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Supplemental information

***Silc1* long noncoding RNA is an immediate-early
gene promoting efficient memory formation**

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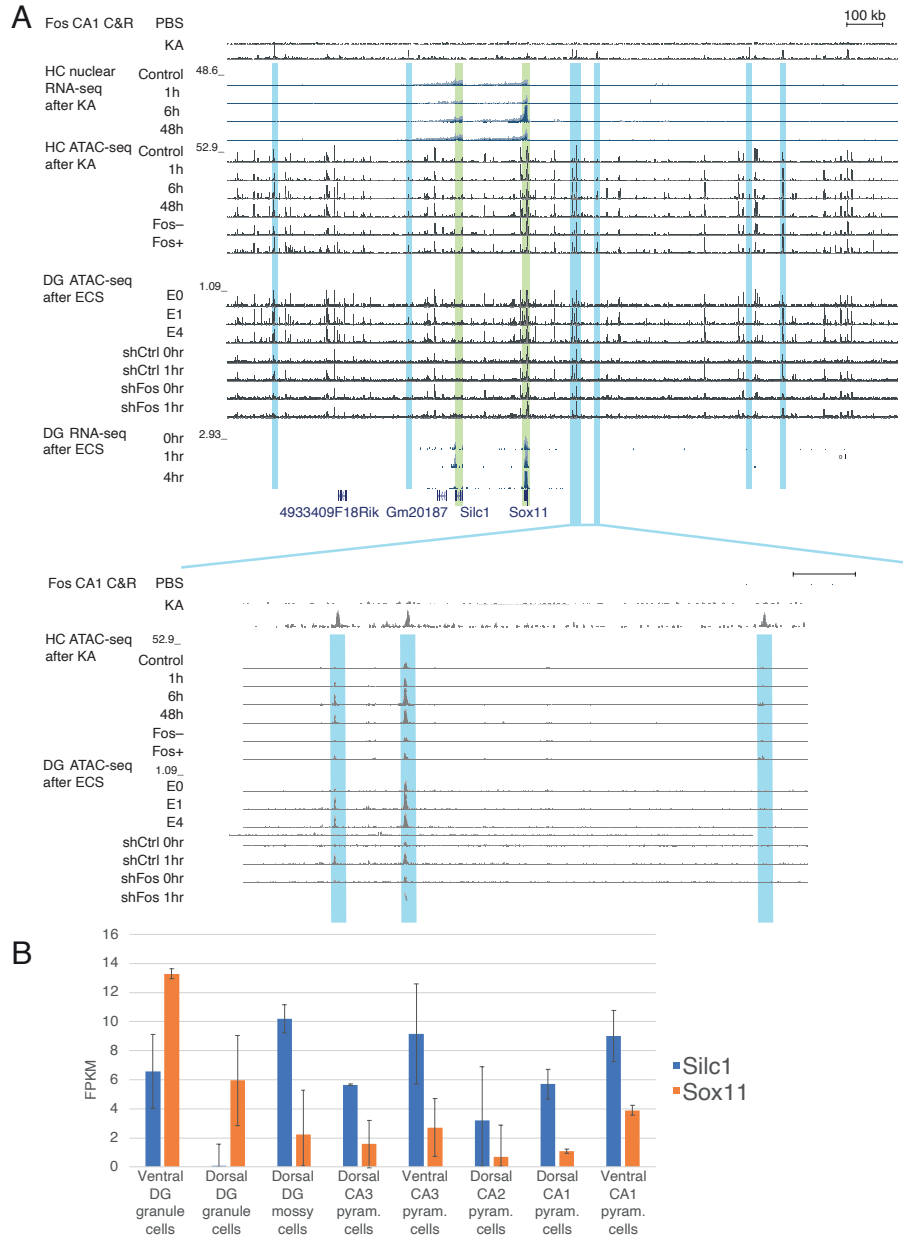


Figure S1. Transcriptomic and epigenomic characterization of the broad Sox11 domain. Related to Figure 1.

