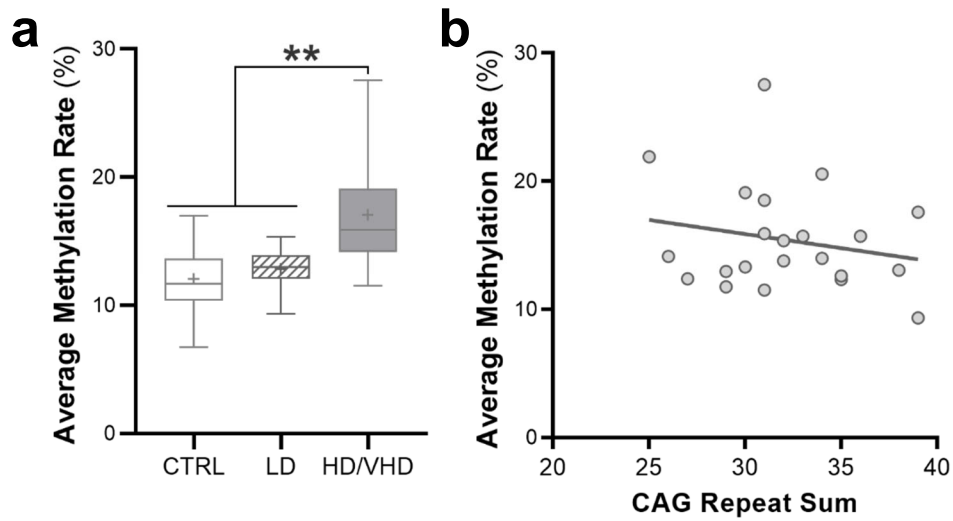


# Supplemental Figure 1



**Supplemental Fig. 1.** Averaged methylation rate (%) grouped by drinking class for all CpGs in male and female rhesus macaques. **a** The methylation rate was significantly higher in the HD/VHD (N = 15) compared to control (N = 13) and LD (N = 9) macaques ( $F(2, 34) = 9.860$ ,  $p = 0.0004$ , Tukey post-hoc,  $**p < 0.01$ ). **b** Lack of correlation between average methylation rate and CAG repeat sum ( $p = 0.3563$ )

