Supporting Information

for Environmental Science & Technology

Reactivity of bromine radical with dissolved organic matter moieties and monochloramine: Effect on bromate formation during ozonation

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This **Supporting Information** includes 18 texts, 10 tables, 21 figures, and 1 scheme in 45 pages.

Table of contents

| Text S1. Chemical reagents and stock solutions | S6 |
|--|----------------|
| Text S2. Preparation of DOM stock solution | S6 |
| Text S3. Preparation of chloride-free monochloramine (NH ₂ Cl) stock solutions | S6 |
| Text S4. Dosimetry | S7 |
| Text S5. Sample preparation for kinetic experiments performed with γ-radiolysis | S8 |
| Text S6. Dissolved O_2 in γ -radiolysis samples | S9 |
| Text S7. Masking bromide by Ag ⁺ | S9 |
| Text S8. Adapted competition kinetics for determining k_{Br} of DOM and NH ₂ Cl | S10 |
| Text S9. Sample preparation for γ-radiolysis for phenol-Br• reaction experiments | S10 |
| Text S10. Chromatographic methods | S11 |
| Text S11. Ozonation experiment | S12 |
| Text S12. Quantum chemical calculation methods | S12 |
| Text S13. Validation of quantum chemical calculation methods | S13 |
| Text S14. Calculation of free energy of activation by the Marcus theory | \$13 |
| Text S15. LC-HRMS/MS evidences supporting the formation of $C_6H_3BrO_3$ and $C_6H_5BrO_3$ for t | the phenol-Br• |
| reaction | S14 |
| Text S16. Formation pathway of 4-bromophenol from the phenol-Br [•] reaction | S15 |
| Text S17. Principles of the chlorine-ammonia pretreatment as a bromate mitigation strate | gy of |
| ozonation | S16 |
| Text S18. Calculation of the fractions of Br*-related reactions during ozonation | S16 |
| Table S1. Chemical reagents | S18 |
| Table S2. Compounds analyzed by HPLC and LC-HRMS/MS and the corresponding analytica | al condition |
| and LOQ. | S19 |
| Table S3. Detected exact masses and mass deviation by LC-HRMS/MS | S19 |
| Table S4. Theoretically calculated $\Delta G_{ m solv,calc}$ values and experimental values of halides and i | halogen |
| radicals in kcal/mol. n is the number of explicit water molecule(s) | S20 |

Table S10. Scavenging rates ($k_{species}$ ', s^{-1}) of e^- , ${}^{\circ}OH$, ${}^{\circ}H$, Br° , and ${}^{\circ}tBA$ (t-butanol derived radical) by the reaction species in γ -radiolysis kinetic experiments (1,2-dibromoethane (1,2-DBE), Ag⁺, t-butanol, a model compound, phosphate ions ($H_2PO_4^-/HPO_4^{2-}$ for pH 7), and dissolved O_2), based on the reported or assumed second-order rate constants ($k_{species}$, $M^{-1}s^{-1}$) and the applied concentrations (conc., M). ... S27

Figure S6. Slopes obtained by linear regression of data points of the competition kinetics plots (shown in Figure S5) as a function of a reciprocal of the DOC or NH₂Cl concentration, to derive k_{Br} for NH₂Cl, DOM, and oxidized DOM (by 0.8 gO₃/gC or 1.5 gO₃/gC). See Text S8 for the corresponding rate expression.S34

Figure S9. Correlation between the measured k_{Br} of the selected organic model compounds and

aqueous phase free energies for the formation of adducts ($\Delta G^{\text{adduct}}_{aq}$)......S35

Figure S10. Concentrations of phenol and the identified products as a function of the γ -radiolysis time during the reaction of phenol with Br[•], for the condition with 22 μ M phenol, 0.7 mM 1,2-dibromoethane, 40 mM *t*-butanol, and 50 mM phosphate buffer (pH 7.1). See Figure 2 (main text) for

Figure S12. Trends of the peak intensity as a function of the γ -radiolysis time for the exact masses of 200.9193 (blue circles, corresponding to C₆H₂⁷⁹BrO₃ as M-H) and 202.9349 (orange triangles, corresponding to C₆H₄⁷⁹BrO₃ as M-H) during the reaction of Br[•] with phenol (left) or *p*-benzoquinone (right).

Figure S13. Left: Filtered chromatograms of a γ -radiolysis sample (t = 40 min); right: synthesized substituted *p*-benzoquinone with the exact masses of 200.9193, 202.9172, 202.9349, and 204.9329 (from top to bottom of the chromatograms) as [M-H], corresponding to the molecular formulas C₆H₃⁷⁹BrO₃, C₆H₃⁸¹BrO₃, C₆H₅⁷⁹BrO₃, and C₆H₅⁸¹BrO₃. Retention times for C₆H₃BrO₃, and C₆H₅BrO₃ are 7.8

and 6.1 min for the γ-radiolysis sample (left) and 11.9 and 8.7 min for the synthesized chemical (right).

Figure S17. Resonance structures of the phenoxyl radical......S39

Figure S18. Calculated fractions of Br[•] reacting with DOM (green line), ozone (red line), bromide (blue line), or NH₂Cl (orange line) as a function of the ozone concentration in (a) absence or (b) presence of NH₂Cl. The selected concentrations were 5 mgC/L DOC, 1.3 μ M Br⁻ (100 μ g/L Br⁻), and 15 μ M NH₂Cl.S40

Text S1. Chemical reagents and stock solutions

The chemical reagents used in this study are summarized in Table S1. Hypochlorite solution was standardized spectrophotometrically ($\epsilon_{290nm} = 350 \text{ M}^{-1} \text{ cm}^{-1}$).¹ Dissolved organic matter (DOM) and monochloramine (NH₂Cl) stock solutions were prepared as described in Text S2 and Text S3, respectively. All solutions were prepared in ultrapure water with a resistivity of 18.2 M Ω ·cm. Lake Water Zurich was collected at Zurich Water Supply (Lengg) and filtered on the same day of collection with a cellulose nitrate filter (0.45 µm, Sartorius Stedim Biotech). General water quality parameters of the filtered Lake water were analyzed by the AuA laboratory (Eawag, Dübendorf) and summarized as follows: pH 8.3, alkalinity 2.69 mM, hardness 1.49 mM, bromide < 0.05 mg/L, nitrate 0.7 mg/L as NO₃⁻-N, ammonium < 5.0 µg/L as NH₄⁺-N, nitrite < 1.0 µg/L as NO₂⁻-N, and DOC 1.4 mg/L.

Text S2. Preparation of DOM stock solution

A 50 mgC/L DOM stock solution was prepared by dissolving the corresponding amount of Suwannee River Fulvic Acid II (2S101F, IHSS, 52.34%(w/w), <u>https://humic-substances.org/</u>) in 20 mM phosphate buffer at pH 7. The stock solution was standardized based on UV absorbance at 254nm after diluting 5 times.² An ozonated DOM stock solution was prepared with two ozone doses (0.8 gO₃/gC and 1.5 gO₃/gC) and ozone was completely consumed over night. Due to the dilution, the final concentrations of the oxidized DOM stock solutions were 33 mgC/L (0.8 gO₃/gC) and 26 mgC/L (15 gO₃/gC). To control the extent of oxidation, the electron donating capacity (EDC) of the DOM stock solutions was measured before and after ozonation by an EDC assay using the radical cation of 2,2'-azino- bis(3ethylbenzothiazoline-6-sulfonate) (ABTS^{*+}) as described by Walpen et al. (2020).³ The decrease in EDC of the DOM stock solution after ozonation was (31 ± 5)% for 0.8 gO₃/gC and (40 ± 4)% for 1.5 gO₃/gC, in a similar range of previously reported EDC of ozonated DOM solutions.^{4,5}

Text S3. Preparation of chloride-free monochloramine (NH₂Cl) stock solutions

Hypochlorite solution contains high concentrations of chloride from the manufacturing process. The presence of chloride in NH₂Cl stock solution would interfere determining k_{Br} of NH₂Cl, because chloride can react fast with Br^{*} as shown in Eq. S1.

