nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

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St	at:	ıct	

n/a	Confirmed
	$oxed{x}$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🗴 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
x	A description of all covariates tested
	🗷 A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
x	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
	$oxed{x}$ For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
x	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	Our web collection an statistics for biologists contains articles an many of the points above

Software and code

Policy information about availability of computer code

Data collection

no software was used for data collection

Data analysis

For genome assembly: Flye v2.9-b1178, Porechop v0.2.4, Guppy, Nextpolish v1.3.1, Juicer v.1.5, 3d-dna v180922. For alignment: lastz v1.1.13, BLASTN 2.6.0. For SNP calls: BWA-memv0.7.17-r1188, GATK 4.0.12.0. For GWAS: Plink v1.90b4. For alignments and phylogenies: CLUSTALW, MEGA11. For plotting: R 4.2.1, last-dotplot Build 1333

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio <u>guidelines for submitting code & software</u> for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our <u>policy</u>

The Oxford Nanopore (ONT) reads generated in this study are deposited in the NCBI BioProject under accession number PRJNA1010806 [https://

www.ncbi.nlm.nih.gov/bioproject/PRJNA1010806]. The Illumina and Hi-C interaction mapping reads generated in this study for genome assembly polishing are deposited in the NCBI BioProject under accession number PRJNA1010212 [https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1010212]. The population sequence capture array Illumina reads generated in this study have been deposited in the NCBI BioProject under accession number PRJNA1009225 [https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1009225]. The genome assembly of male and female Salix exigua have been deposited in the NCBI BioProject under accession number PRJNA1009227 [https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1009230 [https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1009230]

Research involving humar	participants, their	r data, or bio	ological materia	ıl
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and sexual orientation and ra	ace, ethnicity and racism.
Reporting on sex and gender	n/a
Reporting on race, ethnicity, o other socially relevant grouping	
Population characteristics	n/a
Recruitment	n/a
Ethics oversight	n/a
Note that full information on the	e approval of the study protocol must also be provided in the manuscript.
Field-specific	reporting
Please select the one below t	that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.
Life sciences	Behavioural & social sciences Ecological, evolutionary & environmental sciences
For a reference copy of the documen	nt with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf
Ecological, ev	olutionary & environmental sciences study design
	these points even when the disclosure is negative.
Study description	We mapped sex determination in Salix exigua and analyzed the movement of sex chromosomes in the genus.
ŗ	We sampled 48 Salix exigua individuals - 24 males and 24 females - for mapping sex determination regions in the Rio Bonito copulation. This sample size has been shown sufficient for mapping SDRs in many studies. We chose to sample S. exigua because it represents a previously unsampled taxonomic section of Saix. The population sample is meant to represent sex chromosomes in the species,
s t r	Plants were marked in the field and sampled arbitrarily along a linear transect following the bank of the Rio Bonito. Arbitrary sampling was reasonable because we did not know the genotypes of individuals other than by knowing their sexes. To avoid sampling the same clone, we collected from trees that were separated by >10m or were obviously different individuals because one was a male and other other was a female. We sampled 24 males and 24 females. This sample size had been shown in previous studies (Sanderson et al. 2021 Heredity) and in independent studies (Muyle et al. 2016 GBE) to be sufficiently large for detecting sex determination regions with high power (0.8).
Data collection	Field data were recorded on the sexes of individuals in a field notebook by Nan Hu and Haley Hale.
	eaf samples from 48 individuals were collected for sequencing in early April 2017. All samples were collected along a ~2 km transect. Samples for genome assemblies were collected from the same population on October 20, 2018.
Data exclusions	no data were excluded
	The study question of identifying the sex chromosome does not require replicated studies. These GWAS studies are not usually reproduced.
	Genotypes were assumed to be randomly distributed among the collected samples. A posteriori analyses of genetic diversity supported this assumption.
Blinding	Not applicable. No studies with manipulated treatments that would require blinding ere preformed for this research.
Did the study involve field	work? 🗶 Yes No

Field work, collection and transport

Field conditions	We collected during one day. It was sunny.
Location	33º31'39.33"N 150º26'21.05"W on the Rio Bonito on Fort Stanton-Snowy River Cave National Conservation Area, Bureau of Land Management USA
Access & import/export	In 2017, Haley Hale obtained permission for sampling from Michael McGee at the the main office at the Fort Stanton-Snowy River Cave National Conservation Area, Bureau of Land Management (BLM) USA. No samples were imported or exported across national borders. The BLM land where leaf tissues were collected has no restrictions on plant collecting.
Disturbance	We did not disturb the area.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems		Methods		
n/a	Involved in the study	n/a	Involved in the study	
X	Antibodies	×	ChIP-seq	
x	Eukaryotic cell lines	×	Flow cytometry	
x	Palaeontology and archaeology	x	MRI-based neuroimaging	
x	Animals and other organisms			
×	Clinical data			
×	Dual use research of concern			
	x Plants			

Dual use research of concern

Alter the host range of a pathogen

▼ Enable evasion of diagnostic/detection modalities

Enable the weaponization of a biological agent or toxin

Any other potentially harmful combination of experiments and agents

Policy information about <u>dual use research of concern</u>

Hazards

Could the accidental, deliberate or reckless misuse of agents or technologies generated in the work, or the application of information presented in the manuscript, pose a threat to:
No Yes
Public health
■ National security
Crops and/or livestock
Ecosystems Ecosystems
Any other significant area
experiments of concern
Does the work involve any of these experiments of concern:
No Yes
Demonstrate how to render a vaccine ineffective
Confer resistance to therapeutically useful antibiotics or antiviral agents
Enhance the virulence of a pathogen or render a nonpathogen virulent
Increase transmissibility of a pathogen