



Supplemental Figure S1. Elimination of gut microbiota by broad-spectrum antibiotics (ABX) prevents the manifestation of crypt base AB-PAS<sup>+</sup> cells in *Phb1* deficient mice. (A) Bacterial elimination by ABX shown by reads per sample by 16S rRNA sequencing. (B) AB-PAS staining of ileum. Bar = 100  $\mu$ m. Dashed line denotes crypt base. (C) Number of AB-PAS<sup>+</sup> cells/crypt base across 50 crypts. AB-PAS, alcian blue-periodic acid Schiff. *n* = 7-13 each group. \*\*\*\**P* < 0.0001.

## Co-housed across genotype



Supplemental Figure S2. Co-housed *Phb1*<sup> $\Delta PC$ </sup> mice exhibit similar ileal microbiota composition as *Phb1*<sup>fl/fl</sup> littermates. 16S RNA sequencing of luminal ileal content of co-housed *Phb1*<sup>fl/fl</sup></sup> and*Phb1* $<sup><math>\Delta PC$ </sup> mice. (A) Alpha diversity measure by Shannon index. (B) Weighted UniFrac PCoA plot of 16S rRNA gene sequences. (C-D) Relative abundance of bacteria at phylum level (C) in individual mice and (D) combined mice by genotype. (E-F) Relative abundance of bacteria at genus level (E) in individual mice and (F) combined mice by genotype. *n* = 9 each genotype.</sup>

## Co-housed across genotype



Supplemental Figure S3. Co-housed *Phb1*<sup>ΔPC</sup> mice manifest PC abnormalities and ileitis. (A) H&E staining showing Paneth cells (pink granules) in the ileal crypts. Bar = 50  $\mu$ m; boxed pullout bar = 200  $\mu$ m. (B) Immunofluorescent-staining for Iysozyme (red), muc2 (green), and DAPI (nucleus, blue) in ileal crypts (dashed line). Arrows denote Paneth cells with normal Iysozyme packaging into granules. Bar = 50  $\mu$ m. (C) Average number of Iysozyme<sup>+</sup> cells per crypt per mouse. A minimum of 50 crypts per mouse were quantitated. *n* = 13 *Phb1*<sup>ΔPC</sup> or 10 *Phb1*<sup>M/#</sup> littermates. (D) Paneth cell Iysozyme allocation patterns. A minimum of 50 crypts per mouse were quantitated. *n* = 13 *Phb1*<sup>ΔPC</sup> or 10 *Phb1*<sup>M/#</sup> littermates. (D) Paneth cell Iysozyme allocation patterns. A minimum of 50 crypts per mouse were quantitated. *n* = 13 *Phb1*<sup>ΔPC</sup> or 10 *Phb1*<sup>M/#</sup> littermates. (E) mRNA quantification in ileum by qRT-PCR. *n* = 14 each genotype. (F) AB-PAS staining of ileum and number of AB-PAS<sup>+</sup> cells/crypt base. Bar = 100  $\mu$ m. Dashed line denotes crypt base. *n* = 13 *Phb1*<sup>ΔPC</sup> or 10 *Phb1*<sup>M/#</sup> littermates. (G) Representative H&E-stained ileum and histological inflammation scoring of ileum. Bar = 100  $\mu$ m. *n* = 21 *Phb1*<sup>ΔPC</sup> or 10 *Phb1*<sup>M/#</sup> littermates. (H) mRNA quantification in ileum by qRT-PCR. *n* = 14 each genotype. AB-PAS, alcian blue-periodic acid Schiff; Ang4, angiogenin 4; H&E, hematoxylin & eosin; Ifng, interferon gamma; II1b, interleukin 1 beta; muc2, mucin2; Tnfa, tumor necrosis factor alpha. Results are presented as individual mice ± SD. \**P* < 0.001, \*\*\*\**P* < 0.0001.

Phb1

Phb1

Phb1

## Co-housed across genotype





Supplemental Figure S4. Phb1<sup>i \[] EC</sup> mice co-housed with Phb1<sup>fl/fl</sup> littermates exhibit Paneth cell defects and spontaneous ileitis. (A) H&E staining showing Paneth cells (pink granules) in the ileal crypts. Bar = 50 μm. (B) Immunofluorescent-staining for lysozyme (red), muc2 (green), and DAPI (nucleus, blue) in ileal crypts (dashed line). Arrows denote Paneth cells with normal lysozyme packaging into granules. Bar = 50  $\mu$ m. (C) Average number of lysozyme<sup>+</sup> cells per crypt per mouse. *n* = 14 *Phb1<sup>fl/fl</sup>* and 18 *Phb1<sup>ΔPC</sup>* mice. (D) Paneth cell lysozyme allocation patterns. n = 16 each genotype. (E) mRNA quantification in ileum by qRT-PCR. n = 15 Phb1<sup>##</sup> and 14 Phb1<sup>ΔPC</sup> mice. Outliers were identified by ROUT test (Q = 1%) and were removed from Phb1<sup>fl/fl</sup> Ang4 and Phb1<sup>ΔPC</sup> Cryptdin3, Cryptdin 5. (F) AB-PAS staining of ileum and number of AB-PAS<sup>+</sup> cells/crypt base. Bar = 100 µm. Dashed line denotes crypt base. n = 15 each genotype. (G) Histological inflammation scoring of ileum. Arrow denotes infiltrating immune cells. Bar = 100 µm. n = 15 each genotype. AB-PAS, calcian blue-periodic acid Schiff; Ang4, angiogenin 4; H&E, hematoxylin & eosin; muc2, mucin2. Results are presented as individual mice ± SD. \*\**P* < 0.01, \*\*\**P* < 0.001, \*\*\*\**P* < 0.0001.







С





В

D

Supplemental Figure S5. Butyrate supplementation prevents Paneth cell defects in *Phb1*-deficient mice. (A-B) Immunofluorescent-staining for lysozyme (red), muc2 (green), and DAPI (nucleus, blue) in ileal crypts (white outline) of *Phb1*<sup> $\Delta PC$ </sup> mice and *Phb1*<sup>fMI</sup> littermates (A) or *Phb1*<sup> $i\Delta IEC</sup></sup> mice and$ *Phb1*<sup><math>fMI</sup> littermates (B). Arrows denote Paneth cells with normal Lysozyme packaging into granules. Asterisk denotes Muc2 colocalization with Lysozyme. Bar = 50 µm. (C-D) AB-PAS staining of ileum. Bar = 100 µm. Dashed line denotes crypt base. AB-PAS, alcian blue-periodic acid Schiff; muc2, mucin2; Veh, vehicle</sup>





Supplemental Figure S6. Butyrate protects Phb1-deficient ileal enteroids from Phb1<sup>i/JEC</sup> microbiota-induced death. (A) Western blots for total and cleaved Caspase 3 indicating apoptosis. (B) Oxygen consumption relative to Phb1<sup>#/#</sup> vehicle enteroids. As a positive control, enteroids were treated with 0.1 µM Antimycin A 15 min prior to oxygen measurements. n = 3 for all treatments. (C) Mitochondrial superoxide level as measured by MitoSOX fluorescence intensity. n = 3 for all treatments, results are representative of 2 separate experiments. C: Phb1<sup>fl/fl</sup> microbiota filtrate, KO: Phb1<sup>i\triangleC</sup> microbiota filtrate. n = 3 for all treatments. AntiA, Antimycin A. Results are presented as individual mice  $\pm$  SD. \**P* < 0.05, \*\**P* < 0.01, \*\*\**P* < 0.005, \*\*\*\**P* < 0.0001.