							
AO-365/43300823	SC26		X																					
AO-365/43300935	SC27																							
AO-365/43474517	SC28																							
AO-365/43486487	SC29																							
AO-476/15509049	SC30																							
AP-501/43179291	SC31																							
AP-263/40720502	SC32																							
AP-123/40765218	SC33																							
AP-263/43418357	SC34																							
AP-845/42065522	SC35																							
AP-853/42655770	SC36																							
AP-853/43386817	SC37																							

AQ-086/43467809	SC38														
AT-057/43469212	SC39														
AT-057/43485534	SC40														
AT-583/41299652	SC41														
AH-487/41949628	SC42														
AG-690/12094002	SC43														
AI-204/31688027	SC44														
AG-690/12413358	SC45														
AI-899/21033027	SC46														

5. Bioactivity results of selected test compounds from the polymerisation inhibition assay and similarity prediction

Table S4. Overview of the tubulin polymerisation inhibition screening results.

Specs ID-number	ID	% Inhibition (30 µM) ± SD *20 µM	% Inhibition (10 µM) ± SD	IC50 (µM)
AA-504/21163091	SC1	28.8 ± 22.7	n.d	n.d
AB-323/13887441	SC2	≤ 0	n.d	n.d
AC-776/41252593	SC3	Insoluble	≤ 0	n.d
AF-399/14738025	SC4	7.0 ± 8.1	n.d	n.d
AG-205/13184005	SC5	≤ 0	n.d	n.d
AG-205/21054014	SC6	38.4 ± 12.6	n.d	n.d
AG-205/36265044	SC7	insoluble	insoluble	n.d
AG-389/15452424	SC8	Insoluble	≤ 0	n.d
AG-401/30827061	SC9	Insoluble	Insoluble	n.d
AG-690/09788017	SC10	Insoluble	Insoluble	n.d
AJ-292/21122017	SC11	≤ 0	n.d	n.d
AJ-916/12583009	SC12	insoluble	insoluble	n.d
AK-087/42718249	SC13	≤ 0	n.d	n.d
AK-087/42718376	SC14	insoluble	43.9 ± 23.9	n.d
AK-693/40962733	SC15	n.d	≤ 0	n.d
AK-968/41922685	SC16	n.d	≤ 0	n.d
AM-807/13614425	SC17	n.d	1.7 ± 3.1	n.d
AN-329/43211079	SC18	n.d	n.d	n.d
AN-465/14334022	SC19	n.d	≤ 0	n.d
AN-465/41674183	SC20	19. 2 ± 24.9	n.d	n.d
AN-648/42098983	SC21	n.d	1.29 ± 1.28	n.d
AN-652/12103469	SC22	54.7 ± 2.5	n.d	n.d
AO-022/43514129	SC23	99.3 ± 3.72	86.5 ± 2.7	Yes
AO-313/21215019	SC24	insoluble	*95.50 ± 2.5	n.d
AO-365/43300818	SC25	25.5 ± 14.6	n.d	n.d
AO-365/43300823	SC26	insoluble	37.6 ± 35.9	n.d
AO-365/43300935	SC27	32.87 ± 1.69	36.0 ± 4.4	n.d
AO-365/43474517	SC28	Insoluble	2.33 ± 1.8	n.d
AO-365/43486487	SC29	8.69 ± 4.42	n.d	n.d
AO-476/15509049	SC30	Insoluble	Insoluble	n.d
AP-501/43179291	SC31	Insoluble	≤ 0	n.d
AP-263/40720502	SC32	73.9 ± 0.2	n.d	n.d
AP-123/40765218	SC33	≤ 0	n.d	n.d

AP-263/43418357	SC34	37.8 ± 5.7	n.d	n.d
AP-845/42065522	SC35	Insoluble	7.6 ± 8.0	n.d
AP-853/42655770	SC36	Insoluble	≤ 0	n.d
AP-853/43386817	SC37	100.3 ± 0.4	71.3 ± 6.4	Yes
AQ-086/43467809	SC38	Insoluble	6.7 ± 1.0	n.d
AT-057/43469212	SC39	Insoluble	≤ 0	n.d
AT-057/43485534	SC40	Insoluble	3.9 ± 7.3	n.d
AT-583/41299652	SC41	≤ 0	n.d	n.d
AH-487/41949628	SC42	≤ 0	n.d	n.d
AG-690/12094002	SC43	Insoluble	11.3 ± 3.5	n.d
AI-204/31688027	SC44	8.84 ± 1.45	n.d	n.d
AG-690/12413358	SC45	Insoluble	17.7 ± 1.0	n.d
AI-899/21033027	SC46	8.3 ± 2.4	n.d	n.d
MERCK Group	Hit47	≤ 0	n.d	n.d
TCI Chemicals	Colchicine	98.8 ± 5.2	83.2 ± 1.7	YES

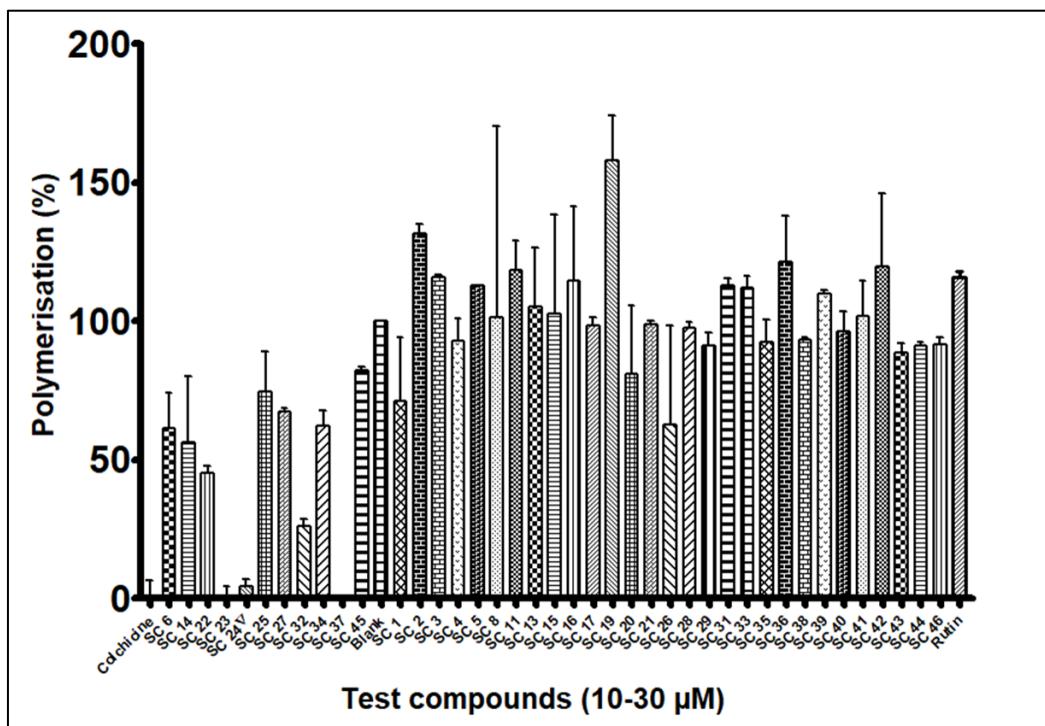


Figure S18. Graphical results of the active/inactive preliminary screening. Corresponding numerical values are shown above in Table S4.

