

Supporting Information

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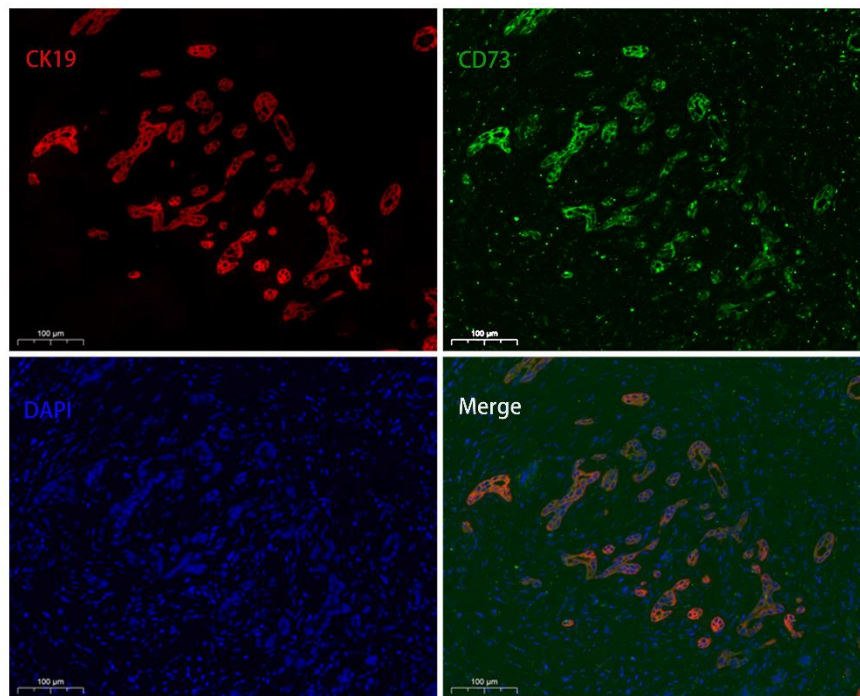
Tumor Microenvironment Responsive CD8⁺ T Cells and Myeloid-Derived Suppressor Cells to Trigger CD73 Inhibitor AB680-Based Synergistic Therapy for Pancreatic Cancer

Qiangda Chen, Hanlin Yin, Junyi He, Yuqi Xie, Wenquan Wang, Huaxiang Xu, Lei Zhang, Chenye Shi, Jun Yu, Wenchuan Wu, Liang Liu, Ning Pu* and Wenhui Lou**

