

Supplemental information

**Three-dimensional genome architecture
coordinates key regulators of lineage
specification in mammary epithelial cells**

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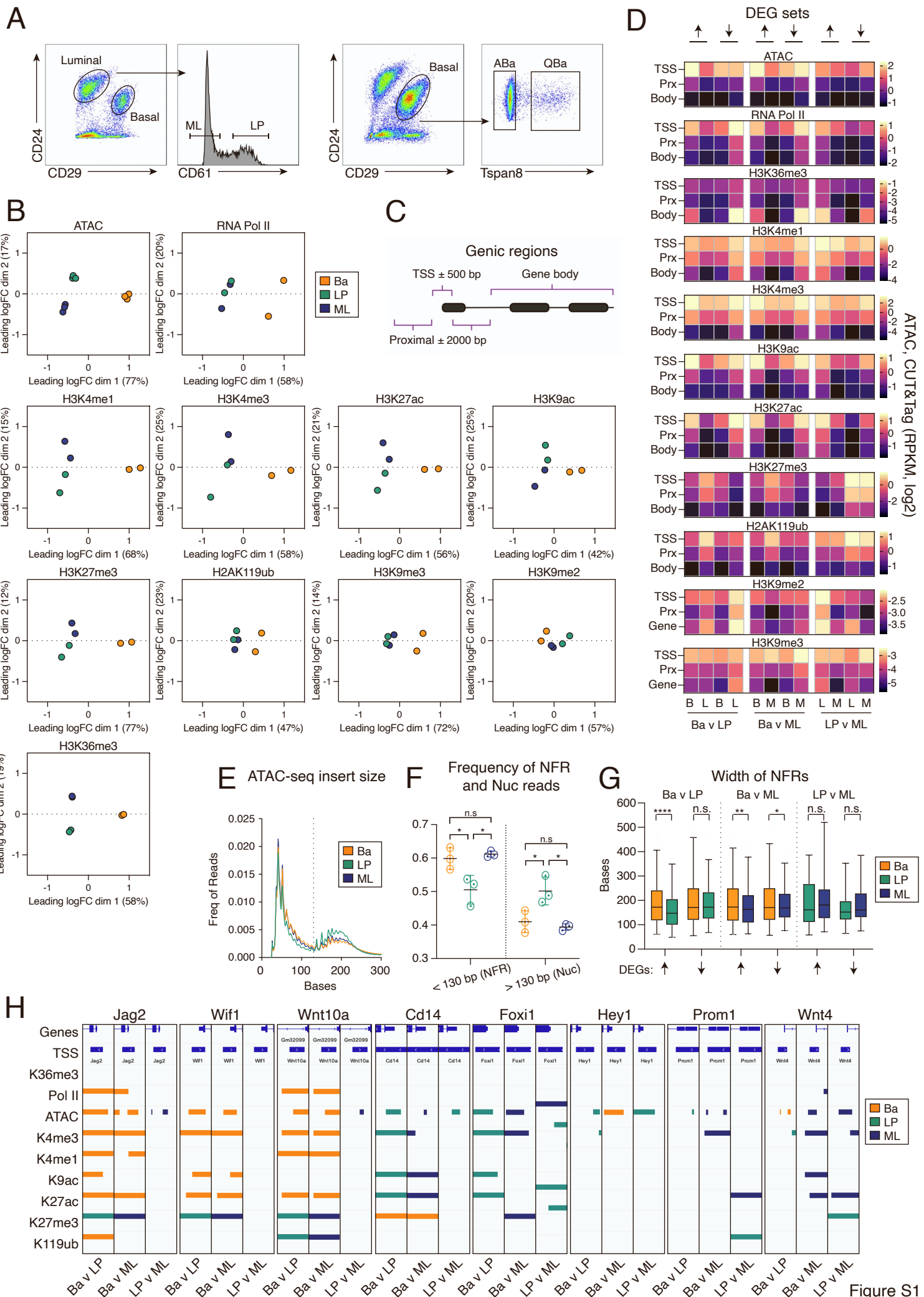


Figure S1

Figure S1 (Related to Figure 1). **Patterns of histone modifications amongst the mammary epithelial populations.** **(A)** FACS purification strategy for mammary epithelial cells. Following gating for single, live cells by 7-AAD, lineage-positive (CD31⁺/CD45⁺/TER-119⁺) cells were excluded and basal (CD29^{hi}/CD24⁺), LP (CD29^{lo}/CD24⁺/CD61⁺) or ML/HS (CD29^{lo}/CD24⁺/CD61⁻) cells were sorted; alternatively, basal Tspan8⁺ (QBa) or Tspan8⁻ (ABa) cells were isolated. **(B)** Multi-dimensional scaling plots (MDS) of normalized, filtered and batch-corrected ATAC and CUT&Tag data. **(C)** Diagram depicting the genic regions to which CUT&Tag and ATAC reads were assigned. TSS region spans 1 kb in length; proximal refers to a 4 kb region lacking the TSS; the gene body is specific to each gene, but in all instances has reads mapping to the TSS and proximal regions subtracted. **(D)** Heatmap of ATAC-seq and CUT&Tag for DE genes. **(E)** ATAC-seq read length insert size, mapped as a frequency of the total reads from merged biological replicate libraries. Dotted line represents the 130 bp cutoff used to assign NFR and nucleosome-associated (Nuc) reads. **(F)** Frequency of ATAC-seq reads. **(G)** Boxplot showing the width of NFRs at the TSSs of DEGs, median, quartiles and 5th and 95th percentiles. Two-tailed t-tests with Welch's correction: n.s.= not significant, * P < 0.05, ** P < 0.01, **** P < 0.0001. **(H)** Significant differential regions as determined by csaw (FDR < 0.05) in ATAC and CUT&Tag data of basal (Ba), LP and ML gene promoter regions. Color indicates the enrichment direction.