Br[•] + Cl⁻
$$\leftrightarrow$$
 BrCl^{•-}
 $k_{+} = 2.3 \times 10^{8} \text{ or } 1.0 \times 10^{10} \text{ M}^{-1} \text{ s}^{-1}$ Eq. S1 ^{6,7}
 $k_{-} = 6.1 \times 10^{4} \text{ or } 8.5 \times 10^{7} \text{ s}^{-1}$

To prevent this interference, chloride in prepared NH₂Cl stock solution was removed by a following description. A primary NH₂Cl stock solution was prepared by mixing 15 mM of sodium hypochlorite (at

pH 9.5 by NaOH) with 7.5 mM of diammonium sulfate at a mixing ratio of 1:1 by a syringe pump with a speed of 1 mL/min. The resulting solution produced (6.9 ± 0.1) mM NH₂Cl as a main species with a negligible impurity of NHCl₂ (up to 0.1 mM), determined by UV absorbance at 245 and 295 nm and the corresponding molar absorptivity coefficients ($\epsilon_{245,NH2Cl}$ = 445 M⁻¹ cm⁻¹, $\epsilon_{245,NHCl2}$ = 208 M⁻¹ cm⁻¹, $\epsilon_{295,NH2Cl}$ = 14 M⁻¹ cm⁻¹, $\epsilon_{295,NHCl2}$ = 267 M⁻¹ cm⁻¹).⁸

To prepare a chloride-free NH_2Cl stock solution, the amount of chloride (Cl^-) present in the primary NH_2Cl stock solution was first determined by diluting the stock solution to 0.4 – 0.5 mM NH_2Cl , subsequently reducing NH₂Cl to Cl⁻ with sulfite, and measuring a total Cl⁻ concentration (as a sum of Cl⁻ present in the diluted stock and Cl⁻ reduced from NH₂Cl) by ion chromatography (Text S10). NH₂Cl was completely reduced by 1.0 - 1.5 molar equivalent of sulfite relative to NH₂Cl (Figure S1). The total [Cl⁻] was 1.4 - 1.6 mM in the diluted NH₂Cl stock solutions and therefore the [Cl⁻] already present in the solutions was 1.0 - 1.2 mM. After determining [Cl⁻], the primary NH₂Cl stock solution was again diluted to 0.4, 1.7, and 3.5 mM NH₂Cl and a 1.1 molar equivalent of silver (Ag⁺) with regard to the inherent [Cl⁻] was added to the diluted NH₂Cl stock solutions to remove Cl⁻. The silver-spiked solutions were allowed to sediment the precipitate (AgCl) for > 6 h. Afterwards, the supernatant was taken to measure [NH₂Cl] by a DPD method⁹ to check an effect of the silver addition on $[NH_2CI]$. The determined $[NH_2CI]$ by DPD were within \pm 3% from the theoretical [NH₂Cl] (except for the stock solution with the lowest [NH₂Cl], 0.4 mM, deviated by 27%) (Figure S2a), and thus the effect was negligible. The supernatant was also taken to measure total [Cl⁻] by IC, by diluting the 0.4, 1.7, and 3.5 mM NH₂Cl stock solutions to 0.2 mM NH₂Cl and reducing NH₂Cl by sulfite. The measured [Cl-] were 0.18 - 0.19 mM, close to the theoretical [NH₂Cl] (0.2 mM) (Figure S2b), indicating that only limited Cl^- is present in the supernatant of the NH₂Cl stock solutions. As a result, the supernatant of the silver-spiked NH₂Cl stock solution was used for determining $k_{\rm Br}$.

Text S4. Dosimetry

The dose rate was determined by an air-saturated 1 mM formate solution prepared in 2 mM phosphate buffer at pH 6.5 where all reactive species (solvated electron (e^-), *OH, H*) are converted to hydrogen peroxide.¹⁰ Hydrogen peroxide formed in the solution over time (0 – 15 min) was quantified spectrophotometrically by the Allen's reagent¹¹ and used to calculate the dose rate. The calculated dose rate was 0.13 kGy/h, corresponding to the electron formation rate ($fr(e^-)$) of 9.7 nM/s and the Br* formation rate ($fr(Br^*)$) of 9.7 nM/s by assuming a 100% conversion of e^- to Br* (from the reaction of e^- with 1,2-dibromoethane).

Text S5. Sample preparation for kinetic experiments performed with y-radiolysis

γ-radiolysis of aqueous solution produces e^- , 'OH, H', and H₂O₂ as reactive species with e^- and 'OH being the major species. For generating Br', 1,2-dibromoethane was used, which forms Br' by the reaction with e^- , according to the Eqs. S2 and S3. The other major reactive species, 'OH, was quenched by *t*-butanol to avoid interferences for the oxidation of probe compounds and guarantee a high yield of Br' from the reaction of 1,2-dibromoethane with e^- . The scavenging of e^- by 1,2-dibromoethane was at least 84% (depending on the dissolved O₂ concentration) of the total scavenging rate of e^- (Table S10). BrCH₂CH₂Br + e^- → BrCH₂CH^{*}₂ + Br⁻ $k = 1.4 \times 10^{10}$ M⁻¹s⁻¹ Eq. S2¹²

BrCH₂CH₂ \rightarrow CH₂CH₂ + Br[•] $k = (0.6 - 3.8) \times 10^{6} \text{ s}^{-1}$ Eq. S3 ¹³⁻¹⁵

The formed Br[•] is in equilibrium with hydroxide (BrOH^{•-} \leftrightarrow Br[•] + HO⁻, $K_{eq} = 3.2 \times 10^{-4}$ M)¹⁶ and therefore affected by pH. Under the pH conditions we applied (pH 7 and 10), the equilibrium clearly favors Br[•], with the fraction of Br[•] as 100% and 76%, respectively. The fraction of Br[•] as a function of pH calculated based on the reported K_{eq} is shown in Figure S21.

Sample preparation is illustrated in Figure S3 for a setup image and in Figure S4 as flow charts. Samples for most organic model compounds (except *p*-benzoquinone, benzene, toluene, and naphthalene) were prepared one day prior to γ -radiolysis. 100 mL of solution containing 4 μ M of ibuprofen, 4 μ M of an organic model/target compound, and 50 mM phosphate buffer (pH 7.1) or 50 mM borate buffer (pH 10.2) was saturated with Ar gas. The Ar-saturated solution was then delivered to six 1.5 mL amber vials containing 0.21 mL of air-saturated solution of 5 mM of 1,2-dibromoethane and 29 mM of *t*-butanol, via a transferring tubing under N₂ gas flow, roughly until the neck of each vial (see Figure S3 for setup image). By mixing of Ar- and air-saturated solutions a certain level of dissolved oxygen (see below) could be achieved for the reasons described in Text S6. After the transfer, the vials were immediately closed with crimp caps to minimize gas exchange with the atmosphere. Mixed concentrations of the reactants in the vials were 3.4 μ M ibuprofen, 3.4 μ M target compound, 0.7 mM 1,2-dibromoethane, and 4 mM *t*-butanol. Each vial represented a designated γ -radiolysis time (0, 4, 8, 12, 16, and 20 min).

Samples for experiments with *p*-benzoquinone were prepared according to the description above, except that *p*-benzoquinone was added to the sample solution on site just before γ -radiolysis to minimize its loss. 10 µL of a 0.6 mM *p*-benzoquinone solution was added to roughly 1.5 mL of the premixed solution containing ibuprofen, 1,2-dibromoethane, and *t*-butanol, using a glass syringe through the septum of the crimp caps. The weight of the vials was determined before and after adding *p*-

benzoquinone, to account for deviations in the final concentration of *p*-benzoquinone, caused by differences in sample volumes across the vials.

Samples for benzene, toluene, and naphthalene were prepared on site. 90 mL of a solution containing 3.3 μ M ibuprofen and 50 mM phosphate buffer (pH 7.1) was placed in a 150 mL reaction vessel and saturated with Ar. 10 mL of an air-saturated solution containing 5 mM 1,2-dibromoethane and 20 mM *t*-butanol and 300 μ L of 1 mM organic compound (benzene, toluene, or naphthalene) were added to the Ar-saturated solution and immediately placed in the γ -radiolysis source. The final concentrations of the reactants were 3 μ M ibuprofen, 3 μ M target compound, 0.5 mM 1,2dibromoethane, and 2 mM *t*-butanol. During γ -radiolysis, 1 mL samples were taken every 4 min up to 20 min for further analyses.

Text S6. Dissolved O_2 in γ -radiolysis samples

 O_2 concentrations were kept at a level to guarantee quenching of carbon-centered radicals derived from the reaction of *t*-butanol with *OH (*tBA) during the radiolysis, which may oxidize target aromatic compounds,¹⁷ but still low enough not to quench too much *e*⁻. The desired O_2 level was achieved by mixing Ar- and air-saturated solutions with a volumetric ratio of 6:1. O_2 concentrations were measured by an optical O_2 sensor (PreSens, Oxygen Microsensor NTH-PSt7). The average of O_2 concentrations measured immediately after the mixing were $(2.7 \pm 0.3) \% O_2$ saturation (equivalent to $(35 \pm 4) \mu M O_2$), which slightly increased to $(4.0 \pm 1.1) \% O_2$ saturation (or $(52 \pm 14) \mu M O_2$) after 15 hours storage at room temperature. For the O_2 concentration of ~70 μM (highest possible O_2), the scavenging of *e*- by O_2 was 11% of the total scavenging rate of *e*-, while the scavenging of *tBA by O_2 was 92% of the total scavenging rate of *tBA (Table S10). The mixed solution was treated by γ -radiolysis within 24 hours to minimize reintroduction of O_2 .