(A) Top: broad ~2Mb region flanking Sox11; Bottom: zoom-in on the indicated region. Shown are (top to bottom): Fos Cut&Run data in the hippocampus (HC) CA1 region ³⁷; HC nuclear RNA-seq data at the indicated time after KA treatment ³⁷; ATAC-seq data from the same study; including from sorted Fos-positive and Fos-negative cells; ATAC-seq data from the DG after ECS ³⁷, including a time course and the Fos knockdown experiment; RNA-seq data from the same study. Fos-bound and apparently Fos-regulated regions are shaded. **(B)** Expression of Sox11 and Silc1 in data of sorted populations from the HippoSeq dataset (n=3).

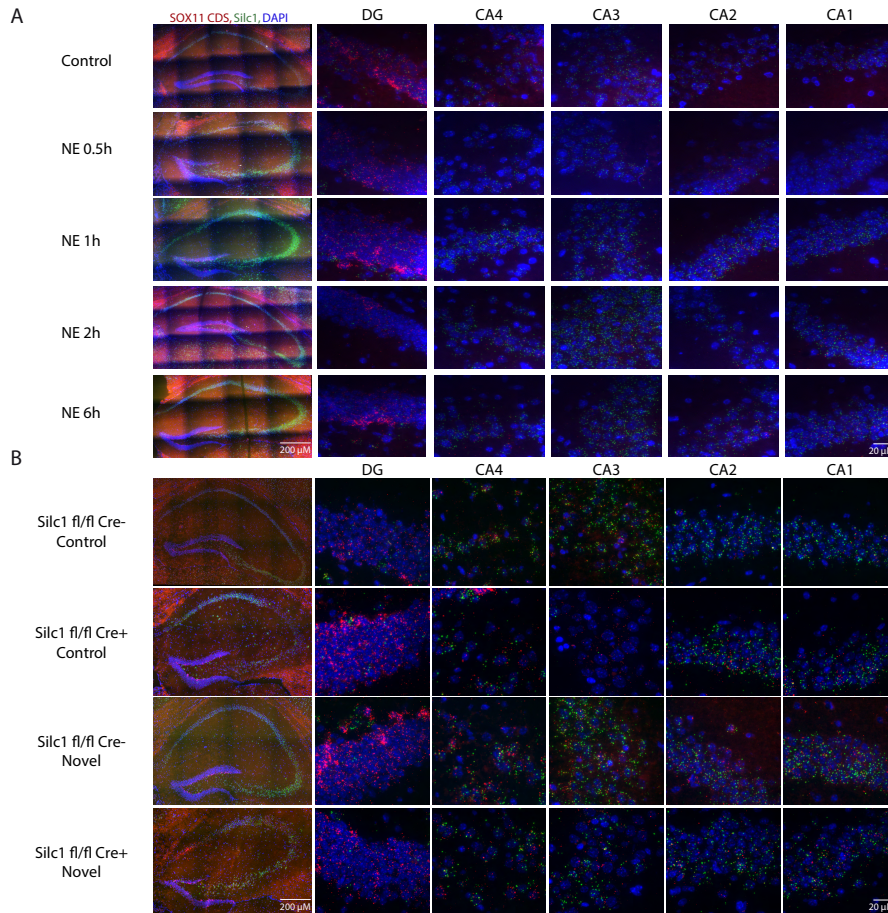


Figure S2. Expression of *Silc1* and *Sox11* at several time points following novel environment (NE) conditions in *Silc1* conditional knockout mice. Related to Figure 2.

(A) RNAscope FISH assay on hippocampal sections from WT mice. Tissues were hybridized with *Silc1* (green) and *Sox11* CDS (red) probes and counterstained with DAPI (blue), and imaged using 20X (Scale bar 200 μm) and 100X oil-immersion objectives (Scale bar 20 μm). Novel environment (NE) performed using the Barnes maze setting and the hippocampus was extracted for coronal sections 0.5, 1, 2 and 6 hr upon stimulus. **(B)** As in A for hippocampus sections from *Silc1*^{fl/fl} mice that were stereotaxic injected in the CA3 region using AAV9 Cre-GFP or AAV9 GFP. Two weeks after injections the mice were exposed to Novel environment (NE) and the hippocampus was extracted for coronal sections after 1 hr of NE.