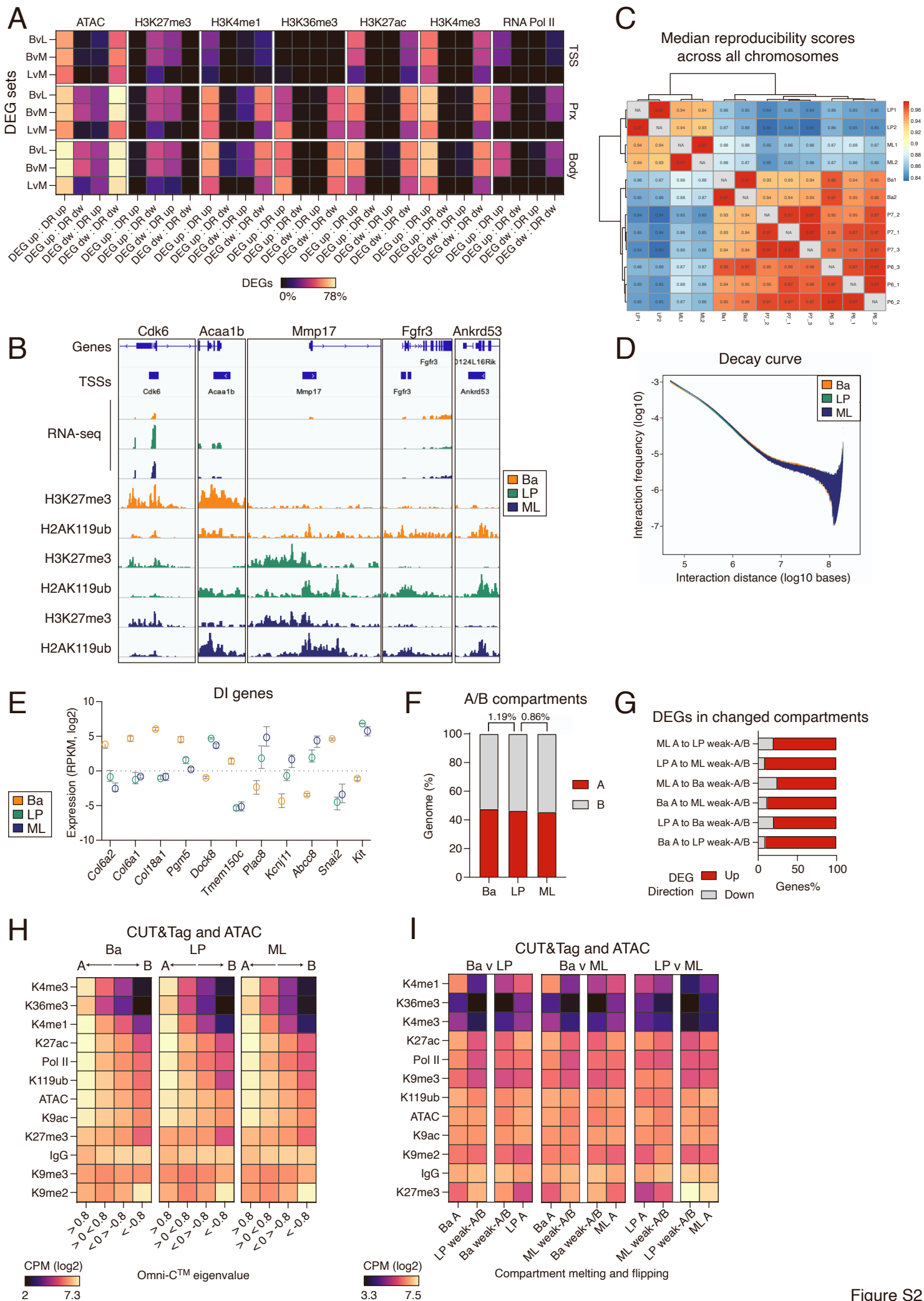


Figure S2

Figure S2 (Related to Figure 1 and 2). **Analysis of histone modifications and chromatin compartments in the different mammary epithelial subsets.** **(A)** Heatmap of DEGs with an overlapping significant differential region (DR) in the TSS, TSS proximal (Prx) regions and gene-bodies (body) for pairwise comparisons. **(B)** Coverage track-plots of genes with divergent H3K27me3 and H2AK119ub profiles. **(C)** Heatmap of the median reproducibility scores from the Omni-C libraries at 50 kb resolution. Basal Tspan8⁺ and Tspan8⁻ cells are listed as P7 and P6, respectively. **(D)** Decay curves of Omni-C data of read-pair interactions as a function of the interaction distance, indicating typical interaction profiles. **(E)** Mean expression of genes located within DI loci. Error bars indicate the range of the biological replicates. **(F)** Genome determined as an A or B compartment, shown above is the percentage of change from basal to LP to ML cells. **(G)** DEGs whose expression is up or down as the compartment changes from A to weak-A/B, between pairwise comparisons. **(H)** Average CUT&Tag and ATAC reads in A (eigenvalues >0) and B (eigenvalues <0) compartments within each cell type. Reads are mapped to 100 kb genomic bins as counts per million (CPM). **(I)** Average CUT&Tag and ATAC reads in compartments that significantly change between pairwise comparisons of basal, LP and ML cells. CPM of reads to each 100 kb bin are indicated.

