



Vitamin B₅ supports MYC oncogenic metabolism and tumor progression in breast cancer

In the format provided by the authors and unedited

Supplementary Table 2. PA control diet (TestDiet 57W5)

INGREDIENTS (%)

Corn Starch	39.7486
Casein - Vitamin Tested	20.0000
Maltodextrin	13.2000
Sucrose	10.0000
Soybean Oil	7.0000
Powdered Cellulose	5.0000
AIN 93G Mineral Mix	3.5000
AIN 93 Vitamin Mix	1.0000
L-Cystine	0.3000
Choline Bitartrate	0.2500
t-Butylhydroquinone	0.0014

NUTRITIONAL PROFILE

Protein, % **18.3**

Arginine, %	0.70
Histidine, %	0.52
Isoleucine, %	0.96
Leucine, %	1.73
Lysine, %	1.45
Methionine, %	0.52
Cystine, %	0.37
Phenylalanine, %	0.96
Tyrosine, %	1.01
Threonine, %	0.77
Tryptophan, %	0.22
Valine, %	1.14
Alanine, %	0.55
Aspartic Acid, %	1.29
Glutamic Acid, %	4.08
Glycine, %	0.39
Proline, %	2.36
Serine, %	1.10
Taurine, %	0.00

Fat, % **7.1**

Cholesterol, ppm	0.00
Linoleic Acid, %	3.58
Linolenic Acid, %	0.55
Arachidonic Acid, %	0.00
Omega-3 Fatty Acids, %	0.55
Total Saturated Fatty A	1.05
Total Monounsaturated Fatty Acids, %	1.54
Polyunsaturated Fatty Acids, %	3.78

Fiber (max), % **5.0**

Carbohydrates, % **63.2**

Minerals

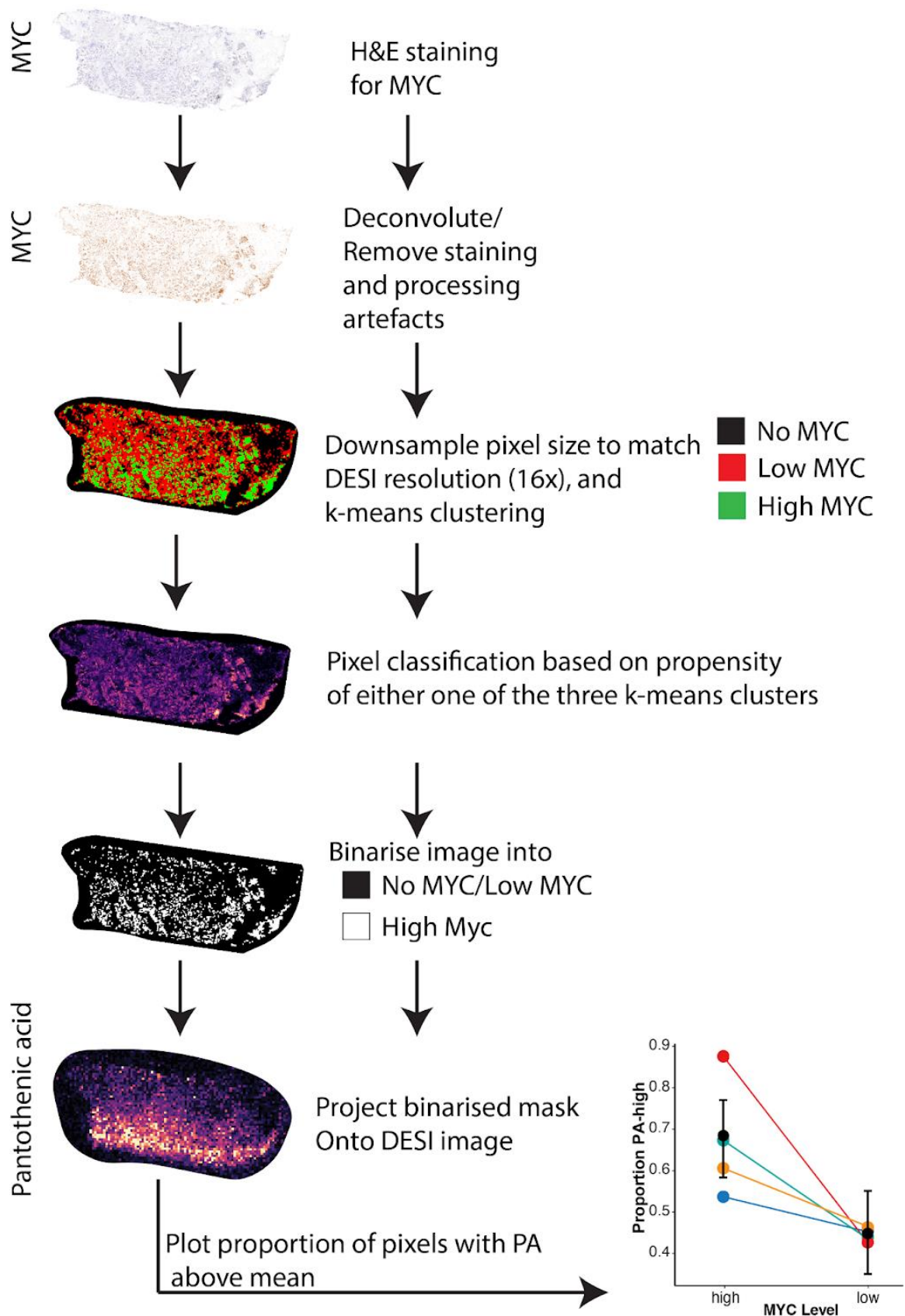
Calcium, %	0.51
Phosphorus, %	0.32
Potassium, %	0.36
Magnesium, %	0.05
Sodium, %	0.13
Chloride, %	0.22
Fluorine, ppm	1.0
Iron, ppm	40
Zinc, ppm	35
Manganese, ppm	11
Copper, ppm	6.0
Cobalt, ppm	0.0