Text S7. Masking bromide by Ag⁺

The bromine radical (Br[•]) was generated from the reaction of 1,2-dibromoethane with e^- (Eqs. S2 and S3). The reaction inherently forms Br⁻ as a side product (Eq. S2) which could quench Br[•] very efficiently (Eq. S4). To prevent this undesired Br[•] quenching, Br⁻ was masked by the fast reaction with silver(I) (Ag⁺, Eq. S5) by injecting 24 µL of 0.135 mM silver nitrate solution with a Hamilton glass syringe through a rubber septum of the 1.5 mL amber vials, after every 4 min of γ -radiolysis treatment (total 11 µM Ag⁺ added for 20 min of γ -radiolysis, equivalent to the expected concentration of Br⁻ formed). Ag⁺ was added in a distributed way to reduce a possibility of quenching Br[•] by Ag⁺ (note that $k_{Br•}$ of Ag⁺ is not

available, but assumed high according to the $k_{\bullet OH}$ of Ag⁺ as high as $10^{10} \text{ M}^{-1} \text{ s}^{-1}$).¹⁸ After the designated time (0 – 20 min with 4 min intervals), samples were taken out from the γ -radiation source and 40 μ L of 26.5 mM sodium chloride was added to the vial to quench potentially residual Ag⁺. The same bromide-masking protocol was applied for the product analysis samples prepared in 11 mL vials, but with higher concentrations of the silver nitrate and sodium chloride stock solutions, to account for the larger volumes of the vials (i.e., 1 mM silver nitrate and 192 mM sodium chloride).

Br[•] + Br⁻ ↔ Br[•]₂

$$k_{+} = 1.2 \times 10^{10} \text{ M}^{-1} \text{ s}^{-1}$$

 $k_{-} = 3.1 \times 10^{4} \text{ s}^{-1}$
Br⁻ + Ag⁺ → AgBr (s)
 $k = 1.5 \times 10^{10} \text{ M}^{-1} \text{ s}^{-1}$
Eq. S5 ²⁰

Text S8. Adapted competition kinetics for determining k_{Br} of DOM and NH₂Cl

 k_{Br} of DOM was determined by measuring a decrease in ibuprofen over time under varying conditions with ibuprofen (1 µM) and DOC concentration variation in the range of 2 – 15 mgC/L. To derive k_{Br} of DOM, a steady-state concentration of Br[•] ([Br[•]]_{ss}) was assumed, where the formation rate ($fr(Br^{•})$, governed by the dose rate of the γ -radiation source (Text S4)), is equal to the consumption rate as expressed in Eq. S6, with X as DOM. Eq. S6 can be rearranged for [Br[•]]_{ss}, which is then approximated to Eq. S7 for the condition where $k_{Br\bullet Ibu}$ [Ibu] << k_{Br} .X[X] (X = DOM). Finally, k_{Br} of DOM is derived by plotting a slope obtained by a linear regression of the In ([Ibu]/[Ibu]_0) over time data (as shown in Figure S5) as a function of 1/[DOM] (as shown in Figure S6), based on Eq. S8. k_{Br} of NH₂Cl was determined accordingly but with varying [NH₂Cl] in the range of 0.01 – 0.4 mM and with X as NH₂Cl in Eqs S6 – S8.

$$fr(Br^{\bullet}) = k_{Br^{\bullet},Ibu}[Br^{\bullet}]_{ss}[Ibu] + k_{Br^{\bullet},X}[Br^{\bullet}]_{ss}[X]$$
 Eq. S6

$$[Br^{\bullet}]_{ss} = \frac{fr(Br^{\bullet})}{k_{Br^{\bullet},Ibu}[Ibu] + k_{Br^{\bullet},X}[X]} \approx \frac{fr(Br^{\bullet})}{k_{Br^{\bullet},X}[X]}$$
Eq. S7

$$\ln\left(\frac{[\text{Ibu}]}{[\text{Ibu}]_0}\right) = -k_{\text{Br},\text{Ibu}}[\text{Br}^*]_{\text{ss}}t = -\frac{k_{\text{Br},\text{Ibu}}}{k_{\text{Br},\text{X}}}\frac{fr(\text{Br}^*)t}{[X]}$$
Eq. S8

Text S9. Sample preparation for y-radiolysis for phenol-Br[•] reaction experiments

Samples for product analyses for the reaction of phenol with Br[•] were prepared one day prior to γ radiolysis similarly to the kinetic experiments but in larger volumes. 100 mL solution containing 25 μ M of phenol and 50 mM phosphate buffer (pH 7.1) was saturated with Ar and brought to four 11 mL clear vials with 1.54 mL of air-saturated solution containing 5 mM of 1,2-dibromoethane and 286 mM of *t*- butanol, via the setup described above. The final concentrations of the reactants in the vials were 22 μ M phenol, 0.7 mM 1,2-dibromoethane, and 40 mM *t*-butanol. Ag⁺ for masking bromide was added as described in Text S7. Samples were taken after 12, 20, and 40 min and analyzed by HPLC-DAD (or HPLC-FLD) and LC-HRMS/MS within 24 hours.

Text S10. Chromatographic methods

All kinetic and product samples from γ -radiolysis were filtered by a syringe filter (Nylon, 0.45 μ m, 4 mm) prior to analyses. Most organic model compounds and phenol-Br[•] products (except those mentioned below) were analyzed by HPLC (Ultimate 3000, Thermo) equipped with diode array detector (DAD) and fluorescence detector (FLD) with a COSMOSIL 5C18-MS-II (3.0 × 150 mm, 5 μm) column using a gradient method with methanol and 10 mM H₃PO₄ as eluents. The gradient started at 45% methanol (i.e., 55% H₃PO₄), gradually increased to 95% methanol for 20 min, stayed at 95% methanol for 4 min, returned to 45% methanol in 5 min, and remained at 45% methanol for 5.5min (total analyzing time = 30 min). For hydroquinone and catechol, the same C18 column was used but with a different gradient: the gradient started at 5% methanol, increased to 95% for 25 min, stayed at 95% for 8.5 min, returned to 5% in 1 min, and remained at 5% for 5.5 min (total analyzing time = 40 min). For 3-phenylpropionic acid, the same C18 column was used but with different eluents (acetonitrile and 10 mM H₃PO₄) and a different gradient: the gradient started at 30% acetonitrile, increased to 60% for 13 min, stayed at 60% for 15 min, returned to 30% in 0.5 min, and remained at 30% for 6.5 min (total analyzing time = 36 min). For benzylamine and N,N-dimethylbenzylamine, a hydrophilic interaction liquid chromatography column (XBridge[™] BEH HILIC XP, 2.5 µm particle size, 3.0 × 150 mm, Waters) was used with an acetonitrile mixture (95:3:2 acetonitrile:methanol:water) and water as eluents. The gradient started at a 100% acetonitrile mixture for the first 2 min, decreased to 5% for 10 min, stayed at 5% for 5 min, returned to 100% in 1 min, and remained at 100% for 19 min (total analyzing time = 37 min). The flow rate was set at 0.4 mL/min for all analyses and the injection volume was 25 μ L. The detector setup is summarized in Table S2.

Identified and suspected products for the reaction of phenol with Br[•] were analyzed by LC coupled with high resolution tandem mass spectrometer (Q Exactive, Thermo Scientific). LC separation was carried out with the same C18 column as above with a gradient method using ultrapure water and methanol (both with 0.1% formic acid) as eluents A and B, respectively. The gradient started with 5% B, gradually increased to 95% B for 25 min, held at 95% B for 8.5 min, returned to 5% B for 1 min, and was maintained at 5% B for 5.5 min. The total analyzing time was 40 min. The flow rate was 0.4 mL/min and

the injection volume was 25 μ L. The MS detector was with electrospray ionization with either a full MS¹ scan mode (R = 35,000, mass range = 60 – 600 for all modes) with data-dependent MS² (R = 35,000) or a targeted MS² mode with an inclusion list specifying the masses of interest (corresponding to suspected phenol oxidation products) and a range of applied collision energies (15, 45, and 60 as (N)CE). Both modes were operated in positive or mostly negative polarity with a spray voltage of 4000 V (positive) or 3000 V (negative), a capillary temperature of 350 °C, a sheath gas flow rate of 40, and an auxiliary gas flow rate of 10. Detected exact masses of the phenol-Br[•] products and mass deviations are summarized in Table S3.

Samples containing chloride and bromate were first diluted with ultrapure water (100 times for chloride, 10 times for bromate) and analyzed by ion chromatography coupled with conductivity detector (Dionex[™] Integrion[™] HPIC[™] System) with an anion exchange IC column (Dionex[™] IonPac[™] AS19-4µm IC Column 2 × 250mm with AG19-4µm Guard Column 2 × 50mm) with a gradient of KOH as eluent, starting at 10 mM for 10 min, gradually increasing to 30 mM for 20 min, increasing to 100 mM for 0.1 min, staying at 100 mM for 6.9 min, decreasing to 10 mM for 0.1 min, and maintaining at 10 mM for the last 4.9 min. The total analyzing time was 30 min. The flow rate was 0.25 mL/min and the injection volume was 50 µL. Retention times were 9.6 min for chloride and 7.9 min for bromate. LOQ were ~0.02 µM for chloride and ~0.01 µM for bromate.