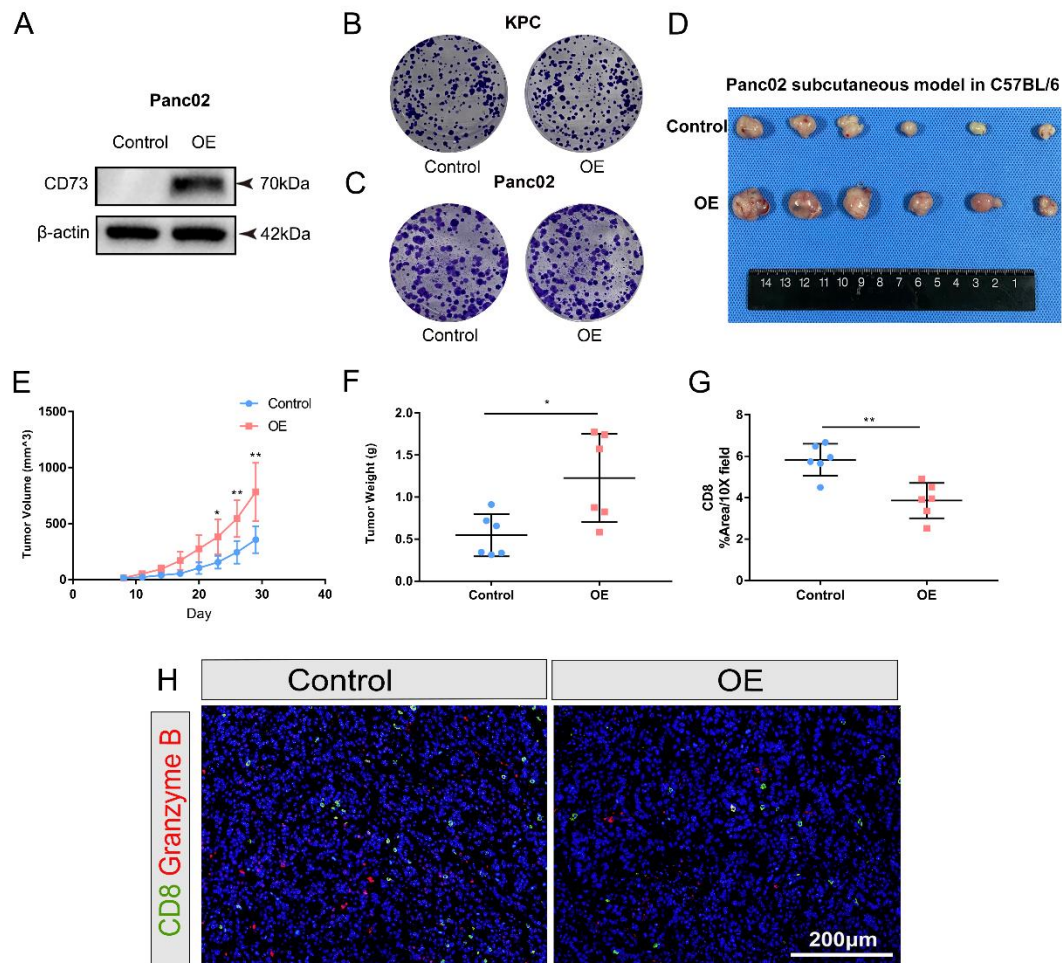
Supporting Information

Tumor microenvironment responsive CD8⁺ T cells and myeloid-derived suppressor cells to trigger CD73 inhibitor AB680-based synergistic therapy for pancreatic cancer