Table S5. SwissTarget Similarity prediction scores for the tubulin inhibitors

Inhibitor	Tubulin-Ligand Probability Score	Best Target Probability score	CHEMBL Target ID
SC6	none	Carbonic anhydrase II (0.04)	CHEMBL205
SC14	CHEMBL195840 (0.62)	-	CHEMBL1915
SC22	none	Phosphodiesterase 5A (0.11)	CHEMBL1827
SC23	none	Acetyl-CoA carboxylase 2 (0.11)	CHEMBL4829
SC24	CHEMBL363063 (0.90)	-	CHEMBL1915
SC25	none	NAD-dependent deacetylase sirtuin 1 (0.14)	CHEMBL4506
SC27	none	NAD-dependent deacetylase sirtuin 1 (0.21)	CHEMBL4506
SC32	none	Metabotropic glutamate receptor 5 (0.11)	CHEMBL3227
SC34	none	Microtubule-associated protein tau (0.79)	CHEMBL1293224
SC37	none	Monoamine oxidase B (0.11)	CHEMBL2039
SC45	none	Histone deacetylase 1 (0.12)	CHEMBL325

6. Active compound characterization provided by Specs

Table S6. List of compound characterization data for the found active inhibitors provided by SPECS.

ID	Specs ID	QC Method	Purity	MW	SMILES
SC6*	AG-205/210 54014	LC-MS	>95%	730.7	CC1=CC(=O)Oc2cc(ccc12)OC6OC(C(=O)C(=O)c3cccc3)C(=O)c4cccc4)C6(OC(=O)c5cccc5)
SC14	AK-087/427 18376	H-NMR,LC-MS	>95%	346.38	Oc3ccc(C(=O)C=Cc2cccc2(OCc1cccc1))c(O)c3

SC22	AN-652/121 03469	H-NMR,LC-MS	95%	401.46	<chem>CCOC(=O)c2ccccc2(NC(=O)c1cc(OCC)c(OCC)c(c1)OCC)</chem>
SC23	AO-022/435 14129	LC-MS	>95%	445.4	<chem>O=C(CC1NC(=O)c2ccccc2(NC1(=O)))NCCn3c4ccccc4(nc3C(F)(F)F)</chem>
SC24	AO-313/212 15019	H-NMR,LC-MS	95%	551.98	<chem>COc1cc(cc(OC)c1(OC))C5c2cc6OCOc6(cc2C(NC(=O)c3ccccc3Cl)C4CO C(=O)C45)</chem>
SC25	AO-365/433 00818	LC-MS	>95%	328.35	<chem>COc1ccc(cc1)OCC3=Nn2c(nnc2S3)c4ccco4</chem>
SC27	AO-365/433 00935	H-NMR,LC-MS	95%	298.33	<chem>COc1ccccc1C3=Nn2c(nnc2S3)c4ccco4</chem>
SC32	AP-263/407 20502	LC-MS	>95%	449.12	<chem>CC(=O)Nc2ccccc2(OS(=O)(=O)c1cc(ccc1Br)Br)</chem>
SC34	AP-263/434 18357	LC-MS	>95%	348.2	<chem>COc1c(ccc(c1Cl)Cl)S(=O)(=O)Nc2cccc(O)c2</chem>
SC37	AP-853/433 86817	H-NMR,LC-MS	95%	389.82	<chem>O=C(CSc1nnnc(o1)c2cccc(c2)Cl)Nc3ccc4OCOc4(c3)</chem>
SC45	AG-690/124 13358	H-NMR,LC-MS	>95%	525.4	<chem>COc1ccc(cc1)NC(=O)c2ccc(cc2)Nc4nc3ccc(cc3c(n4)c5ccccc5)Br</chem>

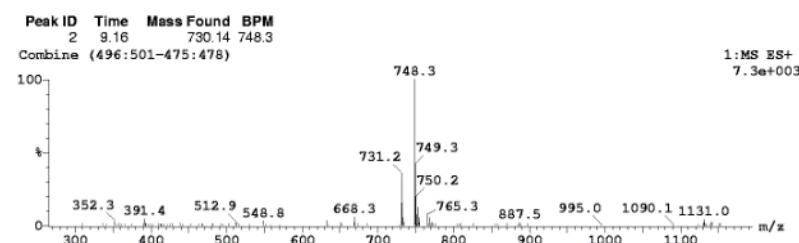
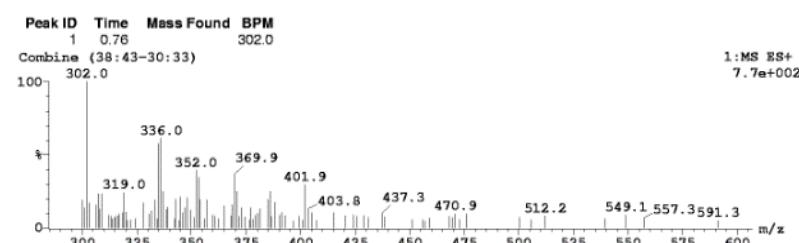
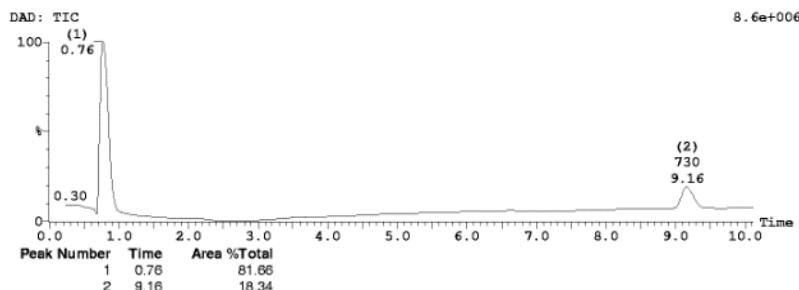
* Stereochemistry undefined

7. LC-MS and ^1H -NMR data provided by Specs

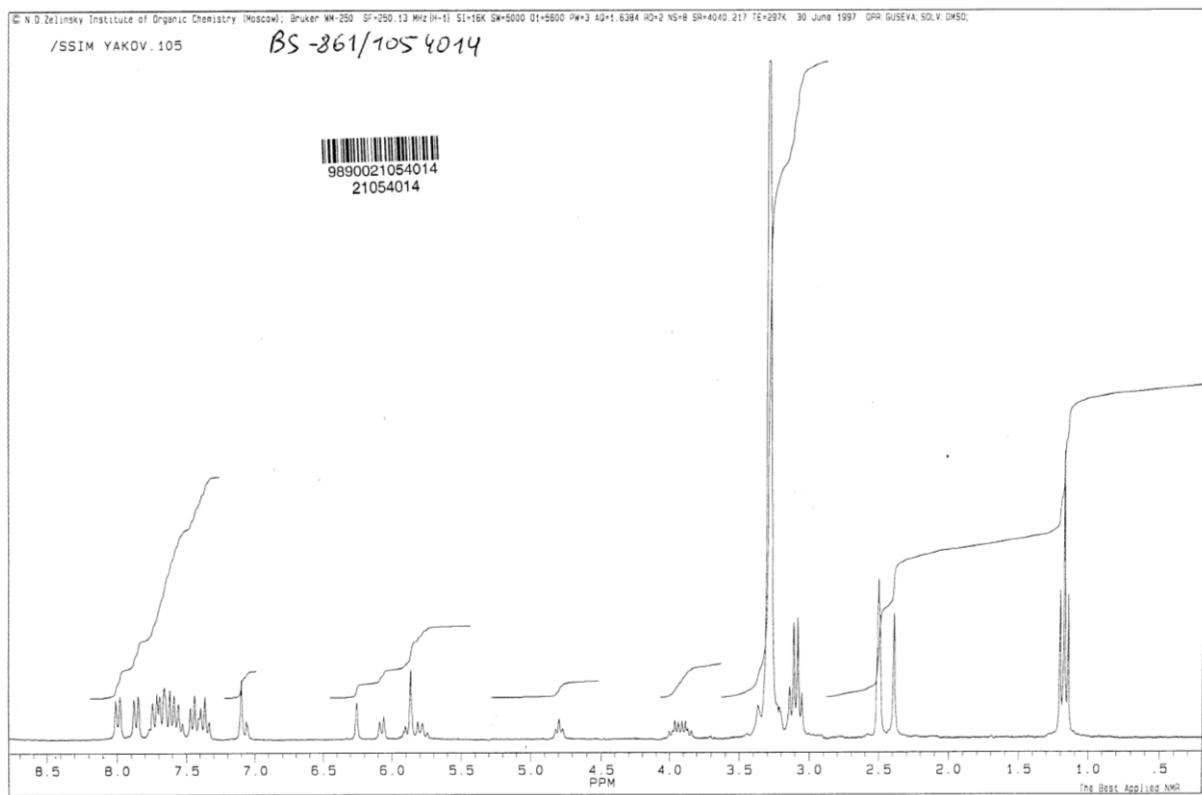
Compound **SC6**, LC-MS:

Printed: Thu Nov 29 09:26:51 2001

Sample Report (continued):



Compound SC6, ¹H-NMR:

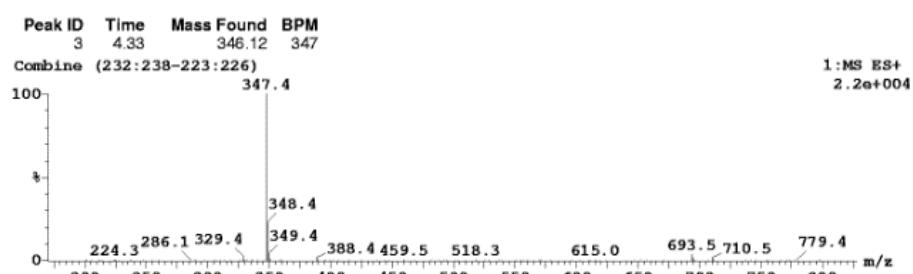
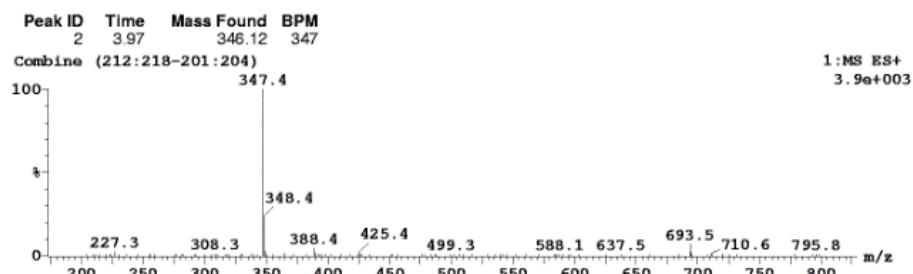
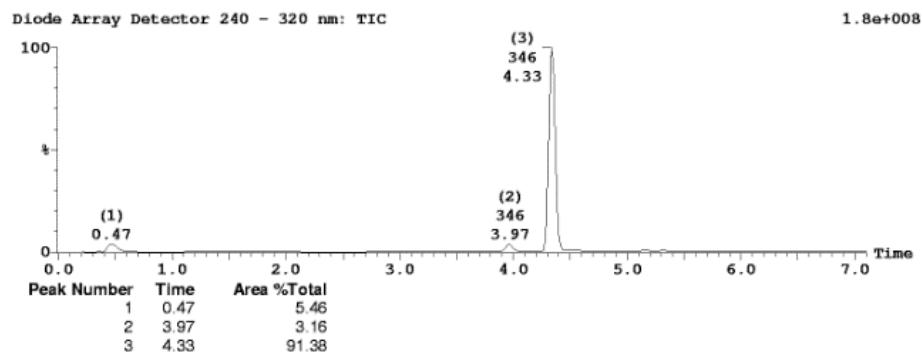


Compound **SC14**, LC-MS:

Specs
 File:9900142718376 ID:AK-087/42718376
 Instrument:LC/MS A Vial:1.89
 Description:C22H18O4
 Date:26-Feb-2004

Printed: Thu Feb 26 16:06:21 2004

Sample Report (continued):

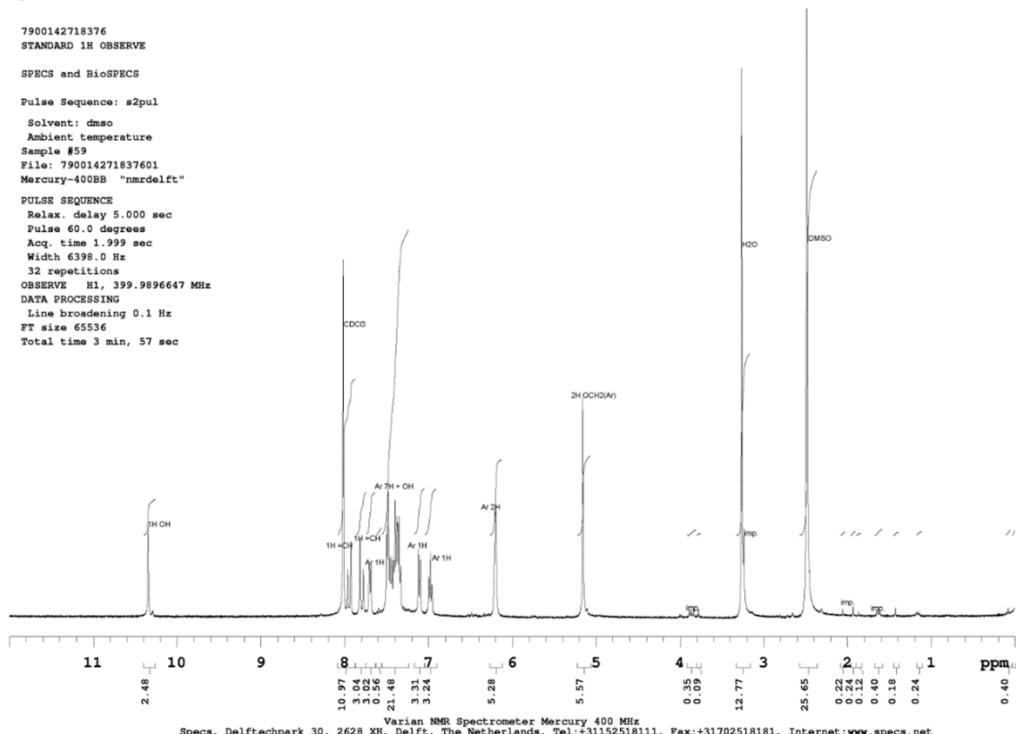


Compound **SC14**, ¹H-NMR:

Specs

Date: Thu Apr 1, 2004

7900142718376
STANDARD 1H OBSERVE
SPECs and BioSPECs
Pulse Sequence: s2pul
Solvent: dmso
Ambient temperature
Sample #59
File: 790014271837601
Mercury-400B "nmmdelft"
PULSE SEQUENCE
Relax. delay 5.000 sec
Pulse 60.0 degrees
Acq. time 1.999 sec
Width 6398.0 Hz
32 repetitions
OBSERVE H1, 399.9896647 MHz
DATA PROCESSING
Line broadening 0.1 Hz
FT size 65536
Total time 3 min, 57 sec