Text S11. Ozonation experiment

Ozone stock solutions were prepared by sparging ozone-containing oxygen gas in ultrapure water cooled with ice. The ozone/oxygen gas mixture was produced by an ozone generator (BMT 803 BT, BMT Messtechnik, Berlin) fed with > 99.995% oxygen. Ozone concentrations in the stock solution were typically 1.6 - 1.8 mM determined by UV absorbance ($\epsilon_{260nm} = 3200 \text{ M}^{-1} \text{ cm}^{-1}$).²¹

Text S12. Quantum chemical calculation methods

The aqueous-phase free energy of formation, G_{aa} , of all species was obtained based on equation S9:

$$G_{\rm aq} = \Delta G_{\rm solv,calc} + G_{\rm corr,gas} + E_{0,gas}$$
 Eq. S9

 $\Delta G_{\rm solv,calc}$ is the solvation free energy of the species calculated using an implicit solvation model (SMD)²² in the absence or presence of explicit water molecules. $G_{\rm corr,gas}$ is the gaseous phase correction to the free energy of the species solvated by explicit water molecules if explicit water molecules are

present. $E_{0,gas}$ is the electronic energy of the species solvated by explicit water molecules. The $\Delta G_{solv,calc}$ and $G_{corr,gas}$ values were calculated at the level of M06-2X/Aug-cc-pVDZ.²³ In particular, the $\Delta G_{solv,calc}$ values were calculated following the procedure of a continuum solvation method.²⁴ The $E_{0,gas}$ value was calculated at the level of M06-2X/Aug-cc-pVTZ.

Text S13. Validation of quantum chemical calculation methods

Theoretically calculated $\Delta G_{\text{solv,calc}}$ in the absence and presence of explicit water molecules (n=1-3) of Br[•] and Br⁻ were validated with experimentally measured or derived values. In the validation process, we included other halogen radicals and halides (i.e., F⁻, Cl⁻ and l⁻). Table S4 summarizes the theoretically calculated $\Delta G_{\text{solv,calc}}$ values and experimental values of halides and halogen radicals. The experimentally derived values of $\Delta G_{\text{solv,calc}}$ for halogen radicals (X[•], where X is Cl or Br) were determined by the equation below²⁵ where IE(gas) and IE(aq) are the adiabatic gaseous and aqueous-phase ionization energy of X⁻, respectively.

$$\Delta G_{\text{solv.calc}} (X^{\bullet}) = -\text{IE}(\text{gas}) + \Delta G_{\text{solv.calc}} (X^{\bullet}) + \text{IE}(\text{aq})$$
Eq. S10

The use of the hybrid DFT method, M06-2X, by validating the experimental one electron reduction potential of Br^{+}/Br^{-} by other quantum mechanical methods including CCSD(T)²⁶ and CBS-QB3^{27,28} and MP2 as summarized in Table S5. In addition to the validation of the one electron reduction potential for Br^{+}/Br^{-} , one the electron reduction potentials for benzene, toluene, and anisole are validated Table S6.

Text S14. Calculation of free energy of activation by the Marcus theory

Based on the validation, we used the G_{aq} values obtained at the level of M06-2X/Aug-cc-pVDZ//M06-2X/Aug-cc-pVTZ with 3 explicit water molecules. We used Spartan 18 (Wavefunction, Irvine, CA)²⁹ to search the conformers of an adduct comprised of Br[•] and a target organic compound using a modified MMFF method.³⁰ The aqueous-phase free energy of adduct formation, $\Delta G^{\text{adduct}}_{aq,calc}$, was calculated as below:

$$\Delta G^{\text{adduct}}_{\text{aq,calc}} = G_{\text{adduct,aq}} - (G_{\text{R,aq}} + G_{\text{Br},\text{aq}})$$
Eq. S11

where $G_{adduct,aq}$, $G_{R,aq}$, and $G_{Br,aq}$ are the aqueous-phase free energies of formation of an adduct, of the target compound, R, and Br, respectively.

The single electron transfer (SET) reaction of Br[•] with an organic compound is not a barrierless interior reaction. Thus, we used the Marcus theory³¹ to calculate the aqueous phase free energy of activation, $\Delta G^{\text{act}}_{\text{aq,SET}}$, in equation as no bond cleavage is involved in the SET reaction of Br[•] (i.e., dissociative electron transfer).

$$\Delta G^{\text{act}}_{\text{aq,SET}} = (1 + \Delta G^{\text{react}}_{\text{aq,SET}})^2 / 4\lambda$$
 Eq. S12

Where $\Delta G^{\text{react}}_{\text{aq,SET}}$ is the standard state aqueous phase free energy of reaction, kcal/mol. The reorganization energy, λ , has two components: (1) the inner-sphere part that represents the change in the structure of solute and (2) the outer-sphere part that represents the change in the structure of the surrounding solvent. The λ_{in} values were calculated as:

$$\lambda_{in} = \Delta E^{\text{react}}_{aq,\text{SET}} - \Delta G^{\text{react}}_{aq,\text{SET}}$$
 Eq. S13

$$\Delta E^{\text{react}}_{\text{aq,SET}} = \Delta E^{\text{react}}_{\text{aq,vertical}} - \Delta E^{\text{react}}_{\text{aq,reactants}}$$
Eq. S14

where $\Delta E^{\text{react}}_{aq,\text{SET}}$ is the standard state aqueous phase reaction energy change between reactants and products, $\Delta E^{\text{react}}_{aq,\text{vertical}}$ is the standard state energy change for vertical product and $\Delta E^{\text{react}}_{aq,\text{reactants}}$ is the standard state energy of reactants. We calculated the λ_{out} values based on the two-sphere model in a continuum medium proposed by Marcus:³¹

$$\lambda_{\text{out}} = \Delta e^2 N_{\text{A}} \left(\frac{1}{2r_1} + \frac{1}{2r_2} - \frac{1}{R} \right) \left(\frac{1}{\varepsilon_{\text{o}}} - \frac{1}{\varepsilon_{\text{s}}} \right)$$
Eq. S15

where Δe is the amount of charge transferred, N_A is the Avogadro's number; r_1 and r_2 are the ionic radii of the reactant molecules 1 and 2, respectively, and $R = r_1 + r_2$, ε_0 and ε_s are the optic (i.e., 78.39) and static (i.e., 1.77) dielectric constants of water, respectively, at 25 °C. Upon the calculations of the chloramine and the reaction product, radical cation, we used the λ_{out} obtained by treating the first solvation sphere explicitly with three water molecules. This way, the term λ_{in} also accounted for the outer-sphere reorganization of the first sphere in going from Br[•] to Br⁻, since the solvation pattern is different in these two species.

The calculated results for the organic model compounds and NH₂Cl are summarized in Table S8.

Text S15. LC-HRMS/MS evidences supporting the formation of $C_6H_3BrO_3$ and $C_6H_5BrO_3$ for the phenol-Br[•] reaction

Based on the LC-HRMS/MS analyses of the γ -radiolysis of phenol, two suspected products were detected containing Br (C₆H₃BrO₃ and C₆H₅BrO₃) with masses of 200.9193 and 202.9349 as [M-H]⁻

corresponding to $C_6H_3^{79}BrO_3$ and $C_6H_5^{79}BrO_3$, respectively, with mass deviation < 1 ppm (Figure S11, Table S3). The presence of Br in the suggested molecular formulas was confirmed by the detection of the ⁸¹Br isotopes with similar peak height, corresponding to the natural abundance of the Br isotope of ⁷⁹Br:⁸¹Br = 1:0.97. Neither of them appeared in blank samples, but showed a gradual increase over time during γ -radiolysis experiments (Figure S12). Based on the molecular formula, *p*-benzoquinone substituted by a hydroxy group and a Br was suggested as a possible molecular structure for C₆H₃BrO₃ (Scheme 1, main text). To confirm this, the substituted *p*-benzoquinone was synthesized by brominating 1,2,4-benzenetriol by N-bromosuccinimide and subsequently oxidizing the hydroxy groups by the Fétizon's reagent (Ag₂CO₃ on celite) to a quinone group. The synthesized product was analyzed by the same LC-HRMS/MS method and its MS spectrum was compared to those obtained from the γ -radiolysis samples containing phenol. The same mass was observed in the synthesized chemical and in the γ radiolysis sample, with a similar MS² spectral pattern, but at a different retention time (difference of 4.1 min, Figure S13). This indicates that the synthesized chemical is likely an isomer of the suspected product formed during the γ -radiolysis. The masses corresponding to C₆H₃BrO₃ and C₆H₅BrO₃ were also detected in the γ -radiolysis sample containing p-benzoquinone (instead of phenol) as a starting compound, showing a similar formation trend (Figure S12). This additionally supports that C₆H₃BrO₃ and $C_6H_5BrO_3$ are likely to have a quinone structure.

