

## Supplementary material

**Supporting Information****Paecilins Q and R: Antifungal Chromanones Produced by the Endophytic Fungus *Pseudofusicoccum stromaticum* CMRP4328**

**Jucélia Iantas,<sup>1, 2, 3\*</sup> Daiani Cristina Savi,<sup>4, 5</sup> Larissa V. Ponomareva,<sup>2, 3</sup> Jon S. Thorson,<sup>2, 3</sup> Jürgen Rohr,<sup>2</sup> Chirlei Glienke,<sup>1, 4</sup> Khaled A. Shaaban<sup>2, 3\*</sup>**

\*These two authors contributed equally to this work.

**Affiliations**

<sup>1</sup>Postgraduate Program in Microbiology, Department of Pathology, Federal University of Paraná (UFPR), Curitiba, Brazil

<sup>2</sup>Department of Pharmaceutical Sciences, College of Pharmacy, University of Kentucky, Lexington, Kentucky, United States

<sup>3</sup>Center for Pharmaceutical Research and Innovation, College of Pharmacy, University of Kentucky, Kentucky, United States

<sup>4</sup>Postgraduate Program in Genetics, Department of Genetics, Federal University of Paraná (UFPR), Curitiba, Brazil

<sup>5</sup>Department of Biomedicine, Centro Universitário Católica de Santa Catarina, Joinville, Brazil

## Supplementary material

**Correspondence*****Chirlei Glienke, PhD***

Department of Genetics

Federal University of Paraná

Av. Cel. Francisco H. dos Santos, 100

81531-990, Curitiba

Brazil

Phone: + 55 413 361 1562

Fax: +55 413 361 1783

[ch.glienke@gmail.com](mailto:ch.glienke@gmail.com)

***Khaled A. Shaaban, PhD***

Department of Pharmaceutical Sciences and

Center for Pharmaceutical Research and Innovation

University of Kentucky

Lexington, Kentucky 40536

United States

Phone: +1 859 218 0916

Fax: +1 859 323 0204

[khaled\\_shaaban@uky.edu](mailto:khaled_shaaban@uky.edu)

## Supplementary material

Table of Contents:	Page
<b>EXPERIMENTAL SECTION</b>	S4
<b>General Experimental Procedures</b>	S4
<b>Cancer Cell Line Viability Assay</b>	S5
<b>Physicochemical Properties of Compounds 3-10</b>	S5-S8
<b>Supplementary References</b>	S8
<b>Fig. 1S</b> Workup scheme of the metabolites produced by the fungus <i>Pseudofusicoccum stromaticum</i> CMRP4328.	S9
<b>Fig. 2S A</b> <sup>1</sup> H, <sup>1</sup> H-COSY (—) and selected HMBC (→) correlations of compounds <b>6-10</b> . <b>B</b> TOCSY (—) and selected NOESY (↔) correlations of compounds <b>6-7</b> and <b>9</b> .	S10
<b>Fig. 3S</b> UV-vis (MeOH) spectra of compounds <b>1-5</b> .	S11
<b>Fig. 4S</b> Chemical structures of the reported natural products, paecilin B ( <b>11</b> ), paecilin E ( <b>12</b> ) along with the synthetic ( <i>R</i> )-5-hydroxy-2-(hydroxymethyl)-2-([2 <i>R</i> ,3 <i>R</i> ]-3-methyl-5-oxotetrahydrofuran-2-yl)chroman-4-one ( <b>13</b> ).	S11
<b>Fig. 5S.</b> Antifungal activity of compounds <b>2</b> , <b>5</b> , and <b>7-10</b> against the phytopathogen <i>Phyllosticta citricarpa</i> .	S12
<b>Table 1S</b> <sup>13</sup> C and <sup>1</sup> H (500 MHz) NMR spectroscopic data for cytochalasin H ( <b>6</b> ) and cytochalasin J ( <b>7</b> ) in CD <sub>3</sub> OD (δ in ppm).	S13
<b>Figs. 6S-111S</b> HPLC/UV, HPLC/MS, HRMS, and NMR spectra of the compounds.	S14- S119

## Supplementary material

**EXPERIMENTAL SECTION****General Experimental Procedures**

Optical rotation measurements were recorded on a Jasco DIP-370 Digital Polarimeter (Jasco). UV spectra were recorded on an Ultrospec 8000 spectrometer (GE). Ultraviolet-visible (UV-VIS) spectra of compounds **6-10** were taken directly from analytical HPLC-PDA runs and show relative intensities. NMR spectra were measured using Varian Vnmr 500 ( $^1\text{H}$ , 500 MHz;  $^{13}\text{C}$ , 125.7 MHz) and Vnmr 400 ( $^1\text{H}$ , 399.8 MHz;  $^{13}\text{C}$ , 100.5 MHz) spectrometers where  $\delta$ -values were referenced to respective solvent signals [ $\text{CDCl}_3$ ,  $\delta_{\text{H}}$  7.24 ppm,  $\delta_{\text{C}}$  77.23 ppm;  $\text{CD}_3\text{OD}$ ,  $\delta_{\text{H}}$  3.31 ppm,  $\delta_{\text{C}}$  49.15 ppm; acetone- $d_6$ ,  $\delta_{\text{H}}$  2.05 ppm,  $\delta_{\text{C}}$  29.92/206.68; DMSO- $d_6$ ,  $\delta_{\text{H}}$  2.50 ppm,  $\delta_{\text{C}}$  39.51 ppm]. High-resolution electrospray ionization (HRESI) mass spectra were recorded on a Thermo Scientific ([www.thermoscientific.com](http://www.thermoscientific.com)) Q Exactive (orbitrap mass spectrometer), with sample introduction by direct infusion at 3  $\mu\text{L}/\text{min}$ . Full-scan mass spectra were recorded in positive and negative ion modes (Instrument parameters included spray voltage: 3.8 kV “for positive ion mode”, 3.2 kV “for negative ion mode”; capillary temperature: 225°C; nominal resolution: 140000). HPLC-UV/MS analyses were accomplished with an Agilent InfinityLab LC/MSD mass spectrometer (MS Model G6125B; Agilent Technologies) equipped with an Agilent 1260 Infinity II Series Quaternary LC system and a Phenomenex NX-C18 column (250  $\times$  4.6 mm, 5  $\mu\text{m}$ ) [Method: solvent A:  $\text{H}_2\text{O}/0.1\%$  formic acid, solvent B:  $\text{CH}_3\text{CN}$ ; flow rate: 0.5  $\text{mL min}^{-1}$ ; 0-30 min, 5-100% B (linear gradient); 30-35 min, 100% B; 35-36 min, 100%-5% B; 36-40 min, 5% B]. HPLC-UV analyses were carried out in a Agilent 1260 system equipped with a photodiode array detector (PDA) and a Phenomenex  $\text{C}_{18}$  column (Phenomenex; 250  $\times$  4.6 mm, 5  $\mu\text{m}$ ; solvent A:  $\text{H}_2\text{O}/0.1\%$  TFA, solvent B:  $\text{CH}_3\text{CN}$ ; flow rate: 1.0  $\text{mL min}^{-1}$ ; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100%-5% B; 36-40 min, 5% B). Semipreparative HPLC was

## Supplementary material

accomplished using Phenomenex C<sub>18</sub> column (10 × 250 mm, 5 μm) on a Varian ProStar Model 210 equipped with a PDA detector and a gradient elution profile (solvent A: 0.05% TFA/H<sub>2</sub>O, solvent B: CH<sub>3</sub>CN; flow rate: 5.0 mL min<sup>-1</sup>; 0-2 min, 25% B; 2-15 min, 25-100% B; 15-17 min, 100% B; 17-18 min, 100%-25% B; 18-19 min, 25% B). All solvents used were of ACS grade and purchased from the Pharmco-AAPER. C<sub>18</sub>-functionalized silica gel (40 ~ 63 μm) was purchased from Material Harvest Ltd.. Amberlite XAD16N resin (20-60 mesh) was purchased from Sigma-Aldrich. Size exclusion chromatography was performed on Sephadex LH-20 (25-100 μm; GE Healthcare). A549 and PC3 cells were obtained from ATCC. All other reagents used were reagent grade and purchased from Sigma-Aldrich.

**Cancer Cell Line Viability Assay**

Mammalian cell line cytotoxicity [A549 (non-small cell lung) and PC3 (prostate) human cancer cell lines] assays were accomplished in triplicate following our previously reported protocols [1-4] Actinomycin D and H<sub>2</sub>O<sub>2</sub> (A549 and PC3) was used as positive controls. To assess the viability of human lung non-small cell carcinoma A549 and prostate adenocarcinoma PC3 cells against compounds **1-10**, the conversion of resazurin (7-hydroxy-10-oxido-phenoxazin-10-ium-3-one) to its fluorescent product resorufin was monitored. DMEM/F-12 Kaighn's modification media (Life Technologies) with 10% heat-inactivated fetal bovine serum (FBS), 100 U/mL penicillin, 100 μg/mL streptomycin, and 2 mM L-glutamine was used to grow A549 and PC3 cells (ATCC). Cells were seeded at a density of 5 × 10<sup>3</sup> cells per well in 96-well clear bottom culture plates, incubated 24 h at 37°C in a humidified atmosphere containing 5% CO<sub>2</sub> and were subsequently exposed to known toxins (1.5 mM hydrogen peroxide or 10 μg/mL actinomycin D, positive control) and test compounds for 72 h. To assess cell viability, 150 μM of resazurin (Sigma) were added to each well, plates were shaken briefly for 10 s, and incubated for another 3 h at 37°C to allow

## Supplementary material

viable cells to convert resazurin into resorufin. The fluorescence intensity for resorufin was detected on a scanning microplate spectrofluorometer FLUOstar Omega (BMG Labtech) using an excitation wavelength of 560 nm and an emission wavelength of 590 nm.

