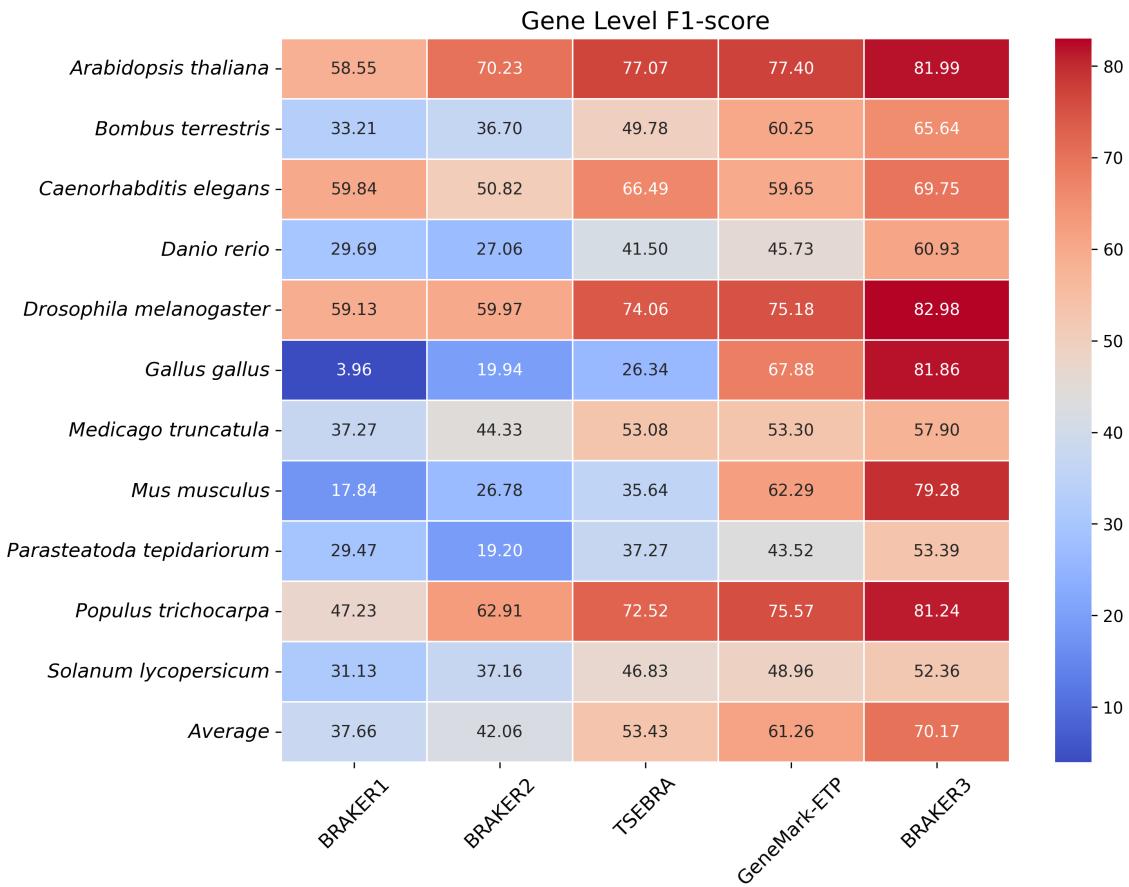
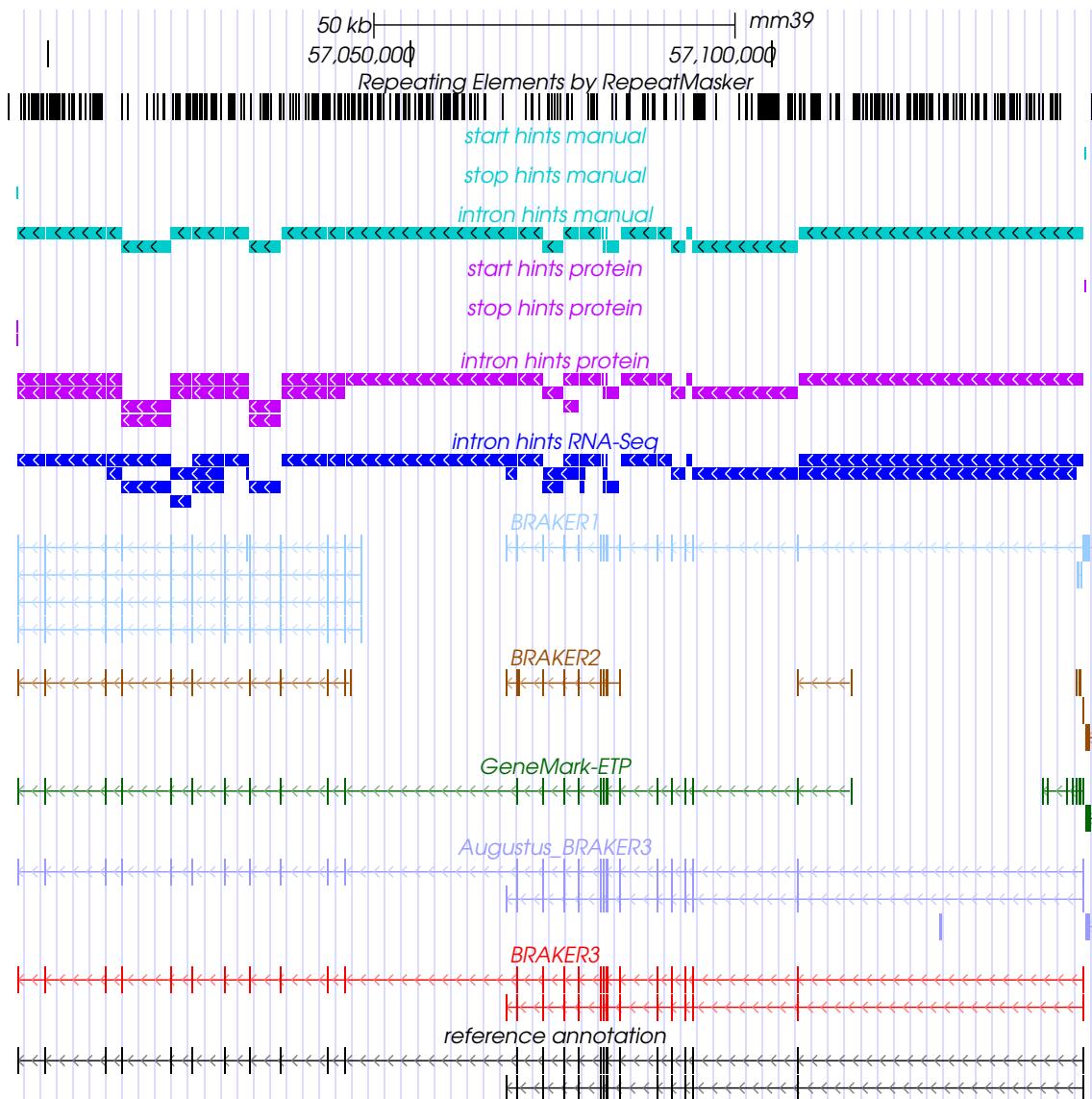