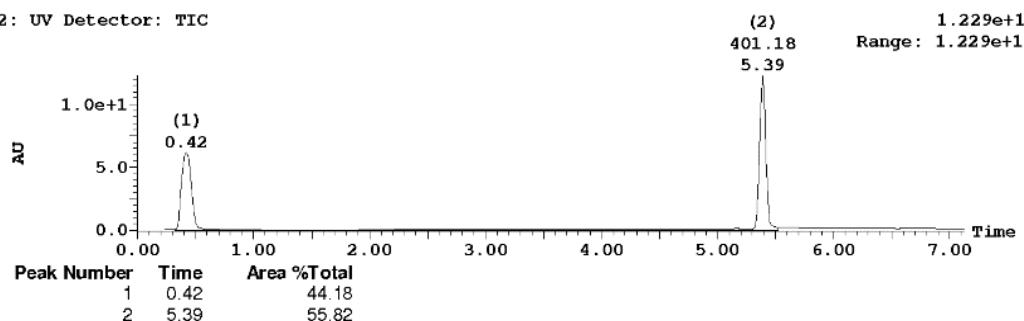
Varian NMR Spectrometer Mercury 400 MHz
Specs, Delfttechpark 30, 2628 XH, Delft, The Netherlands, Tel.:+31152518111, Fax:+31702518181, Internet:www.specs.net

Compound **SC22**, LC-MS:

Specs info@specs.net, www.specs.net
File:9900112103469 ID:AN-652/12103469
Vial:1:105 Date:26-Jul-2006

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Description:C22H27NO6
Time:04:42:24

2: UV Detector: TIC

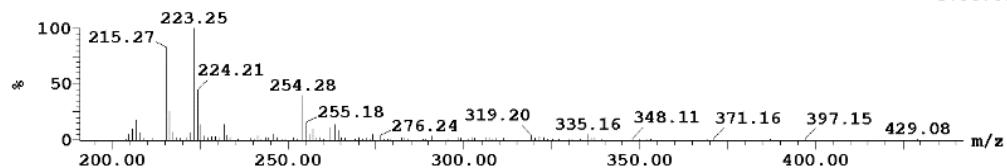


Peak ID Time Mass Found BPM

1	0.42	223.25
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Combine (18:25-9:13)

1:MS ES+
3.5e+004

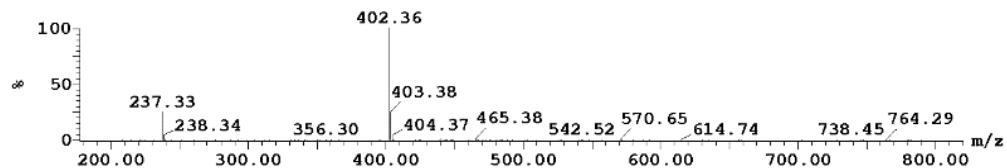


Peak ID Time Mass Found BPM

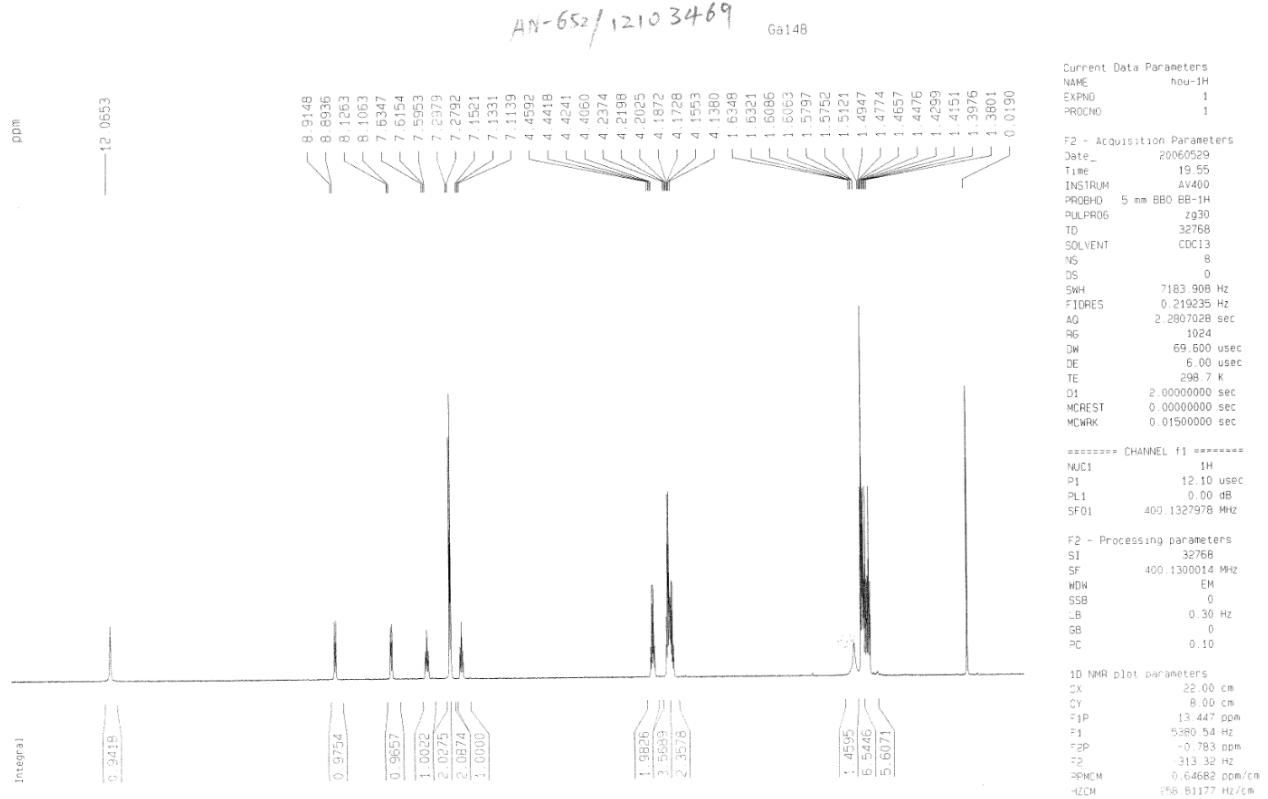
2	5.39	401.18
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Combine (289:296-281:285)

1:MS ES+
1.2e+005

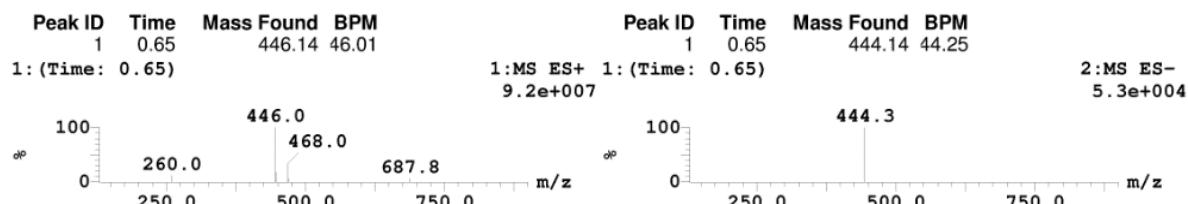
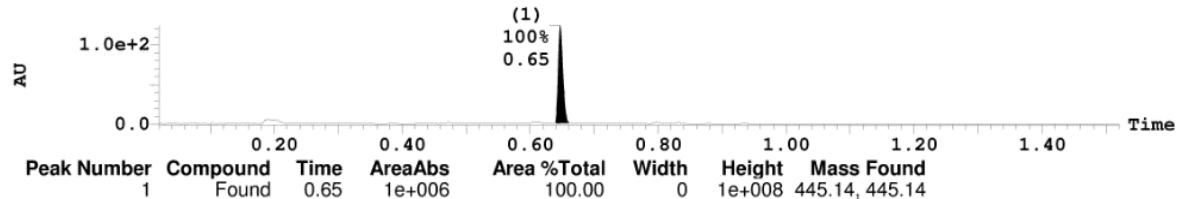