**Physicochemical Properties of Compounds 3-10:**

**Phomoxanthone A (Pomopxanthone A; 3).**  $C_{38}H_{38}O_{16}$  (750); pale yellow solid; *HPLC*- $R_t$  = 33.35 min (Supplementary Fig. 30S-31S); UV/vis (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 203 (4.31), 259 (4.05), 337 (4.36) nm (Supplementary Figs. 3S);  $^1H$  NMR ( $CDCl_3$ , 500 MHz) and  $^{13}C$  NMR ( $CDCl_3$ , 100 MHz), see Tables 1 and 2; (–)-HRESI-MS:  $m/z$  749.2092 [M - H] $^-$  (calcd. for  $C_{38}H_{37}O_{16}$ , 749.2087); (+)-HRESI-MS:  $m/z$  751.2234 [M + H] $^+$  (calcd. for  $C_{38}H_{39}O_{16}$ , 751.2233), 773.2047 [M + Na] $^+$  (calcd. for  $C_{38}H_{38}O_{16}Na$ , 773.2052) (Supplementary Figs. 32S-40S).

**Phomoxanthone B (4).**  $C_{38}H_{38}O_{16}$  (750); pale yellow solid; *HPLC*- $R_t$  = 33.92 min (Supplementary Fig. 41S); UV/vis (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 203 (4.36), 260 (3.95), 339 (4.25) nm (Supplementary Fig. 3S);  $^1H$  NMR ( $CDCl_3$ , 500 MHz) and  $^{13}C$  NMR ( $CDCl_3$ , 100 MHz), see Tables 1 and 2; (–)-HRESI-MS:  $m/z$  749.2083 [M - H] $^-$  (calcd. for  $C_{38}H_{37}O_{16}$ , 749.2087); (+)-HRESI-MS:  $m/z$  751.2209 [M + H] $^+$  (calcd. for  $C_{38}H_{39}O_{16}$ , 751.2233), 773.2022 [M + Na] $^+$  (calcd. for  $C_{38}H_{38}O_{16}Na$ , 773.2052) (Supplementary Figs. 42S-50S).

**Dicerandrol C (5).**  $C_{38}H_{38}O_{16}$  (750); pale yellow solid; *HPLC*- $R_t$  = 32.91 min (Supplementary Fig. 51S); UV/vis (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 204 (4.29), 260 (4.04), 339 (4.40) nm (Supplementary Fig. 3S);  $^1H$  NMR ( $CDCl_3$ , 500 MHz) and  $^{13}C$  NMR ( $CDCl_3$ , 100 MHz), see Tables 1 and 2; (–)-HRESI-MS:  $m/z$  749.2098 [M - H] $^-$  (calcd. for  $C_{38}H_{37}O_{16}$ , 749.2087); (+)-HRESI-MS:  $m/z$  751.2238 [M + H] $^+$  (calcd. for  $C_{38}H_{39}O_{16}$ , 751.2233), 773.2050 [M + Na] $^+$  (calcd. for  $C_{38}H_{38}O_{16}Na$ , 773.2052) (Supplementary Figs. 52S-60S).

**Cytochalasin H [also known as Cytochalasin O, Kodocytochalasin 1, Paspalin P I] (6).**  $C_{30}H_{39}NO_5$  (493); white solid; *HPLC*- $R_t$  = 23.85 min (Supplementary Fig. 61S);

## Supplementary material

UV/vis  $\lambda_{\max}$  210 nm (sh, end-absorption);  $^1\text{H}$  NMR ( $\text{CD}_3\text{OD}$ , 500 MHz) and  $^{13}\text{C}$  NMR ( $\text{CD}_3\text{OD}$ , 100 MHz), see **Table 1S**; (-)-ESI-MS:  $m/z$  492  $[\text{M} - \text{H}]^-$ ; (+)-ESI-MS:  $m/z$  476  $[(\text{M}-\text{H}_2\text{O}) + \text{H}]^+$ , 434  $[(\text{M}-\text{COCH}_3-\text{H}_2\text{O}) + \text{H}]^+$  (Supplementary **Figs. 62S-71S**).

**Cytochalasin J (7)**.  $\text{C}_{28}\text{H}_{37}\text{NO}_4$  (451); white solid; *HPLC*- $R_t$  = 20.89 min (Supplementary **Fig. 72S**); UV/vis  $\lambda_{\max}$  210 nm (sh, end-absorption);  $^1\text{H}$  NMR ( $\text{CD}_3\text{OD}$ , 500 MHz) and  $^{13}\text{C}$  NMR ( $\text{CD}_3\text{OD}$ , 125 MHz), see **Table 1S**; (-)-ESI-MS:  $m/z$  496  $[\text{M} + \text{HCOO}]^-$ ; (+)-ESI-MS:  $m/z$  452  $[\text{M} + \text{H}]^+$ , 434  $[(\text{M}-\text{H}_2\text{O}) + \text{H}]^+$ , 416  $[(\text{M}-2\text{H}_2\text{O}) + \text{H}]^+$  (Supplementary **Figs. 73S-80S**).

**8-Hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (8a)**.  $\text{C}_{15}\text{H}_{10}\text{O}_5$  (270); pale yellow solid; *HPLC*- $R_t$  = 24.58 min (Supplementary **Fig. 81S**); UV/vis  $\lambda_{\max}$  200, 230, 250, 285, 365 nm;  $^1\text{H}$  NMR ( $\text{CD}_3\text{OD}$ , 500 MHz)  $\delta$  7.86 (dd, H,  $J$  = 8.5, 7.3, Hz, 3-H), 7.61 (dd, H,  $J$  = 8.5, 1.0 Hz, 4-H), 7.36 (dd, H,  $J$  = 7.3, 1.0 Hz, 2-H), 6.85 (d, H,  $J$  = 0.6, 5-H), 6.65 (d, H,  $J$  = 0.8 Hz, 7-H), 2.44 (s, 3H, 6- $\text{CH}_3$ );  $^1\text{H}$  NMR (acetone- $d_6$ , 500 MHz)  $\delta$  12.24 (brs, 1H, 8-OH), 7.94 (dd, H,  $J$  = 8.5, 7.3, Hz, 3-H), 7.67 (dd, H,  $J$  = 8.5, 1.1 Hz, 4-H), 7.44 (dd, H,  $J$  = 7.3, 1.1 Hz, 2-H), 6.89 (d, H,  $J$  = 0.7, 5-H), 6.67 (d, H,  $J$  = 0.7 Hz, 7-H), 2.45 (s, 3H, 6- $\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CD}_3\text{OD}$ , 100 MHz)  $\delta$  182.0 (C-9), 173.4 (1-COOH), 162.7 (C-8), 157.6 (C-4a), 157.4 (C-10a), 151.2 (C-6), 136.7 (CH-3), 136.5 (C-1), 123.8 (CH-2), 120.3 (CH-4), 118.2 (C-9a), 112.6 (CH-7), 108.7 (CH-5), 108.0 (C-8a), 22.7 (6- $\text{CH}_3$ );  $^{13}\text{C}$  NMR (acetone- $d_6$ , 100 MHz)  $\delta$  181.6 (C-9), 169.8 (1-COOH), 162.2 (C-8), 157.1 (C-4a), 156.8 (C-10a), 150.7 (C-6), 138.2 (C-1), 136.5 (CH-3), 123.8 (CH-2), 120.0 (CH-4), 119.0 (C-9a), 112.2 (CH-7), 108.4 (CH-5), 108.0 (C-8a), 22.5 (6- $\text{CH}_3$ ); (+)-ESI-MS:  $m/z$  271  $[\text{M} + \text{H}]^+$ , 563  $[2\text{M} + \text{Na}]^+$ . This compound was isolated along with traces ~30% of its isomer monodictyoxanthone (**8b**) (Supplementary **Figs. 82S-94S**).