1 Supplemental Material

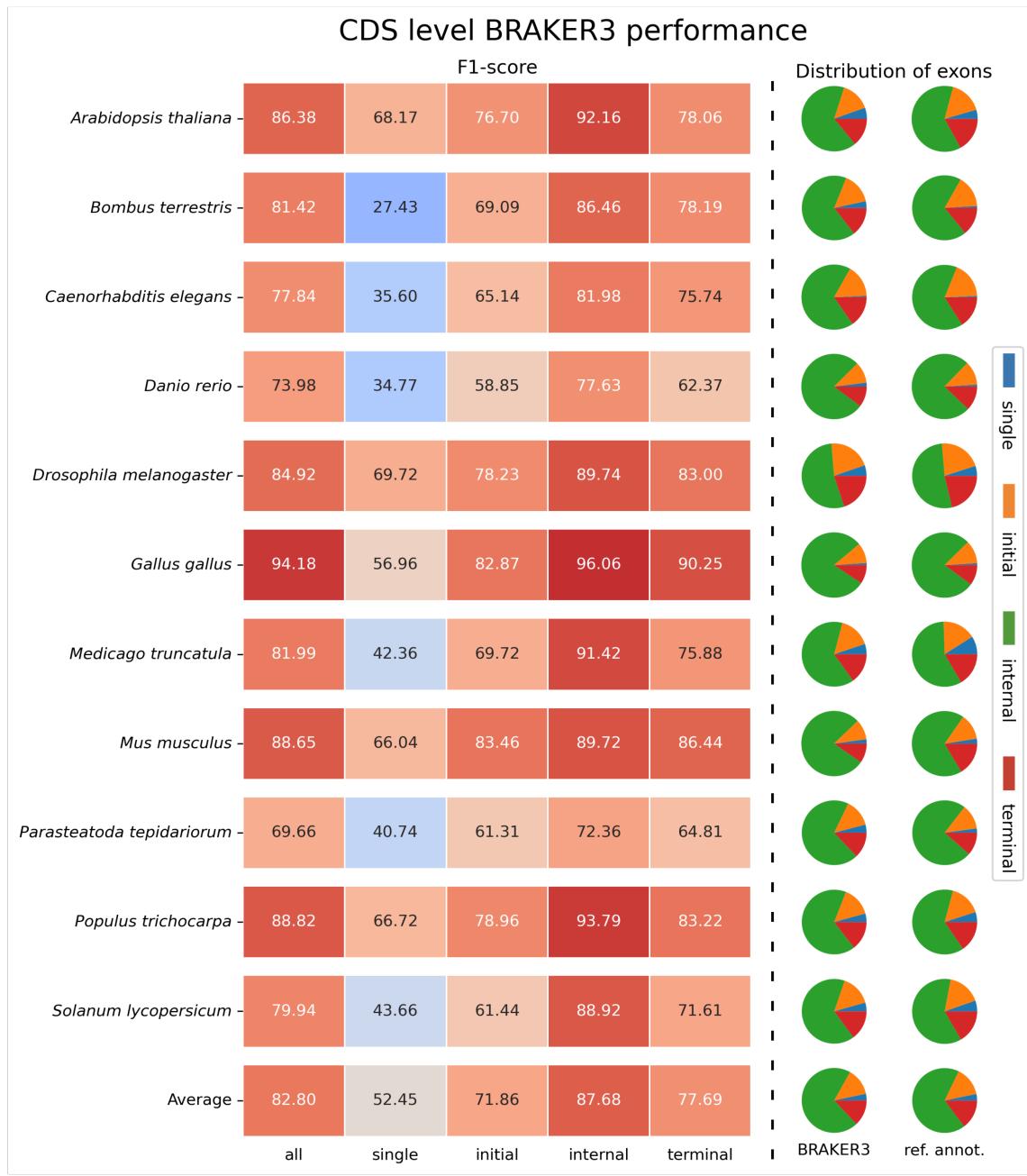
1.1 Supplemental Figures



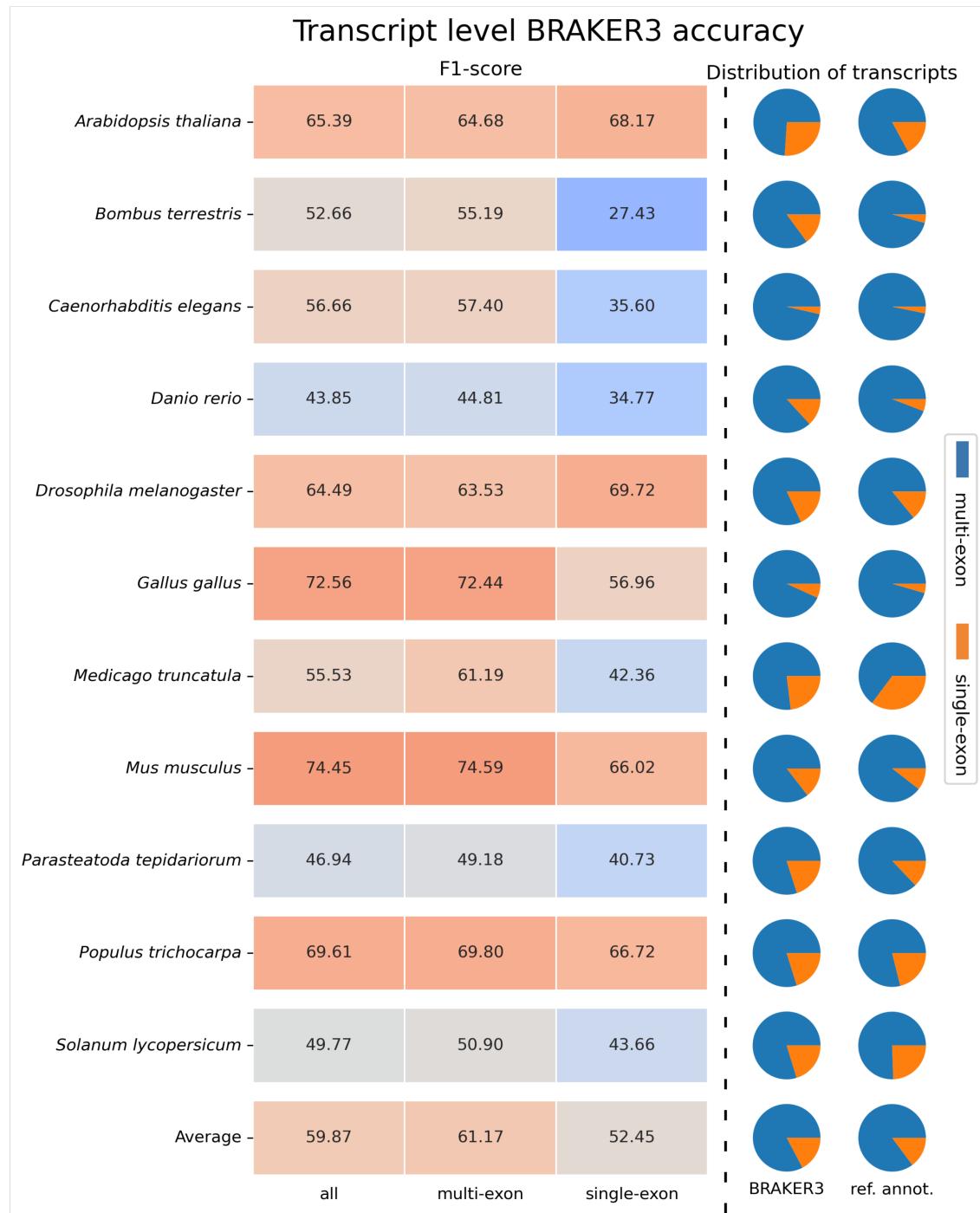
Supplementary Figure S1: Heatmap of F1-scores of pipelines being input short-read RNA-Seq libraries and a protein database (with proteins of species from the respective **order excluded**). The last row shows the averages for the 11 different species.