Compound SC22, ¹H-NMR:



Compound **SC23**, LC-MS:

3: UV Detector: TIC Smooth (SG, 2x2) 1.237e+2
Range: 1.237e+2

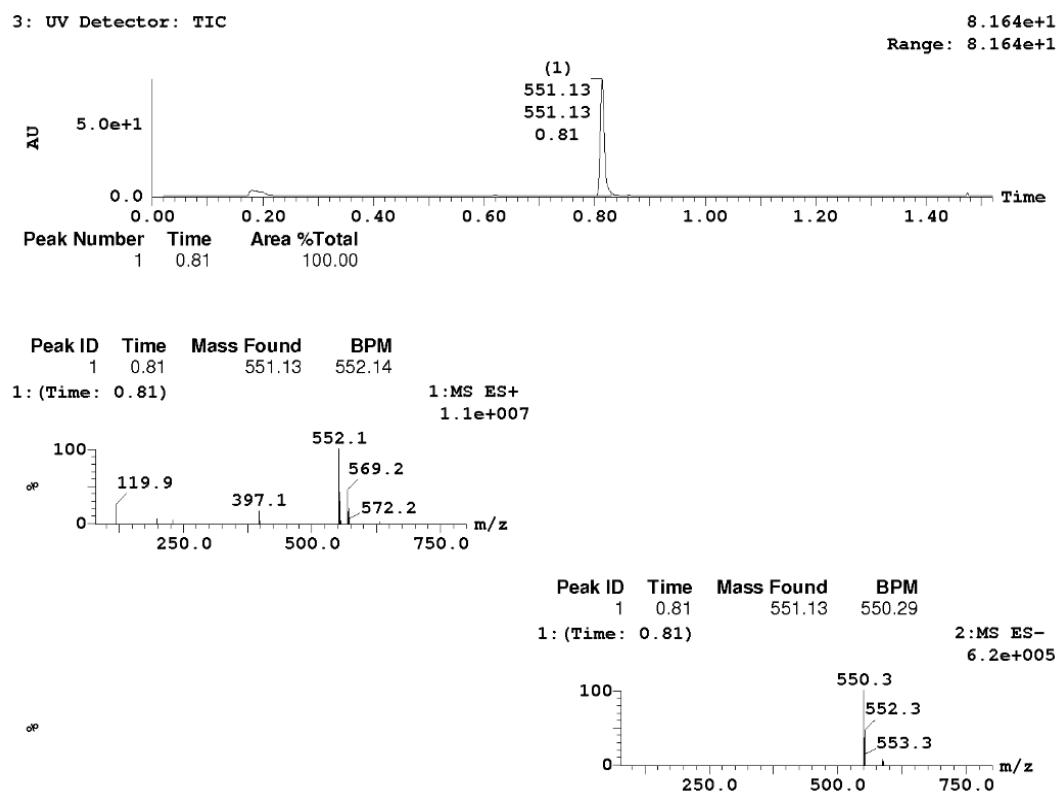


Compound **SC24**, LC-MS:

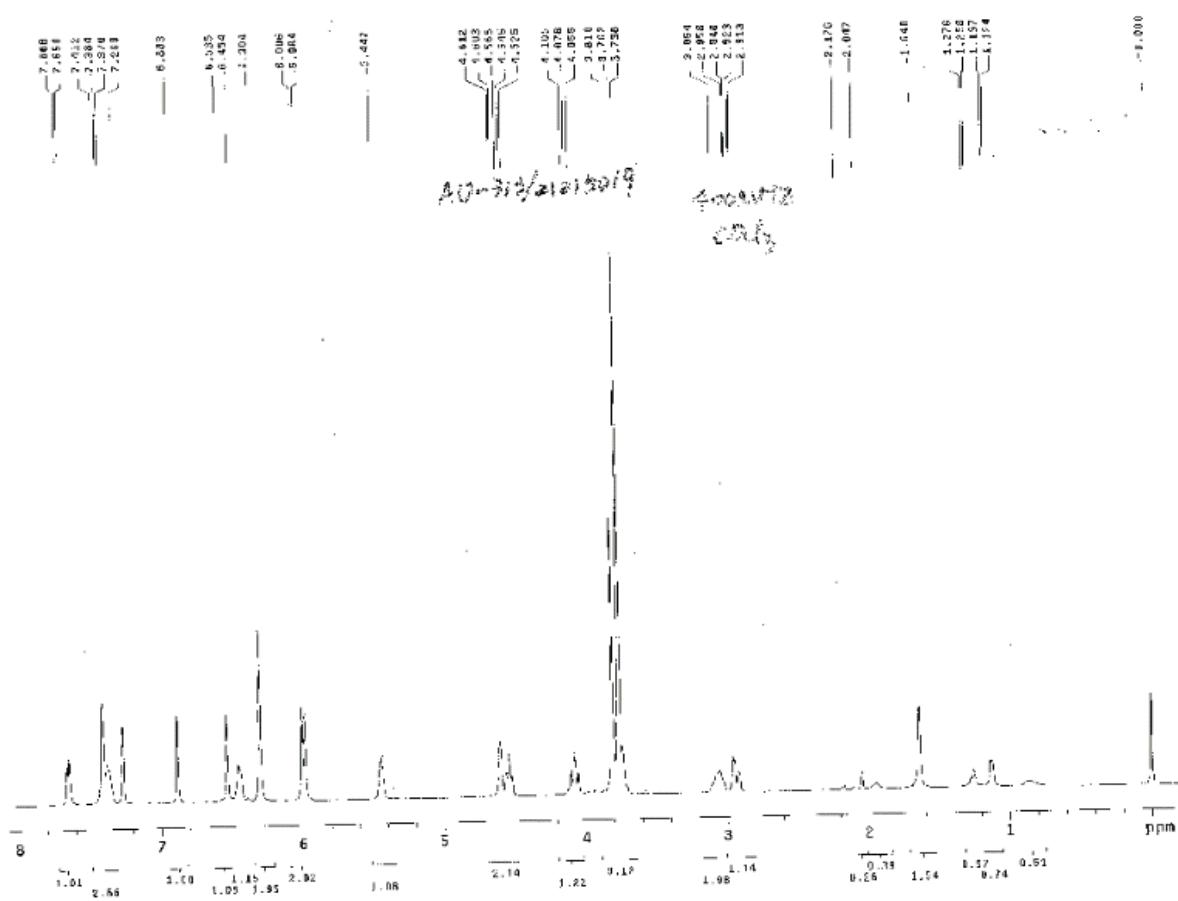
Specs info@specs.net, www.specs.net
File:9920121215019 ID:AO-313/21215019
Vial:7:F,12 Date:19-Feb-2008

Description:C29H26ClNO8
Time:19:52:49

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Compound SC24, ¹H-NMR:



Compound **SC25**, LC-MS:

Specs info@specs.net, www.specs.net

File:6600143300818

Vial:2:64

ID:AO-365/43300818

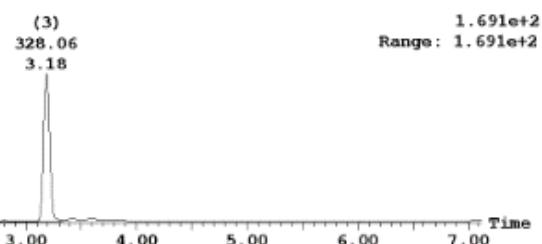
Date:27-Oct-2006

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Description:C15H12N4O3S

Time:17:20:46

2: UV Detector: TIC



1.691e+2

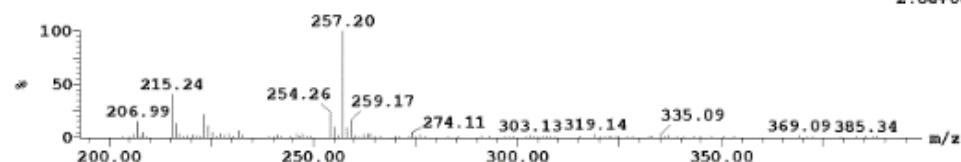
Range: 1.691e+2

Peak ID Time Mass Found BPM

257.20

Combine (19:25-10:14)