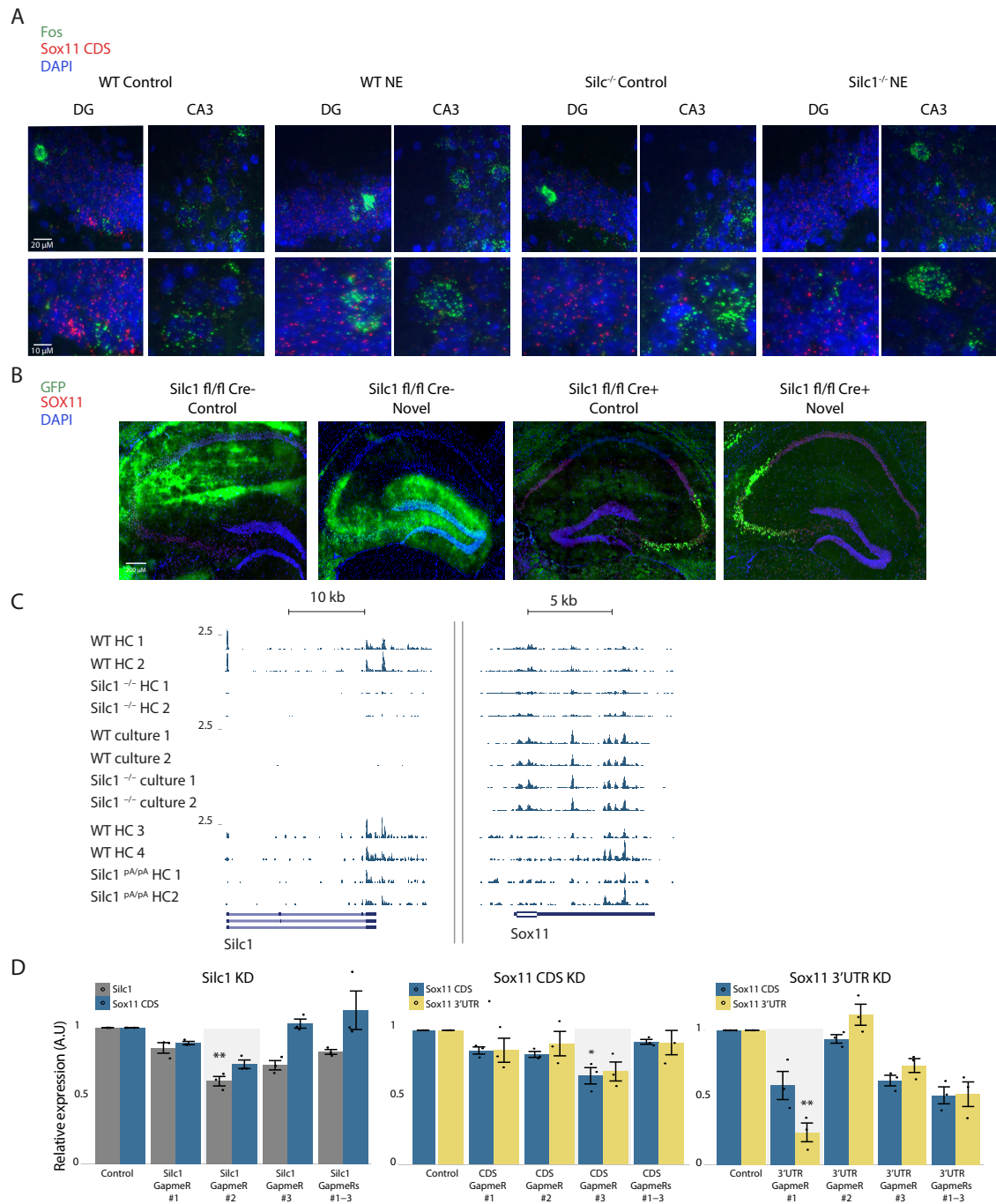


Figure S3. Characterization of gene expression changes in different mouse models. Related to Figure 3.