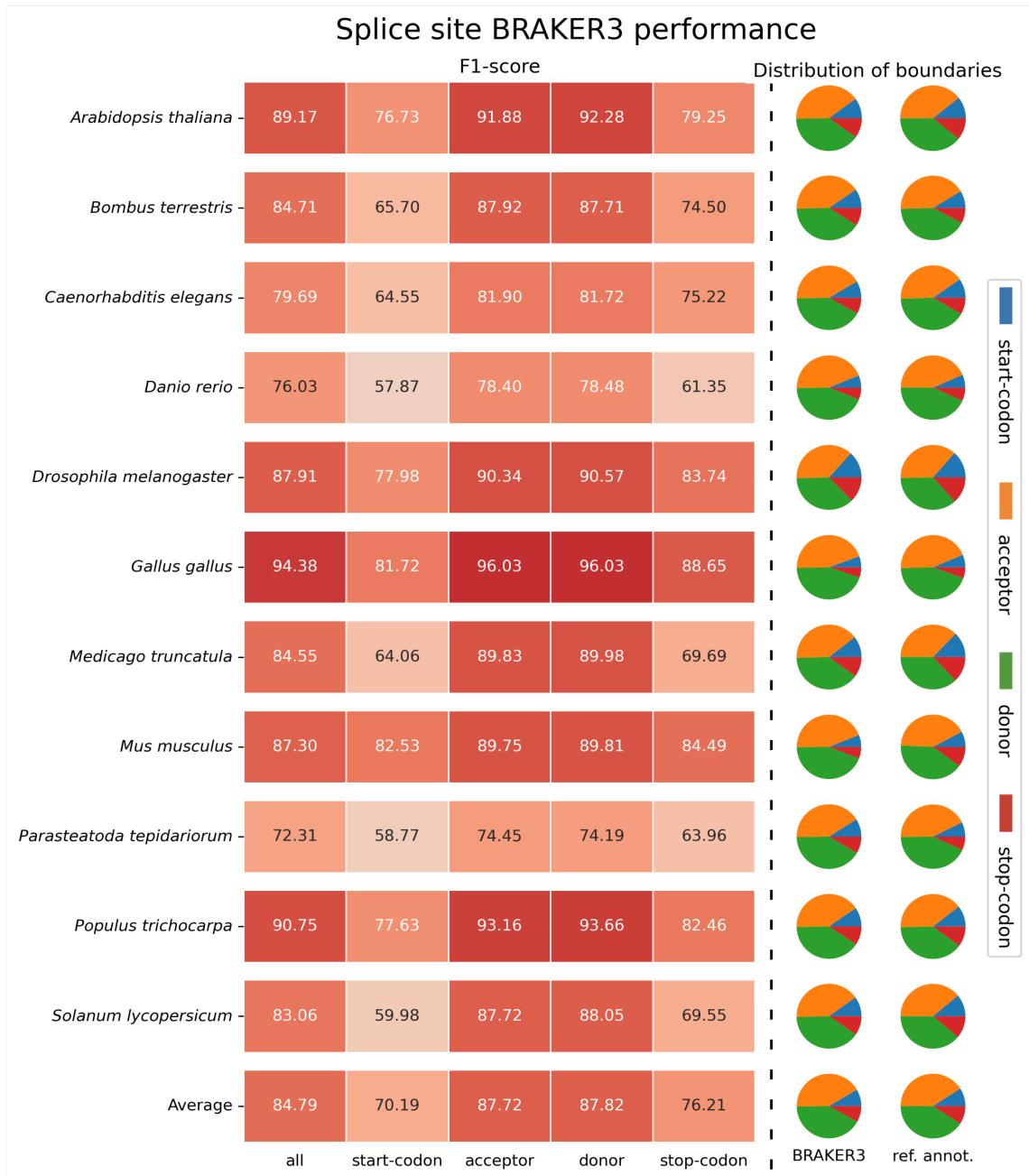
Supplementary Figure S2: A gene locus of *Mus musculus* visualized with the UCSC Genome Browser (Kent et al. [2002], <http://genome.ucsc.edu>). Tracks for the gene prediction of pipelines obtaining short-read RNA-Seq libraries and a protein database (respective *order excluded*) as input are displayed. Tracks highlighting exon boundaries based on extrinsic evidence are shown at the top. Manual hints represent high confidence hints that are highly weighted during AUGUSTUS prediction. In this example, TSEBRA selects two correct transcripts from AUGUSTUS run as part of BRAKER3 (AUGUSTUS_BRAKER3) and filters out three false transcripts from GeneMark-ETP and two from AUGUSTUS.



Supplementary Figure S3: The F1 score when predicting the protein-coding (parts of) exons, broken down by type of exon. Single: unspliced coding region, initial/terminal: first/last coding region, respectively, of a gene with spliced coding sequence, internal: coding region bordered by a splice site on both sides. The pie charts visualize the proportions of exon types in the BRAKER3 and reference annotations.



Supplementary Figure S4: Performance of BRAKER3 broken down by whether the coding sequence of a transcript is spliced (multi-exon) or unspliced (single-exon).



Supplementary Figure S5: F1-score for predicting the signal at the boundaries of coding regions, start- and stop codons and acceptor/donor splice sites, downstream/upstream of an intron, respectively.