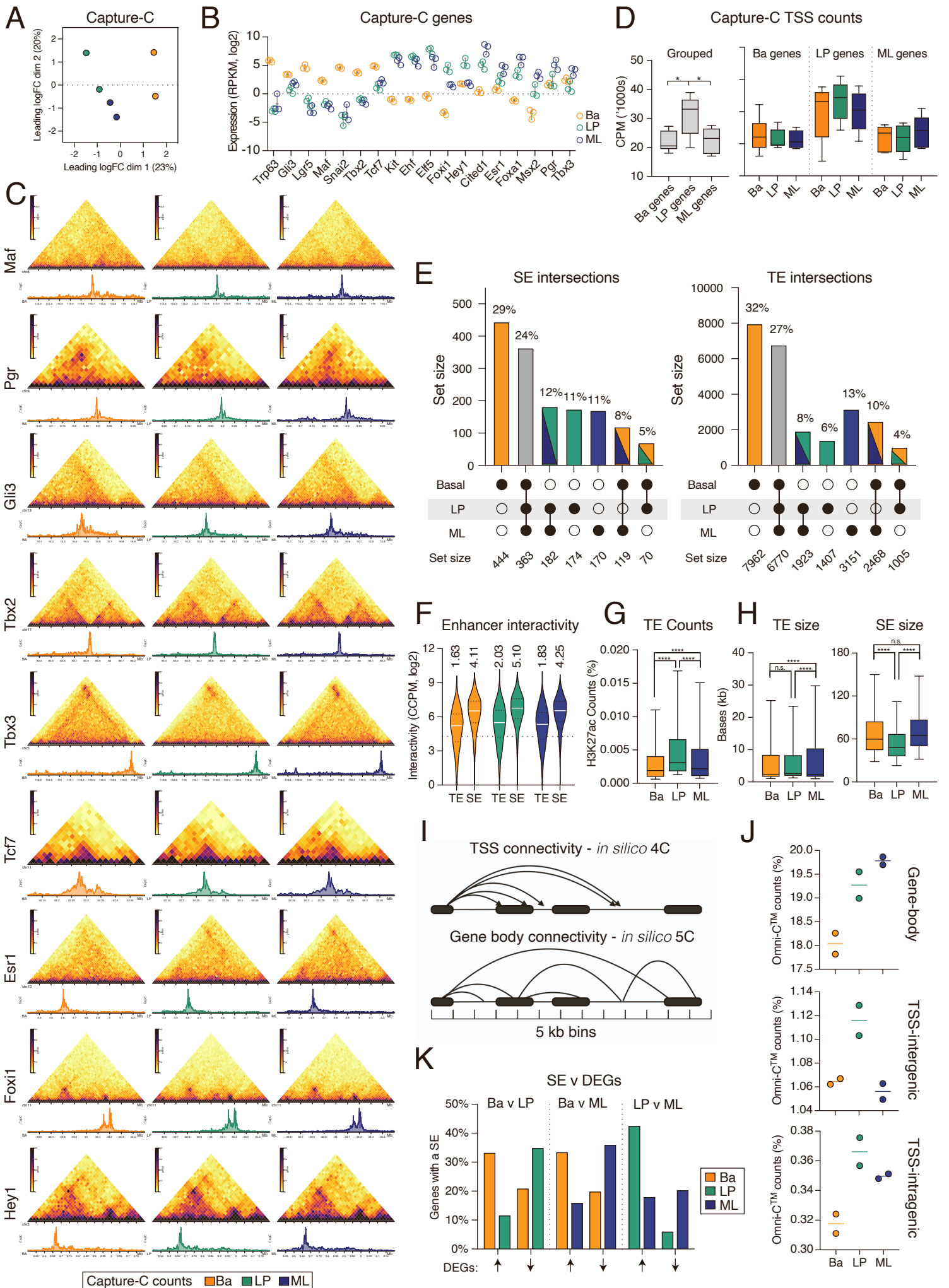


Figure S3

Figure S3 (Related to Figure 3). **Identification of chromatin interactions involving key lineage-restricted genes and analysis of super-enhancers.** **(A)** MDS plot of the normalized and filtered Capture-C data from basal, LP and ML cells. **(B)** Expression of genes targeted with Capture-C. **(C)** Omni-C normalized contact matrices at 20 kb resolution and Capture-C CPM. Each capture site is associated with the major peak in the Capture-C tracks. **(D)** CPM of all Capture-C genes by gene group (basal, LP and ML) and lineage enrichment. **(E)** Upset plots of SEs and TEs showing shared (≥ 500 bp overlap) and unique regions for the cell types. **(F)** Interactivity from the Omni-C data as measured by FitHiChIP, shown as contact counts per million (CCPM) for TSS to TE or SEs. Highlighted above each plot is the fold-increase in CCPM over the expected background (dotted line). **(G)** H3K27ac counts of TEs, shown as a percentage of the merged biological