Text S16. Formation pathway of 4-bromophenol from the phenol-Br[•] reaction

A minor pathway of the reaction of phenol with Br[•] leads to the formation of 4-bromophenol with 4% yield (Scheme 1, main text). This implies a possible addition mechanism where a bromo-hydroxy-cyclohexadienyl radical formed by the addition of Br[•] to phenol is oxidized to a monobromophenol by any oxidant available in reaction solution, such as *p*-benzoquinone (Scheme S1(3)). This would be comparable to the formation of quinones from dihydroxy-cyclohexadienyl radicals observed for the oxidation of phenol by °OH.³² Alternatively, 4-bromophenol can be also formed from PhO[•] by a radical-radical coupling of PhO[•] and Br[•]. The ΔG^{react}_{aq} of PhO[•] with Br[•] for all possible resonance structures of PhO[•] (Figure S17) were calculated as -28.1 and -31.1 kcal/mol for the formation of 2-bromophenol and 4-bromophenol, respectively, and 6.3 kcal/mol of ΔG^{act}_{aq} . The coupling reaction on both *ortho*- and *para*-positions of PhO[•] are similarly favorable according to the calculated ΔG^{react}_{aq} , which however disagrees with the experimental result where only 4-bromophenol was detected. The dominance of the *para*-product is different from the reaction of phenol with HOBr, for which the *ortho*- and *para*-positions are similarly susceptible.^{33,34} The distinction may come from the different nature of the PhO[•]-Br[•] reaction

mechanism (radical-radical coupling) compared to phenol-HOBr (electrophilic aromatic substitution),³³ but more information is needed to clarify this.

Text S17. Principles of the chlorine-ammonia pretreatment as a bromate mitigation strategy of ozonation

The chlorine-ammonia pretreatment is a proven strategy to mitigate bromate during ozonation.²¹ It was originally designed to pre-oxidize bromide to HOBr, which can then be quenched by addition of ammonium.^{35,36} The formed monobromamine reacts moderately with ozone to bromide and nitrate.³⁷ Furthermore, the residual chlorine also reacts with ammonia to NH₂Cl. NH₂Cl has an additional benefit, because it also scavenges 'OH ($k_{\cdot OH,NH2Cl} = 5.2 \times 10^8 \text{ M}^{-1} \text{ s}^{-1}$ or $5.7 \times 10^8 \text{ M}^{-1} \text{ s}^{-1}$).^{38,39} Overall, the chlorine-ammonia process blocks the initial steps for bromate formation by: (1) masking bromide and preventing it from being oxidized to HOBr by ozone and to Br[•] by 'OH (i.e., blocking reactions 1 and 2, main text) and (2) scavenging 'OH.

Text S18. Calculation of the fractions of Br*-related reactions during ozonation

Ozone, DOM, and Br⁻ are considered the main Br[•] consumers during ozonation. They produce bromine monoxide (BrO[•], a transient species in bromate formation during ozonation),¹⁰ DOM oxidation products (e.g., quinones (section 3.2 in the main text), and Br₂^{•-}, respectively, as a result of the reaction with Br[•]. In addition, a Br[•]-specific quencher such as NH₂Cl tested in this study consumes Br[•] as well. The fraction of Br[•] reacting with each consumer (e.g., f(Br[•]+NH₂Cl) for the fraction reacting with NH₂Cl) was calculated (Eqs. S16 – S18). The calculated f(Br[•]+X) as a function of varying conditions are shown in Figure 3 (main text) and in Figures S18 and S19.

$$-\frac{\mathrm{d}[\mathrm{Br}^{\bullet}]}{\mathrm{dt}} = k_{Br^{\bullet},O_{3}}[\mathrm{Br}^{\bullet}][O_{3}] + k_{Br^{\bullet},DOM}[\mathrm{Br}^{\bullet}][\mathrm{DOM}] + k_{Br^{\bullet},Br^{-}}[\mathrm{Br}^{\bullet}][\mathrm{Br}^{-}] + k_{Br^{\bullet},\mathrm{NH}_{2}\mathrm{Cl}}[\mathrm{Br}^{\bullet}][\mathrm{NH}_{2}\mathrm{Cl}]$$
Eq. S16

$$-\frac{\mathrm{d}[\mathrm{Br}^{\bullet}]}{[\mathrm{Br}^{\bullet}]} = \left(k_{Br^{\bullet},O_{3}}[O_{3}] + k_{Br^{\bullet},DOM}[\mathrm{DOM}] + k_{Br^{\bullet},Br^{-}}[\mathrm{Br}^{-}] + k_{Br^{\bullet},\mathrm{NH}_{2}\mathrm{Cl}}[\mathrm{NH}_{2}\mathrm{Cl}]\right)\mathrm{dt} \qquad \mathsf{Eq. S17}$$

$$f(Br^{\bullet} + NH_2Cl) = \frac{k_{Br^{\bullet}, NH_2Cl}[NH_2Cl]}{\left(k_{Br^{\bullet}, O_3}[O_3] + k_{Br^{\bullet}, DOM}[DOM] + k_{Br^{\bullet}, Br^{-}}[Br^{-}] + k_{Br^{\bullet}, NH_2Cl}[NH_2Cl]\right)}$$
Eq. S18

The selected k_{Br} for the f(Br⁺+X) calculation were 1.5×10^8 M⁻¹ s⁻¹ for ozone, 10 1.7×10^4 (mgC/L)⁻¹ s⁻¹ for DOM (Table 1, main text), 4.2×10^7 M⁻¹ s⁻¹ for Br⁻ (see below), and 4.4×10^9 M⁻¹ s⁻¹ for NH₂Cl (Table 1, main text). The k_{Br} of 4.2×10^7 M⁻¹ s⁻¹ for Br⁻ is an apparent k_{Br} calculated by taking into account the

reverse reaction of Br[•] and Br⁻ (Eq. S19) and a subsequent reaction of Br₂^{•-} with DOM (Eq. S20) for [DOM] = 1.4 mgC/L. Details on deriving the apparent $k_{Br•}$ are shown below.

Br[•] + Br⁻
$$\leftrightarrow$$
 Br₂^{•-}
Br₂^{•-} + DOM \rightarrow products $k_{20} = 78 \text{ (mgC/L)}^{-1} \text{ s}^{-1} (\text{for SRFA II})^{40}$ Eq. S19
Eq. S19
Eq. S19

According to the Eqs. S19 and S20, the rate expression for Br[•] and Br₂^{•–} are as follows:

$$-\frac{d[Br^{\bullet}]}{dt} = k_{+19}[Br^{\bullet}][Br^{-}] - k_{-19}[Br_{2}^{\bullet-}]$$
Eq. S21

$$-\frac{d[Br_2]}{dt} = k_{20}[Br_2^{\bullet-}][DOM] + k_{-19}[Br_2^{\bullet-}] - k_{+19}[Br^{\bullet}][Br^{-}]$$
Eq. S22

Assuming Br2^{•-} is in steady-state, Eq. S22 can be expressed in terms of the concentration of Br2^{•-}:

$$[Br_2^{\bullet-}]_{ss} = \frac{k_{\pm 19}[Br^{\bullet}][Br^{-}]}{(k_{20}[DOM] + k_{\pm 19})}$$
Eq. S23

By substituting Eq. S21 with Eq. S23, the rate expression for Br[•] is converted to Eq.S24 and finally k_{app} is obtained as a value dependent on DOM concentration (e.g., $k_{app} = 4.2 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$ for 1.4 mgC/L DOM).

$$-\frac{d[Br^{\bullet}]}{dt} = \frac{k_{\pm 19}k_{20}[DOM]}{(k_{20}[DOM] + k_{\pm 19})}[Br^{\bullet}][Br^{-}] = k_{app}[Br^{\bullet}][Br^{-}]$$
Eq. S24