**5-Carbomethoxymethyl-2-heptyl-7-hydroxychromone [also known as (2'S)-7-hydroxy-2-(2-hydroxypropyl)-5-methylchromone; 2-(2'-hydroxypropyl)-5-methyl-7-**

## Supplementary material

**hydroxychromone] (9).** C<sub>13</sub>H<sub>14</sub>O<sub>4</sub> (234); pale yellow solid; *HPLC*-*R*<sub>t</sub> = 17.54 min (Supplementary Fig. 95S); UV/vis λ<sub>max</sub> 227, 244, 251, 300 nm; <sup>1</sup>H NMR (CD<sub>3</sub>OD, 500 MHz) δ 6.67 (brd, 1H, *J* = 2.3 Hz, 8-H), 6.64 (brd, 1H, *J* = 2.3 Hz, 6-H), 6.07 (s, 1H, 3-H), 4.19 (m, 1H, 2'-H), 2.73 (s, 3H, 5-CH<sub>3</sub>), 2.72 (dd, 1H, *J* = 14.4, 5.0 Hz, 1'-H<sub>a</sub>), 2.66 (dd, 1H, *J* = 14.4, 7.9 Hz, 1'-H<sub>b</sub>), 1.28 (d, *J* = 6.3 Hz, 3H, CH<sub>3</sub>-3'); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 100 MHz) δ 182.1 (C-4), 167.3 (C-2), 163.3 (C-7), 162.8 (C-8a), 143.8 (C-5), 118.2 (CH-6), 116.0 (C-4a), 112.7 (CH-3), 101.9 (CH-8), 66.5 (CH-2'), 44.4 (CH<sub>2</sub>-1'), 23.7 (CH<sub>3</sub>-3'), 23.3 (5-CH<sub>3</sub>); (-)-ESI-MS: *m/z* 233 [M - H]<sup>-</sup>; (+)-ESI-MS: *m/z* 235 [M + H]<sup>+</sup> (Supplementary Figs. 96S-103S).

**Maltol [also known as 3-hydroxy-2-methyl-4h-pyran-4-one; 2-methyl-3-hydroxy-4-pyrone; e 636; larixic acid larixin; larixin; larixinic acid; maltol; methylmaltol; NSC 2829; NSC 404458; palatone; veltol (plant growth regulator)] (10).** C<sub>6</sub>H<sub>6</sub>O<sub>3</sub> (126); pale yellow solid; *HPLC*-*R*<sub>t</sub> = 12.87 min (Supplementary Fig. 104S); UV/vis λ<sub>max</sub> 227, 280 nm; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 500 MHz) δ 8.81 (brs, 1H, 3-OH), 8.02 (d, 1H, *J* = 5.5 Hz, 6-H), 6.33 (d, 1H, *J* = 5.5 Hz, 5-H), 2.24 (s, 3H, 2-CH<sub>3</sub>); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz) δ 172.5 (C-4), 154.6 (CH-6), 149.2 (C-2), 142.9 (C-3), 113.5 (CH-5), 14.0 (2-CH<sub>3</sub>); (+)-ESI-MS: *m/z* 127 [M + H]<sup>+</sup> (Supplementary Figs. 105S-111S).

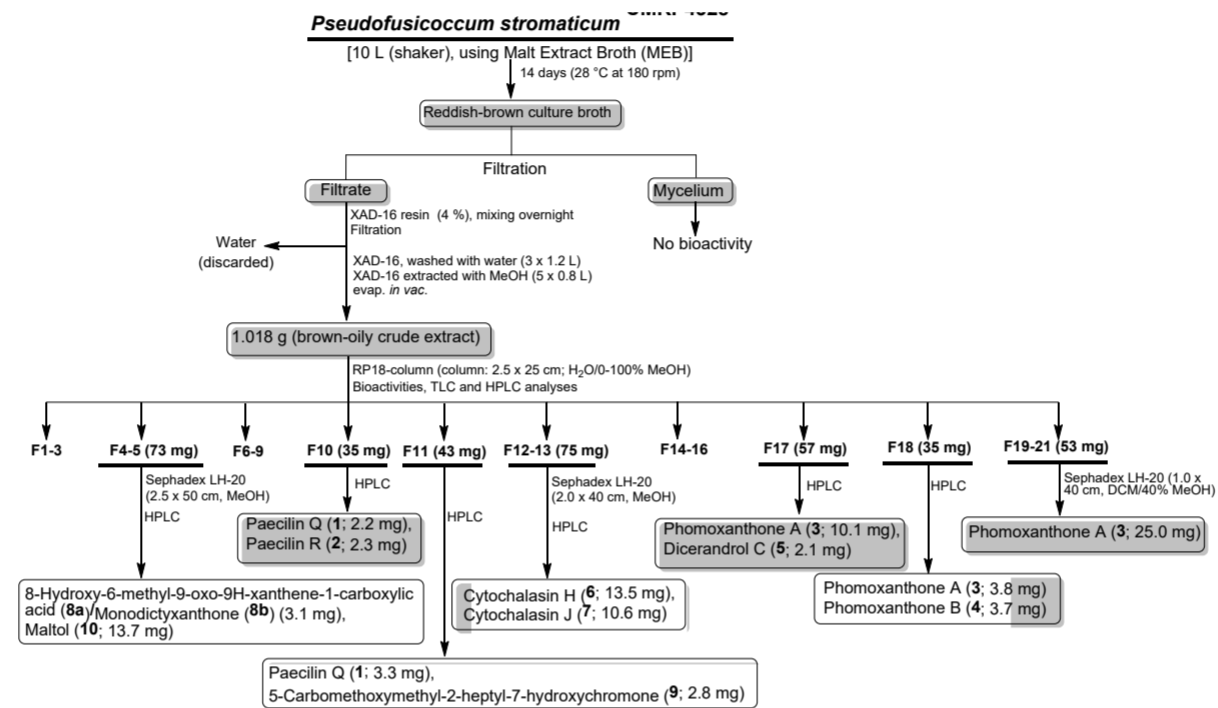
## Supplementary References

- (1) Shaaban KA, Wang X, Elshahawi SI, Ponomareva LV, Sunkara M, Copley GC, Hower JC, Morris AJ, Kharel MK, Thorson JS. *J Nat Prod* 2013; 76: 1619-1626
- (2) Wang X, Shaaban KA, Elshahawi SI, Ponomareva LV, Sunkara M, Zhang Y, Copley GC, Hower JC, Morris AJ, Kharel MK, Thorson JS. *J Nat Prod* 2013; 76: 1441-1447
- (3) Shaaban KA, Elshahawi SI, Wang X, Horn J, Kharel MK, Leggas M, Thorson JS. *J Nat Prod* 2015; 78: 1723-1729
- (4) Savi DC, Shaaban KA, Gos F, Ponomareva LV, Thorson JS, Glienke C, Rohr J. *Sci Rep* 2018; 8: 3122
- (5) El-Elimat T, Figueroa M, Raja HA, Graf TN, Swanson SM, Falkinham III JO, Wani MC, Peaerce CJ, Oberlies NH. *Eur J Org Chem* 2015; 1: 109-121
- (6) Kumla D, Shine Aung T, Buttachon S, Dethoup T, Gales L, Pereira JA, Inácio A, Costa PM, Lee M, Sekeroglu N, Silva AMS, Pinto MMM, Kijjoa A. *Mar Drugs* 2017; 15: 375
- (7) Valdomir G, Senthilkumar S, Ganapathy D, Zhang Y, Tietze LF. *Chem Eur J* 2018; 24: 8760-8763