replicate libraries. **(H)** Size of TEs and SEs. **(I)** Diagram depicting the Omni-C *in silico* 4C and 5C analyses used to connect the TSS or gene-body respectively with other interactions: only genes >100 kb were considered. **(J)** Omni-C library counts for the 5C and 4C analysis. The TSS analysis was split into TSS-to-intergenic or TSS-to-intragenic interactions. **(K)** DEGs that are connected to a SE. Genes were considered connected with a SE if they overlapped their gene-body or if their TSS interacted with a SE as detected by FitHiChIP through chromatin interactions or gene-body-overlap. * $P < 0.05$, **** $P < 0.0001$ (t-test with Welch's correction). Boxes shows median, quartiles and 5th and 95th percentiles.

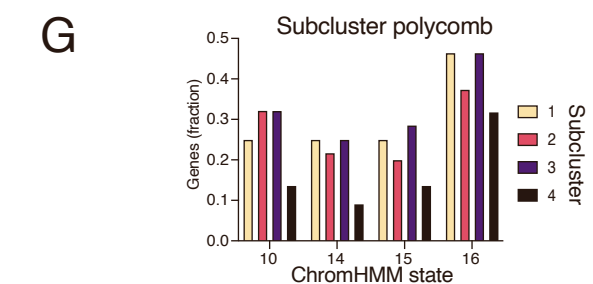
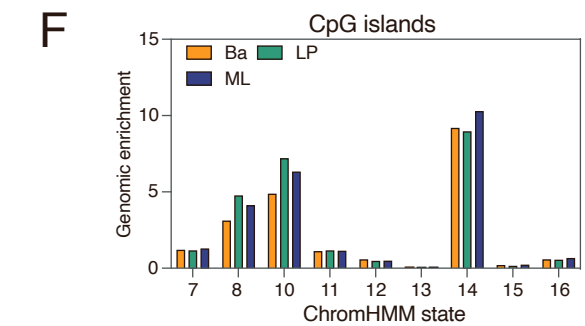
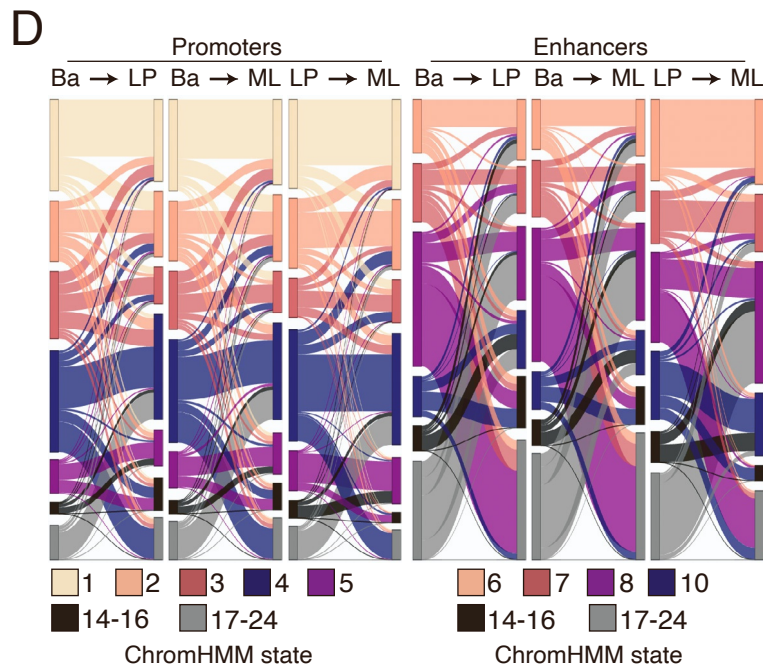
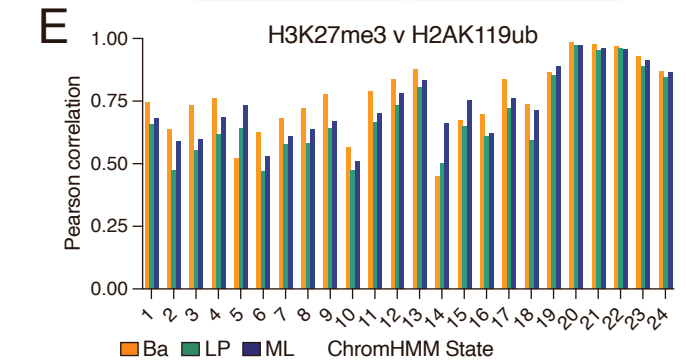
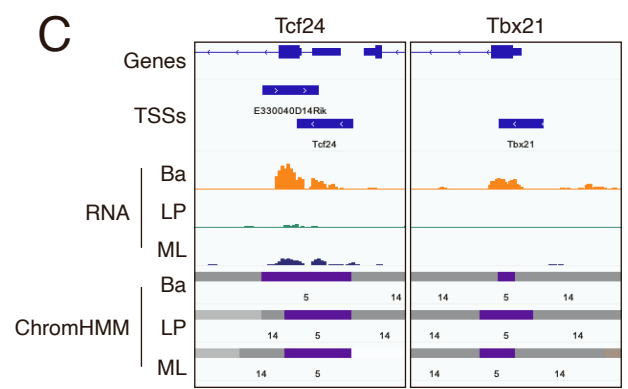
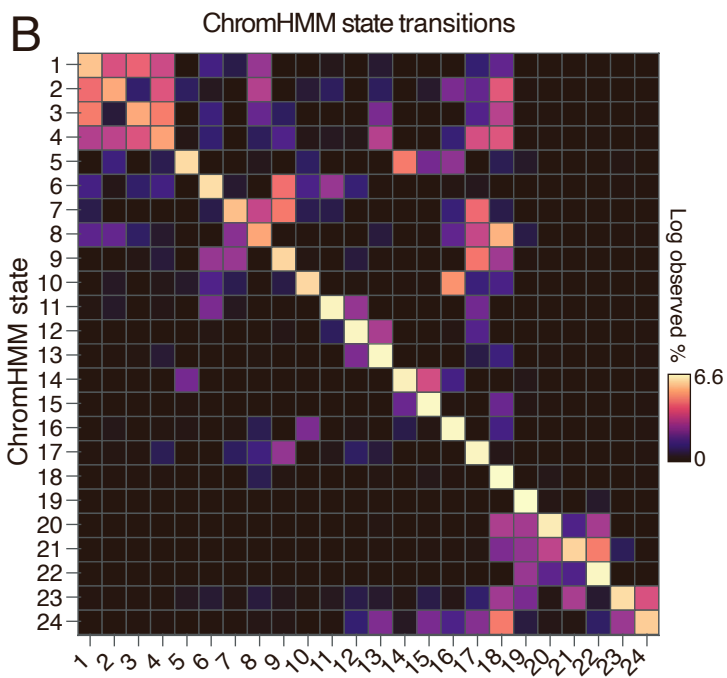
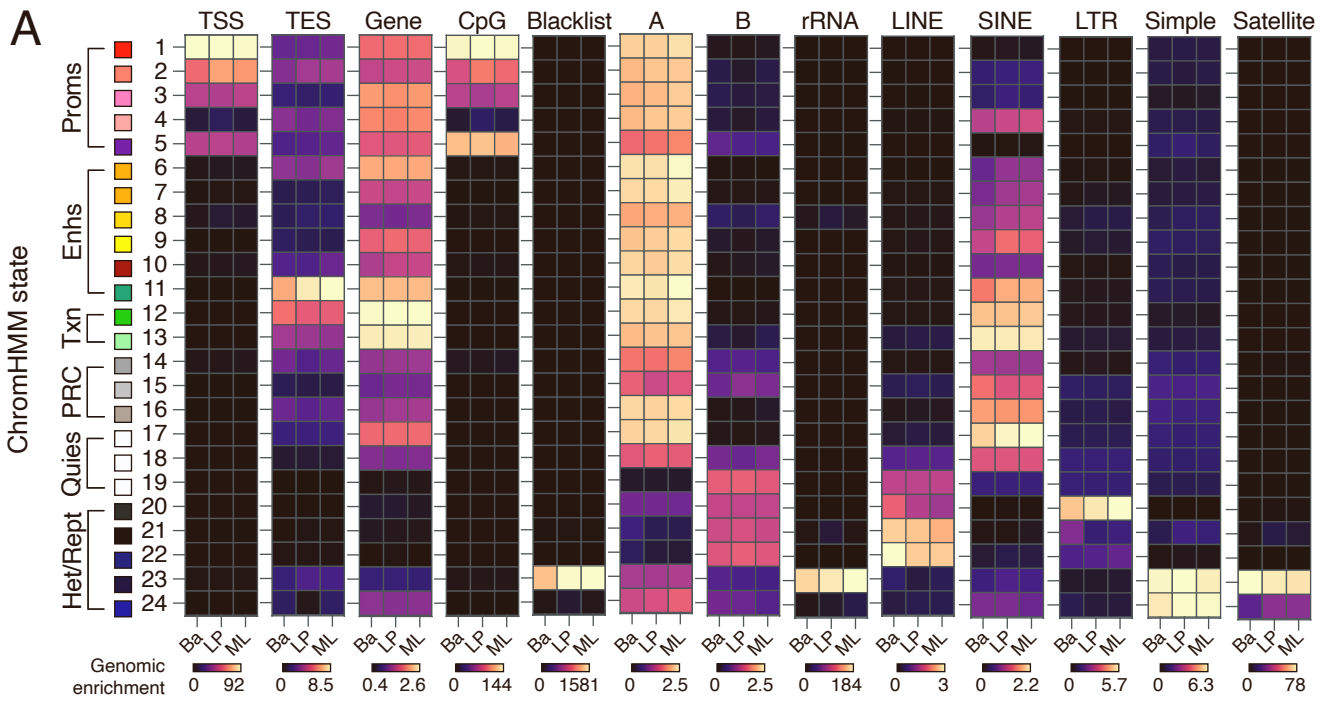