Table S1. Chemical reagents

| Compound | Supplier | Catalog no. | Purity |
|-------------------------|----------------|-------------|----------------------|
| 1,2-Dibromoethane | Sigma-Aldrich | 240656 | ≥ 99% |
| 2,2'-Azino-bis(3- | Sigma-Aldrich | 10102946001 | > 98% |
| ethylbenzothiazoline-6- | | | |
| sulfonic acid) | | | |
| diammonium salt (ABTS) | | | |
| 3-Phenylpropionic acid | Alfa Aesar | A14908 | 99% |
| 4-Chlorobenzoic acid | Sigma-Aldrich | 135585 | 99% |
| 4-Bromophenol | Fluk | 18010 | ≥ 98% |
| 4-chlorophenol | Riedel-de Haën | 35826 | 99.8% |
| 4-iodophenol | Fluka | 58020 | ≥ 98% |
| Ammonium sulfate | Sigma-Aldrich | A5132 | ≥ 99% |
| Anisole | Sigma-Aldrich | 123226 | 99% |
| Benzene | Sigma-Aldrich | 32212 | ≥ 99% |
| Benzoic acid | Sigma-Aldrich | 33047 | ≥ 99% |
| <i>p</i> -Benzoquinone | Sigma-Aldrich | B10358 | ≥ 98% |
| Benzylamine | Sigma-Aldrich | 185701 | 99% |
| N,N-Dimethylbenzylamine | Sigma-Aldrich | 185582 | ≥ 99% |
| Formic acid | Sigma-Aldrich | 1.00264 | 90-100% |
| Ibuprofen | Sigma-Aldrich | 11892 | ≥ 98% |
| Naphthalene | Fluka | 70211 | ≥ 99% |
| Phenol | Fluka | 77610 | ≥ 99% |
| Sorbic acid | Fluka | 85510 | ≥ 99% |
| Toluene | Merck | 1.08325 | ≥ 99% |
| trans-cinnamic acid | Sigma-Aldrich | C80857 | ≥ 99% |
| tert-butanol | Sigma-Aldrich | 19460 | ≥ 99% |
| Sodium hypochlorite | Sigma-Aldrich | 13440 | 6-14% active |
| solution | | | chlorine |
| Suwannee River Humic | IHSS | 2S101F | 52.34% carbon as |
| Acid Standard II | | | dry, ash-free sample |
| Silver nitrate | Merck | 1512 | ≥ 99% |
| Hydroquinone | Sigma-Aldrich | H9003 | ≥ 99% |
| Catechol | Merck | 82261 | ≥ 99% |

| Compounds | Column | Detection ¹ | RT, min | LOQ, μM | Measuring range |
|-------------------------|--------|---------------------------|-------------------------|-------------|------------------------------|
| Benzylamine | HILIC | 200nm (Em) /270nm (Ex) | 6.9 | 1.2 | 2 – 3 µM |
| N,N-Dimethylbenzylamine | HILIC | 200nm (Em) /270nm (Ex) | 8.0 | 0.6 | 2 – 3 µM |
| 4-Bromophenol | C18 | 220nm or MS | 10.4 (UV), 18.5 (MS) | 0.3 (MS) | 1 – 3 µM |
| 4-Chlorophenol | C18 | 220nm | 9.3 | 0.6 | 0.6 – 3 μM |
| 4-lodophenol | C18 | 220nm | 11.8 | 0.6 | 0.7 – 3 μM |
| Anisole | C18 | 267nm | 10.5 | 0.3 | 1 – 3 µM |
| Benzene | C18 | 200nm (Em) /270nm (Ex) | 11.0 | 2.5 | 3 – 4 µM |
| Benzoic acid | C18 | 230nm | 6.7 | 0.3 | 2 – 3 µM |
| p-Chlorobenzoic acid | C18 | 230nm | 11.5 | 0.6 | 2 – 3 µM |
| Ibuprofen | C18 | 220nm | 18.1 | 0.2 | 0.3 – 4 μM |
| Naphthalene | C18 | 220nm | 16.9 | 0.2 | 0.2 – 2 μM |
| Phenol | C18 | 200nm (Em) /310nm (Ex) | 5.1 | 0.2 | 2 – 3 µM |
| Toluene | C18 | 210nm | 14.5 | 1.5 | 2 – 3 µM |
| 3-Phenylpropionic acid | C18 | 210nm | 7.8 | 0.3 | 2 – 3 µM |
| <i>p</i> -Benzoquinone | C18 | 254nm | 3.0 | 0.6 | 0.8 – 4 μM |
| Sorbic acid | C18 | 267nm | 6.6 | 0.2 | 0.3 – 3 μM |
| trans-Cinnamic acid | C18 | 267nm | 9.3 | 0.1 | 1 – 3 μM |
| Hydroquinone | C18 | 220nm | 5.6 | 0.6 | 0.1 – 0.6 μM |
| Catechol | C18 | MS | 8.4 | 0.02 | 0.0 <mark>2 – 0.1 μ</mark> Μ |

Table S2. Compounds analyzed by HPLC and LC-HRMS/MS and the corresponding analytical condition and LOQ.

¹Diode-array detector, fluorescence detector, or mass spectrometer

| Table S3. Detected exact masses and | d mass deviation by | LC-HRMS/MS. |
|-------------------------------------|---------------------|-------------|
|-------------------------------------|---------------------|-------------|

| Compounds | Molecular formula (M) | Polarity, Adduct | Theoretical exact mass | Detected mass | Mass deviation, ppm |
|--|--|---------------------|---------------------------|------------------|---------------------------|
| Phenol | C ₆ H ₆ O | pos, M+H | 95.0491 | 95.0491 | 0.0 |
| <i>p</i> -Benzoquinone | $C_6H_4O_2$ | pos, M+H | 109.0284 | 109.0285 | 0.9 |
| 4 Promonhonol | C ₆ H ₅ ⁷⁹ BrO | neg, M-H | 170.9451 | 170.9454 | 1.8 |
| 4-Bromophenoi | $C_6H_5^{81}BrO$ | | 172.9431 | 172.9429 | 1.2 |
| Catechol | $C_6H_6O_2$ | neg, M-H | 109.0295 | 109.0299 | 3.7 |
| | C ₆ H ₃ ⁷⁹ BrO ₃ | neg, M-H | 200.9193 | 200.9194 | 0.5 |
| C6H3BIO3 (Suspected) | $C_6H_3^{81}BrO_3$ | | 202.9172 | 202.9174 | 1.0 |
| C-H-PrO- (suspected) | $C_6H_5^{79}BrO_3$ | neg, M-H | 202.9349 | 202.9352 | 1.5 |
| C ₆ H ₅ BrO ₃ (suspected) | $C_6H_5^{81}BrO_3$ | | 204.9329 | 204.9328 | 0.5 |
| $C_6H_4O_3$ (suspected) | $C_6H_4O_3$ | neg, M-H | 123.0088 | 123.0090 | 1.6 |
| C ₆ H ₆ O ₃ (suspected) | C ₆ H ₆ O ₃ | neg, M-H | 125.0244 | 125.0246 | 1.6 |
| C ₆ H ₆ O ₄ (suspected) | C ₆ H ₆ O ₄ | neg, M-H | 141.0193 | 141.0196 | 2.1 |

| C ₁₂ H ₁₀ O ₂ (suspected) | $C_{12}H_{10}O_2$ | neg, M-H | 185.0608 | 185.0609 | 0.5 | |
|--|-------------------|----------|----------|----------|-----|--|
|--|-------------------|----------|----------|----------|-----|--|

Table S4. Theoretically calculated $\Delta G_{\text{solv,calc}}$ values and experimental values of halides and halogen radicals in kcal/mol. n is the number of explicit water molecule(s).

| X⁻ or X• | $\Delta G_{ m solv,calc}$ | $\Delta G_{ m solv,calc}$ | $\Delta G_{ m solv,calc}$ | $\Delta G_{ m solv, calc}$ | $\Delta G_{ m solv,exp}$ |
|----------------|---------------------------|---------------------------|---------------------------|----------------------------|--------------------------|
| | (n=0) | (n=1) | (n=2) | (n=3) | |
| F ⁻ | -86.9 | -96.0 | -99.2 | -104.6 | -104.3 ⁴¹ |
| Cl | -65.1 | -69.1 | -69.3 | -71.0 | -75.5 ²⁵ |
| Br⁻ | -53.3 | -58.7 | -60.1 | -63.0 | -66.3 ⁴² |
| F | -64.1 | -66.8 | -66.0 | -67.7 | -57.4 ⁴² |
| Cl• | 0.1 | -4.0 | -3.7 | -4.3 | -4.3 ^a |
| Br• | -0.8 | -3.4 | -1.1 | -1.4 | 0.11 ^a |

^a experimentally derived value in Eq. S10.