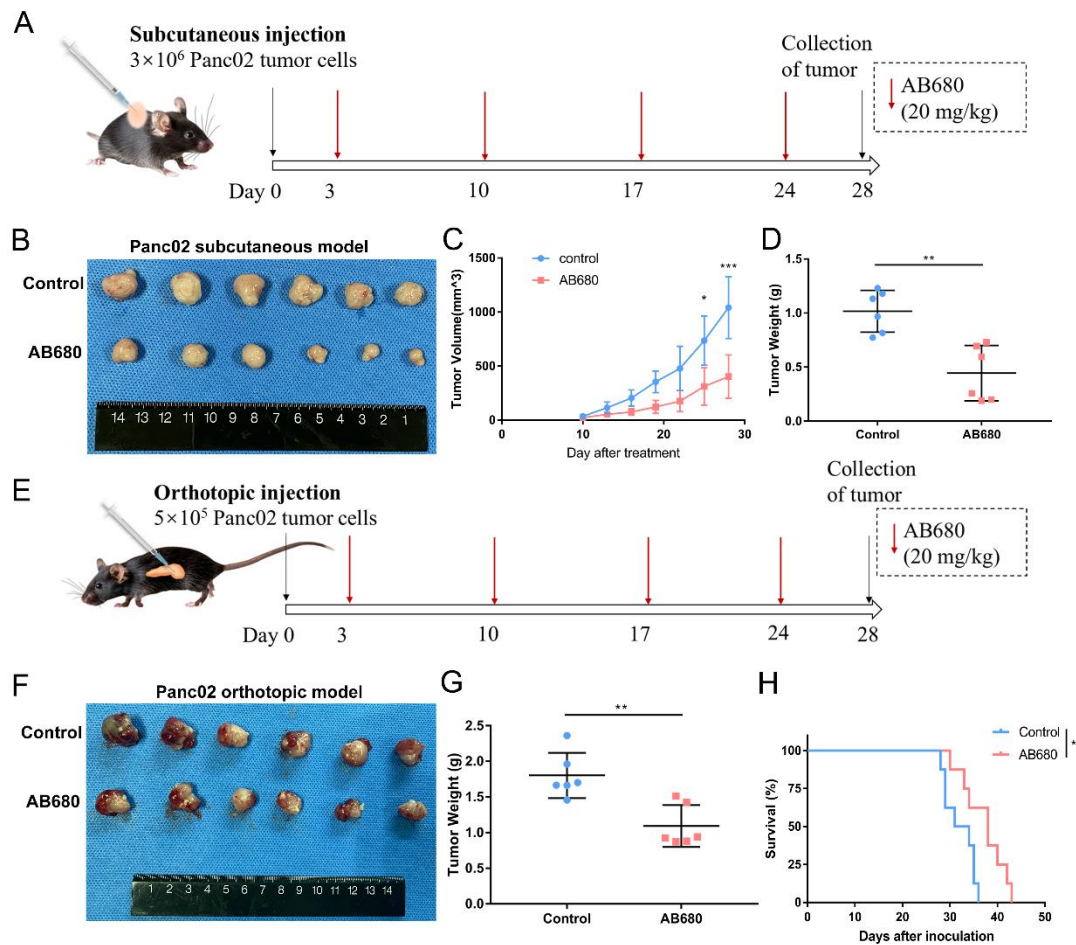
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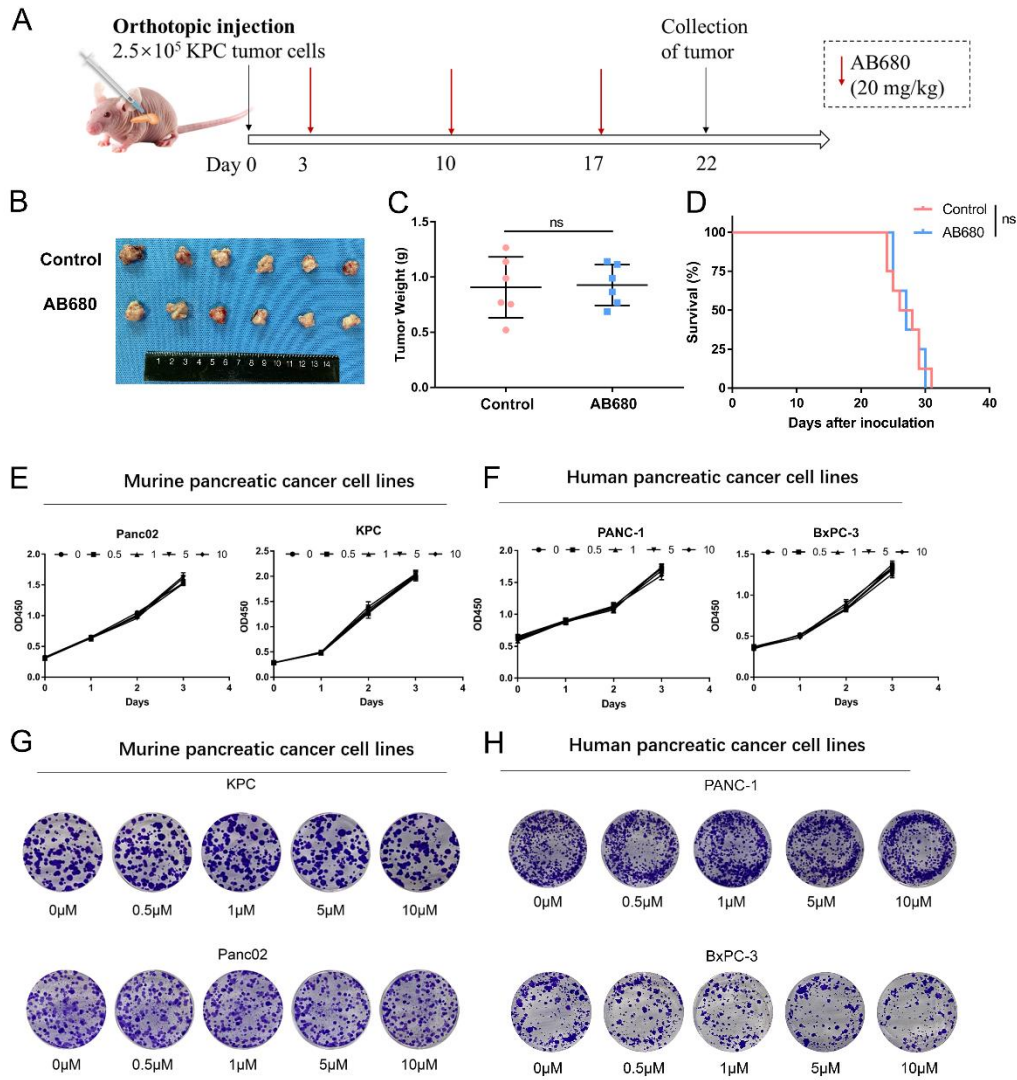
Supplementary Figure 1. Representative immunofluorescence staining of CD73 and CK19. CK19: red; CD73: green; DAPI: blue. Scale bar: 200μm.