Figure S5

Figure S4 (Related to Figure 4). **Alternative use of transcription start sites associates with histone modification and chromatin looping.** **(A)** Expression and connectivity of differentially expressed TSSs between cell types. Expression values are Z-scored RNA-seq log₂ RPKM, graphed as median and range. Connectivity values are Z-scored counts from *in silico* 4C analysis determining chromatin interactions between the TSSs and gene-bodies, graphed as mean and range. **(B)** Mean coverage (RPKM) of CUT&Tag and ATAC data across TSSs identified as either 'off' and 'on'. **(C)** Diagram depicting the analysis used to determine differentially expressed alternative transcriptional start sites (DEATSS) and differentially expressed transcriptional start sites (DETSS). **(D)** Coverage track-plots of expression (RPKM log₂) for *Eya2* and *Lrrfip1* showing alternative TSSs, expression and H3K27ac (RPKM) for basal, LP and ML cells and known isoforms from RefSeq annotation.

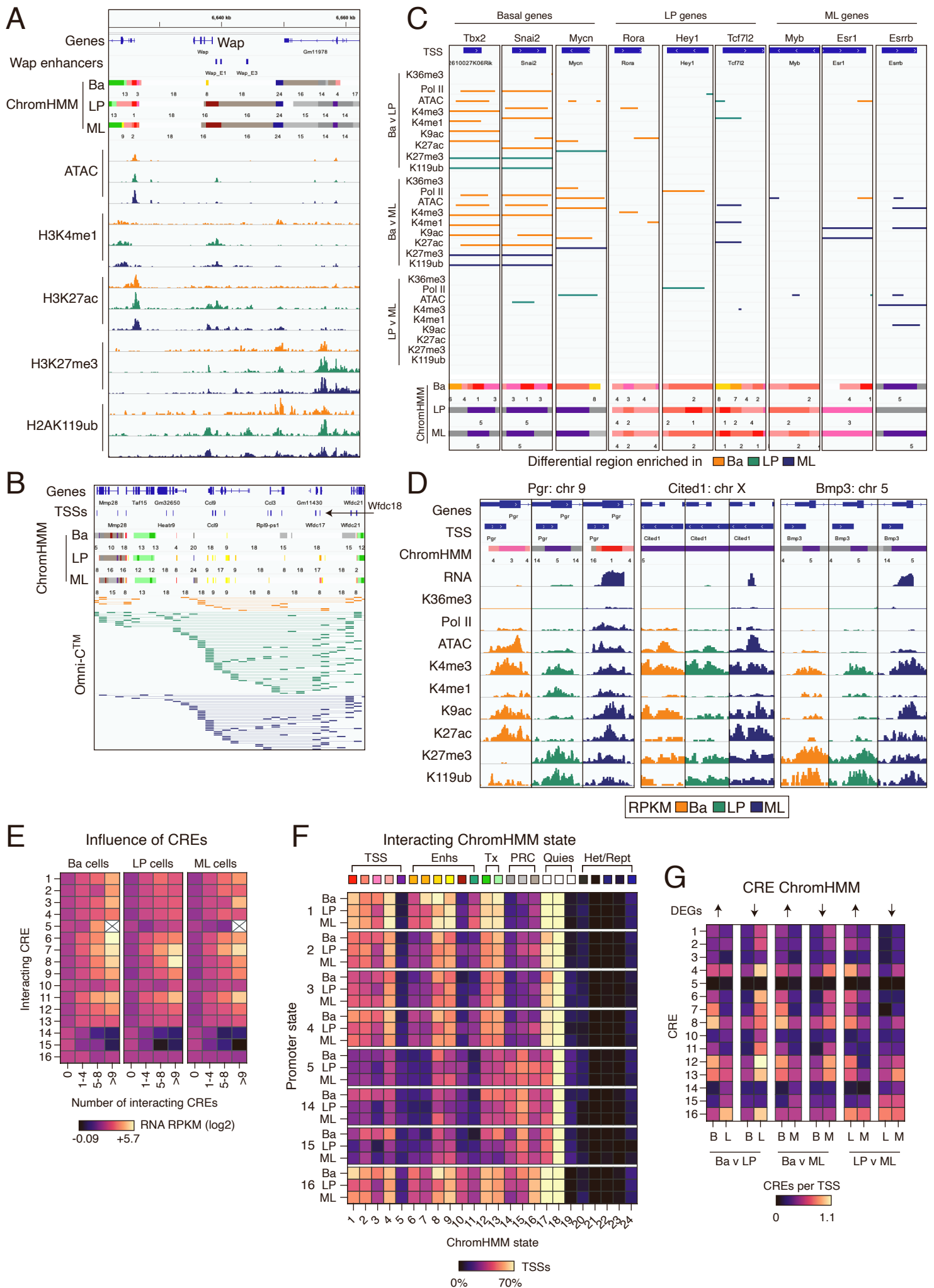


Figure S6

Figure S5 (Related to Figure 5). **Derivation of a chromatin model for the different mammary epithelial lineages.** **(A)** The enrichment of genomic features for the 24-state ChromHMM model. Shown are the enrichments in basal (Ba), LP and ML cells for TSS, transcriptional end sites (TES), gene-bodies (Gene), CpG islands, genome sequencing blacklisted sites (Blacklist), A and B chromatin compartments as determined by Omni-C and repeat element classes: ribosomal RNAs (rRNA)s, long-interspersed elements (LINE), short interspersed elements (SINE), long terminal repeats (LTR), simple repeats (Simple) and satellite regions (Satellite). **(B)** Transition frequencies between ChromHMM states. **(C)** Coverage track-plots for *Tcf24* and *Tbx21*. **(D)** Sankey transition plots showing the proportion of promoter and enhancer chromatin states that change between pairwise comparisons. States 14-16 were grouped as PRC and states 17-24 grouped as heterochromatin/silent. **(E)** Correlation of CUT&Tag H3K27me3 and H2AK119ub (RPKM, log2) across 24 ChromHMM state models in basal, LP and ML cells. **(F)** Genomic enrichment of CpG islands for different ChromHMM states. **(G)** Genes in scRNA-seq subclusters (Figure 5G) with an interacting polycomb state.

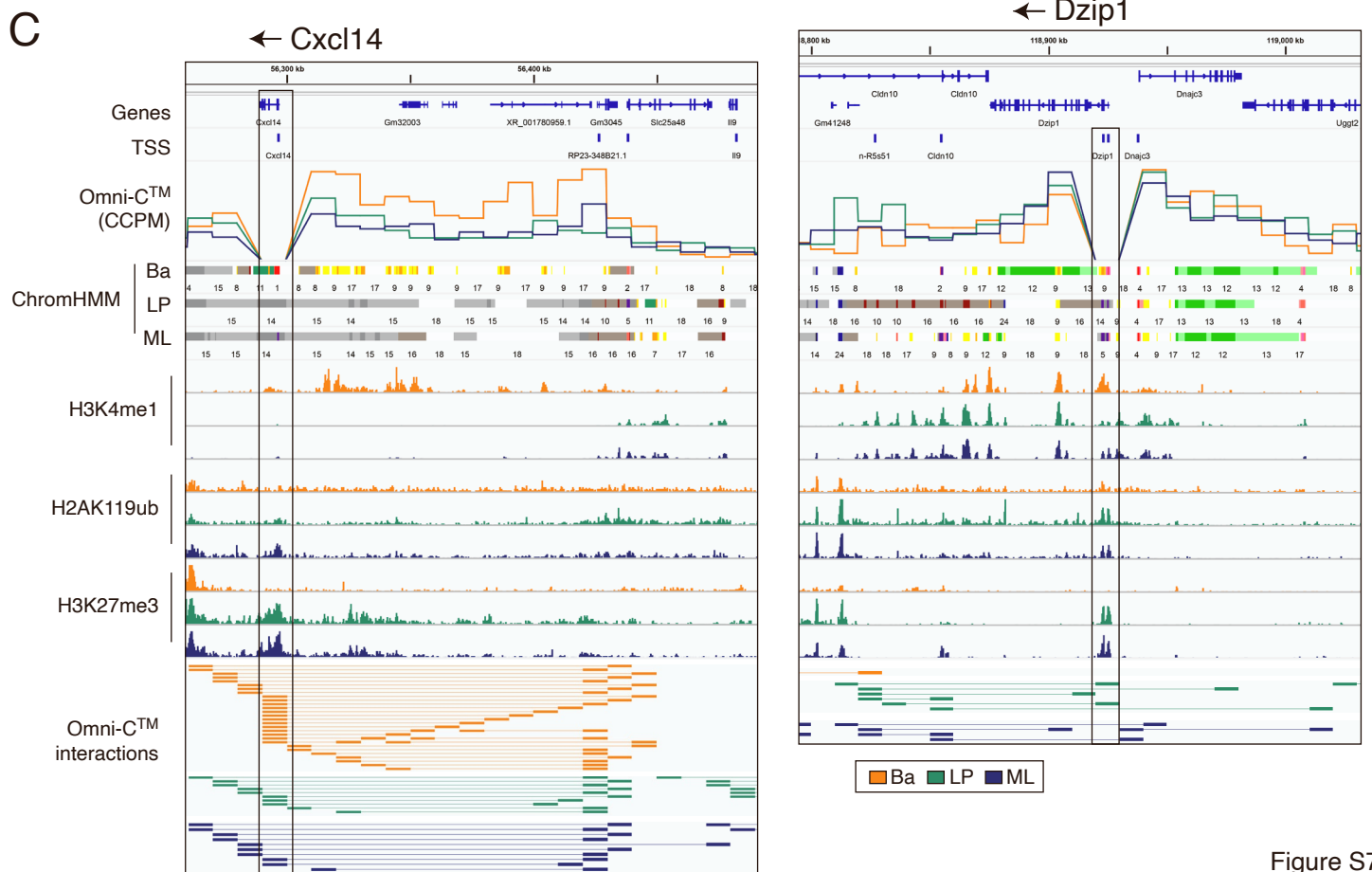
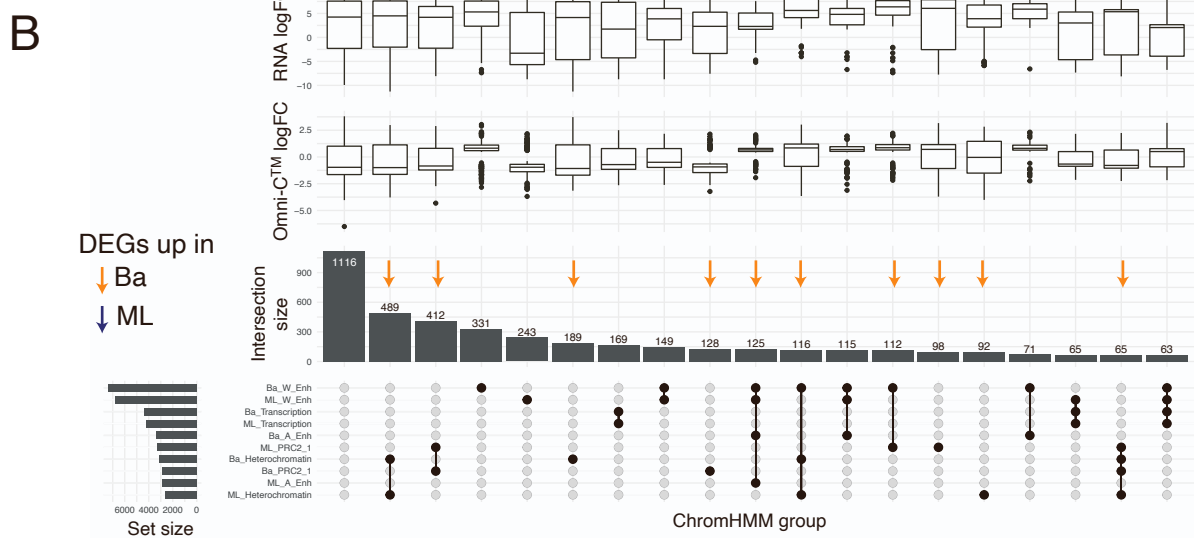
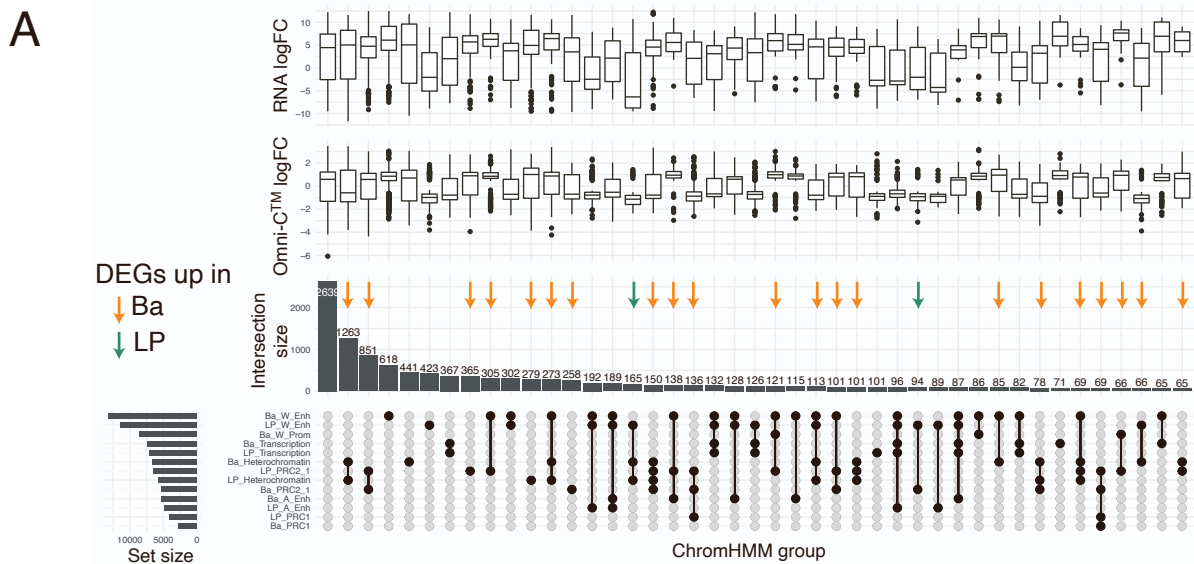