# Supplemental Figure 2

	Hs 154,869,051	CCTTTGAGTAC	154,869,041
	Rm 129,130,369	CCTTTGAGTAC	129,130,359
	Mm 89,521,405	CCTTTGAGTAC	89,521,395
Hs 154,869,042	AAACCCCAAGAGAGCT <sup>*</sup> CGGTCTCTATCACCATAACAACAATTCCAAACATCCCAAAAATC		154,869,101
Rm 129,130,360	AAACCCCAAGAGAGCT <sup>*</sup> CGGTCTCTATCACCATAACAACAATTCCAAACATCCCAAAAATC		129,130,419
Mm 89,521,394	AAACCCCAAGACAGTT <sup>*</sup> CGGTCTCTATCACCATAACAACAATTCCAAACATCCCAAAAATC		89,521,335
Hs 154,869,102	AGAGCATAGTCACTCAGT <sup>*</sup> CGCTTTCTCTTTTCAAACAGGGCCCTCCTGTGTCCAGCTTA		154,869,161
Rm 129,130,420	AGAGCATAGTCACTCAGT <sup>*</sup> CGCTTTCTCTTTTCAAACAGGGCCCTCCTGTGTCCAGCTTA		129,130,479
Mm 89,521,334	AGAGCATAGTCACTCAGT <sup>*</sup> CGCTTTCTCTTTTCAAACAGGGCCCTCCTGTGTCCAGCTTA		89,521,275
Hs 154,869,162	TAGCCAATGTTTTGGTTTTT <sup>*</sup> CGCTTGTTGGCTTTGGGGAAGGTGGTGCTGCTGG <sup>*</sup> CGGTG		154,869,221
Rm 129,130,480	TAGCCAATGTTTTGGTTTTT <sup>*</sup> CGCTTGTTGGCTTTGGGGAAGGTGGTGCTGCTGG <sup>*</sup> CGGTG		129,130,539
Mm 89,521,274	TAGCCAATGTTTTGGTTTTT <sup>*</sup> CGCTTGTTGGCTTTGGGGAAGGTGGTGCTGCTGG <sup>*</sup> CGGTG		89,521,215
Hs 154,869,222	GTGC <sup>*</sup> CGGCATGCTGGTGGTTGTGGGTGGCATTAGGGTGATGGAGCAGGGTCTGGTGGGCA		154,869,281
Rm 129,130,541	GTGC <sup>*</sup> CGGCATGCTGGTGGTTGTGGGTGGCATTAGGGTGATGGAGCAGGGTCTGGTGGGCA		129,130,599
Mm 89,521,214	GTGC <sup>*</sup> CGGCATGCTGGTGGTTGTGGGTAGC <sup>*</sup> TTGGGGTGATGGAGCAGAGTCTGGTGGGCA		89,521,155
Hs 154,869,282	TGGTTGTCTCC <sup>*</sup> CGGGAGGAGATGAC <sup>*</sup> CGATCTC <sup>*</sup> CGGGGGGTTGCTAGGGCTGAAAAGCTGG		154,869,341
Rm 129,130,600	TGGTTGTCTCC <sup>*</sup> CGGGAGGAGATGAC <sup>*</sup> CGATCTC <sup>*</sup> CGGGGGGTTGCTAGGGCTGAAAAGCTGG		129,130,659
Mm 89,521,154	TGGTTATCTCC <sup>*</sup> CGGGAGGAGATGATAATCTCTGGGGGGTTGCTGGGACTGAAGAGCTGG		89,521,095
Hs 154,869,342	AGGGGTTGGCCCTCAGTCT <sup>*</sup> CGGCCT <sup>*</sup> CGATGAGGTTCTC <sup>*</sup> CGGGAGG <sup>*</sup> CGCTGAGG <sup>*</sup> CGGCTG		154,869,401