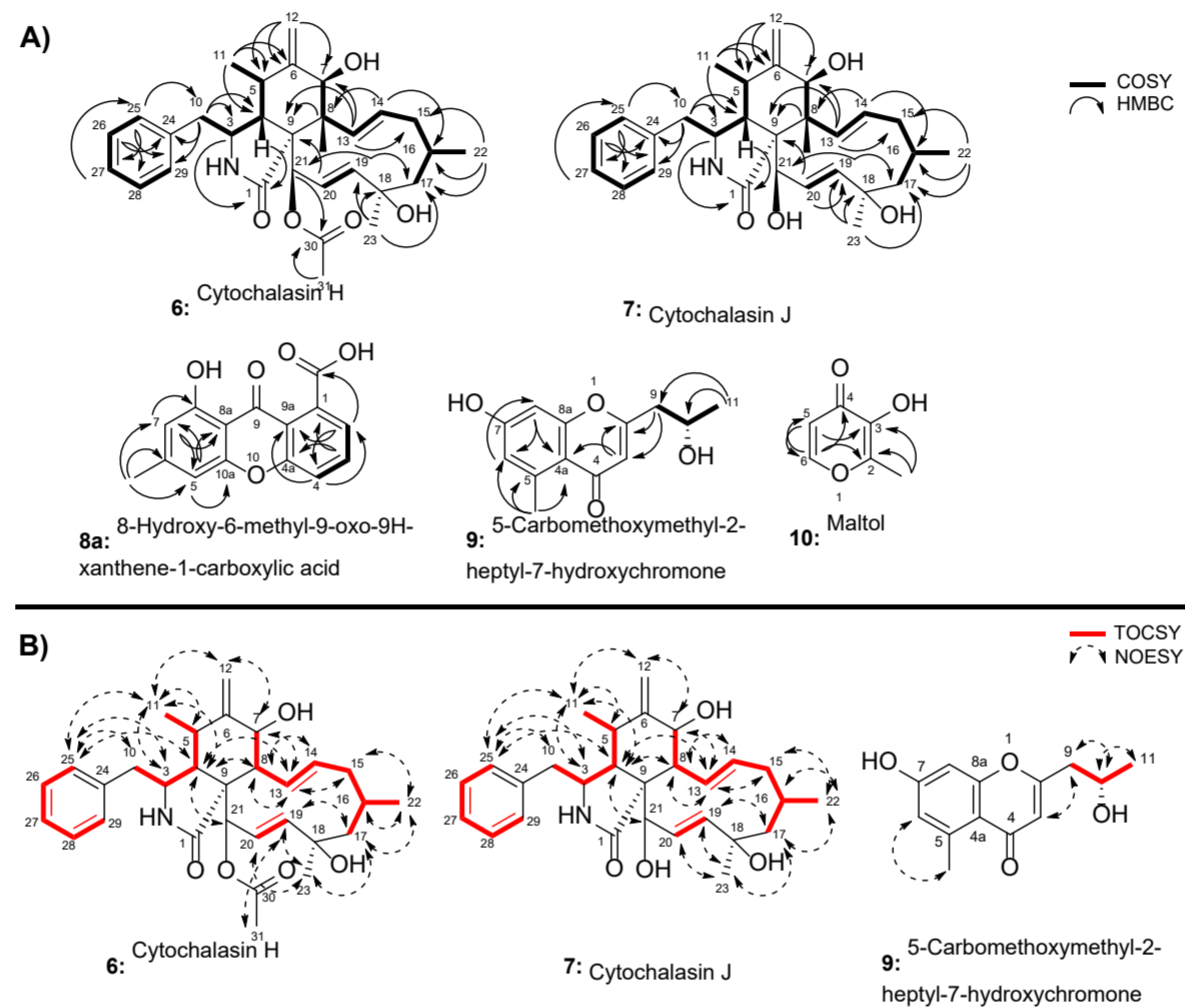
Supplementary material

## Supplementary material



**Fig. 1S** Workup scheme of the metabolites produced by the fungus *Pseudofusicoccum stromaticum* CMRP4328.

## Supplementary material



**Fig. 2S A**  $^1\text{H}, ^1\text{H}$ -COSY (—) and selected HMBC (↷) correlations of compounds **6-10**. **B**

TOCSY (—) and selected NOESY (---) correlations of compounds **6-7** and **9**.

## Supplementary material

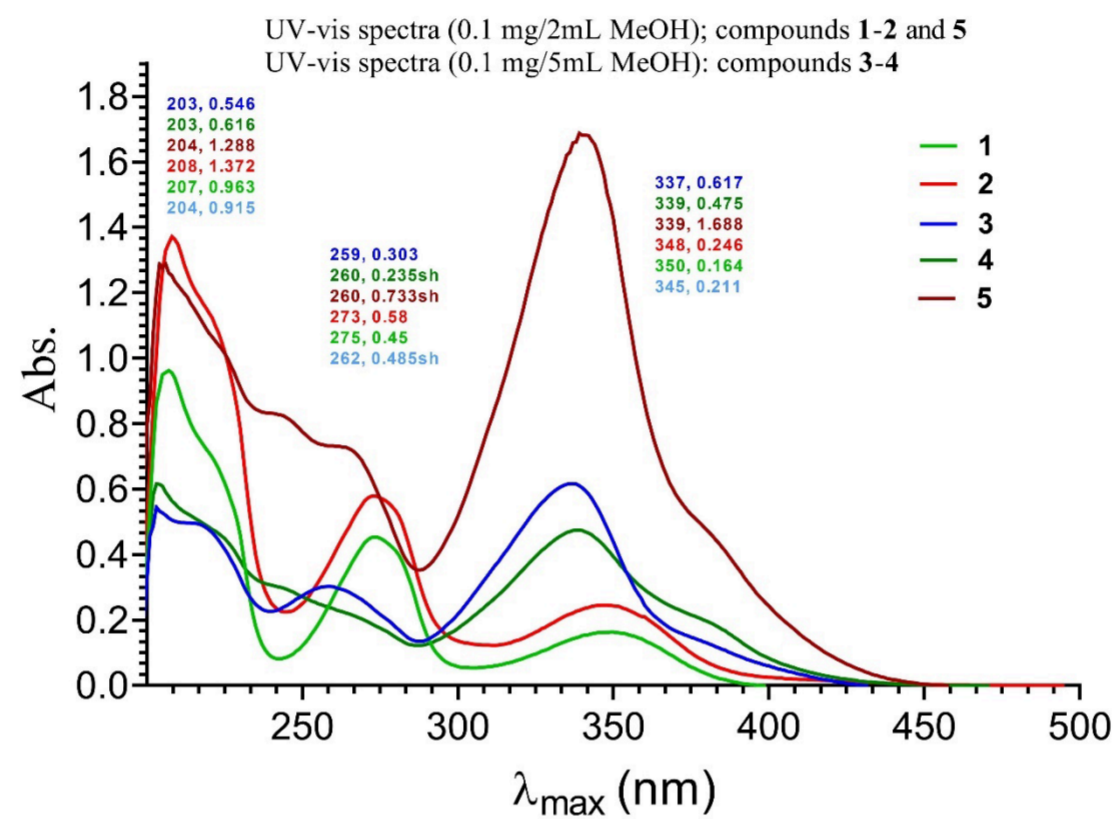


Fig. 3S UV-vis (MeOH) spectra of compounds 1-5.

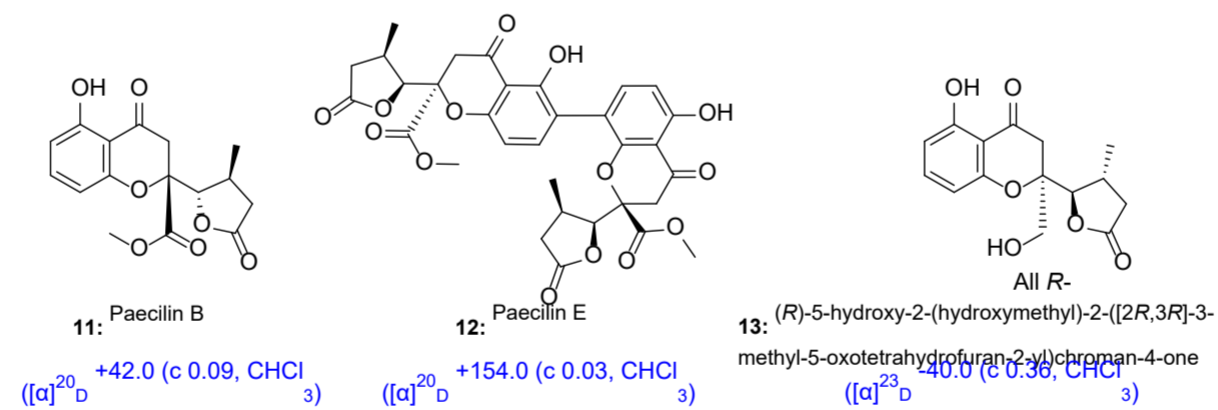
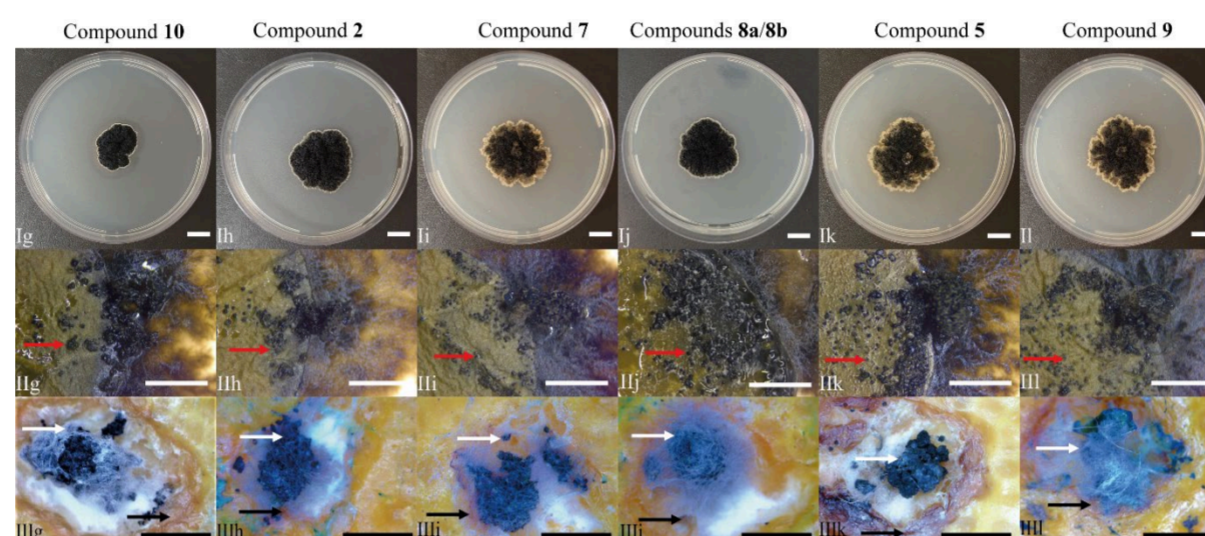


Fig. 4S Chemical structures of the reported natural products, paecilin B (**11**)<sup>5</sup>, paecilin E (**12**)<sup>6</sup> along with the synthetic (R)-5-hydroxy-2-(hydroxymethyl)-2-([2R,3R]-3-methyl-5-oxotetrahydrofuran-2-yl) chroman-4-one (**13**).<sup>7</sup>

## Supplementary material



**Fig. 5S** Antifungal activity of compounds **2**, **5**, and **7-10** (10 mg/mL) against the phytopathogen *Phyllosticta citricarpa*. **Ig-l** Evaluation of mycelial growth; **IIg-l** Development of pycnidia in citrus leaves, **IIIg-l** citrus black spot (CBS) lesions in detached fruits. **g** Maltol (**10**); **h** Paecilin R (**2**); Cytochalasin J (**7**); **j** 8-Hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**)/Monodictyxanthone (**8b**); **k** Dicerandrol C (**5**) and **l** 5-Carbomethoxymethyl-2-heptyl-7-hydroxychromone (**9**); **Table 3**. Red arrow (**II**): pycnidia of *P. citricarpa*. Black arrow (**III**): necrotic zone. White arrow (**III**): fungal growth. — Scale bars: I = 10 mm; II = 3 mm; III = 5 mm.