Figure S7

Figure S6 (Related to Figure 5). **The influence of promoter and distal CREs on lineage-restricted expression.** **(A)** Coverage track-plots of the *Wap* locus with the *Wap* enhancers identified by Shin et al. 2016.¹ **(B)** Coverage track-plot of the *Wfdc18* locus: shown are the genes, TSSs, ChromHMM states and Omni-C interactions as determined by FitHiChIP (FDR < 0.1). **(C)** Chromatin states of gene promoters, shown are the significant differential regions as determined by csaw (FDR < 0.05) in ATAC and CUT&Tag data of basal (Ba), LP and ML gene promoter regions (1 kb TSS region shown). Color indicates the enrichment direction in the comparison. Shown below are the ChromHMM states for Ba, LP and ML cells. **(D)** Coverage track-plots of ML gene promoters. Shown are CUT&Tag (RPKM), ATAC-seq (RPKM) and the ChromHMM states. Each TSS site is 1 kb for reference. **(E)** Heatmap of the average expression of genes with 0 or increasing numbers of interacting CREs in each population as determined by states showing overlapping interactions from the FitHiChIP analysis. **(F)** Heatmap of TSSs interacting with each chromatin state, as determined by overlapping with interactions from the FitHiChIP analysis. **(G)** Average number of CREs interacting with TSSs of DEGs from pairwise comparisons between basal, LP and ML cells.