Rm 129,130,660	AGGGGTTGGCCCTCAGTCT <sup>*</sup> CGGCCT <sup>*</sup> CGATGAGGTTCTC <sup>*</sup> CGGGAGG <sup>*</sup> CGCTGAGG <sup>*</sup> CGGCTG		129,130,719
Mm 89,521,094	AGGGGTTGGCCCTCAGGCTCAGCCT <sup>*</sup> CGATAAGGTTCTC <sup>*</sup> CGAGAGG <sup>*</sup> CGCTGAGG <sup>*</sup> CGGCTG		89,521,035
Hs 154,869,402	AGGGGCTTCATGACCCAC <sup>*</sup> CGCTATACTTGCAGGAGCTCATGG <sup>*</sup> CGATCTC <sup>*</sup> CGTGAAGGGG		154,869,461
Rm 129,130,720	AGGGGCTTCATGACCCAC <sup>*</sup> CGCTATACTTGCAGGAGCTCATGG <sup>*</sup> CGATCTC <sup>*</sup> CGTGAAGGGG		129,130,779
Mm 89,521,034	AGGGGCTTCATGACCCAC <sup>*</sup> CGCTGATTTGCAGGAGCTCATAGCTATCTC <sup>*</sup> CGTGAAGGGA		89,520,975
Hs 154,869,462	TTGCTGTCC <sup>*</sup> CGCCGGTGCACCAGGGGGCTGGCCTGT <sup>*</sup> CGGTGCC <sup>*</sup> CGGCTGCCTC <sup>*</sup> CGCCAGGC		154,869,521
Rm 129,130,780	TTGCTGTCC <sup>*</sup> CGCCGGTGCACCAGGGGGCTGGCCTGT <sup>*</sup> CGGTGCC <sup>*</sup> CGGCTGCCTC <sup>*</sup> CGCCAGGC		129,130,839
Mm 89,520,974	TTGCTGTCC <sup>*</sup> CGCCGGTGCACCAGGGGGCTGGCCTGT <sup>*</sup> CGGTGCC <sup>*</sup> CGGCTGCCTC <sup>*</sup> CGCCAGGC		89,520,915
Hs 154,869,522	CCAATTGTAGCTGTGGAACCTGGAGAGTGGCCAAGCAAGTGGTCATTGAGATTGAGCTGG		154,869,581
Rm 129,130,840	CCAATTGTAGCTGTGGAACCTGGAGAGTGGCCAAGCAAGTGGTCATTGAGATTGAGCTGG		129,130,899
Mm 89,520,914	CCAATTGTGGCTGTGGAACCTGGAGAGTGGCCAAGCAAGTGGTCATTGAGATTGAGCTGG		89,520,855
Hs 154,869,582	CTGCCTTGCCTGGAGGAAGGGTGGAGGATGG <sup>*</sup> CGGTGGAGTTGGAC <sup>*</sup> CGAAGGGGGGGCCCTG		154,869,641
Rm 129,130,900	CTGCCTTGCCTGGAGGAAGGGTGGAGGATGG <sup>*</sup> CGGTGGAGTTGGAC <sup>*</sup> CGAAGGGGGGGCCCTG		129,130,959
Mm 89,520,854	CTGCCTTGCCTGGAGGAAGGGTGGAGGATGG <sup>*</sup> CGGTAGAGTTGGCTGAAGTGGGGGGCCCTG		89,520,795
Hs 154,869,642	AAAG <sup>*</sup> CGGTGGGAGAGGAGTGCAGCAGGCCAGGGTGGACGGGCTGGCTCTGGAGTTGGGCG		154,869,701
Rm 129,130,960	AAAG <sup>*</sup> CGGTGGGAGAGGAGTGCAGCAGGCCAGGGTGGACGGGCTGGCTCTGGAGTTGGGCG		129,131,019
Mm 89,520,794	AACGCGTGGGAGAAGAGTGCACAGACC <sup>*</sup> AGGATGGAC <sup>*</sup> AAGCTGGCTCTGGAGTTGGGCA		89,520,735