Iodine, ppm	0.21
Chromium (added), ppm	1.0
Molybdenum, ppm	0.14
Selenium, ppm	0.24

Vitamins

Vitamin A, IU/g	4.0
Vitamin D-3 (added), IU/g	1.0
Vitamin E, IU/kg	81.6
Vitamin K, ppm	0.75
Thiamin, ppm	4.8
Riboflavin, ppm	6.7
Niacin, ppm	30
Pantothenic Acid, ppm	16
Folic Acid, ppm	2.1
Pyridoxine, ppm	5.8
Biotin, ppm	0.2
Vitamin B-12, mcg/kg	28
Choline Chloride, ppm	1,250
Ascorbic Acid, ppm	0.0

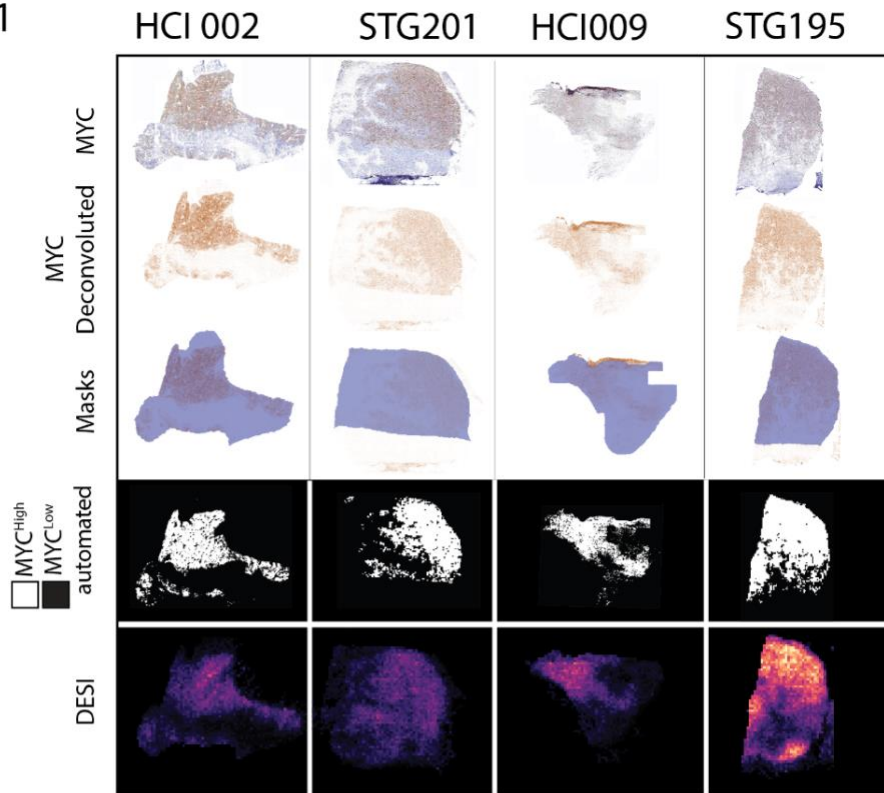
Flow Chart for automated k-means based tissue segmentation



