nature portfolio

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Last updated by author(s):	Oct 22, 2023		

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
X	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	A description of all covariates tested
\boxtimes	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
\boxtimes	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
X	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection

Data was collected using custom Matlab code (v2023a) running on a BPod State Machine (r2, Sanworks).

Data analysis

Data was analyzed with code written in Matlab (v2023a) that is publicly available at https://github.com/constantinoplelab/published/tree/main/rat_behavior. This information has been included in the Data and Code Availability section of the paper.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Data is publicly available on Zenodo at https://zenodo.org/records/10031483. This information has been included in the Data and Code Availability section of the paper.

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Policy information a and sexual orientati		with <u>human participants or human data</u> . See also policy information about <u>sex, gender (identity/presentation),</u> thnicity and racism.			
Reporting on sex and gender		Data was collected for both male and female rats. We found no differences in their behavior (Supplemental Figure 12), so all figures include data from both male and female rats.			
Reporting on race, ethnicity, or other socially relevant groupings		N/A			
Population charac	cteristics	N/A			
Recruitment		N/A			
Ethics oversight		New York University Animal Welfare Committee (UAWC #2021-1120)			
Note that full informat	tion on the appro	oval of the study protocol must also be provided in the manuscript.			
Field-spe	cific ro	norting			
· · ·		the best fit for your research. If you are not sure, read the appropriate sections before making your selection.			
X Life sciences		ehavioural & social sciences			
_		all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>			
Life scien	icas sti	ıdy design			
		points even when the disclosure is negative.			
Sample size		arge sample size (N = 291) in order to capture as much behavioral variability as possible across a large population of rats.			
Data exclusions	times were rem	ude behavioral sessions where wait time and log(reward) were not linearly related, for at least 2 consecutive sessions. Wait loved on a session-by-session basis if they were greater than 2 standard deviations above the mean. Trial initiation times were were above the 99th percentile of cumulative trial initiation times.			
Replication		Il rats received the exact same training protocol. Where applicable, we plotted data from individual rats, to confirm that the results were enerally reproduced across the population of rats.			
Randomization	For Figures 1-4, there were no experimental manipulations so randomization was not necessary. For Figure 5, rats were randomly selected by drawing a small subset from the currently running rats. Their data was similar to the population data collected thus far.				
Blinding	For Figures 1-4, there were no experimental manipulations so blinding was not necessary. For Figure 5, experimenters were not blinded, but all data was processed using a common pipeline developed for Figures 1-4 before the pre-initiation cue experiments were performed.				
We require informatio	on from authors a	Decific materials, systems and methods about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.			
Materials & exp					
n/a Involved in the Antibodies	e study	n/a Involved in the study ChIP-seq			
Eukaryotic o	cell lines	Flow cytometry			
	itology and archaeology MRI-based neuroimaging				
Animals and	s and other organisms				
Clinical data	a				
Dual use res	use research of concern				

Animals and other research organisms

Policy information about <u>studies involving animals</u>; <u>ARRIVE guidelines</u> recommended for reporting animal research, and <u>Sex and Gender in Research</u>

Laboratory animals Long-evans rats (184 male, 107 female) between ages of 6 and 24 months were used for this study (Rattus norvegicus)

Wild animals This study did not involve wild animals.

Reporting on sex Data was collected for both male and female rats. We found no differences in their behavior (Supplemental Figure 12), so all figures

include data from both male and female rats.

Field-collected samples This study did not contain samples collected from the field.

Ethics oversight Animal use procedures were approved by the New York University Animal Welfare Committee (UAWC #2021-1120) and carried out

in accordance with National Institutes of Health standards.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

off-target gene editing) were examined.

Plants

Seed stocks

Report on the source of all seed stocks or other plant material used. If applicable, state the seed stock centre and catalogue number. If plant specimens were collected from the field, describe the collection location, date and sampling procedures.

Novel plant genotypes

Describe the methods by which all novel plant genotypes were produced. This includes those generated by transgenic approaches, gene editing, chemical/radiation-based mutagenesis and hybridization. For transgenic lines, describe the transformation method, the number of independent lines analyzed and the generation upon which experiments were performed. For gene-edited lines, describe

Describe any authentication procedures for each seed stock used or novel genotype generated. Describe any experiments used to assess the effect of a mutation and, where applicable, how potential secondary effects (e.g. second site T-DNA insertions, mosiacism,

the editor used, the endogenous sequence targeted for editing, the targeting guide RNA sequence (if applicable) and how the editor was applied:

Authentication