1:MS ES+
2.8e+004

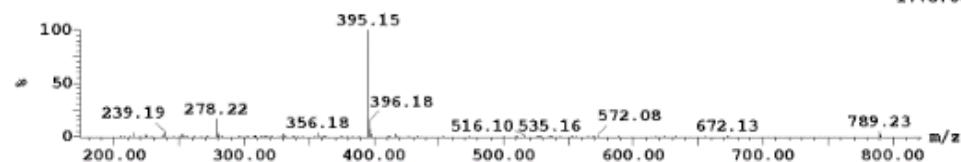


Peak ID Time Mass Found BPM

395.15

Combine (125:132-117:121)

1:MS ES+
1.7e+004

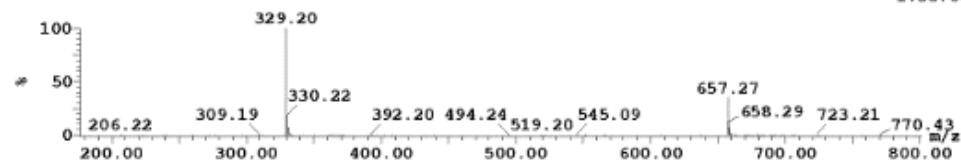


Peak ID Time Mass Found BPM

328.06

Combine (169:175-158:162)

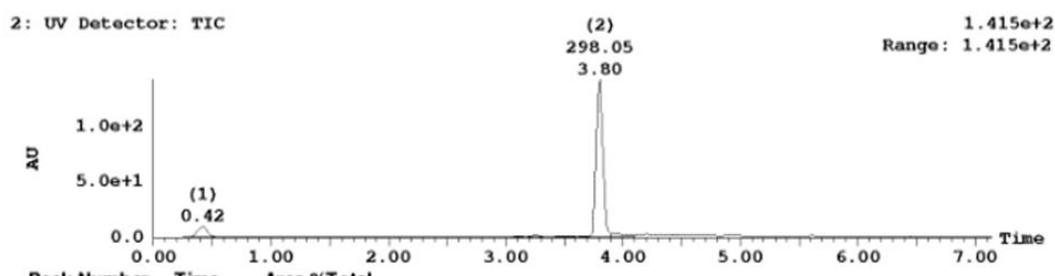
1:MS ES+
1.3e+006



Compound SC27, LC-MS:

Specs info@specs.net, www.specs.net
File:9900143300935 ID:AO-365/43300935
Vial:1:52 Date:31-Aug-2006

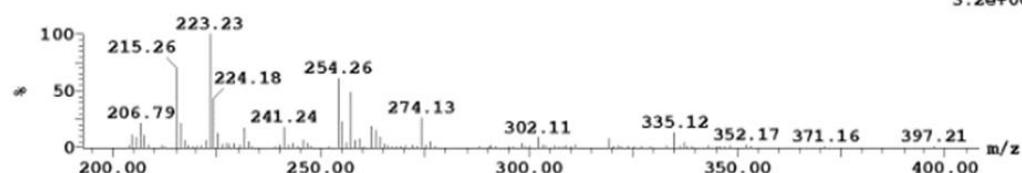
Page 52
Description:C14H10N4O2S
Time:13:50:32



Peak ID Time Mass Found BPM
1 0.42 223.23

Combine (18:25-9:13)

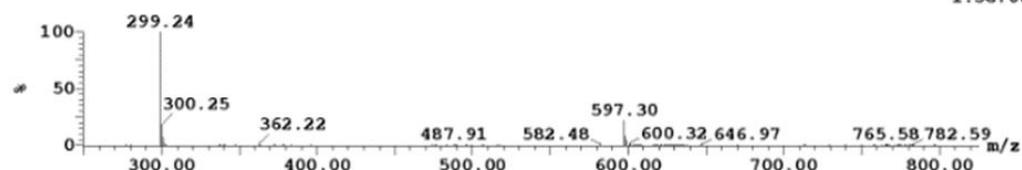
1:MS ES+
3.2e+004



Peak ID Time Mass Found BPM
2 3.80 298.05 299.24

Combine (203:209-190:194)

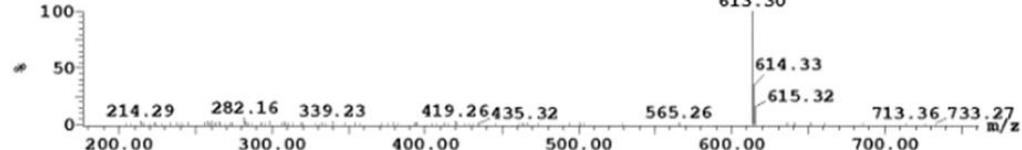
1:MS ES+
1.5e+006



Peak ID Time Mass Found BPM
3 3.95 613.30

Combine (211:217-204:209)

1:MS ES+
1.7e+004

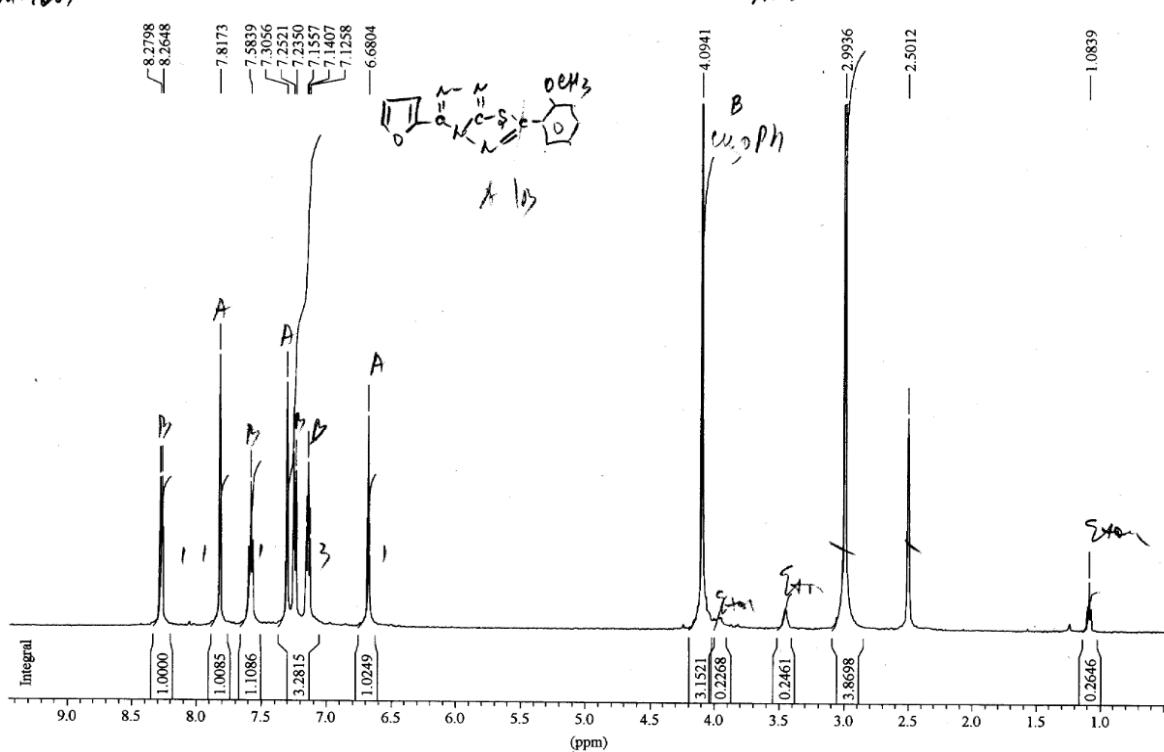


Compound SC27, ¹H-NMR:

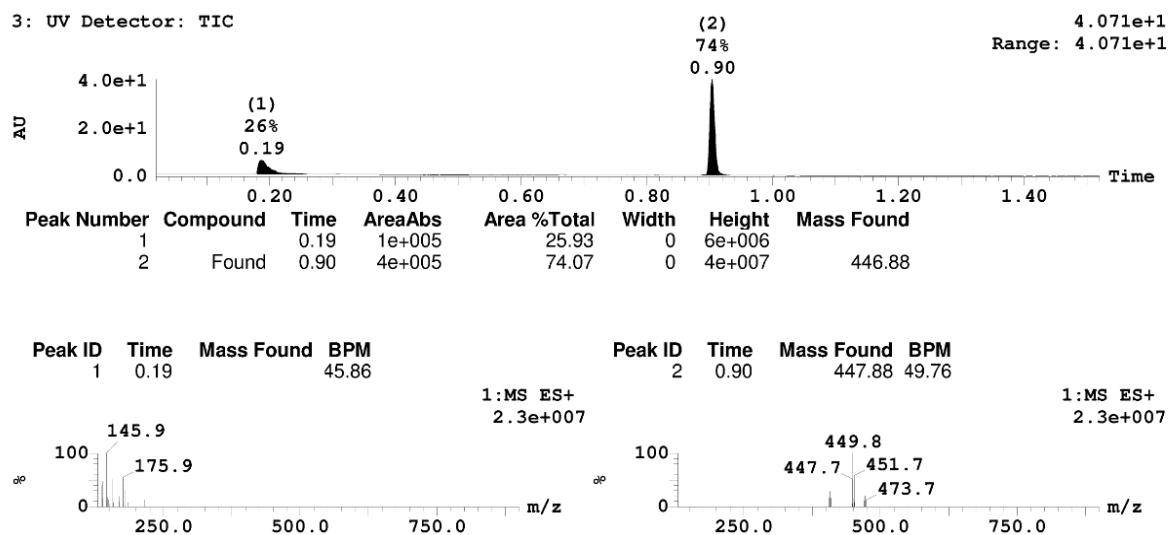
B-644
on 160Y

δ. 22 / 664

+
A0-365/43300935



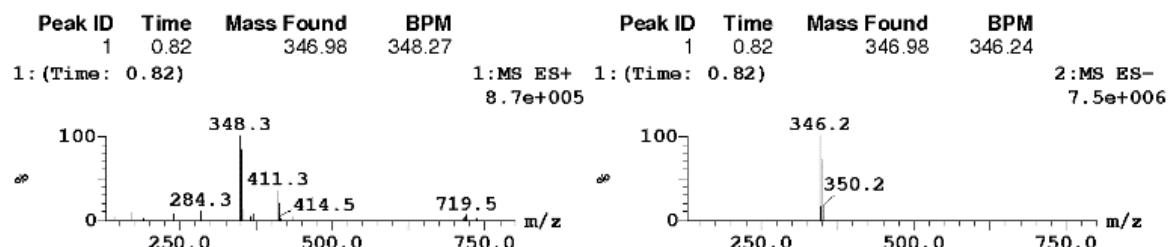
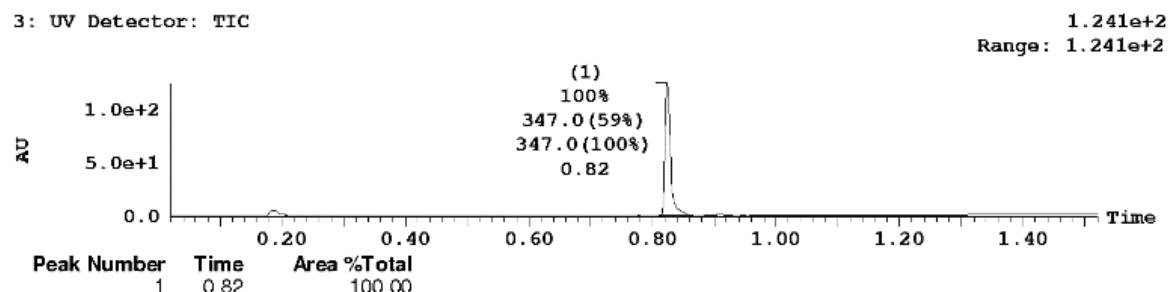
Compound **SC32**, LC-MS:



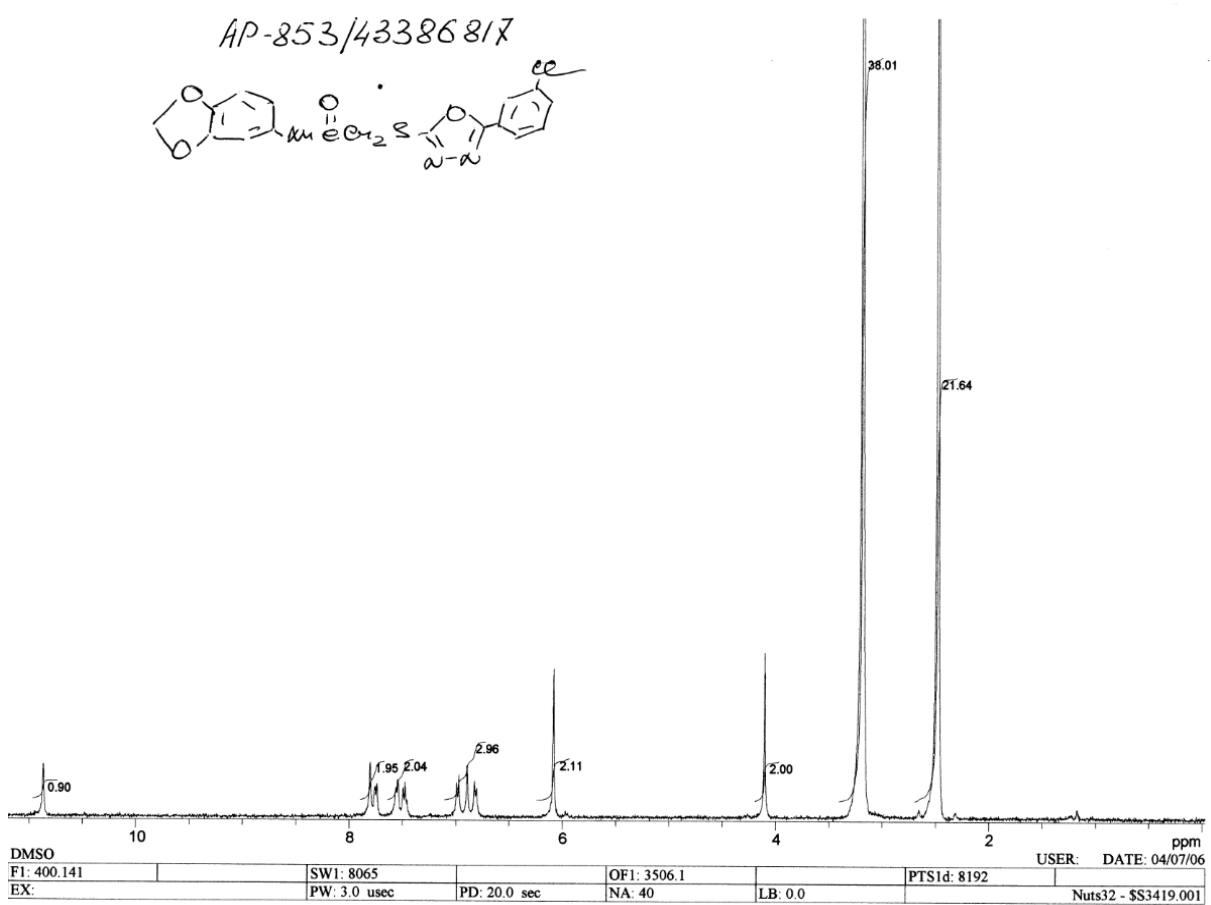
Compound **SC34**, LC-MS:

Specs info@specs.net, www.specs.net
File:9900143418357 ID:AP-263/43418357
Vial:5:11,F Date:09-Mar-2010

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Description:C13H11Cl2NO4S
Time:04:53:06



Compound SC37, ¹H-NMR:



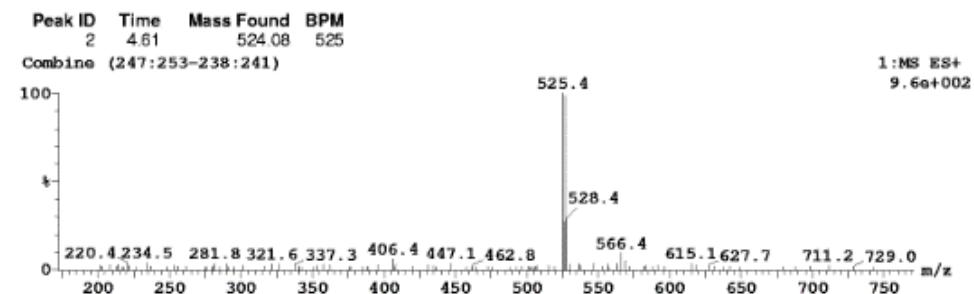
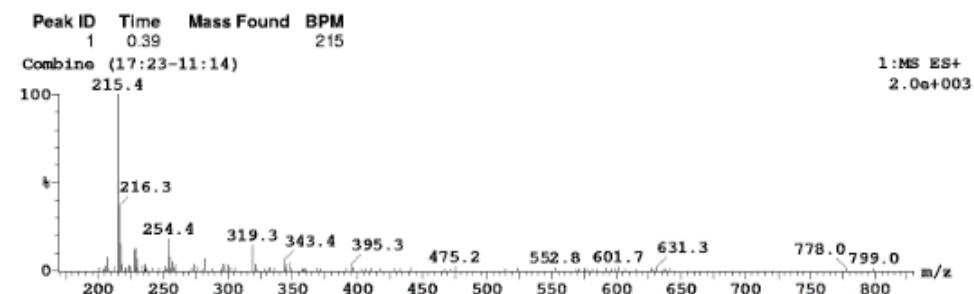
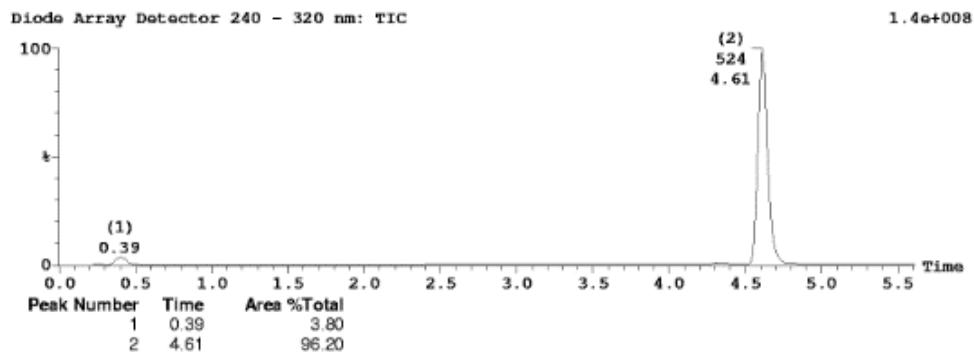
Compound **SC45**, LC-MS:

Specs
File:5000112413358 ID:AG-690/12413358 Description:C28H21BrN4O2
Instrument:LC/MS A Vial:1:11 Date:13-Apr-2004

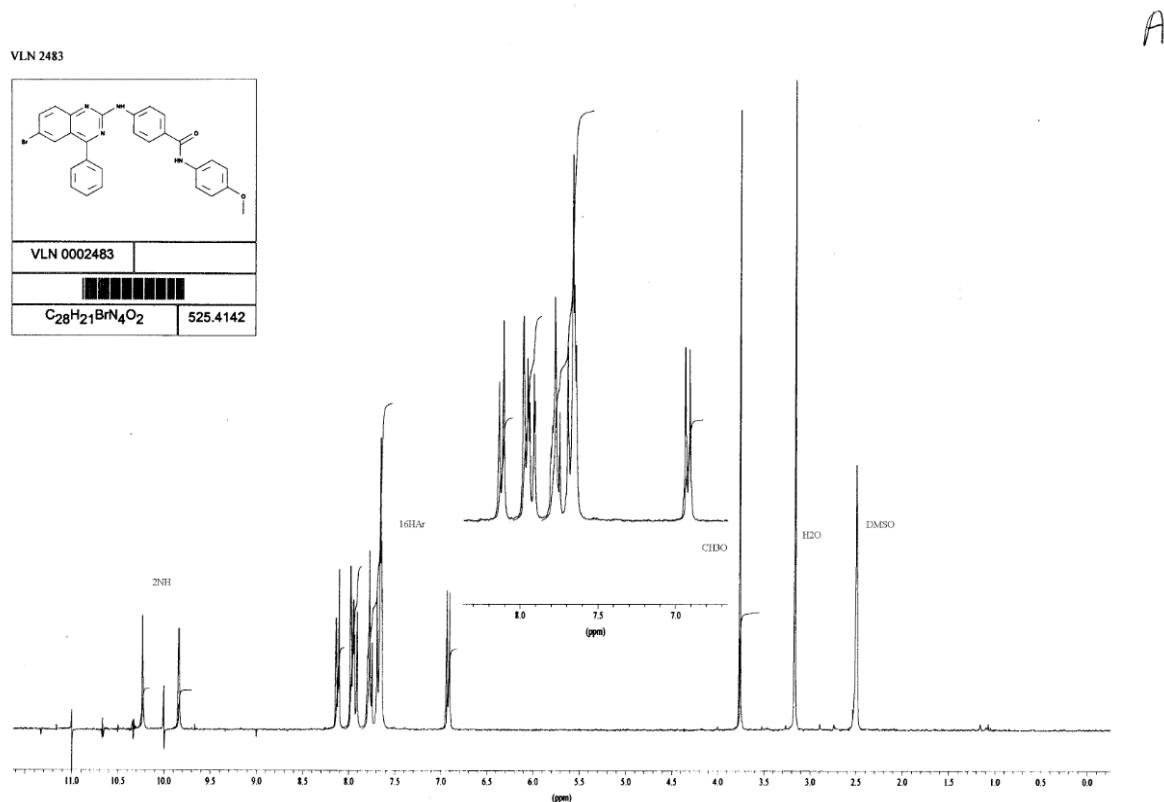
Printed: Wed Apr 14 13:04:05 2004

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Sample Report (continued):



Compound SC45, ^1H -NMR:



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