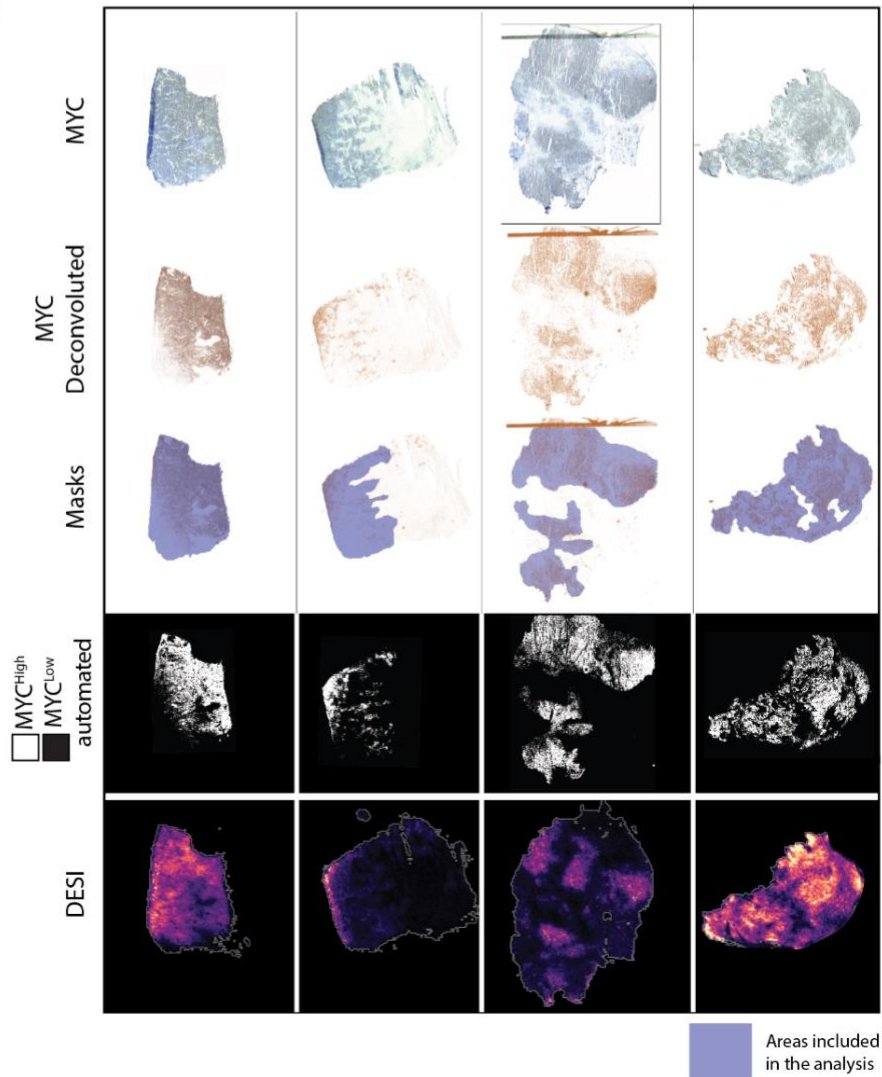
Supplementary Figure 1. An automated k means-based tissue segmentation.

Levels of MYC were identified by Immunohistochemistry in PDX samples and human biopsies after removal of areas containing staining artefacts. These areas were also excluded from downstream analysis. IHC signal was deconvoluted using the inbuilt DAB staining deconvolution algorithm of the QuPath software package. The pixels void of any DAB staining from the deconvoluted IHC images were then removed by thresholding the green image channel at an intensity of 230. The remaining pixels were subsequently assigned as MYC positive. The image was then re-binned by a factor of 16 by summing the total number of MYC positive pixels in any given 16x16 area using the Matlab function "blockproc" (Mathworks, Image processing toolbox). The resulting images (the MYC percentage proportions figures) were then clustered using the k-means clustering algorithm using Euclidean distance and $k = 2$ to differentiate regions that are high in MYC stain vs. low. The binary image of the cluster that was highest in MYC signal vs those with low and no MYC was then extracted and registered to the MSI image. Within each tumour the average difference of mean levels of PA between regions of high and low MYC was assessed by a linear mixed effect model fitted with the `glmmTMB` package for R (random intercepts were considered for run and tissue ID).