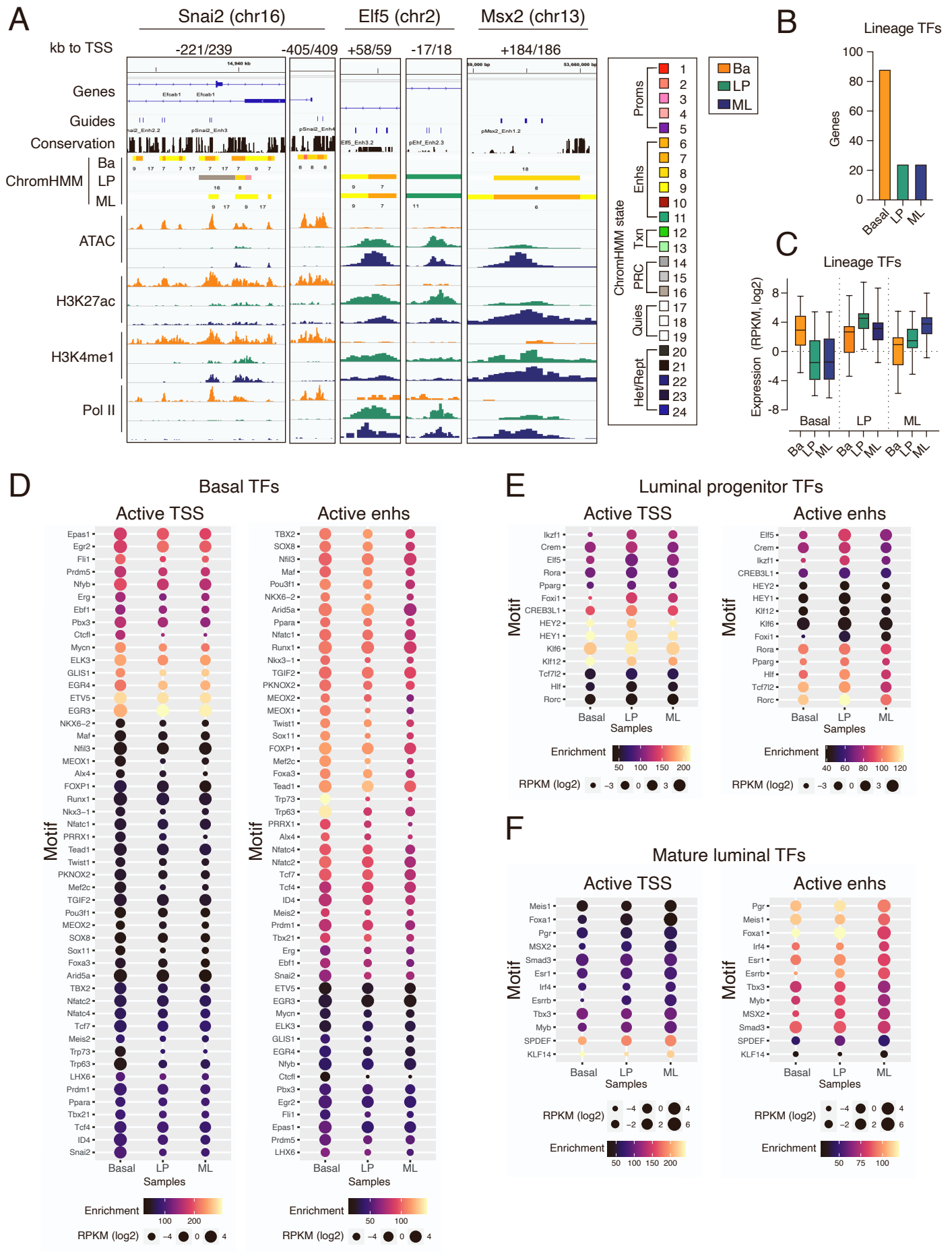


Figure S8

Figure S7 (Related to Figure 5) **Silencers primarily influence gene expression in luminal cells. (A and B)** Upset plot analysis of DEGs overlapping with DI regions between basal (Ba) and LP and basal and ML cells, respectively. Shown are the log fold-change expression (RPKM, log2) and log-FC chromatin interactivity (Omni-C) for basal versus LP and basal versus ML respectively, intersection size and ChromHMM combinations. ChromHMM states were summarized with state 1 = A_Prom, states 2-4 = W_Prom, states 5, 14 and 15 = PRC2_1, states 6 and 7 = A_Enh, states 8, 9 and 11 = W_Enh, states 10 and 16 = PRC1, states 12 and 13 = transcription, states 17-19 = silent chromatin and states 20-24 = heterochromatin. Arrows indicate the interactions that contain a repressive element (PRC1, PRC2 and or heterochromatin) and the color indicates the cell in which the DEG is more expressed. **(C)** Coverage track-plots for *Cxcl14* and *Dzip1*. Omni-C reads show regions connected to TSSs, whilst Omni-C interactions are those determined by FitHiChIP.

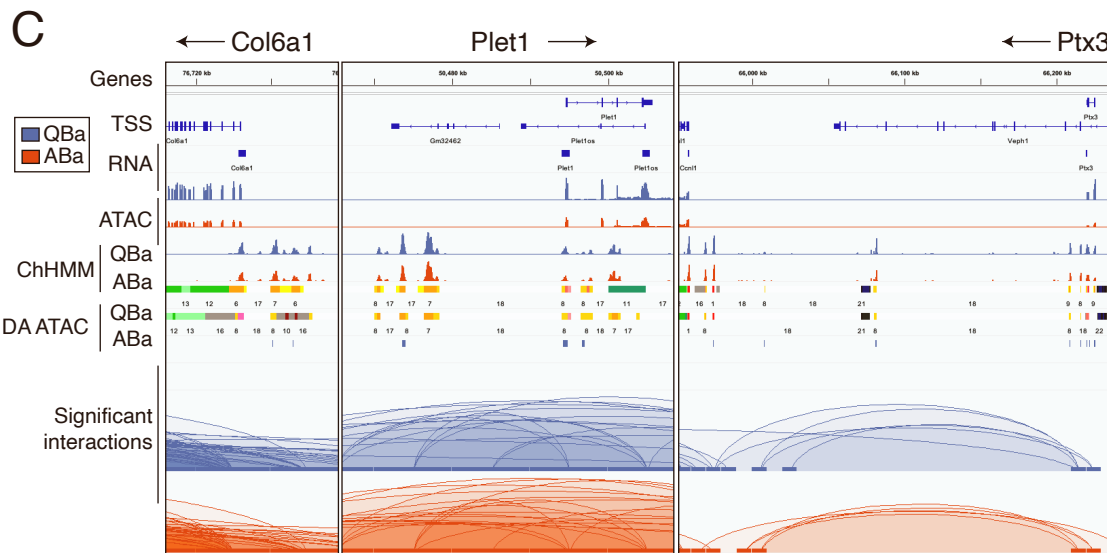
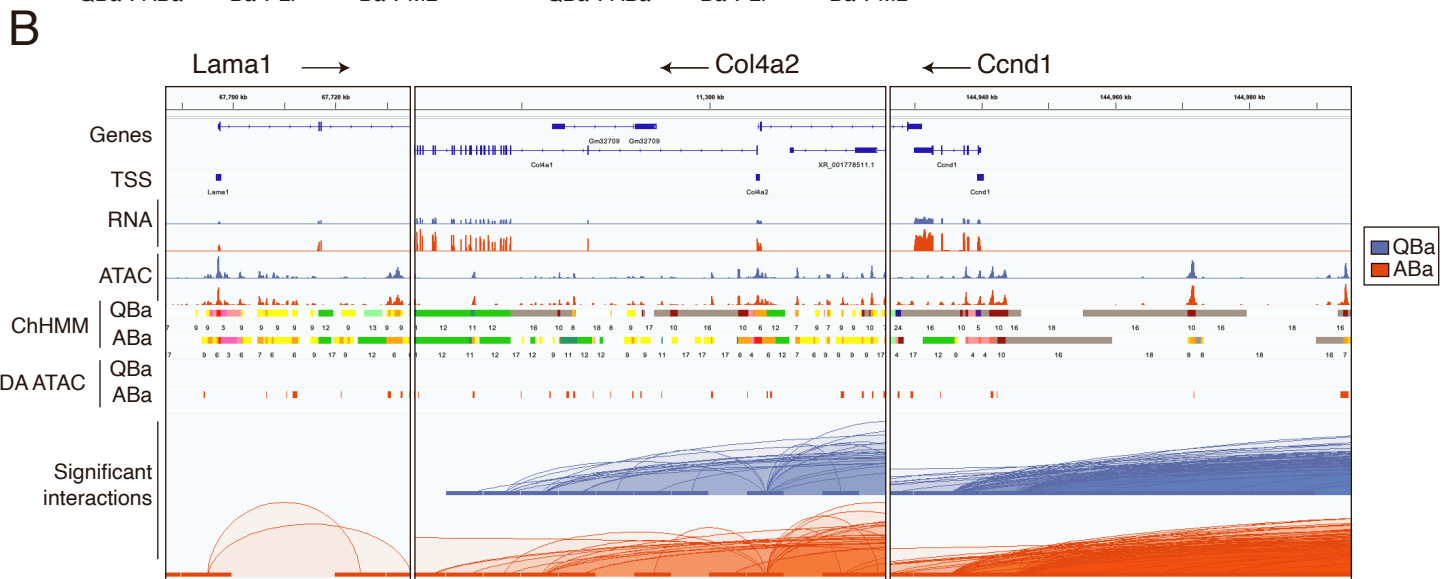
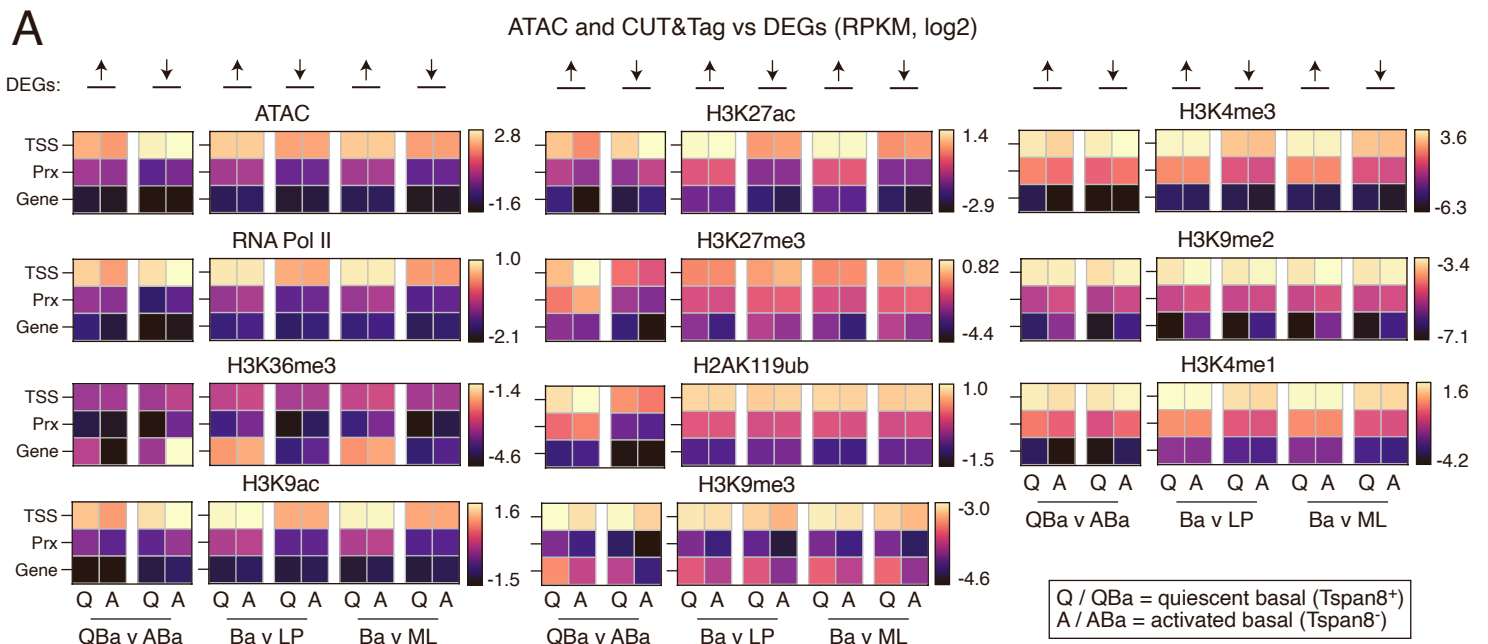