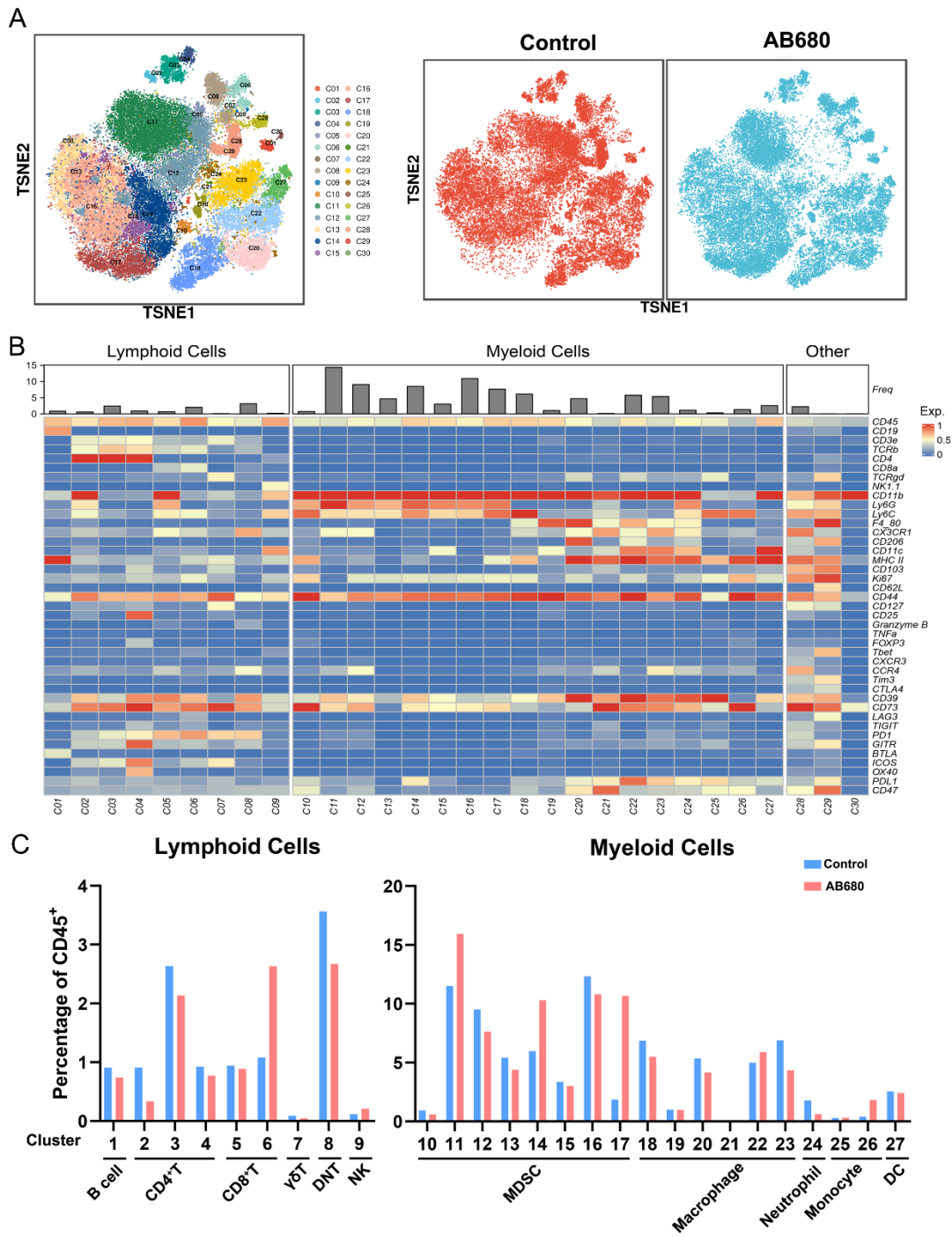
Supplementary Figure 2. Overexpression of CD73 in murine tumor cells promotes tumor growth of subcutaneous tumor-bearing mice. (A) Western blot analysis for CD73 in CD73 overexpressing (OE) and control Panc02 cells. (B-C) Clone formation assay for 2 weeks for cell proliferation after CD73 overexpression in KPC (B) and Panc02 (C) cells. (D-F) Image (D), volumes (E), and weights (F) of the CD73 OE vs Control Panc02 tumors (n=6 samples per group). *p<0.05, **p<0.01 in Student's t-test. Data presented as mean ± SD. (G) Quantification of immunohistochemistry result of CD8⁺ T cells in the CD73 OE vs Control Panc02 tumors (n=6 samples per group). **p<0.01 in Student's t-test. Data presented as mean ± SD. (H) Representative immunofluorescence staining of CD8 and Granzyme B. Granzyme B: red; CD8: green; DAPI: blue. Scale bar: 200μm.