**Supplemental Fig. 2.** Sequence homology of MR-ex 1 (exon 1/intron 1) across human, macaque and mouse. The start and stop of the differentially methylated region spanning 646 base pairs is shown by green and red arrows, respectively. Conserved CpGs within MR-ex1 are shown in bold, red font. CpGs unique to a species are underlined. Nucleotides that are not conserved across species are shown in italic. “\*” and “\*” are CpGs that are significantly different in HD/VHD monkeys and CIE-exposed drinking mice, respectively.

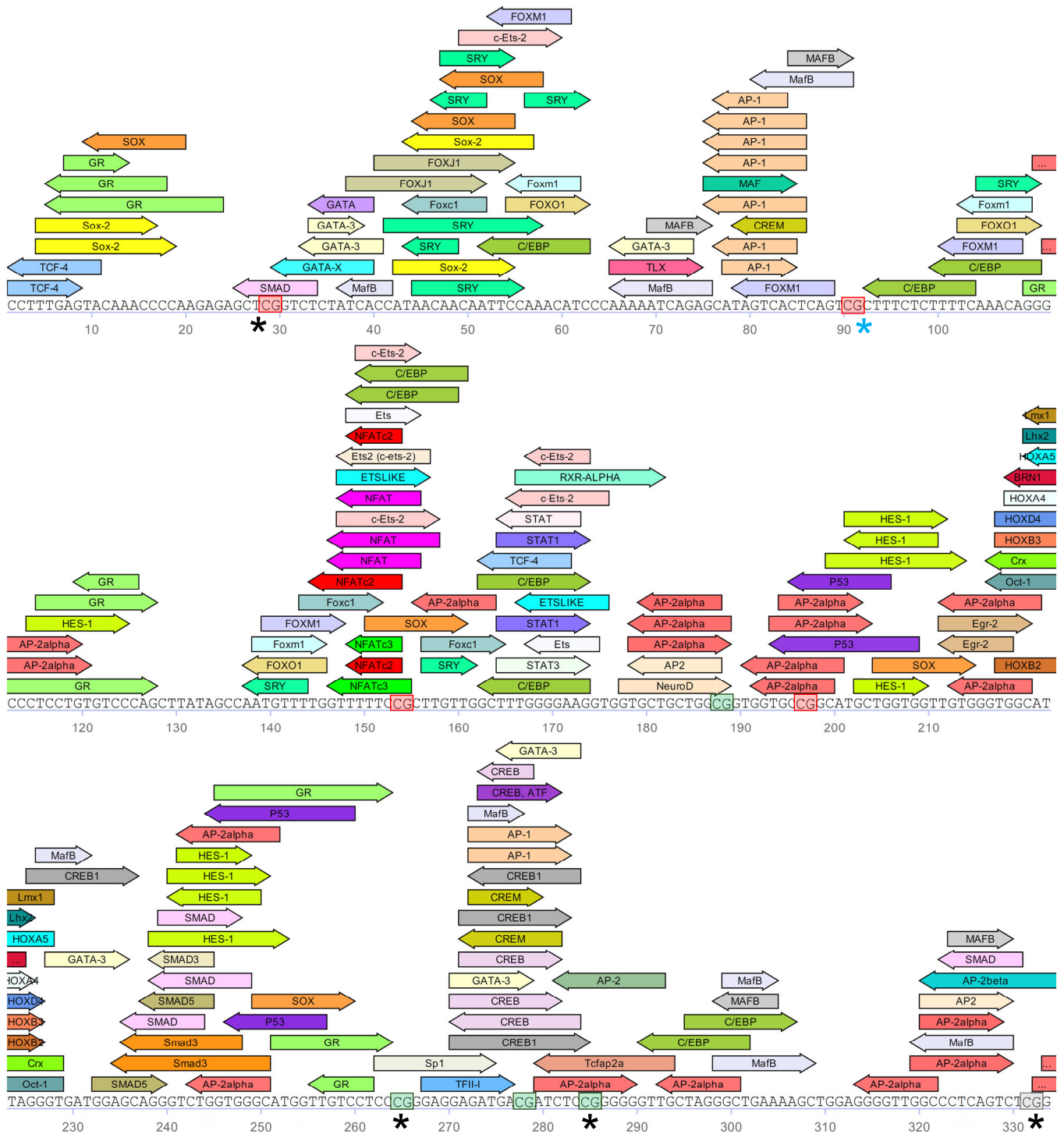
# Supplemental Figure 3

## *KCNN3* MR-ex1



**Supplemental Fig. 3.** Overlap of the macaque homologous methylation region within MR-ex1 on the human Roadmap 25 chromatin states. The MR-ex1 region on the human genome (hg19) is located at chr1: 154,841,507-154,842,152. According to the Roadmap Epigenomic project, the orange color in the 25 chromatin states represents “promoter upstream transcription start site”. Note that the nucleus accumbens is not included in the Epigenomics Roadmap database.