(A) RNAscope Fluorescent *in situ* hybridization (FISH) assay on WT and *Silc1*^{-/-} mice hippocampal sections from HC and NE conditions. Tissues were hybridized with Fos mRNA (green) and Sox11 CDS (red) probes and counterstained with DAPI (blue), and imaged using 100X oil-immersion objectives (Scale bar 20 μ m). **(B)** Immunostaining with anti-SOX11 (red) and DAPI (blue) in hippocampi of *Silc1*^{fl/fl} mice that were stereotaxically

injected in the CA3 region using AAV9 Cre-GFP or AAV9 GFP. The GFP signal marks the site of injection. Imaging using 20X objective (Scale bar 200 μm). (C) RNA-seq read coverage at the *Silc1* (left) and *Sox11* (right) genomic regions, in the adult hippocampus (HC) or in cultured hippocampal neurons ("culture") from the indicated genetic background. (D) Changes in expression of the indicated genes and regions upon the use of the indicated GapmeRs targeting *Silc1* (left) *Sox11* CDS (middle) and *Sox11* 3'UTR (right), as evaluated by qRT-PCR. Levels were normalized to control GapmeR and β -actin for internal control. The GapmeRs selected for further experiments are shaded in gray. 3 biological repeats. Mean \pm SEM is shown. P value calculated using unpaired two-sample t-test, * P < 0.05, ** P < 0.005.