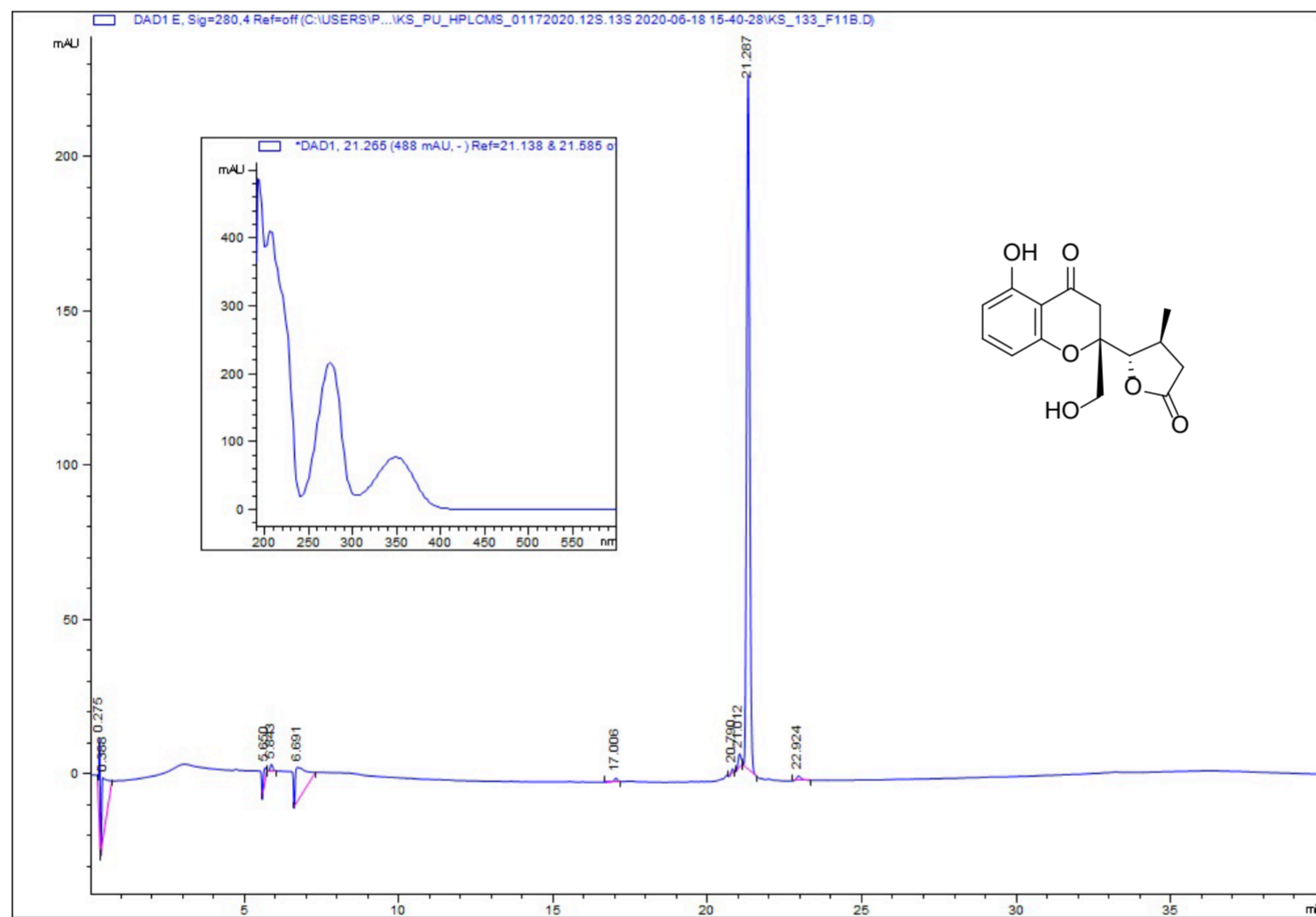
## Supplementary material

**Table 1S**  $^{13}\text{C}$  and  $^1\text{H}$  (500 MHz) NMR spectroscopic data for cytochalasin H (**6**) and cytochalasin J (**7**) in  $\text{CD}_3\text{OD}$  ( $\delta$  in ppm).

Position	Cytochalasin H ( <b>6</b> )		Cytochalasin J ( <b>7</b> )	
	$\delta_{\text{C}}$ , type <sup>(a)</sup>	$\delta_{\text{H}}$ (mult, <i>J</i> in [Hz])	$\delta_{\text{C}}$ , type <sup>(b)</sup>	$\delta_{\text{H}}$ (mult, <i>J</i> in [Hz])
1	177.2, C		178.9, C	
3	55.2, CH	3.28 (ddd, 8.3, 5.4, 2.8)	55.1, CH	3.1 (m)
4	50.0, CH	2.16 (dd, 5.4, 2.8)	50.0, CH	2.60 (dd, 5.2, 3.4)
5	33.6, CH	2.63 (m)	34.1, CH	2.75 (m)
6	151.3, C		152.0, C	
7	72.8, CH	3.81 (dd, 10.5, 1.1)	72.7, CH	3.76 (dd, 10.7, 1.2)
8	48.0, CH	2.92 (t, 10.1)	46.7, CH	2.89 (t, 10.1)
9	53.9, C		55.2, C	
10	45.3, CH <sub>2</sub>	2.87 (dd, 13.0, 5.3) 2.71 (dd, 13.2, 8.3)	45.1, CH <sub>2</sub>	2.81 (dd, 13.4, 6.1) 2.71 (dd, 13.4, 5.8)
11	13.8, CH <sub>3</sub>	0.54 (d, 6.7)	14.2, CH <sub>3</sub>	0.82 (d, 6.7)
12	113.6, CH <sub>2</sub>	5.19 (brd, 1.3) 4.98 (brs)*	113.2, CH <sub>2</sub>	5.22 (brd, 1.4) 5.01 (brd, 1.1)
13	129.4, CH	5.63 (ddd, 15.4, 9.4, 1.4)	129.5, CH	5.56 (ddd, 15.4, 9.5, 1.4)
14	137.9, CH	5.29 (ddd, 15.1, 10.2, 4.9)	137.4, CH	5.25 (ddd, 15.0, 10.7, 5.4)
15	44.7, CH <sub>2</sub>	2.02 (ddd, 11.4, 4.9, 1.7) 1.83-1.66 (m)	44.7, CH <sub>2</sub>	1.97 (dd, 12.5, 4.7) 1.67 (m)
16	29.4, CH	1.83-1.66 (m)	29.5, CH	1.84-1.65 (m)
17	55.2, CH <sub>2</sub>	1.83-1.66 (m) 1.52 (dd, 14.0, 3.0)	55.1, CH <sub>2</sub>	1.77 (m) 1.50 (dd, 14.0, 3.3)
18	74.9, C		75.3, C	
19	139.2, CH	5.52 (dd, 16.6, 2.3)	137.5, CH	5.74 (dd, 16.6, 2.2)
20	127.0, CH	5.74 (dd, 16.5, 2.4)	132.4, CH	5.82 (dd, 16.5, 2.2)
21	78.6, CH	5.43 (t, 2.4)	76.8, CH	3.70 (t, 2.3)
22	26.8, CH <sub>3</sub>	1.02 (d, 6.0)	26.9, CH <sub>3</sub>	1.00 (d, 6.6)
23	30.7, CH <sub>3</sub>	1.28 (s)	31.2, CH <sub>3</sub>	1.26 (s)
24	138.6, C		138.8, C	
25/29	131.1, CH	7.20 (dd, 8.5, 1.5)	131.3, CH	7.25-7.20 (m)
26/28	129.8, CH	7.31 (td, 7.2, 1.2)	129.7, CH	7.31 (td, 7.5, 1.0)
27	128.0, CH	7.23 (td, 7.4, 1.2)	127.9, CH	7.25-7.20 (m)
30	172.0, C			
31	20.9, CH <sub>3</sub>	2.28 (s)		