Figure S9

Figure S8 (Related to Figure 6). **Transcription factors display distinct chromatin binding patterns.** **(A)** Zoomed-in coverage track-plots from Figure 6A for the strongest enhancers of *Snai2*, *Elf5* and *Msx2*. From top to bottom, genes, short-guide-RNAs for CRISPR-activation, conservation across placental mammals, ChromHMM modeling, RPKM for ATAC and CUT&Tag. **(B)** Lineage-restricted TFs identified through signature profiling of basal (Ba, 88), LP (24) and ML (24) cells. **(C)** Gene expression of lineage-specific TFs. Boxes shows median, quartiles and 5th and 95th percentiles. **(D to F)** TOBIAS analysis of TF accessible motif enrichment (observed vs expected, log2, color chart) and TF RNA expression (circle size) for lineage-specific TFs, showing enrichment for active promoters (state 1) and distal active enhancers (state 7).

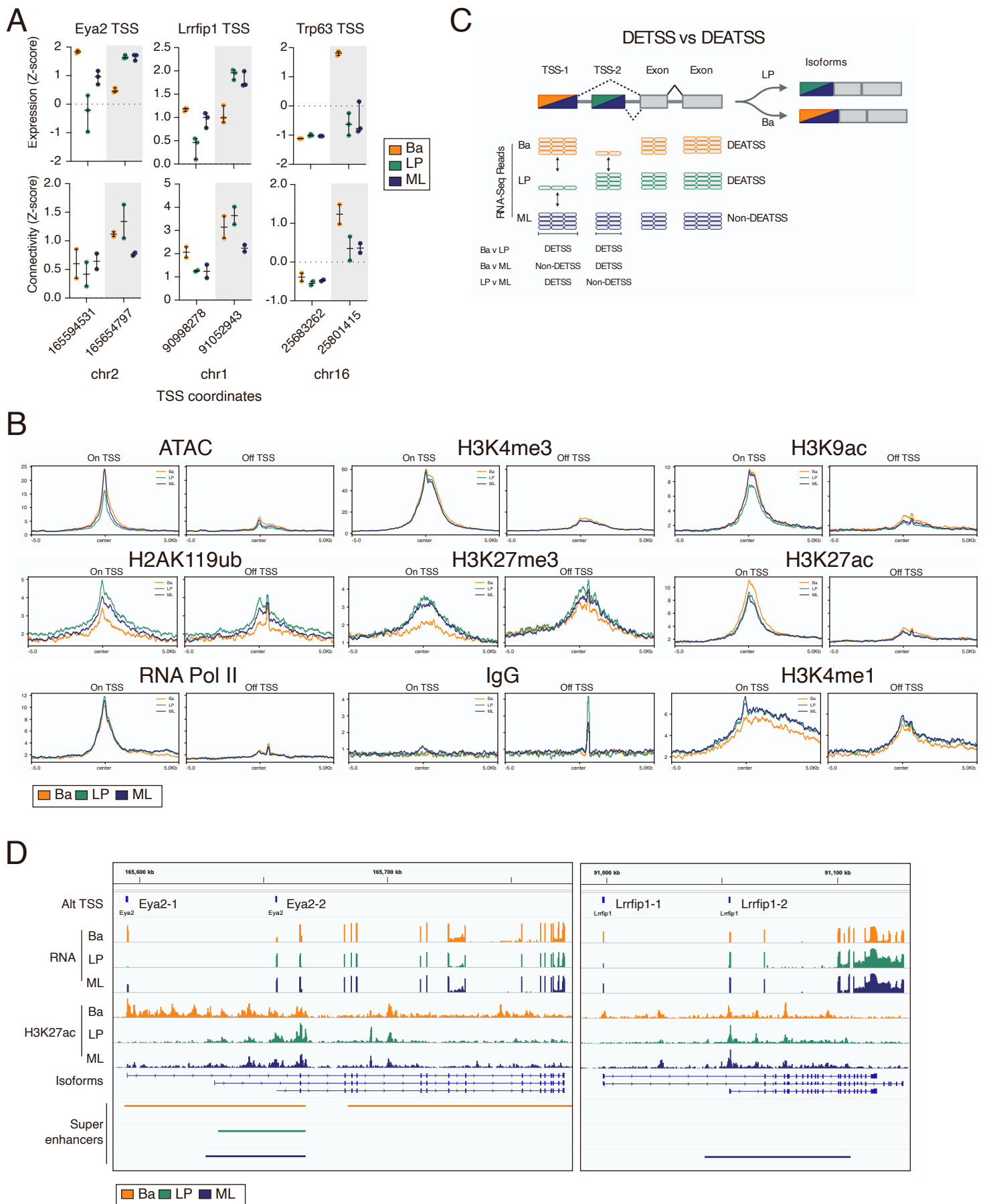


Figure S4

Figure S9 (Related to Figure 7). **Histone modifications and chromatin looping events that define the basal compartment.** **(A)** Average RPKM for ATAC-seq and CUT&Tag across the TSS, proximal (Prx) and gene-body (Gene) regions for DEGs between QBa versus ABa, basal versus LP, and basal versus ML populations. **(B and C)** Coverage track-plots for genes enriched in active basal (ABa, Tspan8⁻) and quiescent basal (QBa, Tspan8⁺) cells, respectively. Shown are the genes, TSSs, expression (RPKM), ATAC-Seq (RPKM), ChromHMM, DA regions, and the interactions determined by FitHiChIP. **(D)** Sankey transition plots showing the proportion of promoter and enhancer chromatin states that change between QBa and ABa cells. States 14-16 were grouped as PRC and states 17-24 grouped H/S.

References

1. Shin, H.Y., Willi, M., HyunYoo, K., Zeng, X., Wang, C., Metser, G., and Hennighausen, L. (2016). Hierarchy within the mammary STAT5-driven Wap super-enhancer. *Nature genetics* 48, 904-911. [10.1038/ng.3606](https://doi.org/10.1038/ng.3606).