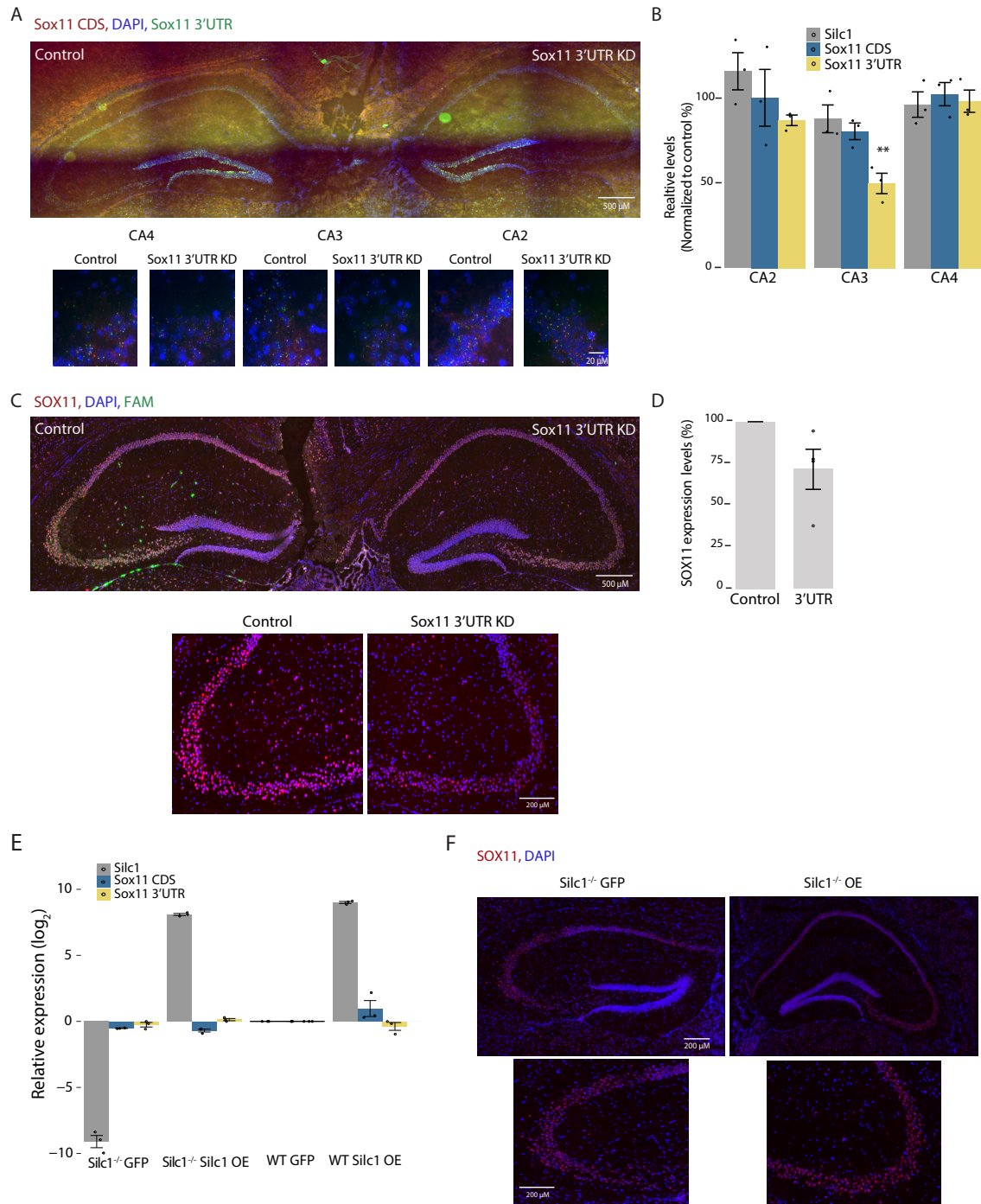


Figure S4. Expression of Sox11 after Sox11 3' UTR KD by injection of GapmeRs into the CA3 region and *Silc1* over-expression in *Silc1*^{-/-} mice. Related to Figure 4.

(A) Control-FAM (left) or Sox11 3' UTR (right) GapmeRs were injected into the CA3 region. 5 days later, the hippocampus was extracted for coronal sections. RNAscope analysis of Sox11 expression using Sox11 3' UTR(green), Sox11 CDS (red) probes, and DAPI. Imaging was done using 20X (Scale bar 200 μ m) and 100X oil-immersion objectives (Scale bar 20 μ m). **(B)** RNAscope quantification of the number of green and

red dots, normalized to control GapmeR, performed using IMARIS software. 12 images of non-overlapping fields per biological repeat were quantified; 3 biological repeats. Mean \pm SEM is shown. P value calculated using an unpaired two-sample t-test, * P < 0.05. **(C)** Immunostaining with anti-SOX11 (red) and DAPI (blue) in hippocampi of *Sox11* 3' UTR KD mice. FAM signal marks the injection site of the control GapmeR. Imaging using 20X objective (Scale bar 200 μ m). **(D)** Quantification of 3 biological repeats of hippocampal staining, normalized to the injection of control GapmeR. Mean \pm SEM. **(E)** qRT-PCR quantifications of *Silc1* and *Sox11* after injection of AAV9-Silc1 or AAV9-GFP into the CA3 region of WT and *Silc1*^{-/-} mice. Levels were normalized to WT mice injected with AAV9-GFP, and β actin was used as an internal control. **(F)** Immunostaining with anti-SOX11 (red) and DAPI (blue) in hippocampi of *Silc1*^{-/-} mice injected with Silc1 OE AAV. Imaging using 20X objective (Scale bar 200 μ m).