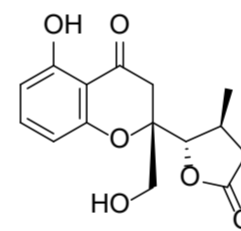
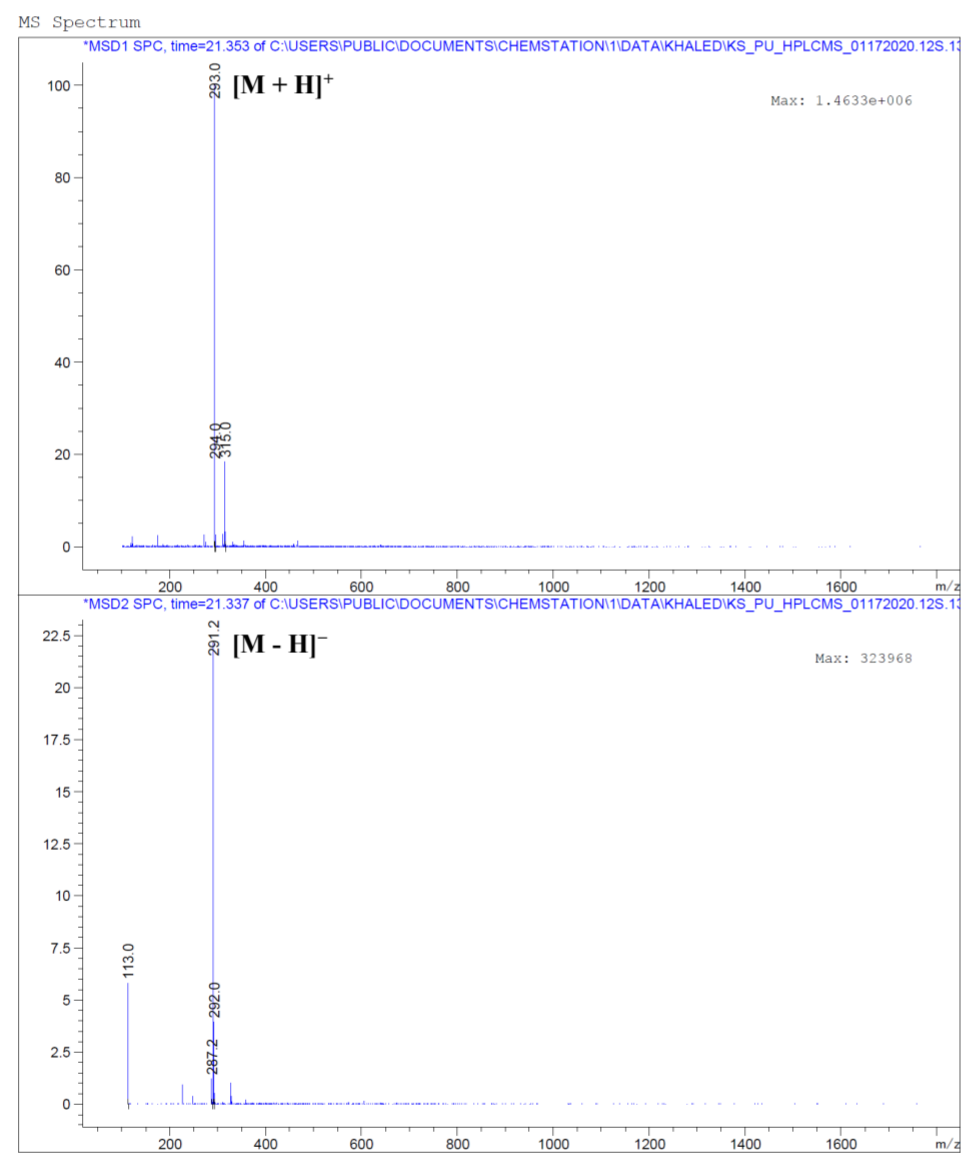
<sup>(a)</sup>100 MHz; <sup>(b)</sup>125 MHz; See Supporting Information for NMR spectra. Assignments supported by 2D HSQC and HMBC experiments. \*Signal buried under water signal in  $^1\text{H}$  (500 MHz) NMR spectrum, and it was displayed as a broad singlet signal (not buried under water signal) in the  $^1\text{H}$  (400 MHz) NMR spectrum.

## Supplementary material



**Fig. 6S** HPLC analysis of paeicilin Q (**1**). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 280 nm. UV-vis inset of full wavelength scan (190-600 nm).