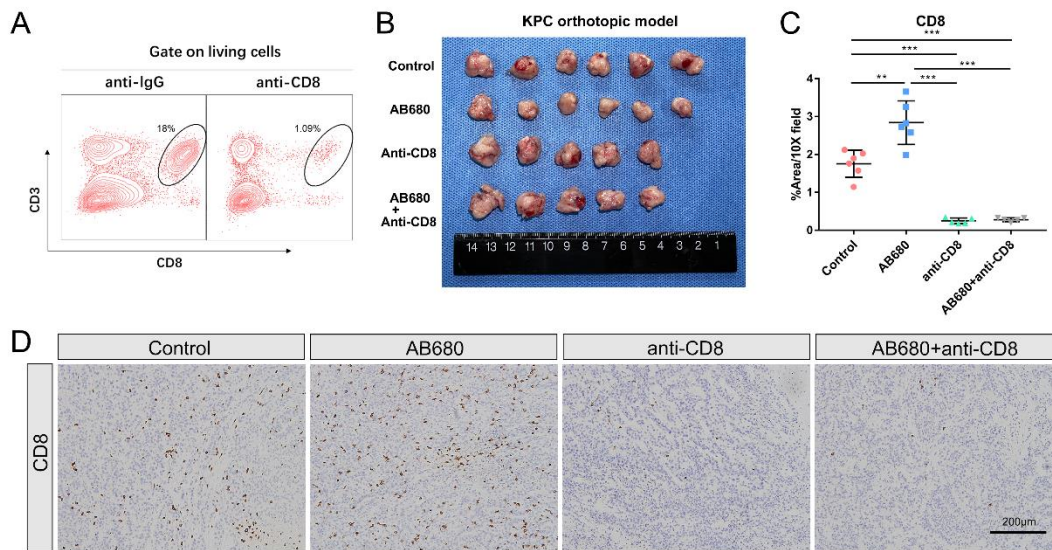
Supplementary Figure 3. AB680 reduces the tumor burden and prolongs survival in Panc02 subcutaneous and orthotopic models. (A) Schematic showing the schedule of CD73 inhibitor AB680 treatment in Panc02 subcutaneous model. (B-D) Image (D), volumes (E), and weights (F) of the AB680-treated vs Control Panc02 subcutaneous tumors (n=6 samples per group). * $p < 0.05$, ** $p < 0.01$ in Student's t-test. Data presented as mean \pm SD. (E) Schematic showing the schedule of AB680 treatment in Panc02 orthotopic model. (F-G) Image (F) and weights (G) of the AB680-treated vs Control Panc02 orthotopic tumors (n=6 samples per group). ** $p < 0.01$ in Student's t-test. Data presented as mean \pm SD. (H) Kaplan-Meier survival analysis of tumor-bearing mice untreated or treated with 20 mg/kg AB680 (n=8 samples per group). * $p < 0.05$ in the log-rank test.