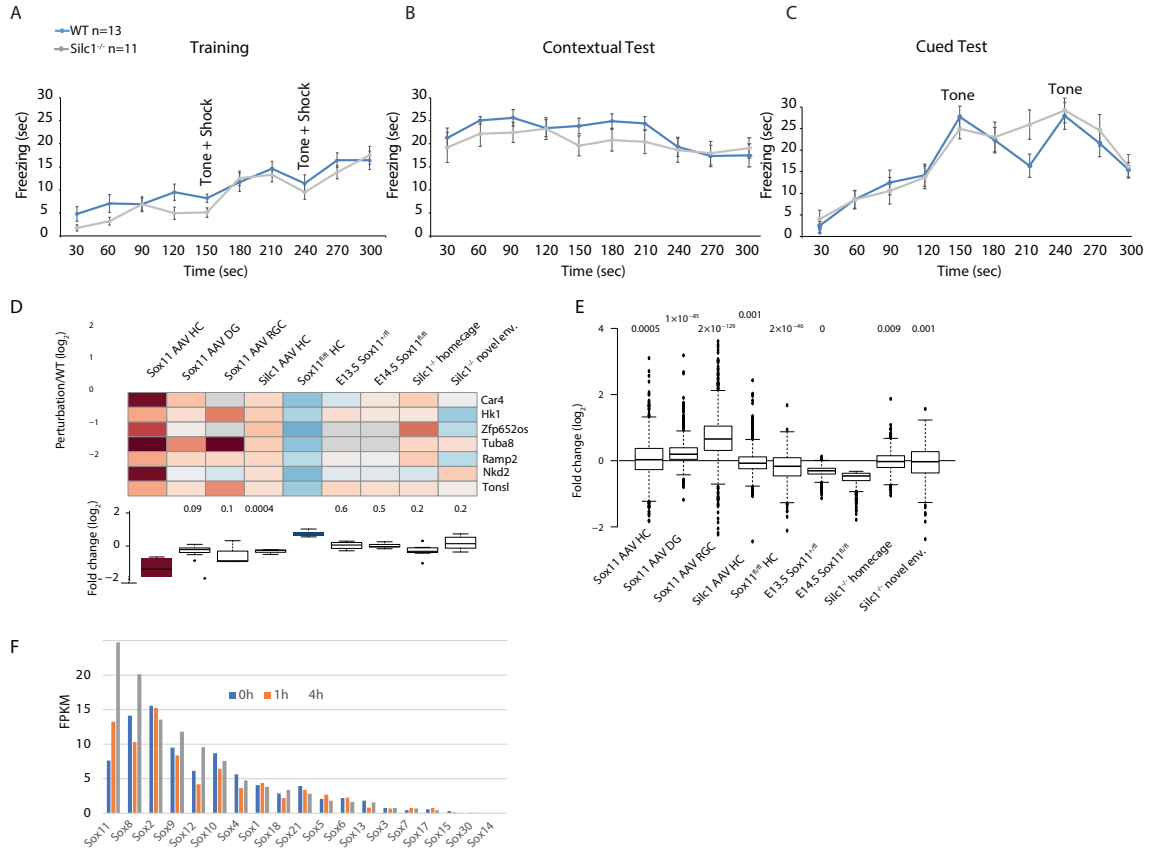


Figure S5. *Silc1*^{-/-} mice showed no alteration in freezing responses during fear conditioning and subsequent recall tests and characterization of genes regulated by *Sox11* and *Silc1* by RNA-seq. Related to Figures 5 and 6.

(A-C) Freezing behavior of *Silc1*^{-/-} (13) mice compared to WT (13) littermates over Fear Conditioning **(A)**, Context Test **(B)** and Cue Test **(C)**. Data represent mean +/- SEM (error bars). Two-way ANOVA, for Gene (Between-Subjects), Time (30 sec intervals; Within-Subjects with repeated measures), and their interaction (Gene × Time) indicated no difference between the genotypes in this type of learning and its recall [Gene (main effect): Conditioning- $F_{(1,24)}=2.294$; $p=0.143$. Context- $F_{(1,22)}=0.839$; $p=0.370$. Cue- $F_{(1,22)}=0.210$; $p=0.839$]. **(D)** Genes negatively regulated by *Sox11* in the hippocampus. As in **Fig. 6**, for the seven genes negatively regulated by *Sox11*. **(E)** Distribution of the changes in gene expression for the comparisons shown in Fig. 7 for the genes that are significantly reduced in the E13.5 *Sox11*^{fl/fl} Cre⁺ dentate neuroepithelium. P-values for each group obtained using Wilcoxon rank-sum test are shown above each boxplot. **(F)** As in Fig. 1C, for the different genes in the *Sox* family of TFs. The RNA-seq data are from the mouse dentate gyrus and the indicated time after ECS.