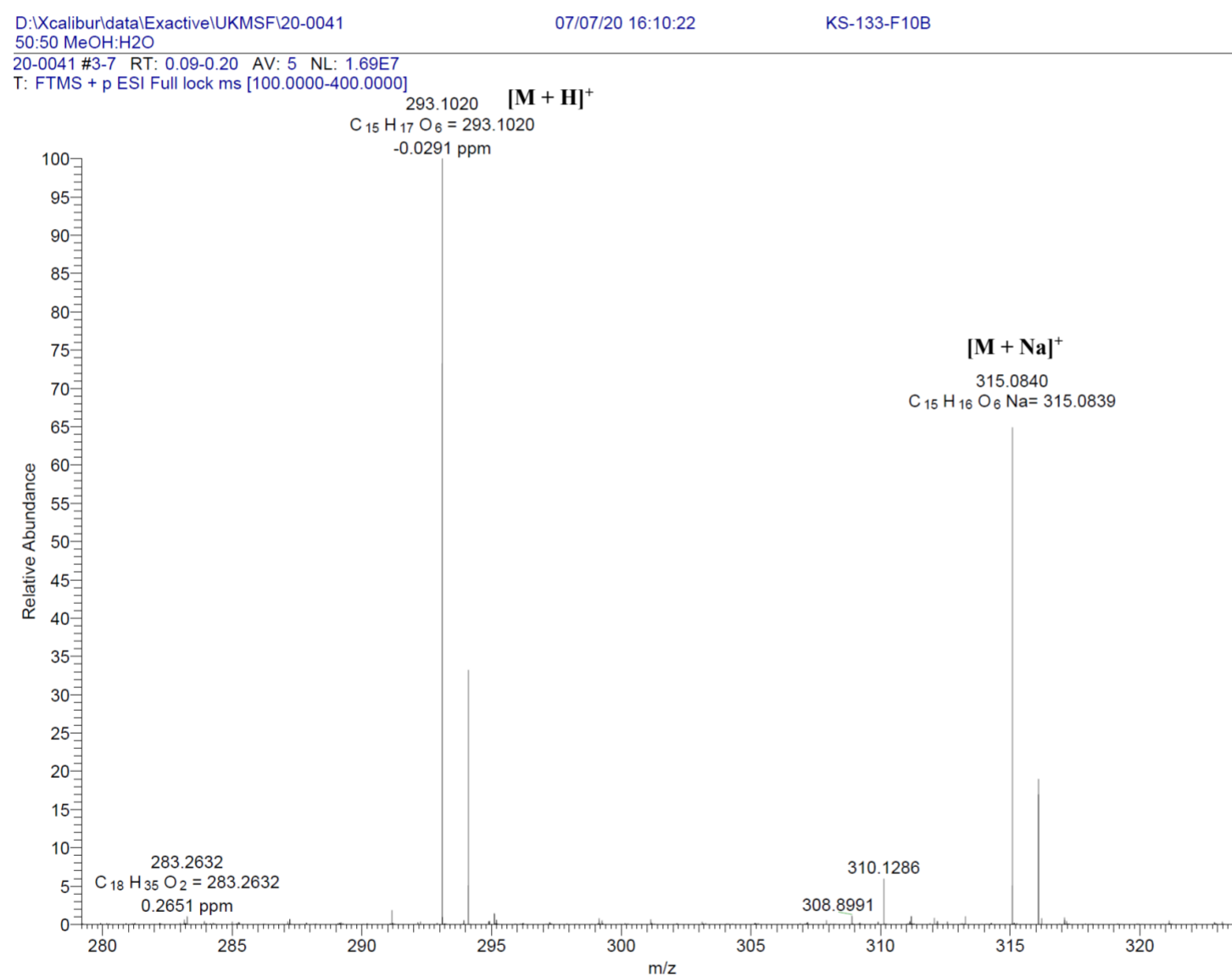
## Supplementary material



**Fig. 7S** (+) and (-)-ESI-MS spectra of paeclin Q (**1**).



## Supplementary material



**Fig. 8S** (+)-HRESI-MS spectrum of paecilin Q (**1**).

## Supplementary material

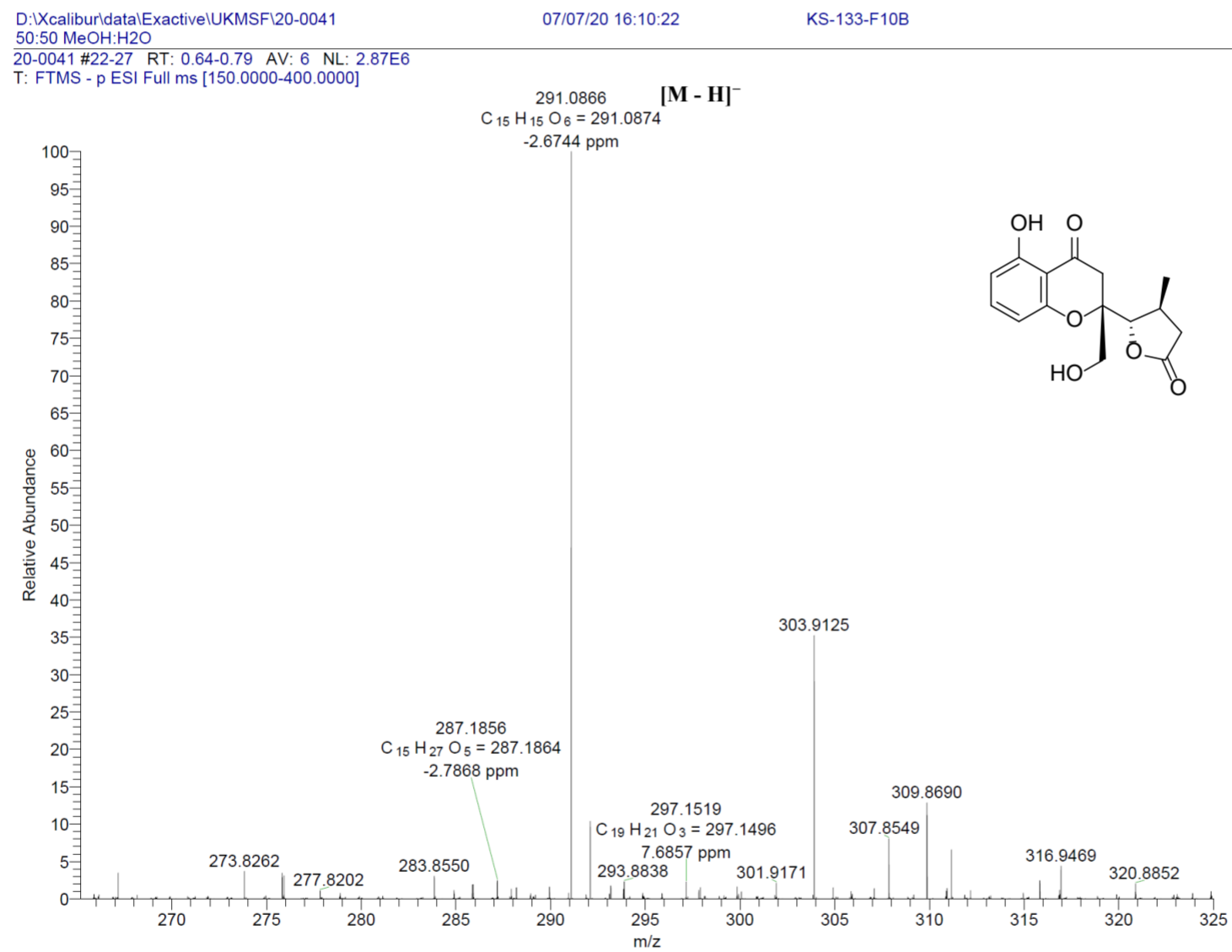
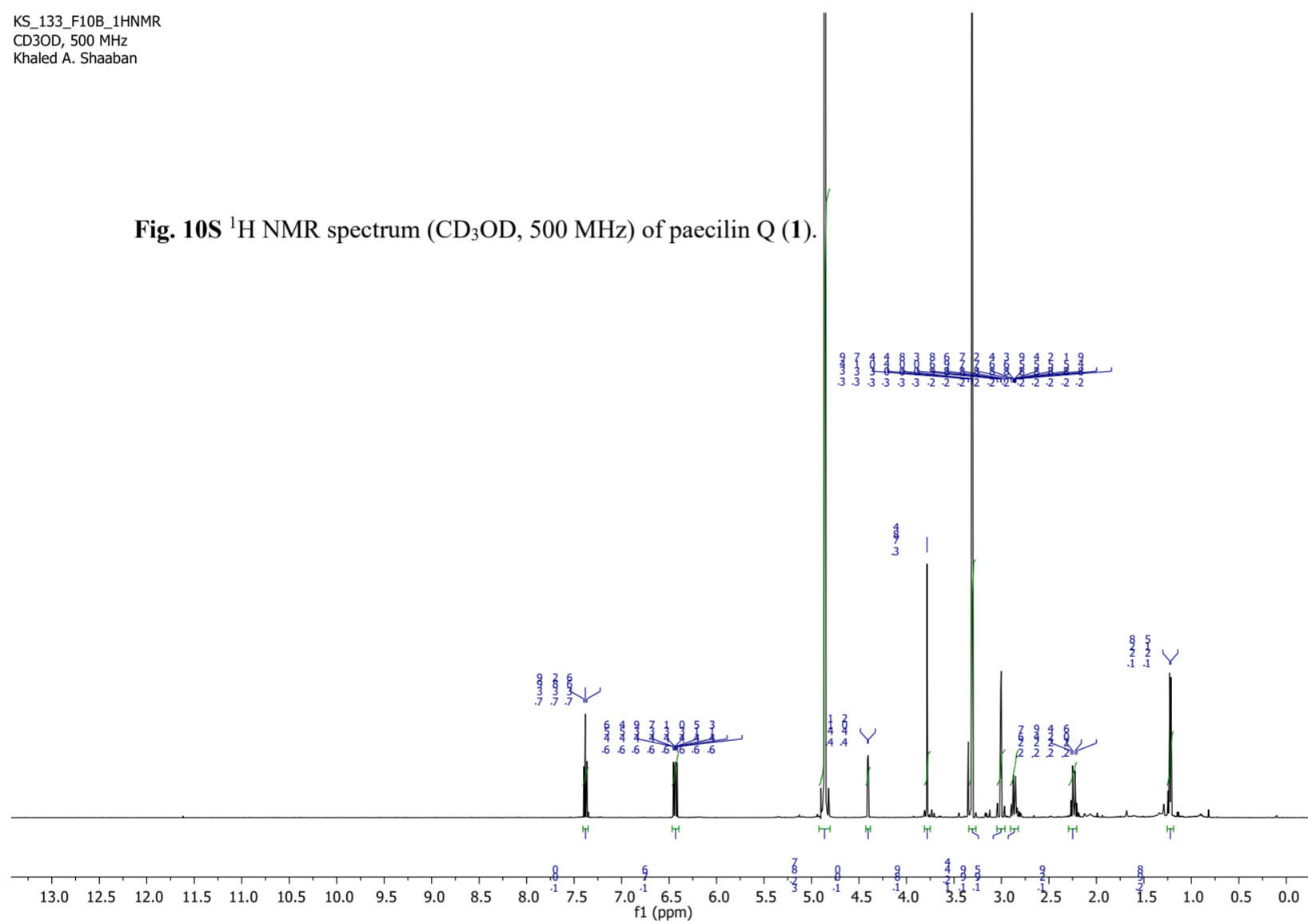
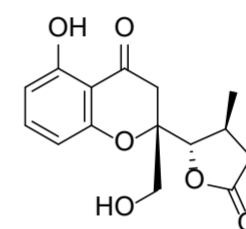


Fig. 9S (-)-HRESI-MS spectrum of paecilin Q (1).

## Supplementary material

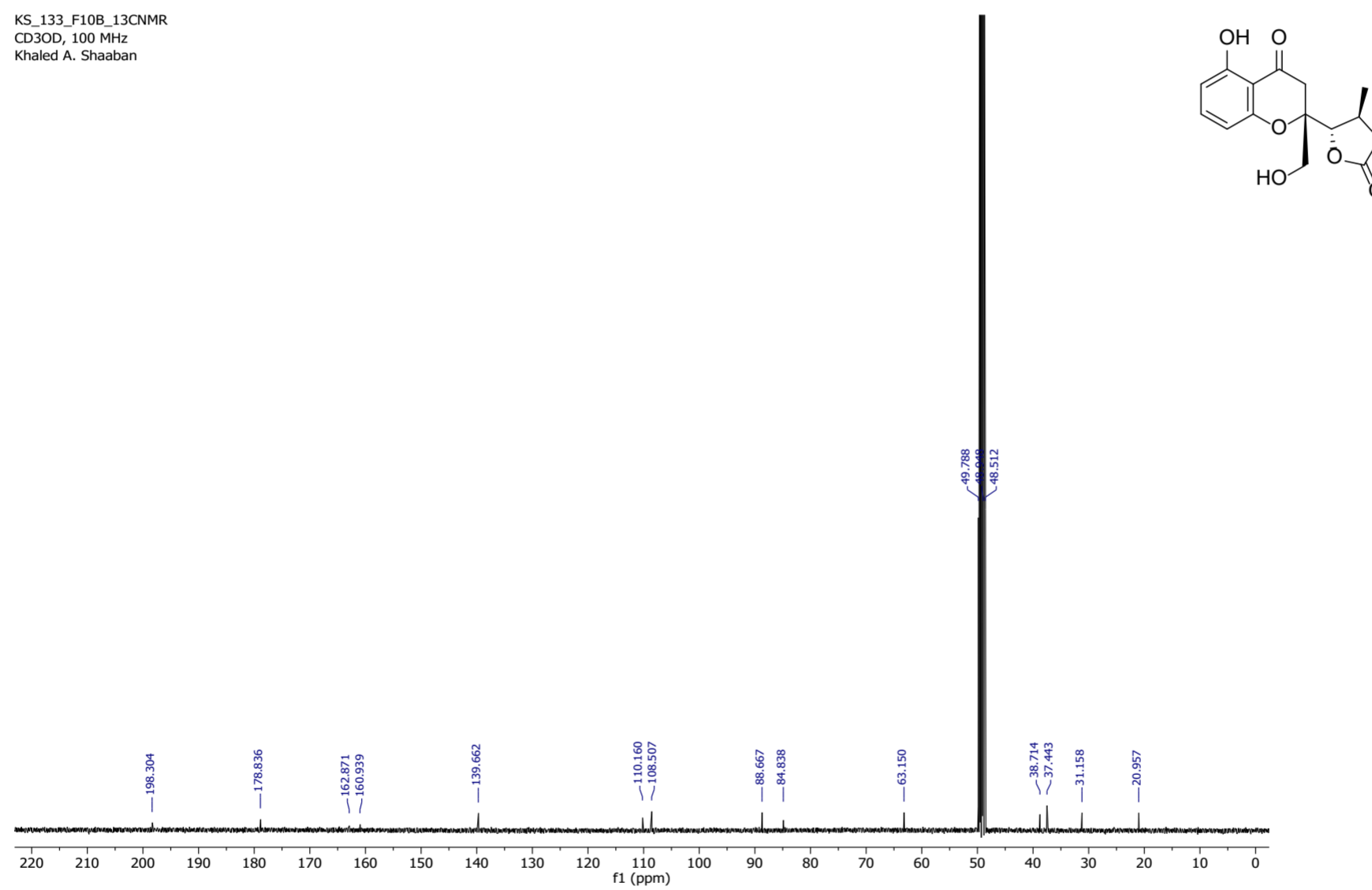
KS\_133\_F10B\_1HNMR  
CD3OD, 500 MHz  
Khaled A. Shaaban

Fig. 10S  $^1\text{H}$  NMR spectrum (CD<sub>3</sub>OD, 500 MHz) of paecilin Q (1).



