## **Supporting Information**



**Figure S1.** Example of plant (marked with red asterisk; ID 003) from the 6-week drought group at (A) 4 weeks and (B) 5 weeks after the start of the experimental drought. Photosynthesis and chlorophyll fluorescence measurements were not measured for droughted-plants in the final two weeks (weeks 5 and 6) of the experiment because they no longer had healthy leaves.



**Figure S2.** Comparison of stem water potential (MPa) measured by each ICT psychrometer (n=4) and a pressure chamber in the afternoon of August 25, 2020 on the day before the start of the experimental drought. This comparison was conducted to ensure that the psychrometer measurements were giving valid stem water potentials. The pressure chamber was used to measure stem water potentials prior to the start of the experimental drought (see Fig. S6), and the psychrometers were used to monitor stem water potentials throughout the experimental drought (see Figs. 2, S7).



**Figure S3**. Iodine-stained starch (%) in (A) xylem ray parenchyma, (B) phloem ray parenchyma, and (C) whole rays at two stem locations, 10% (left column) and 50% (right column) of the total plant length above the soil surface following sequential harvesting. Error bars denote  $\pm 1$  SD of the mean. Statistical results displayed above plots are from two-way ANOVA testing (treatment x stem location). Data are provided in Table S1. Since starch did not differ by stem location, data were averaged across stem locations for visualization and analysis in Fig. 3.



**Figure S4**. MicroCT images of stems from the well-watered control group (A-C), the 2-week drought group (D-F), the 4-week drought group (G-I), and the 6-week drought group (J-L). Ray parenchyma are full of starch granules when appearing bright gray in color, and depleted when appearing dark gray or black in color. All stems were collected at 10% above the soil. Plant IDs are listed in white text in the lower left corner of each image. Scale bar = 1 mm.



**Figure S5.** Fluorescence intensity in the (A) xylem and (B) phloem of stems across drought treatment groups. Fluorescence intensity was quantified using the integrated density measurement in FIJI on FDA-stained images of the stem at 10% and 50% of the total plant length above the soil surface. Error bars denote  $\pm$  SE of the mean. Statistical results displayed above plots are from one-way nested ANOVA testing with stem location treated as technical replicates per plant (treatment:stem) and lowercase letters within the plots indicate whether treatments significantly differed from each other based on Tukey's honest significant difference (HSD) at  $\alpha = 0.05$ 



**Figure S6.** Weekly stem water potential (MPa) measured using a Scholander pressure chamber for each individual plant prior to the start of the experimental drought. Black points represent individual plants (n=12), while red points represent the mean. Three weeks prior to the start of the experimental drought, water potentials were measured on August 7, 2020 between approximately 16:00-16:30, whereas water potentials were measured earlier between approximately 13:30-14:30 in the two weeks that followed (on August 14, 2020 and August 21, 2020).



(A)



**Figure S7**. Hourly stem water potential (in MPa) measured using ICT stem psychrometers for the (A) well-watered control, (B) 2-week drought group, (C) 4-week drought, and (D) 6-week drought groups.

(C)



**Figure S8.** Visualization of (A) starch content and (C) metabolically active cells in the stem of each plant by treatment following drought. Iodine-stained starch was visualized with compound light microscopy and then iodine-stained starch was identified in the xylem (orange) and phloem (pink) ray parenchyma via thresholding in ImageJ in (B). Quantification of this starch thresholding as percent starch for all plants is displayed in Fig. 3. Additionally, fluorescent living cells following FDA staining were visualized with fluorescent microscopy in (C) and autofluorescence was visualized with water in (D). Black box indicates missing image. Images shown are for stem samples at 10% above the soil, but data from 50% above the soil was also collected. Scale bar = 0.5 mm

**Table S1** Percent of starch in the xylem ray parenchyma, phloem ray parenchyma, and whole rays quantified using thresholding in ImageJ as well as starch concentration from enzymatic digestion for each individual plant. Percent starch was quantified at two stem locations, 10% and 50% above the soil. Enzymatically-derived starch concentrations were quantified at 10% above the soil.

plant ID	stem location	treatment	starch in	starch in	starch in	starch
_			xylem rays	phloem rays	whole rays	concentration
			(%)	(%)	(%)	$(mg g^{-1})$
003	50	6-week drought	95.4	4.1	72.5	NA
003	10	6-week drought	95.0	4.7	67.0	8.6
005	50	6-week drought	60.7	0.0	52.0	NA
005	10	6-week drought	75.3	0.0	54.8	5.1
004	50	6-week drought	10.9	0.0	9.1	NA
004	10	6-week drought	77.4	0.0	66.4	0
006	50	4-week drought	68.2	39.2	62.5	NA
006	10	4-week drought	74.9	35.6	66.4	18.8
008	50	4-week drought	84.4	51.9	77.2	NA
008	10	4-week drought	65.0	24.5	57.6	17.1
009	50	4-week drought	79.7	54.1	76.2	NA
009	10	4-week drought	67.8	53.5	65.1	19.4
001	50	2-week drought	66.2	29.2	60.7	NA
001	10	2-week drought	85.1	60.9	80.9	13.7
0010	50	2-week drought	86.4	40.6	75.0	NA
0010	10	2-week drought	89.8	77.8	87.7	19.8
0012	50	2-week drought	89.3	45.5	80.1	NA
0012	10	2-week drought	85.0	26.0	70.8	20.7
002	50	control	95.8	75.5	92.7	NA
002	10	control	97.3	91.0	96.0	21.9
0011	50	control	98.7	98.7	98.7	NA
0011	10	control	97.8	95.5	97.3	30
007	50	control	96.0	91.8	94.8	NA
007	10	control	96.3	87.8	94.4	38.5