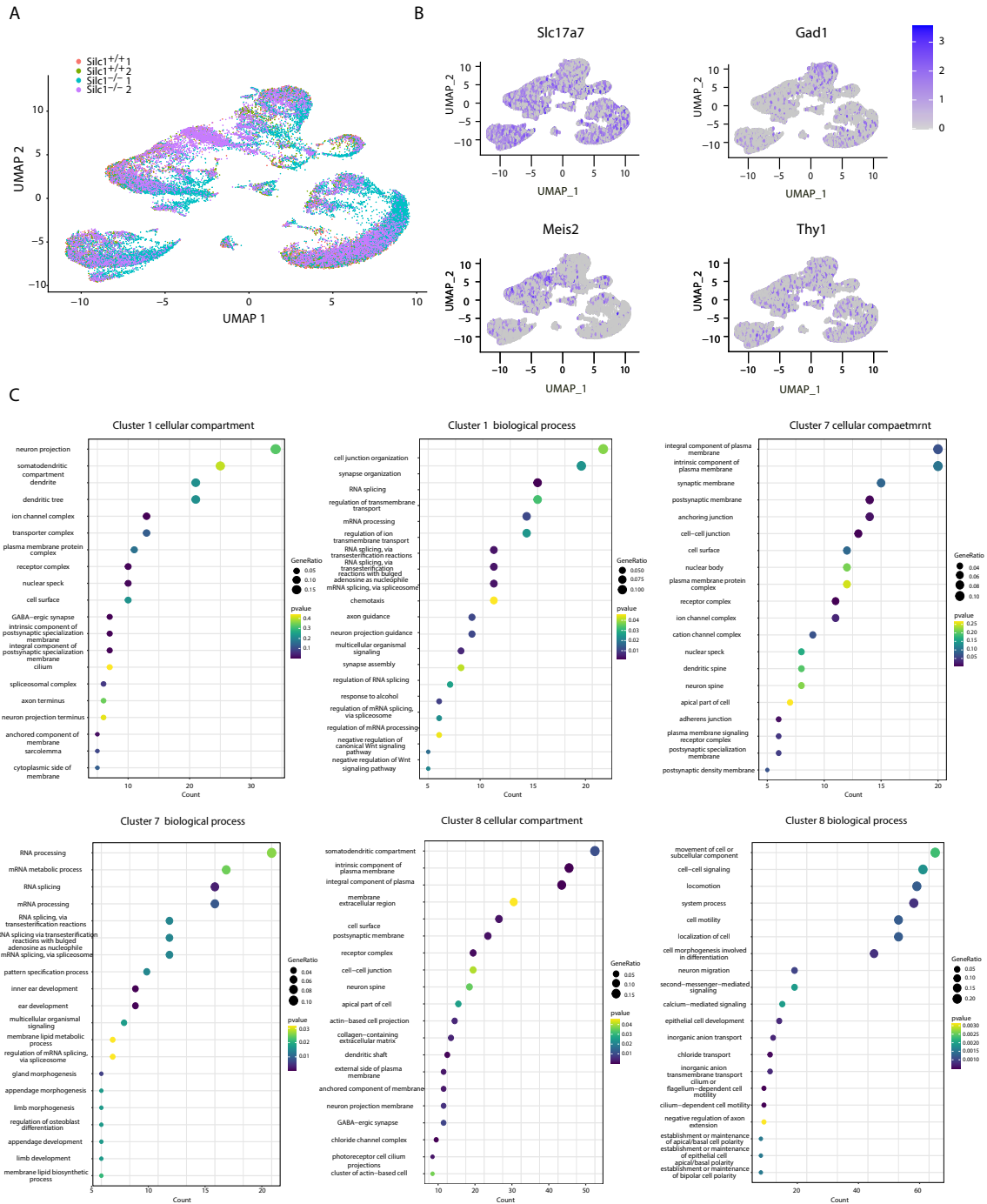


Figure S6. Characterization of genes regulated by *Sox11* and *Silc1* by RNA-seq. Related to Figure 6.

(A) UMAP visualization of the single-nucleus RNA-seq expression data, color-coding the sample from which each cell originated. **(B)** Expression levels of the indicated genes projected onto the UMAP visualization. **(C)** GO cellular compartment and biological process terms enriched in the genes significantly down-regulated in *Silc1*^{-/-} cells compared to WT cells in the indicated clusters.