Slide 1

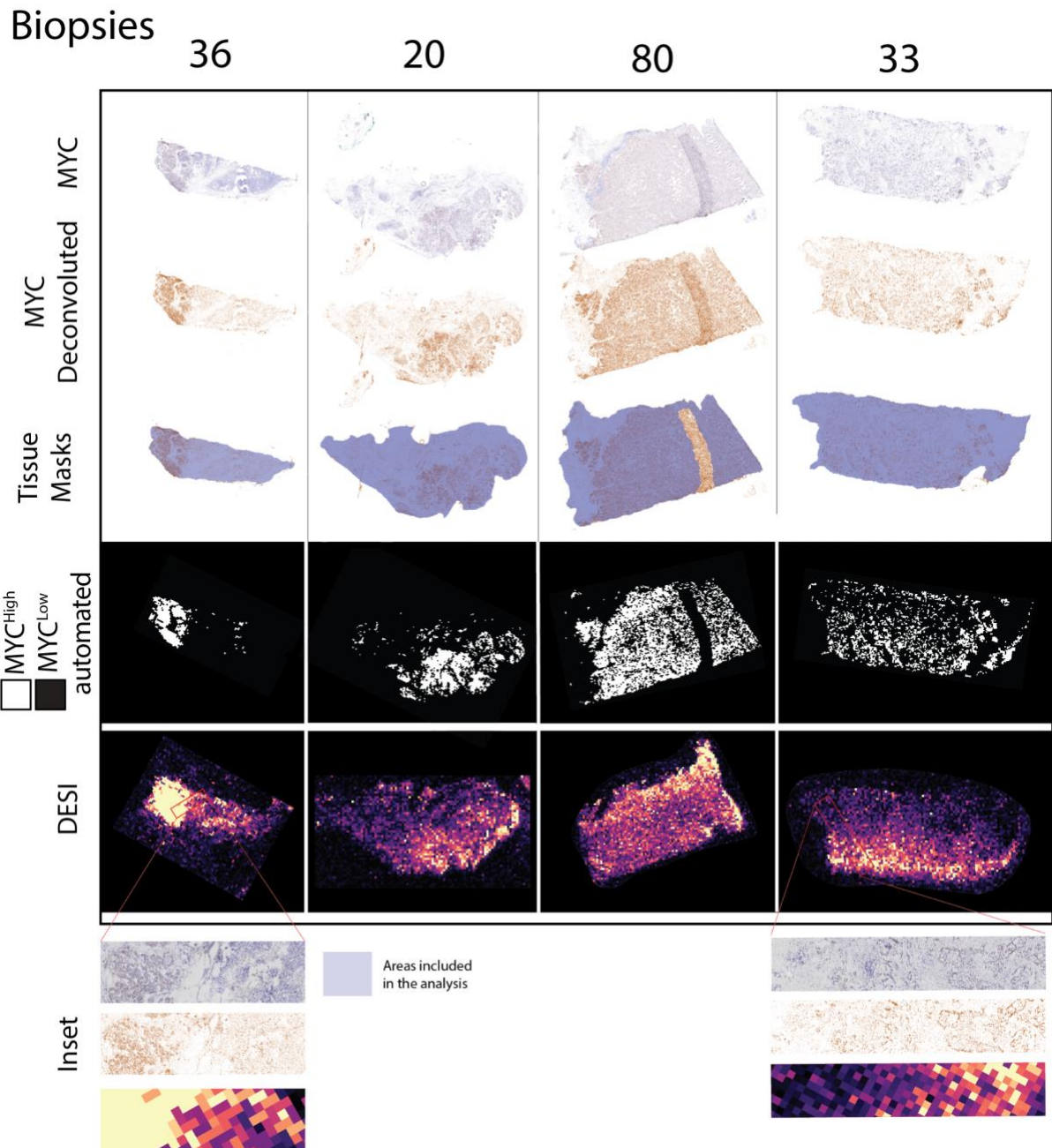


slide 2



Supplementary Figure 2. Outputs of automated segmentation of MYC staining in PDX samples.

Levels of MYC were identified by immunohistochemistry on the DEFFI-imaged section. IHC signal was deconvoluted using the inbuilt DAB staining deconvolution algorithm of the QuPath software package. Areas of overt necrosis and staining artefacts were excluded from the analysis by drawing masks over processing artefacts. The masks were also applied to the single ion images for PA from MSI and within each tumour the average difference of mean levels of PA between regions of high and low MYC was assessed as described above.



Supplementary Figure 3. Outputs of automated segmentation of MYC staining in human breast cancer biopsies.

Levels of MYC were identified by immunohistochemistry on the section consecutive to the DEFFI-imaged section. IHC signal was deconvoluted using the inbuilt DAB staining deconvolution algorithm of the QuPath software package. Areas of overt necrosis and staining artefacts were excluded from the analysis by drawing masks over processing artefacts. The masks were also applied to the single ion images for PA from MSI and within each tumour the

average difference of mean levels of PA between regions of high and low MYC was assessed as described above.