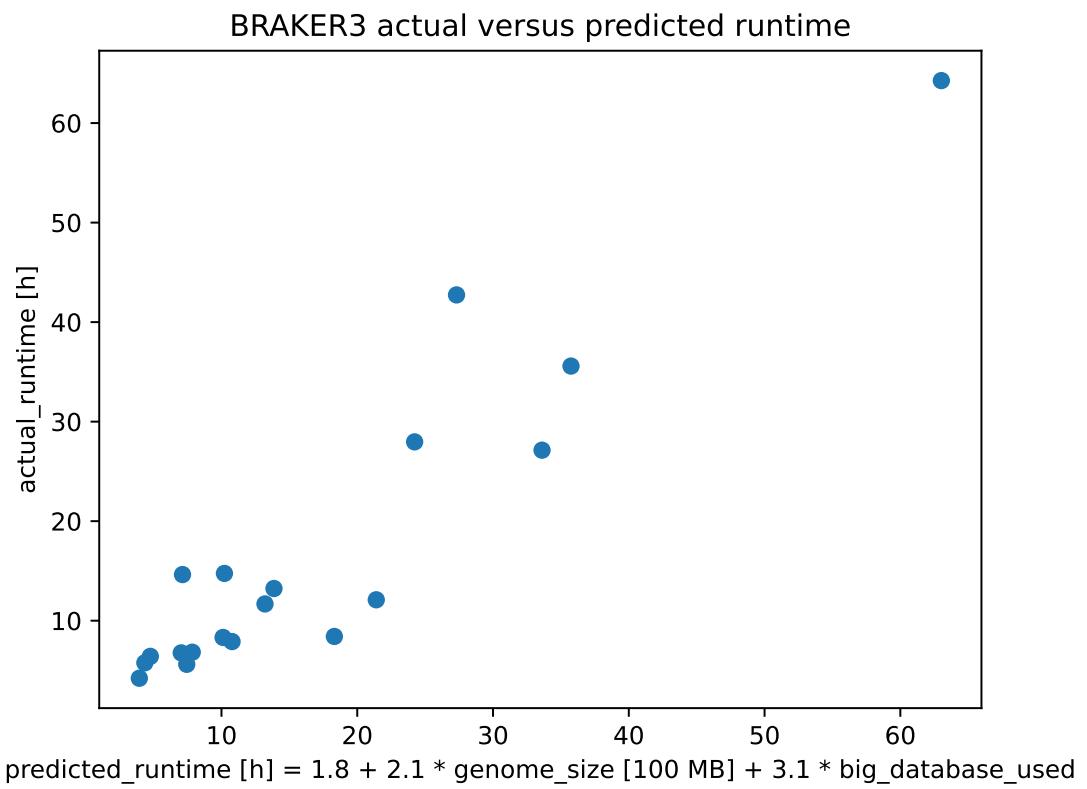


Supplementary Figure S6: The left side of the heat map refers to spliced genes, the right side to unspliced genes. The cells show the percentage of reference transcripts that are correctly predicted by both BRAKER3 and GeneMark-ETP (left columns), by GeneMark-ETP only (middle columns) and by BRAKER3 only (right columns). The pie charts to the right visualize the relative sizes of the predicted transcript sets regardless of whether they are correct.

Comparison of BRAKER3 and GeneMark-ETP gene sets grouped by RNA-Seq expression level

				Transcript level Sn					
	GeneMark-ETP and BRAKER3	only GeneMark-ETP	only BRAKER3	GeneMark-ETP and BRAKER3	only GeneMark-ETP	only BRAKER3	GeneMark-ETP and BRAKER3	only GeneMark-ETP	only BRAKER3
<i>Arabidopsis thaliana</i>	18.22	4.53	3.76	52.16	1.58	4.00	83.70	0.62	2.59
<i>Bombus terrestris</i>	17.11	3.56	5.67	45.24	2.21	7.11	63.33	1.18	6.16
<i>Caenorhabditis elegans</i>	6.60	11.99	3.73	39.58	2.58	13.29	62.74	0.78	11.99
<i>Danio rerio</i>	4.85	1.35	3.48	24.01	1.23	9.78	61.23	0.55	3.64
<i>Drosophila melanogaster</i>	21.12	3.66	6.22	53.61	1.32	8.32	69.86	0.79	7.46
<i>Gallus gallus</i>	27.67	0.80	12.84	56.48	0.33	11.58	83.17	0.24	6.29
<i>Medicago truncatula</i>	16.03	1.35	4.07	50.65	0.43	3.51	68.63	0.49	2.29
<i>Mus musculus</i>	19.32	3.35	12.44	41.97	1.81	18.96	88.15	0.20	5.76
<i>Parasteatoda tepidariorum</i>	16.86	1.70	6.82	39.44	1.27	6.41	55.56	1.00	3.88
<i>Populus trichocarpa</i>	26.00	1.54	6.00	61.97	0.66	3.84	86.27	0.43	1.88
<i>Solanum lycopersicum</i>	15.49	1.32	3.46	49.53	0.31	2.28	64.07	0.23	1.81
Average	17.20	3.19	6.23	46.79	1.25	8.10	71.52	0.59	4.89
	GeneMark-ETP and BRAKER3 lowly expressed	only GeneMark-ETP	only BRAKER3	GeneMark-ETP and BRAKER3 medium expressed	only GeneMark-ETP	only BRAKER3	GeneMark-ETP and BRAKER3 highly expressed	only GeneMark-ETP	only BRAKER3

Supplementary Figure S7: The heat map shows the percentage of reference transcripts, in the first, second and third expression tercile, that are correctly identified by BRAKER3 and GeneMark-ETP or by either program only.



Supplementary Figure S8: Actual runtime versus the runtime predicted for 19 whole genomes. The regression to predict the runtime ($R^2 = 0.87$) considered only the size of the genome and whether an OrthoDB partition was used (`big_database_used=1`) or only the proteomes of a few closely related genomes were used (`big_database_used=0`).

1.2 Supplemental Tables

Species	Annotation version	Size (Mbp)	#Genes	#Transcripts	#CDS	# Sequences in protein database		
						species excluded	order excluded	close relatives included
<i>Arabidopsis thaliana</i>	TAIR Araport 11 (Jun 2016)	119	27,445	48,149	156,731	5,283,100	4,825,128	306,719
<i>Bombus terrestris</i>	NCBI Annotation Release 102 (Apr 2017)	249	10,581	22,091	78,337	4,297,173	3,416,393	180,811
<i>Caenorhabditis elegans</i>	WormBase WS271 (May 2019)	100	20,172	33,624	130,885	15,237,008	15,145,380	115,553
<i>Danio rerio</i>	Ensembl GRCz11.96 (May 2019)	1,345	25,611	42,934	262,325	9,779,764	9,468,332	760,754
<i>Drosophila melanogaster</i>	FlyBase R6.18 (Jun 2019)	138	13,930	30,561	62,841	4,293,925	3,029,616	116,493
<i>Gallus gallus</i>	Ensembl GRCg6a.105 (March 2018)	1,050	17,279	38,534	192,095	9,787,814	9,690,812	220,641
<i>Medicago truncatula</i>	MtrunA17r5.0-ANR-EGN-r1.6 (Feb 2019)	420	44,464	44,464	174,281	5,278,627	4,680,430	134,774
<i>Mus musculus</i>	GENCODE GRCh39 version M28	2,723	22,405	58,318	243,366	9,782,804	9,059,968	510,476
<i>Parasteatoda tepidariorum</i>	NCBI Annotation Release 101 (May 2017)	1,445	18,602	27,516	143,792	4,287,893	4,240,214	1,163,197
<i>Populus trichocarpa</i>	JGI Pttrichocarpa_533_v4.1 (Nov 2019)	389	34,488	52,085	187,170	5,278,879	5,112,927	199,662
<i>Solanum lycopersicum</i>	Consortium ITAG4.0 (May 2019)	773	33,562	33,562	152,352	5,284,979	4,823,899	149,444

Supplementary Table S1: Summary of genomes, annotations and protein databases used for performance evaluation. Data extracted from Table 4 in Brúna et al. [2023b] and computed from raw data of Brúna et al. [2021, 2023b]. Genome versions, repeat masking and annotation processing are documented at <https://github.com/gatech-genemark/EukSpecies-BRAKER2> and at <https://github.com/gatech-genemark/GeneMark-ETP-exp>. The protein databases were generated with orthodb-clades from <https://github.com/tomasbruna/orthodb-clades>.

Supplementary Table S2: Donor proteins used for each species for the close relative included protein set.