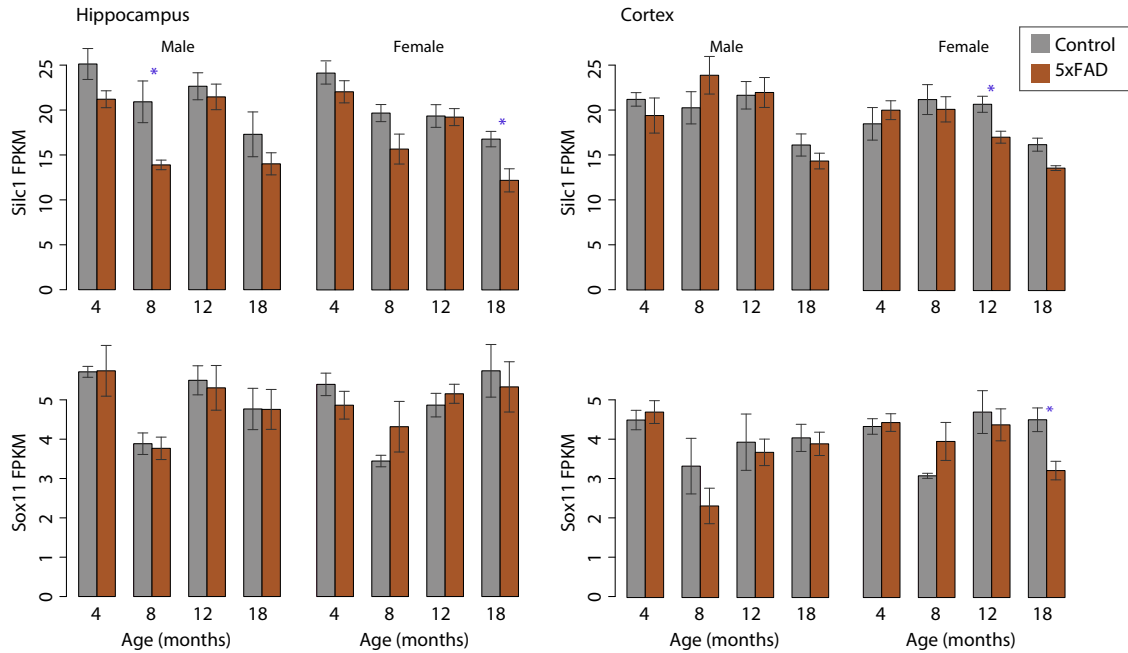


Figure S7. Changes in expression *Silc1* and *Sox11* during aging and in a model of Alzheimer's disease. Related to Discussion.

Expression of *Silc1* (top) and *Sox11* (bottom) in the indicated brain region extracted from control WT mice and from 5xFAD mice at the indicated age, data from⁶⁶. Blue asterisks denote $P < 0.05$ for the comparison between Control and 5xFAD mice, two-sided Wilcoxon rank-sum test.