# nature portfolio

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## **Reporting Summary**

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

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For	all s	tatistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Со	nfirmed
	X	The exact sample size $(n)$ for each experimental group/condition, given as a discrete number and unit of measurement
$\boxtimes$		A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
		The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
$\times$		A description of all covariates tested
$\boxtimes$		A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
		A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
		For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
$\boxtimes$		For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
$\boxtimes$		For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
$\boxtimes$		Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated
		Our web collection on statistics for biologists contains articles on many of the points above

#### Software and code

Policy information about availability of computer code

Data collection

Canu was used to assembly long reads for genome assembly; LTR FINDER (version 1.05), MITE-Hunter (20100819), RepeatScout (version 1.0.5) and PILER-DF (version 2.4) were utilized to generate repeat libraries based on structure-based method and de novo prediction; PASTEClassifier (version 1.0) was applied for repeat classification, then merged with all repeats from the Repbase Database (version 19.06); RepeatMasker was used to predict fungal isolate repeats; EVM was used to integrate all predicted results; Augustus, GlimmerHMM, and SNAP (version 2006-07-28) to scan the repeat-masked genome; PASA were used to predict genes; the 6mA and 4mC base modifications were identified using PacBio SMRT Analysis 2.3.0 with default settings; The program Nucmer in Mummer was used to compare two genomes.

Data analysis

No commercial, open source and custom code used to analyse the data in this study were used.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

#### Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

All supporting data from this study are available in the article and supplementary information files, or from the corresponding author upon reasonable request. The raw V991 and 1cd3-2 genome sequencing in this study have been deposited in the NCBI Bioproject database under accession code PRJNA510201. The V991 and 1cd3-2 genome assembly data generated in this study have been deposited in the Genome Warehouse in National Genomics Data Center, Beijing Institute of Genomics, Chinese Academy of Sciences / China National Center for Bioinformation, under accession number PRJCA020793 and PRJCA020800. The raw RNA-seq data generated in this paper have been deposited in the Genome Sequence Archive (Genomics, Proteomics & Bioinformatics 2021) in the National Genomics Data Center (Nucleic Acids Res 2022), China National Center for Bioinformation/Beijing Institute of Genomics, Chinese Academy of Sciences (GSA: CRA008562), which are publicly accessible.

Policy information about studies with human participants or human data. See also policy information about sex, gender (identity/presentation),

### Research involving human participants, their data, or biological material

and sexual orientatio	<u>n</u> and <u>race, ethnicity and r</u>	<u>racism</u> .		
Reporting on sex a	nd gender N/A	N/A		
Reporting on race, other socially relev groupings	**	r N/A		
Population charact	eristics N/A	N/A		
Recruitment	N/A	N/A		
Ethics oversight	N/A	N/A		
Note that full information	on on the approval of the stud	dy protocol must also be provided in the manuscript.		
Field-spec	ific reportin	ng		
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All studies must discl	ose on these points even v	when the disclosure is negative.		
(	35.7Mb vs. 34.8Mb) (Chen et 8-95 fold (V991 3.8 Mb and 1	nced a 20 kb library to high depth (> 100X) using PacBio RS II . We found the determined genome size was almost 1 Mb larger s. 34.8Mb) (Chen et al., 2017). Scaffold number sharply decreased ca. 13 fold (13 vs. 167), and contig L50 sharply increased by (V991 3.8 Mb and 1cd3-2 3.5 Mb, vs VdLs.17 0.04Mb) . By optimal homologue comparison with chromosomes of VdLs.17 (carried no et al., 2015), we finally determined the sequence of 8 chromosomes of V991 and 1cd3-2, constructed from 13 and 19 contigs.		
Data exclusions	lo data was excluded from th	cluded from the analysis.		
Replication	hese samples were selected	samples were selected for DNA sequencing and do not require replication.		
Randomization [1	These samples were selected for DNA sequencing and do not require Randomization.			
Blinding	These samples were selected for DNA sequencing and do not require Blinding.			

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems	Methods
n/a Involved in the study	n/a Involved in the study
Antibodies	ChIP-seq
Eukaryotic cell lines	Flow cytometry
Palaeontology and archaeology	MRI-based neuroimaging
Animals and other organisms	
Clinical data	
Dual use research of concern	
Plants	
Dual use research of concern	
Policy information about <u>dual use research of concer</u>	<u>n</u>
Hazards	
Could the accidental, deliberate or reckless misuse in the manuscript, pose a threat to:	of agents or technologies generated in the work, or the application of information presented
No   Yes	
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National security	
Crops and/or livestock	
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Any other significant area	
Experiments of concern	
Does the work involve any of these experiments of	concern:
No Yes	
Demonstrate how to render a vaccine ineffective	re
Confer resistance to therapeutically useful antik	piotics or antiviral agents
Enhance the virulence of a pathogen or render	a nonpathogen virulent
Increase transmissibility of a pathogen	
Alter the host range of a pathogen	
Enable evasion of diagnostic/detection modaliti	es
Enable the weaponization of a biological agent	
Any other potentially harmful combination of e	xperiments and agents