Species	Reference Protein File
<i>Arabidopsis thaliana</i>	
<i>Arabidopsis lyrata</i> subsp. <i>lyrata</i>	GCF_000004255.2_v.1.0_protein.faa.gz
<i>Arabidopsis thaliana</i> x <i>Arabidopsis arenosa</i>	GCA_019202795.1_ASM1920279v1_protein.faa.gz
<i>Camelina sativa</i>	GCF_000633955.1_Cs_protein.faa.gz
<i>Arabidopsis suecica</i>	GCA_019202805.1_ASM1920280v1_protein.faa.gz
<i>Capsella rubella</i>	GCF_000375325.1_Caprub1_0_protein.faa.gz
<i>Bombus terrestris</i>	
<i>Bombus vancouverensis</i> nearcticus	GCF_011952275.1_Bvanc_JDL1245_protein.faa.gz
<i>Bombus huntii</i>	GCF_024542735.1_lyBomHunt1.1_protein.faa.gz
<i>Bombus affinis</i>	GCF_024516045.1_lyBomAff1.2_protein.faa.gz
<i>Bombus pyrosoma</i>	GCF_014825855.1_ASM1482585v1_protein.faa.gz
<i>Bombus vosnesenskii</i>	GCF_011952255.1_Bvos_JDL3184-5_v1.1_protein.faa.gz
<i>Bombus bifarius</i>	GCF_011952205.1_Bbif_JDL3187_protein.faa.gz
<i>Bombus impatiens</i>	GCF_000188095.3_BIMP_2.2_protein.faa.gz
<i>Caenorhabditis elegans</i>	
<i>Caenorhabditis auriculariae</i>	GCA_904845305.1_CAUJ_protein.faa.gz
<i>Caenorhabditis bovis</i>	GCA_902829315.1_CBOVIS_v1.1_protein.faa.gz
<i>Caenorhabditis brenneri</i>	GCA_000143925.2_C_brenneri-6.0.1b_protein.faa.gz
<i>Caenorhabditis briggsae</i>	GCF_000004555.2_CB4_protein.faa.gz
<i>Caenorhabditis remanei</i>	GCF_000149515.1_ASM14951v1_protein.faa.gz
<i>Danio rerio</i>	
<i>Cyprinus carpio</i>	GCF_018340385.1_ASM1834038v1_protein.faa.gz
<i>Carassius auratus</i>	GCF_003368295.1_ASM336829v1_protein.faa.gz
<i>Puntigrus tetrazona</i>	GCF_018831695.1_ASM1883169v1_protein.faa.gz
<i>Sinocyclocheilus rhinoceros</i>	GCF_001515625.1_SAMN03320098_v1.1_protein.faa.gz
<i>Sinocyclocheilus anshuiensis</i>	GCF_001515605.1_SAMN03320099_WGS_v1.1_protein.faa.gz
<i>Onychostoma macrolepis</i>	GCA_012432095.1_ASM1243209v1_protein.faa.gz
<i>Carassius gibelio</i>	GCF_023724105.1_carGib1.2-hapl.c_protein.faa.gz
<i>Pimephales promelas</i>	GCF_016745375.1_EPA_FHM_2.0_protein.faa.gz
<i>Labeo rohita</i>	GCF_022985175.1_IGBB_LRoh_1.0_protein.faa.gz
<i>Megalobrama amblycephala</i>	GCF_018812025.1_ASM1881202v1_protein.faa.gz
<i>Sinocyclocheilus grahami</i>	GCF_001515645.1_SAMN03320097_WGS_v1.1_protein.faa.gz
<i>Ctenopharyngodon idella</i>	GCF_019924925.1_HZGC01_protein.faa.gz
<i>Drosophila melanogaster</i>	
<i>Drosophila ananassae</i>	GCF_017639315.1_ASM1763931v2_protein.faa.gz
<i>Drosophila grimshawi</i>	GCF_018153295.1_ASM1815329v1_protein.faa.gz
<i>Drosophila pseudoobscura</i>	GCF_009870125.1_UCI_Dpse_MV25_protein.faa.gz
<i>Drosophila virilis</i>	GCF_003285735.1_DvirRS2_protein.faa.gz
<i>Drosophila willistoni</i>	GCF_018902025.1_UCI_dwil_1.1_protein.faa.gz
<i>Gallus gallus</i>	
<i>Lagopus muta</i>	GCF_023343835.1_bLagMut1_primary_protein.faa.gz
<i>Tympanuchus pallidicinctus</i>	GCF_026119805.1_pur_lepc_1.0_protein.faa.gz
<i>Lagopus leucura</i>	GCF_019238085.1_USGS_WTPPT01_protein.faa.gz
<i>Centrocercus urophasianus</i>	GCF_019232065.1_USGS_Curo_1.0_protein.faa.gz
<i>Centrocercus urophasianus</i>	GCF_019232065.1_USGS_Curo_1.0_protein.faa.gz
<i>Coturnix japonica</i>	GCF_001577835.2_Coturnix_japonica_2.1_protein.faa.gz

<i>Meleagris gallopavo</i>	GCF_000146605.3_Turkey_5.1_protein.faa.gz
<i>Medicago truncatula</i>	
<i>Trifolium pratense</i>	GCF_020283565.1_ARS_RC_1.1_protein.faa.gz
<i>Pisum sativum</i>	GCF_024323335.1_CAA_S_Psat_ZW6_1.0_protein.faa.gz
<i>Cicer arietinum</i>	GCF_000331145.1_ASM33114v1_protein.faa.gz
<i>Mus musculus</i>	
<i>Arvicathis niloticus</i>	GCF_011762505.1_mArvNil1.pat.X_protein.faa.gz
<i>Grammomys surdaster</i>	GCF_004785775.1_NIH_TR_1.0_protein.faa.gz
<i>Mastomys coucha</i>	GCF_008632895.1_UCSF_Mcou_1_protein.faa.gz
<i>Mus pahari</i>	GCF_900095145.1_PAHARI_EIJ_v1.1_protein.faa.gz
<i>Apodemus sylvaticus</i>	GCF_947179515.1_mApoSyl1.1_protein.faa.gz
<i>Mus caroli</i>	GCF_900094665.1_CAROLI_EIJ_v1.1_protein.faa.gz
<i>Rattus rattus</i>	GCF_011064425.1_Rrattus_CSIRO_v1_protein.faa.gz
<i>Rattus norvegicus</i>	GCF_015227675.2_mRatBN7.2_protein.faa.gz
<i>Homo sapiens</i>	GCF_000001405.40_GRCh38.p14_protein.faa.gz
<i>Parasteatoda tepidariorum</i>	
<i>Trichonephila inaurata</i>	GCA_019973955.1_Tnin_1.0_protein.faa.gz
<i>Caerostris extrusa</i>	GCA_021605095.1_Cext_1.0_protein.faa.gz
<i>Caerostris darwini</i>	GCA_021605075.1_Cdar_1.0_protein.faa.gz
<i>Oedothorax gibbosus</i>	GCA_019343175.1_Ogib_1.0_protein.faa.gz
<i>Trichonephila clavata</i>	GCA_019973975.1_Tnct_1.0_protein.faa.gz
<i>Trichonephila clavipes</i>	GCA_019973935.1_Tncv_1.0_protein.faa.gz
<i>Araneus ventricosus</i>	GCA_013235015.1_Ave_3.0_protein.faa.gz
<i>Nephila pilipes</i>	GCA_019974015.1_Npil_1.0_protein.faa.gz
<i>Populus trichocarpa</i>	
<i>Populus tomentosa</i>	GCA_018804465.1_PTtv2_protein.faa.gz
<i>Populus euphratica</i>	GCF_000495115.1_PopEup_1.0_protein.faa.gz
<i>Populus alba</i>	GCF_005239225.1_ASM523922v1_protein.faa.gz
<i>Populus deltoides</i>	GCA_015852605.2_ASM1585260v2_protein.faa.gz
<i>Solanum lycopersicum</i>	
<i>Solanum stenotomum</i>	GCF_019186545.1_ASM1918654v1_protein.faa.gz
<i>Solanum tuberosum</i>	GCF_000226075.1_SolTub_3.0_protein.faa.gz
<i>Solanum verrucosum</i>	GCF_900185275.1_falcon_dt.bn_protein.faa.gz
<i>Solanum pennellii</i>	GCF_001406875.1_SPENN_V200_protein.faa.gz