Table S5. Benchmark calculations of one electron reduction potential of Br*/Br-.

| Method and basis set | E ⁰ red, V |
|--|-----------------------|
| UCCSD(T)/Aug-cc-pVTZ | 1.4 |
| UCCSD(T)/Aug-cc-pVQZ | 1.5 |
| UCCSD(T)/Aug-cc-pV5Z | 1.5 |
| UCCSD(T)/Aug-cc-pVTZ | 1.4 |
| CBS-QB3//M06-2X/Aug-cc-pVDZ | 1.5 |
| MP2(FULL)/Aug-cc-pVDZ//M06-2X/Aug-cc-pVTZ | 1.4 |
| M06-2X/Aug-cc-pVDZ// M06-2X/Aug-cc-pVTZ (2 explicit water molecules) | 1.7 |
| M06-2X/Aug-cc-pVDZ// M06-2X/Aug-cc-pVTZ (3 explicit water molecules) | 1.8 |
| Experimental value 43 | 1.8-2.2 |

Table S6. Validation of one electron reduction potential for selected organic compounds at the M06-2X/Aug-cc-pVDZ//M06-2X/Aug-cc-pVTZ level of theory with the SMD solvation model

| R/R*+ | <i>E</i> ⁰ _{red} , V | Reference | $\Delta G^{\text{react}}_{\text{exp}}$ | $\Delta G^{\text{react}}_{\text{calc}}$ |
|--|--|----------------------------|--|---|
| | | | kcal/mol | kcal/mol |
| $C_6H_6/C_6H_6^{\bullet+}$ | -2.1 | Pearson 1986 ⁴⁴ | 48.4 | 56.9 |
| $C_6H_5CH_3/C_6H_5CH_3^{\bullet+}$ | -1.98 | Pearson 1986 ⁴⁴ | 45.7 | 49.1 |
| C ₆ H ₅ OCH ₃ /C ₆ H ₅ OCH ₃ ^{•+} | -1.62 | Jonsson et al. 1993 45 | 37.4 | 41.0 |

| No. | Compound | Optimized molecular structure of an adduct | $\Delta G^{\text{adduct}}_{\text{aq}}$, kcal/mol |
|-----|------------------------------------|--|---|
| 1 | Benzylamine | | -0.4 |
| 2 | <i>N,N-</i> Dimethylbenzylamine | | -0.4 |
| 3 | <i>N</i> -Methylbenzylamine | | -0.3 |
| 4 | 4-Bromophenol | | -0.8 |
| 5 | 4-lodophenol | | -1.4 |
| 6 | Anisole | 0 33-33-9-9-1 | -2.5 |
| 7 | Benzene | | 4.5 |

Table S7. Calculated free energy of adduct formation ($\Delta G^{\text{adduct}}_{\text{aq}}$) for the reactions of organic model compounds with Br[•] (red sphere) and the optimized molecular structures of the adducts.

| 8 | Benzoate | ٩ | -0.6 |
|----|-----------------------------|---|-------|
| | | • <mark>• • • • • • • • • • • • • • • • • • </mark> | |
| 9 | Naphthalene | | -0.6 |
| 10 | Toluene | | -1.4 |
| 11 | Sorbic acid | | -9.9 |
| 12 | <i>trans</i> -Cinnamic acid | | -1.3 |
| 13 | Ibuprofen | | -1.6 |
| 14 | Phenol | | -1.9 |
| 15 | Phenolate | | -19.8 |

| 16 | 4-Chlorophenol | 9 20.01 | -0.7 |
|----|---|-------------------|-------|
| 17 | 4 Chlorophonolata | • | 19.6 |
| 17 | 4-Chiorophenolate | | -18.0 |
| 18 | Benzylamine (deprotonated) | | -8.4 |
| 19 | N,N- Dimethylbenzylamine (deprotonated) | | -12.2 |
| 20 | N-Methylbenzylamine (deprotonated) | | -10.8 |
| 21 | 4-Bromophenolate | | -17.8 |
| 22 | <i>p</i> -Chlorobenzoate | | -0.6 |
| 23 | 3-Phenylpropionic acid | | -1.2 |

| 24 | <i>p</i> -Benzoquinone | 9 | 1.2 |
|----|------------------------|---|-----|
| | | | |

Table S8. Compiled QC calculation results for the reactions of the selected organic model compounds and of NH₂Cl with Br' ($\Delta G^{\text{react}}_{aq,calc}$ as the standard state aqueous phase free energy of reaction, λ as the reorganization energy, and $\Delta G^{\text{act}}_{aq,SET}$ as the aqueous phase free energy of activation).

| No. | Name | Structure | $\Delta G^{\text{react}}_{\text{aq,calc}}$ | $\Delta G^{act}_{aq,SET}$ | |
|-----|---|--|--|---------------------------|----------|
| | | *H-N | kcal/mol | kcal/mol | kcal/mol |
| 1 | Benzylamine (protonated) | | 20.2 | 26.4 | 20.5 |
| 2 | N,N- dimethylbenzylamine (protonated) | NH* | 19.8 | 27.1 | 20.3 |
| 3 | N-methylbenzylamine (protonated) | "HyM | 18.4 | 28.4 | 19.3 |
| 4 | 3,4-Chlorophenol | | 5.2 | 28.5 | 10.0 |
| 5 | 4-bromophenol | | 2.8 | 28.4 | 8.6 |
| 6 | 4-iodophenol | OH CH | -3.4 | 27.3 | 5.3 |
| 7 | Anisole | | -0.5 | 28.0 | 6.7 |
| 8 | Benzene | \bigcirc | 15.4 | 26.3 | 16.5 |
| 9 | Benzoate | | 8.8 | 34.9 | 13.7 |
| 10 | Naphthalene | | -1.4 | 25.5 | 5.7 |
| 11 | Toluene | | 7.6 | 26.9 | 11.1 |
| 12 | Sorbic acid | , solution of the second secon | -5.7 | 30.3 | 5.0 |
| 13 | Trans-cinnamic acid | C C | 0.5 | 29.3 | 7.6 |
| 14 | Ibuprofen | | -23.9 | 63.7 | 6.2 |

| 15 | Phenol | | 0.9 | 28.0 | 7.5 |
|----|---------------------|--------------------|-------|------|------|
| | | ОН | | | |
| 16 | Phenolate | | -30.5 | 25.2 | 0.3 |
| 17 | 4-chlorophenol | CI | 1.5 | 28.5 | 7.9 |
| | | | | | |
| 18 | 4-chlorophenolate | CI | -29.4 | 25.6 | 0.1 |
| | | | | | |
| 19 | Benzylamine | H ₂ N | -1.9 | 36.0 | 8.1 |
| | (deprotonated) | | | | |
| 20 | N,N- | N N | -17.9 | 40.0 | 3.1 |
| | (deprotonated) | | | | |
| | | | | | |
| 21 | N-methylbenzylamine | HN | -10.4 | 36.3 | 4.6 |
| | (deprotonated) | | | | |
| | | | | | |
| 22 | 4-bromophenolate | 0 | -28.5 | 25.9 | 0.1 |
| | | | | | |
| 23 | p-chlorobenzoate | | 12.5 | 29.3 | 14.9 |
| | | | | | |
| | | | | | |
| 24 | 3-phenyl-propionate | | 4.5 | 37.8 | 11.8 |
| | | | | | |
| 25 | p-benzoquinone | | 39.8 | 27.3 | 41.2 |
| | | | | | |
| | | | | | |
| 26 | monochloramine | NH ₂ Cl | -34.0 | 65.5 | 3.8 |
| | | | | | |

| Perspective | Benzylamine (protonated) | Benzylamine (deprotonated) | | | |
|-------------|--------------------------------------|--|--|--|--|
| A | | | | | |
| В | | | | | |
| Perspective | N,N-Dimethylbenzylamine (protonated) | <i>N,N</i> -dimethylbenzylamine (deprotonated) | | | |
| A | | | | | |
| В | | | | | |

Table S9. Spin density distribution of protonated and deprotonated forms of benzylamine and *N*,*N*-dimethylbenzylamine.

Table S10. Scavenging rates (k_{species} ', s⁻¹) of e^- , °OH, °H, Br°, and °tBA (*t*-butanol derived radical) by the reaction species in γ -radiolysis kinetic experiments (1,2-dibromoethane (1,2-DBE), Ag⁺, *t*-butanol, a model compound, phosphate ions (H₂PO₄⁻/HPO₄²⁻ for pH 7), and dissolved O₂), based on the reported or assumed second-order rate constants (k_{species} , M⁻¹s⁻¹) and the applied concentrations (conc., M).