# Supplemental Figure 4

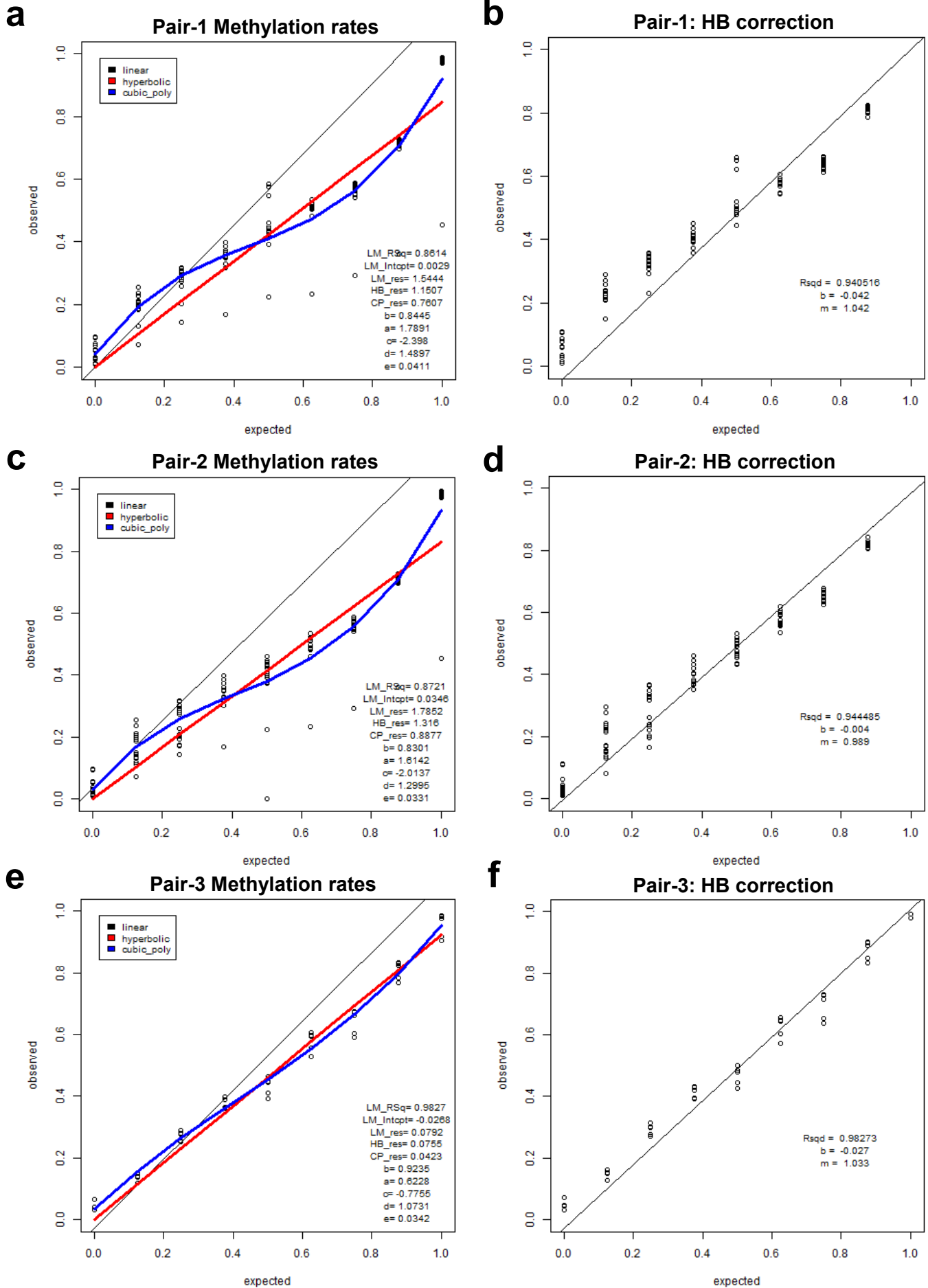


**Supplemental Fig. 4.** Predicted transcription factors that bind to the human MR-ex1 (TRANSFAC). CpGs enclosed in a red box are conserved in human, monkey and mouse; those CpGs unique to human are enclosed in a grey box, and those conserved in human and monkey are enclosed in a green box. '\*' and '\*\*' are CpGs that are significantly different in HD monkeys and CIE-exposed drinking mice, respectively. TRANSFAC conditions were as follows: nerve\_system\_specific matrices, matrix similarity cut-off: 0.9 and core similarity cut-off: 0.95.

# Supplemental Figure 4, cont.



# Supplemental Figure 5



**Supplemental Fig. 5. Titration curves to correct for PCR bias during amplification of the methylation region under study.** Three different amplicons were used to amplify the complete region 1 (a), 2 (c) and 3 (e). PCR-bias was corrected using the hyperbolic function (HB, b, d and f).