	Exon			Gene			Transcript		
	Sn	Sp	F1	Sn	Sp	F1	Sn	Sp	F1
<i>Arabidopsis thaliana</i>									
MAKER2*	78.05	82.02	79.99	60.57	57.69	59.09	40.72	57.69	47.74
Funannotate	82.07	93.21	87.29	75.26	79.58	77.36	50.59	79.58	61.86
BRAKER3	83.03	94.38	88.34	82.93	86.27	84.57	58.70	80.78	67.99
<i>Bombus terrestris</i>									
MAKER2*	74.12	75.22	74.67	50.24	45.57	47.79	31.55	36.75	33.95
Funannotate	76.23	72.72	74.43	51.73	30.80	38.61	32.11	30.85	31.47
BRAKER3	79.39	88.71	83.79	73.07	67.32	70.08	57.86	57.31	57.58
<i>Caenorhabditis elegans</i>									
MAKER2*	73.13	76.63	74.84	41.31	41.10	41.20	29.34	38.56	33.32
Funannotate	79.71	84.19	81.89	49.44	52.16	50.76	35.01	52.18	41.90
BRAKER3	76.39	95.35	84.82	68.97	85.61	76.39	52.52	77.33	62.55
<i>Drosophila melanogaster</i>									
MAKER2*	75.28	73.11	74.18	61.10	52.77	56.63	38.26	52.77	44.36
Funannotate	73.87	82.66	78.02	62.87	62.98	62.92	39.38	62.99	48.46
BRAKER3	81.64	95.18	87.89	83.36	90.55	86.81	59.71	81.58	68.95
<i>Gallus gallus</i>									
MAKER2*	85.98	78.83	82.25	49.42	38.65	43.38	41.99	32.64	36.73
Funannotate	58.75	71.22	64.39	32.23	23.20	26.98	27.19	23.23	25.05
BRAKER3	93.82	93.99	93.90	85.80	78.88	82.19	78.39	66.65	72.04
<i>Medicago truncatula</i>									
MAKER2*	69.63	73.87	71.69	33.48	49.12	39.82	33.48	45.67	38.64
Funannotate	74.71	63.54	68.67	35.26	37.40	36.30	35.26	37.40	36.30
BRAKER3	74.76	90.14	81.73	47.86	75.25	58.51	47.86	68.22	56.25
<i>Populus trichocarpa</i>									
MAKER2*	74.04	71.96	72.99	41.76	42.67	42.21	32.64	38.27	35.23
Funannotate	82.44	79.30	80.84	60.83	54.90	57.71	47.38	54.90	50.86
BRAKER3	84.56	92.94	88.55	76.99	85.60	81.07	63.07	76.49	69.13
<i>Solanum lycopersicum</i>									
MAKER2*	74.16	45.21	56.17	33.88	18.85	24.22	33.88	18.85	24.22
Funannotate	74.62	65.91	70.00	36.12	34.10	35.08	36.12	34.10	35.08
BRAKER3	74.87	85.25	79.72	45.90	61.67	52.63	45.90	53.49	49.41

Supplementary Table S7: Sensitivity, specificity, and F1-score for MAKER2, Funannotate, and BRAKER3 using the **close relatives included** protein databases. *The values for MAKER2 may overestimate the realistic performance on new genomes (see main text).

	Exon			Gene			Transcript		
	Sn	Sp	F1	Sn	Sp	F1	Sn	Sp	F1
<i>Arabidopsis thaliana</i>									
	18.0	50.42	26.53	6.03	18.42	9.09	4.16	16.68	6.66
<i>Caenorhabditis elegans</i>	28.62	50.67	36.58	8.10	12.4	9.80	5.89	11.54	7.80
<i>Danio rerio</i>	15.16	44.12	22.57	3.26	7.17	4.48	1.99	7.06	3.10
<i>Drosophila melanogaster</i>	24.28	53.95	33.49	11.32	23.01	15.17	7.48	21.18	11.06
<i>Medicago truncatula</i>	17.79	48.88	26.09	3.92	16.87	6.36	3.92	15.54	6.26
<i>Populus trichocarpa</i>	26.99	50.07	35.07	7.37	15.34	9.96	5.8	14.75	8.33
<i>Solanum lycopersicum</i>	31.21	43.98	36.51	6.41	12.68	8.52	6.41	10.12	7.85
Average	23.15	48.87	30.98	6.63	15.13	9.05	5.09	13.84	7.29

Supplementary Table S8: Sensitivity, specificity, and F1-score of FINDER for runs using the same input data (genomic sequence + proteins + RNA-Seq) as in the experiments from Table 3. However, FINDER exited with an error for 4 out of the 11 species tested, and we therefore do not report its performance for those species.

	Exon			Gene			Transcript		
	Sn	Sp	F1	Sn	Sp	F1	Sn	Sp	F1
<i>Arabidopsis thaliana</i>	83.4	87.70	85.50	78.78	75.18	76.94	54.18	71.77	61.75
<i>Bombus terrestris</i>	84.49	84.17	84.33	67.38	55.01	60.57	46.56	50.28	48.35
<i>Caenorhabditis elegans</i>	84.97	91.99	88.34	70.36	74.46	72.35	51.99	70.34	59.79
<i>Danio rerio</i>	81.74	75.80	78.66	58.06	36.21	44.6	36.29	33.04	34.59
<i>Drosophila melanogaster</i>	83.49	89.93	86.59	80.87	80.71	80.79	54.09	75.66	63.08
<i>Gallus gallus</i>	96.17	89.11	92.51	81.70	61.03	69.87	71.73	57.34	63.73
<i>Medicago truncatula</i>	81.16	69.84	75.08	48.93	46.70	47.79	48.93	43.68	46.16
<i>Mus musculus</i>	94.51	85.48	89.77	79.46	46.23	58.45	79.46	43.42	56.15
<i>Parasteatoda tepidariorum</i>	78.67	65.55	71.51	48.13	26.32	34.03	39.77	24.53	30.34
<i>Populus trichocarpa</i>	87.70	81.99	84.75	75.77	63.60	69.15	60.80	60.97	60.88
<i>Solanum lycopersicum</i>	79.64	66.54	72.50	46.12	37.27	41.23	46.12	34.58	39.52
Average	85.09	80.74	82.69	66.87	54.79	59.62	53.63	51.42	51.30

Supplementary Table S9: Sensitivity, specificity, and F1-score of the AUGUSTUS predictions made as part of the BRAKER3 pipeline. The results correspond to Table 3 – proteins from the same order as the target species were excluded.

	Exon			Gene			Transcript		
	Sn	Sp	F1	Sn	Sp	F1	Sn	Sp	F1
Funannotate	73.20	74.40	73.79	47.11	44.69	45.87	35.75	44.70	39.73
Funannotate updated	73.20	74.40	73.79	47.11	44.69	45.87	35.75	44.70	39.73
Funannotate -repeats2evm	71.12	77.06	73.97	45.49	45.64	45.56	34.45	45.64	39.27
Funannotate -repeats2evm updated	73.20	74.40	73.79	47.11	44.69	45.87	35.75	44.70	39.73

Supplementary Table S10: Average sensitivity, specificity, and F1-score for four different predictions generated by Funannotate using the **close relatives included** protein databases for the same species as listed in Table 7. The prediction step of Funannotate was run with and without the option to pass gene predictions of repetitive regions to EVidenceModeler (`--repeats2evm`). The resulting predictions were both post-processed using Funannotate's update protocol, which updates the predicted gene models using RNA-Seq data.