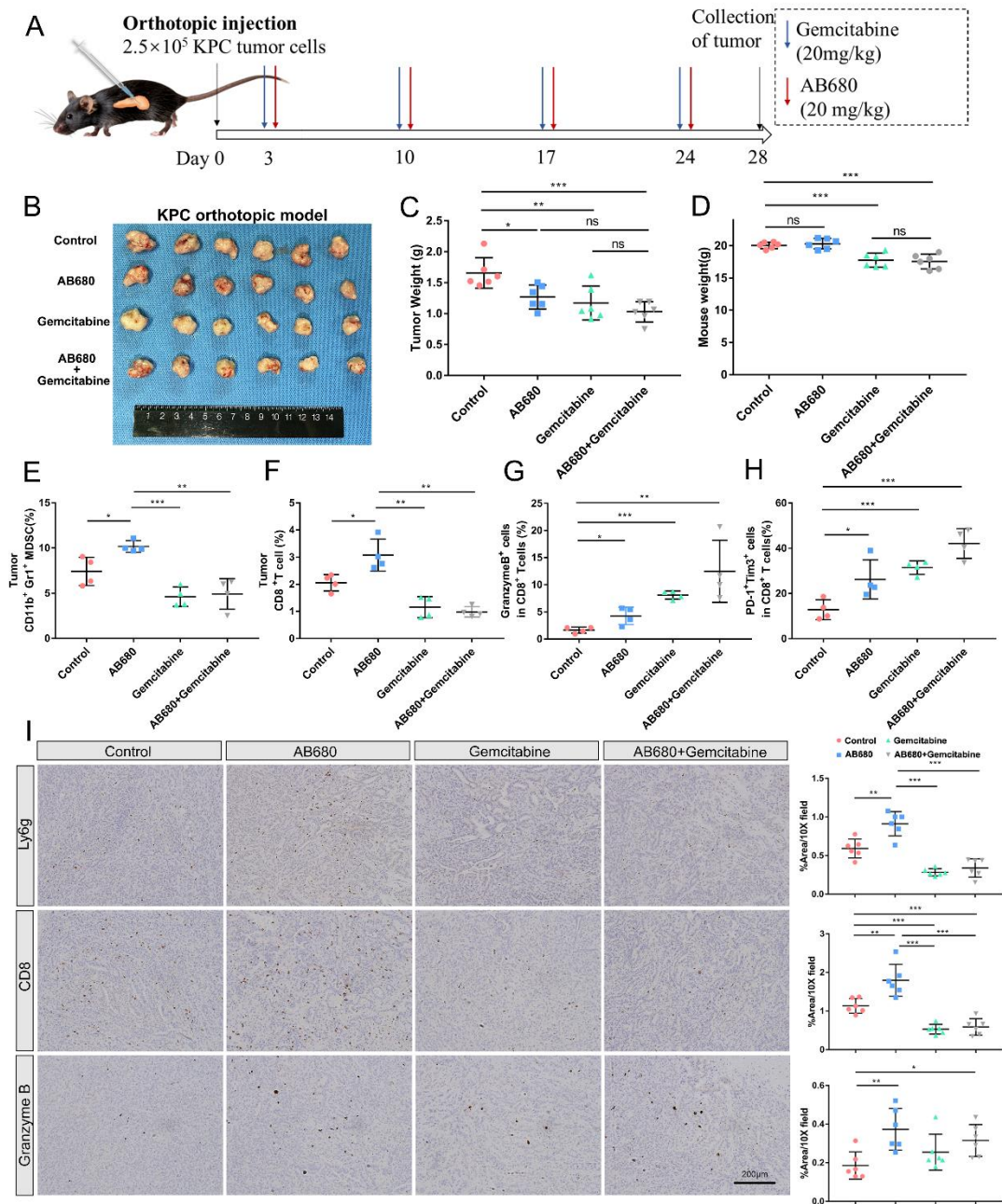
Supplementary Figure 4. AB680 has no anti-tumor effect in nude mice and *in vitro*. (A) Schematic showing the schedule of CD73 inhibitor AB680 treatment in KPC subcutaneous model of nude mice. (B-C) Image (B) and weights (C) of the AB680-treated vs Control KPC orthotopic tumors in nude mice (n=6 samples per group). Data presented as mean ± SD. ns = no significance in Student's t-test. (D) Kaplan-Meier survival analysis of tumor-bearing nude mice untreated or treated with 20 mg/kg AB680 (n=8 samples per group). ns, no significance in the log-rank test. (E-F) CCK-8 assay at 1, 2, and 3 days for cell proliferation after the indicated concentration of AB680 stimulation in murine (E) and human (F) pancreatic cancer cell lines. n=4 samples per group. (G-H) Clone formation assay at 1, 2, and 3 days for cell proliferation after the indicated concentration of AB680 stimulation in murine (G) and human (H) pancreatic cancer cell lines.