| species | conc. | k _{e-} | <i>k</i> e-' | к •он | <i>к</i> •он ' | к •н | <i>k</i> •н ' | kBr∙ | k₀r•' | k ∙tba | k•tba ' |
|--------------------|----------------------|-------------------------|---------------------|-------------------------|---------------------|-------------------------|---------------------|------------------------|---------------------|-----------------------|---------------------|
| 1,2-DBE | 7.0×10 ⁻⁴ | 1.4×10 ^{10 12} | 9.8×10 ⁶ | 2.4×10 ^{8 12} | 1.7×10 ⁵ | 1.0×10 ^{9 b} | 7.0×10 ⁵ | 1.0×10 ^{6 46} | 7.0×10 ² | na | na |
| Ag+ | 2.2×10 ⁻⁶ | 3.7×10 ^{10 47} | 8.0×10 ⁴ | 1.4×10 ^{10 47} | 3.0×10 ⁴ | 2.0×10 ^{10 47} | 4.4×10 ⁴ | 1.0×10 ^{10 b} | 2.2×10 ⁴ | 1.0×10 ^{9 b} | 2.2×10 ³ |
| <i>t</i> -butanol | 4.0×10 ⁻³ | 4.0×10 ^{5 47} | 1.6×10 ³ | 6.0×10 ^{8 47} | 2.4×10 ⁶ | 1.7×10 ^{5 47} | 6.8×10 ² | 3.0×10 ^{4 c} | 1.2×10 ² | na | na |
| model ^a | 3.4×10 ⁻⁶ | 5.1×10 ^{9 47} | 1.7×10 ⁴ | 5.0×10 ^{9 47} | 1.7×10 ⁴ | 3.4×10 ⁹⁴⁷ | 1.2×10 ⁴ | 5.5×10 ^{9 d} | 1.9×10 ⁴ | 1.0×10 ^{9 e} | 3.4×10 ³ |
| phosphate | 5.0×10 ⁻² | 9.6×10 ^{6 47} | 4.8×10 ⁵ | 8.5×10 ^{4 47} | 4.3×10 ³ | 2.8×10 ^{5 47} | 1.4×10 ⁴ | na | na | na | Na |
| O ₂ | 6.8×10 ⁻⁵ | 1.9×10 ^{10 47} | 1.3×10 ⁶ | na | na | na | na | na | na | 1.0×10 ^{9 b} | 6.8×10 ⁴ |
| total ^f | | | 1.2×10 ⁷ | | 2.6×10 ⁶ | | 7.7×10 ⁵ | | 4.1×10 ⁴ | | 6.8×10 ⁴ |

^a naphthalene as an example; ^b assumed; ^c average value of ¹³ and ¹⁴; ^d Table 1 (main text); ^e assumed based on the reaction of [•]tBA with histidine¹⁷; ^f total calculated scavenging rate as a sum of the scavenging rates of all species



Figure S1. Total chloride concentrations ($[Cl-]_{tot}$) of primary NH₂Cl stock solutions after reducing NH₂Cl by sulfite. $[Cl-]_{tot}$ indicates the sum of the chloride concentration already present in the stock solution and chloride formed from the reduction of NH₂Cl with sulfite. The NH₂Cl concentration of the stock solution was 0.38 mM.



Figure S2. Left: Determined and theoretical NH₂Cl concentrations in supernatant of the silver(I)-treated NH₂Cl stock solutions (as 0.4mM, 1.7mM and 3.5mM NH2Cl); right: measured chloride concentrations in 0.2 mM NH₂Cl solutions prepared by diluting 0.4, 1.7, or 3.5 mM NH₂Cl stock solutions and reducing NH₂Cl of each solution by sulfite.



Figure S3. Setup for preparing γ -radiolysis samples for the competition kinetics experiments. To transfer the Ar-saturated solution to the sample vials containing an air-saturated solution (shown as 1.5mL amber vials with crimp neck), the transferring tubing was placed to the bottom of the vials.



Figure S4. Flow charts of the sample preparation for kinetic experiments performed with γ-radiolysis for (a) most organic model compounds, (b) *p*-benzoquinone, and (c) benzene, toluene, and naphthalene. DBE stands for 1,2-dibromoethane. Background shades separate the days on which an individual step (preparation, γ-radiolysis, and analyses) was carried out.







Figure S5. Competition kinetics plots for the reactions of Br[•] with the organic model compounds, NH₂Cl, DOM, and oxidized DOM (by 0.8 gO₃/gC or 1.5 gO₃/gC) at pHs 7.1 or 10.2. Concentrations of the model compounds were fixed at $3.0 - 3.4 \mu$ M. Concentrations of NH₂Cl, DOM, and oxidized DOM are stated in the figure headings. Data points are from a single measurement and k_{Br} was calculated mostly based on an average of experimental duplicates.



Figure S6. Slopes obtained by linear regression of data points of the competition kinetics plots (shown in Figure S5) as a function of a reciprocal of the DOC or NH₂Cl concentration, to derive k_{Br} for NH₂Cl, DOM, and oxidized DOM (by 0.8 gO₃/gC or 1.5 gO₃/gC). See Text S8 for the corresponding rate expression.



Figure S7. Quantitative structure-activity relationship of the measured k_{Br} of the selected aromatic model compounds with (a) Hammett constants and (b) computed activation energies.



Reaction coordinate

Figure S8. Schematic reaction coordinate for the reaction of phenol with Br[•].



Figure S9. Correlation between the measured k_{Br} of the selected organic model compounds and aqueous phase free energies for the formation of adducts (ΔG^{adduct}_{aq}).



Figure S10. Concentrations of phenol and the identified products as a function of the γ -radiolysis time during the reaction of phenol with Br[•], for the condition with 22 μ M phenol, 0.7 mM 1,2-dibromoethane, 40 mM *t*-butanol, and 50 mM phosphate buffer (pH 7.1). See Figure 2 (main text) for relative product formation.



Figure S11. Left: Filtered chromatograms of a blank sample (no γ -radiolysis); right: γ -radiolysis sample (t = 40 min) with exact masses of 200.9193, 202.9172, 202.9349, and 204.9329 (from top to bottom of the chromatograms) corresponding to the molecular formulas of C₆H₃⁷⁹BrO₃, C₆H₃⁸¹BrO₃, C₆H₅⁷⁹BrO₃, and C₆H₅⁸¹BrO₃.



Figure S12. Trends of the peak intensity as a function of the γ -radiolysis time for the exact masses of 200.9193 (blue circles, corresponding to C₆H₂⁷⁹BrO₃ as M-H) and 202.9349 (orange triangles, corresponding to C₆H₄⁷⁹BrO₃ as M-H) during the reaction of Br[•] with phenol (left) or *p*-benzoquinone (right).



Figure S13. Left: Filtered chromatograms of a γ -radiolysis sample (t = 40 min); right: synthesized substituted *p*-benzoquinone with the exact masses of 200.9193, 202.9172, 202.9349, and 204.9329 (from top to bottom of the chromatograms) as [M-H], corresponding to the molecular formulas C₆H₃⁷⁹BrO₃, C₆H₃⁸¹BrO₃, C₆H₅⁷⁹BrO₃, and C₆H₅⁸¹BrO₃. Retention times for C₆H₃BrO₃, and C₆H₅BrO₃ are 7.8 and 6.1 min for the γ -radiolysis sample (left) and 11.9 and 8.7 min for the synthesized chemical (right).



Figure S14. MS² spectra of m/z of 123.0089 (as [M-H]) with a HCD normalized collision energy (unitless) of 15 (top), 45 (middle), or 60 (bottom).



Figure S15. Simulated fragment ions corresponding to the detected major daughter ions of m/z = 95.0140 and 68.9974. Two different suspected structures are shown for the parent ion of 123.0089 as [M-H]. The fragment ions were obtained based on an *in silico* fragmentation simulation by MetFrag.⁴⁸



Figure S16. Top: Measured; bottom: reported MS^2 spectra for the exact masses of 185.0610 and 185.0608, respectively (both corresponding to $C_{12}H_9O_2$ as M-H). The reported MS^2 spectrum was obtained from <u>https://massbank.eu/MassBank/.⁴⁹</u>



Figure S17. Resonance structures of the phenoxyl radical.



Figure S18. Calculated fractions of Br[•] reacting with DOM (green line), ozone (red line), bromide (blue line), or NH₂Cl (orange line) as a function of the ozone concentration in (a) absence or (b) presence of NH₂Cl. The selected concentrations were 5 mgC/L DOC, 1.3 μ M Br⁻ (100 μ g/L Br⁻), and 15 μ M NH₂Cl.



Figure S19. Calculated fractions of Br[•] reacting with NH₂Cl as a function of (a) bromide, (b) ozone, or (c) DOC concentration. Lines indicate modelled NH₂Cl concentrations (1, 2, 5, 10, and 15 μ M) and an asterisk symbol indicates the conditions applied in the ozonation experiment. Bromide, ozone, and DOC concentrations were fixed at 2 μ M, 60 μ M, and 1.4 mgC/L (corresponding to the experimental conditions) when they were not an independent variable of the plot.



Figure S20. (a) *p*CBA abatement as a function of the ozone exposure and (b) ozone decrease as a function of reaction time, for ozonation of Lake Zurich water (1.4 mgC/L DOM, 2 μ M bromide, 1 mM phosphate buffer (pH 7.6), 5 μ M *p*CBA, and an ozone dose of 60 μ M in absence or presence of an additional agent (10 μ M formate, 4 μ M ammonium, 7 μ M NH2Cl, or 15 μ M NH2Cl). The numbers in parentheses in the legend in plot (a) indicate the R_{ct} derived from the slopes of the regression lines.



Figure S21. Fraction of Br[•] in equilibrium with BrOH^{•–} as a function of pH.

(1) Electron transfer (supported by QC data)



(2) H-abstraction (suggested by Merényi and Lind 1994, for alcohols)



(3) Addition (suggested by Guha 1993, for olefins)



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Detailed mechanism of the oxidation step of (3):



Scheme S1. Possible initial reaction pathways for the reaction of phenol with Br*.

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