	<i>Arabidopsis thaliana</i>	<i>Bombus terrestris</i>	<i>Caenorhabditis elegans</i>	<i>Danio rerio</i>					
	Sn	Sp	F1	Sn	Sp	F1	Sn	Sp	F1
BRAKER1	01:23			01:36			10:28		
BRAKER2	06:39			04:57			19:47		
GeneMark-ETP	04:05			03:36			09:54		
BRAKER3	05:37			06:46			27:08		
	<i>Drosophila melanogaster</i>	<i>Gallus gallus</i>	<i>Medicago truncatula</i>	<i>Mus musculus</i>					
BRAKER1	01:55	10:12	03:20	19:41					
BRAKER2	05:28	14:10	10:59	30:24					
GeneMark-ETP	02:37	12:11	07:55	22:13					
BRAKER3	06:50	42:44	13:14	64:16					
	<i>Parasteatoda tepidariorum</i>	<i>Populus trichocarpa</i>	<i>Solanum lycopersicum</i>	Average					
BRAKER1	14:47	04:41	03:47	06:55					
BRAKER2	23:16	08:52	09:19	12:45					
GeneMark-ETP	10:38	07:51	06:37	08:14					
BRAKER3	35:35	11:41	12:06	21:53					

Supplementary Table S11: Runtime of BRAKER1, BRAKER2, GeneMark-ETP, and BRAKER3 for all test species. The runtime is written as hours and minutes. The hardware is described in the caption of Suppl. Figure 6.

Species	Runtime (h:m)
<i>Bombus terrestris</i>	4:00
<i>Drosophila melanogaster</i>	2:06
<i>Gallus gallus</i>	13:54
<i>Medicago truncatula</i>	2:48
<i>Populus trichocarpa</i>	3:36

Supplementary Table S12: Runtime of MAKER2 with *close relatives included* database in MPI mode using LINUX nodes with 96 cores available through the Azure cloud. Training of *ab initio* gene finders and transcriptome assembly are not included in these figures.

	<i>Arabidopsis thaliana</i>	<i>Bombus terrestris</i>	<i>Caenorhabditis elegans</i>	<i>Drosophila melanogaster</i>
Funannotate	05:06	04:59	02:02	02:52
BRAKER3	05:46	14:38	04:13	06:25
	<i>Gallus gallus</i>	<i>Medicago truncatula</i>	<i>Populus trichocarpa</i>	<i>Solanum lycopersicum</i>
Funannotate	41:07	07:11	07:03	09:28
BRAKER3	27:58	07:54	08:19	08:25
	Average			
Funannotate	09:59			
BRAKER3	10:27			

Supplementary Table S13: Runtime of Funannotate and BRAKER3 for the experiments of Table 7 (*close relatives included*). The runtime is written as hours and minutes. The hardware is described in the caption of Suppl. Figure 6.

Species	SRA ID	#spots	#bases	Date
<i>A. tha.</i>	SRR8714016	26,518,474	8,008,579,148	2020-02-27
	SRR8759751	21,388,481	6,458,011,735	2019-04-01
	SRR4010853	25,658,415	6,465,920,580	2017-05-10
	SRR7289569	9,222,308	1,391,511,444	2018-12-06
	SRR12547664	874,772	66,482,672	2021-04-07
	SRR12076896	21,857,417	6,557,225,100	2021-05-24
<i>B. ter.</i>	SRR5125126	28,930,495	5,207,489,100	2017-10-30
	SRR5125123	25,185,030	4,533,305,400	2017-10-30
	SRR16931591	25,056,341	7,516,902,300	2021-11-12
	SRR5125133	26,059,038	4,690,626,840	2017-10-30
	SRR5125134	23,745,975	4,274,275,500	2017-10-30
	SRR8085469	32,768,117	6,553,623,400	2019-03-05
<i>C. ele.</i>	SRR7446944	44,853,824	8,970,764,800	2020-06-01
	ERR2756716	4,785,656	1,445,268,112	2018-08-29
	SRR6815567	5,901,194	890,097,086	2018-03-08
	SRR6474814	34,199,521	8,618,279,292	2018-01-16
	SRR10238291	47,652,843	4,955,895,672	2019-11-07
<i>D. rer.</i>	SRR3179613	9,994,443	2,018,877,486	2017-02-10
	ERR1857957	2,536,494	380,474,100	2017-03-03
	SRR9159941	19,527,035	5,858,110,500	2019-12-01
	SRR9159937	37,156,875	11,147,062,500	2019-12-01
	ERR958944	10,490,607	1,363,778,910	2015-07-17
	SRR8106574	1,281,189	186,371,148	2020-01-20
<i>D. mel.</i>	SRR19416937	21,377,196	6,413,158,800	2022-05-30
	SRR19416947	22,611,252	6,783,375,600	2022-05-30
	SRR19416944	17,754,393	5,326,317,900	2022-05-30
	SRR19446462	40,819,492	4,163,588,184	2022-05-30
	SRR19416948	21,257,788	6,377,336,400	2022-05-30
<i>M. tru.</i>	SRR3735569	13,021,000	2,630,242,000	2016-07-02
	SRR2016009	4,289,979	428,997,900	2016-05-12
	SRR10416790	9,757,388	2,927,216,400	2019-11-23
	SRR3726824	44,634,145	13,390,243,500	2016-06-28
	SRR2015998	7,916,226	791,622,600	2016-05-12
	SRR15462191	21,367,770	6,410,331,000	2021-08-15
<i>P. tep.</i>	SRR5458595	94,367,928	18,873,585,600	2017-06-14
	SRR1824488	33,099,205	6,686,039,410	2015-03-03
	SRR12687629	41,410,278	12,423,083,400	2021-06-22
	SRR8755634	45,919,505	8,852,599,087	2019-03-20
	SRR5602551	9,600,853	2,400,213,250	2017-10-16
	SRR1824489	60,177,256	12,155,805,712	2015-03-03
<i>P. tri.</i>	SRR3019959	32,156,743	4,591,697,381	2016-01-19
	SRR3019957	44,863,324	6,484,994,933	2016-01-19
	SRR3019251	29,773,370	4,305,817,693	2016-01-19
	SRR3019585	36,843,662	5,279,588,065	2016-01-19
	SRR3019304	41,358,002	5,970,634,746	2016-01-19
	SRR12671667	17,429,154	5,263,604,508	2020-09-18
<i>G. gal.</i>	SRR5340686	27,956,970	2,851,610,940	2017-03-15
	ERR2113192	20,988,313	3,148,246,950	2017-09-07
	ERR2113173	21,562,175	3,234,326,250	2017-09-07
	SRR5190436	23,938,529	4,787,705,800	2017-09-06
	SRR1822373	48,874,648	7,819,943,680	2015-05-05
	SRR5437696	35,378,254	5,306,738,100	2017-09-18
<i>M. mus.</i>	SRR5197958	10,577,301	3,184,479,151	2017-08-31
	SRR3094250	1,568,252	316,786,904	2016-11-14
	SRR6067921	5,886,768	888,143,922	2018-08-03
	SRR10115888	44,307,998	8,861,599,600	2020-10-02
	ERR3005082	8,598	1,289,700	2019-01-02
	SRR9202226	5,355,073	546,217,446	2019-08-07

Supplementary Table S14:
This table lists all RNA-Seq libraries used for each species in all experiments. It includes the ID of the library at the Sequence Read Archive, the number of spots, number of bases, and publication date.