Supplementary Figure 5. AB680 alters the tumor microenvironment. (A) The t-distributed stochastic neighbor embedding (t-SNE) plot of CD45⁺ tumor-infiltrating immune cells of the total six samples (left), control and AB680-treated samples (right). (B) Heatmap of 42 markers in each cell cluster. (C) The frequencies of each cell cluster among the CD45⁺ cells in the two groups (n=3 samples per group).

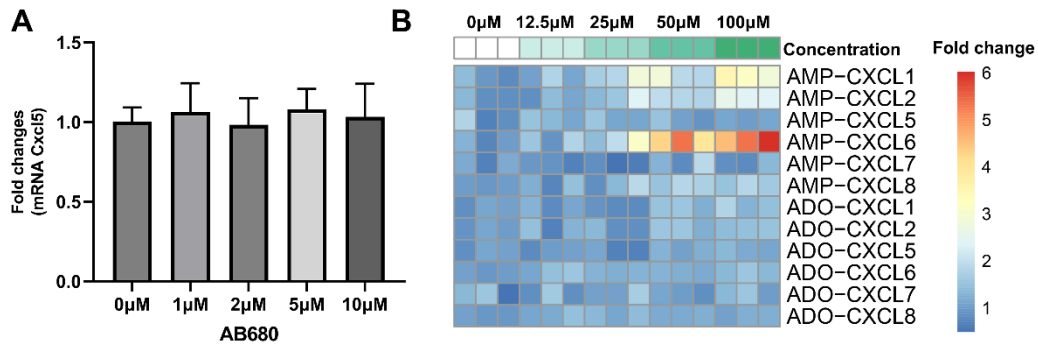


Supplementary Figure 6. CD8⁺ T cells are the key mediators for the antitumor response of AB680. (A) Flow cytometry analysis showing the depletion of CD8⁺ T cells in the spleen undergoing treatment with anti-CD8 antibody. (B) The gross appearance of the orthotopic KPC tumors from the Control(n=6), AB680(n=6), anti-CD8(n=5), and AB680+anti-CD8(n=5) groups. (C) Quantification of CD8 % positive area in the tumors of each group at the endpoint. **p<0.01, ***p<0.001 in Student's t-test. Data presented as mean ± SD. (D) Representative images of immunohistochemistry staining of CD8 in the KPC tumors from the control, AB680, anti-CD8, and AB680+anti-CD8 groups. Scale bar: 200µm.



Supplementary Figure 7. Combination of AB680 and gemcitabine cannot enhance the effect to suppress tumor growth. (A) Schematic representation of the therapy schedule for AB680 and gemcitabine in the KPC orthotopic model. (B-D) Image (B), tumor weight (C), and body weight (D) of KPC orthotopic tumors from the indicated treatment groups (n=6 samples per group). Data presented as mean \pm SD. ns = no significance, *p<0.05, **p<0.01, ***p<0.001 in Student's t-test. (E-H) The percentages of CD11b⁺Gr1⁺MDSCs (E), CD8⁺ T cells (F), Granzyme B⁺ (G), and PD1⁺TIM3⁺ cells (H) among the total CD8⁺ T cells in the tumors of each group at the endpoint determined by flow cytometry (n=4 samples per group). Data presented as mean \pm SD. *p<0.05, **p<0.01, ***p<0.001 in Student's t-test. (I) Representative images and quantification of immunohistochemistry staining of Ly6G, CD8, and Granzyme B in the KPC tumors

from the control, AB680, Gemcitabine, and AB680+Gemcitabine groups. Scale bar: 200 μ m. n=6 samples per group. Data presented as mean \pm SD. * p <0.05, ** p <0.01, *** p <0.001 in Student's t-test.



Supplementary Figure 8. Regulation of the indicated chemokine following different concentrations of drugs. (A) qRT-PCR analysis of Cxcl5 mRNA expression in the KPC cells with different concentrations of AB680 at 0, 1, 2, 5, and 10 μ M (n=3). (B) qRT-PCR analysis of the ligands of CXCR2 (CXCL1, CXCL2, CXCL5, CXCL6, CXCL7, and CXCL8) mRNA expression in the PANC1 cells with different concentrations of AMP and ADO at 0, 10, 20, 50, and 100 μ M (n=3). ADO, adenosine.

6	143Nd	CD366(Tim-3)	RMT 3-23	Biolegend	119702
7	144Nd	T-bet	4B10	BioLegend	644802
8	145Nd	CD152(CTLA-4)	UC10-4B9	Biolegend	106302
9	146Nd	CD206(MMR)	C068C2	Biolegend	141702
10	147Sm	Ly-6G	1A8	Biolegend	127602
11	148Nd	Ly-6C	HK1.4	Biolegend	128002
12	149Sm	CD19	6D5	Biolegend	115502
13	150Nd	CD127(IL-7R α)	A7R34	Biolegend	135002
14	151Eu	CD62L	MEL-14	Biolegend	104402
15	152Sm	CD11c	N418	Biolegend	117302
16	153Eu	CD44	IM7	Biolegend	103002
17	154Sm	CD73	TY/11.8	Biolegend	127202
18	155Gd	CD223 (LAG-3)	C9B7W	Biolegend	125202
19	156Gd	GITR(TNFRSF18)	2375B	R&D	MAB52412-100
20	157Gd	CD39	5F2	Biolegend	135702
21	158Gd	TCR γ / δ	GL3	Biolegend	118140
22	159Tb	F4/80	C1:A3-1	BioRAD	MCA497G
23	160Gd	CD274(B7-H1,PD-L1)	10F.9G2	Biolegend	124302
24	161Dy	TIGIT(VSTM3)	2190A	R&D	MAB72671
25	162Dy	CD183(CXCR3)	CXCR3-173	Biolegend	126502
26	163Dy	CD25	3C7	Biolegend	101902
27	164Dy	CD272	6A6	Biolegend	139117
28	165Ho	CD278(ICOS)	C398.4A	BioLegend	313502
29	166Er	CD194(CCR4)	2G12	Biolegend	131202
30	167Er	CD134(OX40)	OX-86	Biolegend	119429
31	168Er	FOXP3	FJK-16s	eBioscience	14-5773-82
32	169Tm	CD47	Miap301	Biolegend	127502
33	170Er	CD161(NK-1.1)	PK136	Biolegend	108702
34	171Yb	CX3CR1	SA011F11	Biolegend	149002
35	172Yb	CD279(PD-1)	29F.1A12	Biolegend	135202
36	173Yb	Granzyme B Fluidigm	GB11	Fluidigm	3173006B
37	174Yb	IFN- γ	XMG1.2	Bio-Xcell	BE0055
38	175Lu	TCR β chain	H57-597	Biolegend	109202
39	176Yb	TNF- α	MP6-XT22	Biolegend	506302
40	197Au	CD4	RM4-5	Biolegend	100520
41	198Pt	CD8a	53-6.7	Biolegend	100746
42	209Bi	CD11b	M1/70	BioLegend	101202