Species	AUGUSTUS	GeneMark.hmm	SNAP
<i>Arabidopsis thaliana</i>	arabidopsis*	from GeneMark-ES run	<i>A.thaliana.hmm*</i>
<i>Bombus terrestris</i>	bombus.terrestris2*	from GeneMark-EP+ run	trained with genes from ref. annotation
<i>Caenorhabditis elegans</i>	c.elegans.trsk*	from GeneMark-ES run	Ce.hmm*
<i>Drosophila melanogaster</i>	fly*	from GeneMark-ES run	fly*
<i>Gallus gallus</i>	chicken*	medium GC model from GeneMark-ETP run	trained with genes from ref. annotation
<i>Medicago truncatula</i>	trained with genes from ref. annotation	from GeneMark-EP+ run	trained with genes from ref. annotation
<i>Populus trichocarpa</i>	trained with genes from ref. annotation	from GeneMark-EP+ run	trained with genes from ref. annotation
<i>Solanum lycopersicum</i>	tomato*	from GeneMark-EP+ run	trained with genes from ref. annotation

Supplementary Table S15: Configuration of AUGUSTUS, GeneMark.hmm and SNAP gene models using parameter from the AUGUSTUS and the SNAP distribution packages (marked with *) or with manually training using a training gene set.

425 1.3 Supplementary Methods

Running BRAKER

BRAKER v3.0.2 was installed from GitHub (<https://github.com/Gaius-Augustus/BRAKER>) and the BRAKER pipelines were run as follows. The GeneMark-ETP results were taken from the BRAKER3 runs:

430

BRAKER1:

```
braker.pl --genome=genome.softmasked.fasta --threads=48 \
--rnaseq_sets_dirs=/location/of/local/RNA_Seq/Files \
--rnaseq_sets_ids=RNA_Seq_IDS
```

435

BRAKER2:

```
braker.pl --genome=genome.softmasked.fasta --prot_seq=proteins.fa \
--threads=48
```

BRAKER3:

```
braker.pl --genome=genome.softmasked.fasta --prot_seq=proteins.fa \
--rnaseq_sets_dirs=/location/of/local/RNA_Seq/Files \
--rnaseq_sets_ids=RNA_Seq_IDS --threads=48
```

Running Funannotate

Funannotate v1.8.14 was installed using a Singularity container as follows:

445

```
# only once, to get the singularity container
singularity pull docker://nextgenusfs/funannotate
```

```
export GENEMARK_PATH=/path/to/GeneMark-ES-ET-EP_v4.71_lic
```

450

```
species="name of species"
buscoSeedSpecies="name of seed species"
buscodb="name of busco db"
genome="/path/to/genome.fasta.masked"
proteins="/path/to/proteins.fa"
bamfile="/path/to/bamfile.bam"
```

455

```
# calculateGenomeSizeFromFasta.pl adds up the length of all sequences in a fasta
genomeSize=$(perl ~/calculateGenomeSizeFromFasta.pl $genome)
maxIntronLen_f=$(echo "3.6 * sqrt($genomeSize)" | bc -l)
maxIntronLen=$(printf "%0.f" "$maxIntronLen_f")
```

460

```
singularity run funannotate_latest.sif funannotate sort \
--input $genome \
--out genome.sorted.fa \
--simplify
```

465

We produced four different results for each species, using the option `--repeats2evm` during the prediction step and a recommended update step:

470

```
mkdir -p fun tmp
# run prediction
singularity run funannotate_latest.sif funannotate predict \
--input genome.sorted.fa --out fun --species $species \
--busco_seed_species $buscoSeedSpecies --busco_db $buscodb \
--organism other --protein_evidence $proteins \
--rna_bam $bamfile --max_intronlen $maxIntronLen \
--cpus 72 --tmpdir tmp --no-progress \
[--repeats2evm]
```

475

```

# run update step
singularity run funannotate_latest.sif funannotate update \
--input fun/ --left $readL --right $readR \
--max_intronlen $maxIntronLen --species $species \
--memory 50G --cpus 72 --no-progress

```

Running FINDER

FINDER v1.1.0 was installed from GitHub (<https://github.com/sagnikbanerjee15/Finder>) and run as follows:

```

run_finder --protein proteins.fa --framework singularity --output_directory finder --cpu 48 \
485   --metadatafile metadata.csv --genome genome.softmasked.fasta --genemark_license gm_key \
   --genemark_path /location/of/GeneMark-ES/ET/EP --organism_model {VERT,INV,PLANTS,FUNGI}

```

The information in the `metadata.csv` files was manually generated for each species and includes details about the RNA-Seq libraries. The files consist of the following fields:

```

BioProject, SRA Accession, Tissues, Description, Date, Read Length (bp),
490   Ended, RNA Seq, process, Location

```

Running MAKER2

We ran MAKER2 version 3.01.04 using AUGUSTUS version 3.5.0, GeneMark.hmm version 3.68, SNAP version 2.59.5, Exonerate version 2.2.0, BLAST version 2.14.0, Tandem Repeats Finder version 4.09.

We provided MAKER with a GFF file containing the coordinates of repeats, as well as their predicted repeat types from RepeatMasker. To ensure compatibility with MAKER, we reformatted the RepeatMasker output to a specific GFF format using the following script from <https://github.com/gatech-genemark/BRAKER2-exp.git> and command line:

```
rmasker_out2maker_gff.pl < genome.fasta.out > rmasker4maker.gff
```

This script is available from the repository <https://github.com/gatech-genemark/GeneMark-ETP-exp>.

500 The default MAKER2 configuration was adjusted with following settings:

```

genome=genome.fasta
est=transcriptome.fasta
protein=proteindb.fasta
model_org= #empty
505 rm_gff=repeatmasker.gff
snaphmm=snap.model
gmhmm=genemark.mod
augustus_species=model_name
est2genome=1
510 protein2genome=1
keep_preds=0

```

Here, `transcriptome.fasta` contained the same transcriptome assemblies that were used with BRAKER3 and constructed with HISAT2 and StringTie2. MAKER2 was then run with:

```
mpexec.mpich -n 96 maker
```

515 Preparing protein data

OrthoDB v.11 was partitioned into proteins of species from the clade *Arthropoda*, *Metazoa*, *Vertebrata*, and *Viridiplantae*. The partitioning is available from https://bioinf.uni-greifswald.de/bioinf/partitioned_odb11/. Subsequently, two protein sets for each species from Suppl. Table 1 were generated, excluding either only the target species (species-excluded) or all species of the same taxonomic order (order-excluded).

520 These sets were prepared using the orthodb-clades pipeline, downloaded from GitHub (<https://github.com/tomasbruna/orthodb-clades>).

```
snakemake --cores 48
```

Accuracy evaluation

525 The performance measurements were computed using scripts from the BRAKER suite:

```
compute_accuracies.sh ref_annot.gtf pseudo.gff3 gene_set.gtf gene trans cds
```

Measuring expression

530 Expression levels were estimated with `kallisto` version 0.50.0 [Bray et al., 2016]. For each species, the mRNA sequences of the respective reference annotation were quantified using a pool of RNA-Seq libraries. More specifically, all paired libraries that were used for annotation were pooled and TPM (transcript per million) values estimated with the following command line.

```
kallisto quant -i kallisto.idx -o work/ -t 8 --verbose input/*.fastq
```

Subsequently, the transcripts for each species were partitioned in three expression *terciles*: the first with lowest TPM value, the second with medium TPM value and the third with highest TPM value.