## Supplementary material

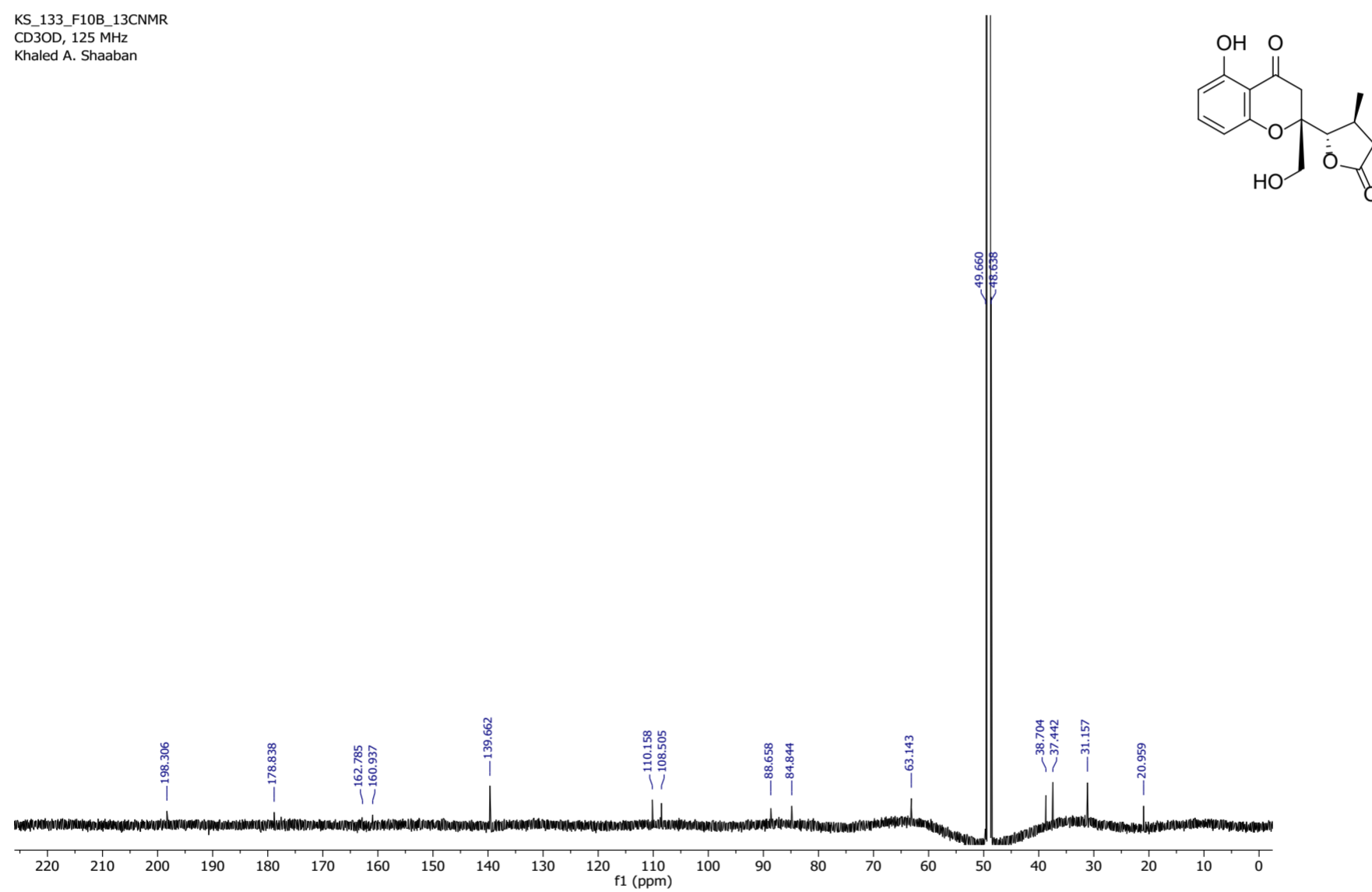
KS\_133\_F10B\_13CNMR  
CD3OD, 100 MHz  
Khaled A. Shaaban



**Fig. 11S** <sup>13</sup>C NMR spectrum (CD<sub>3</sub>OD, 100 MHz) of paecilin Q (**1**).

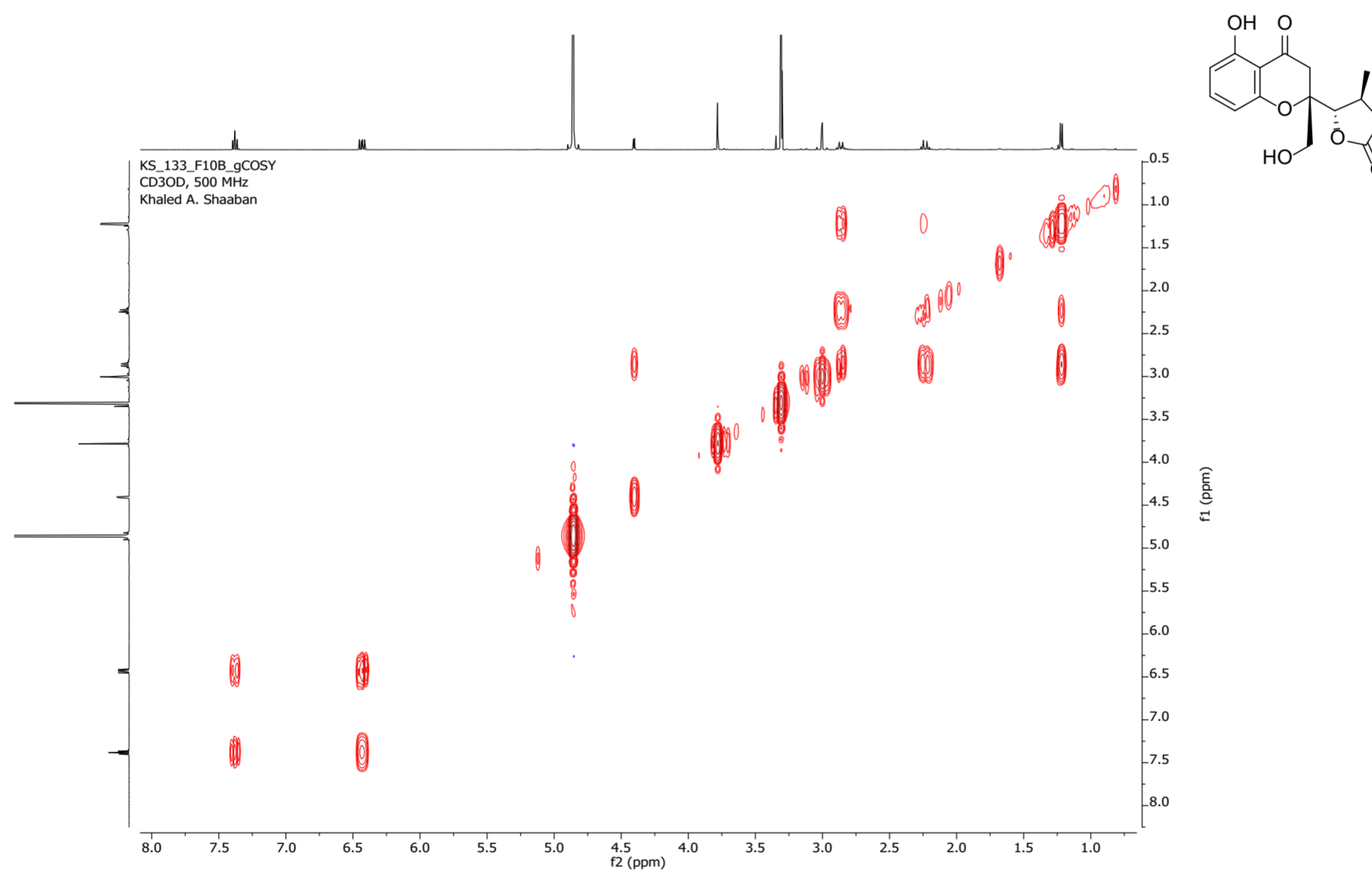
## Supplementary material

KS\_133\_F10B\_13CNMR  
CD3OD, 125 MHz  
Khaled A. Shaaban



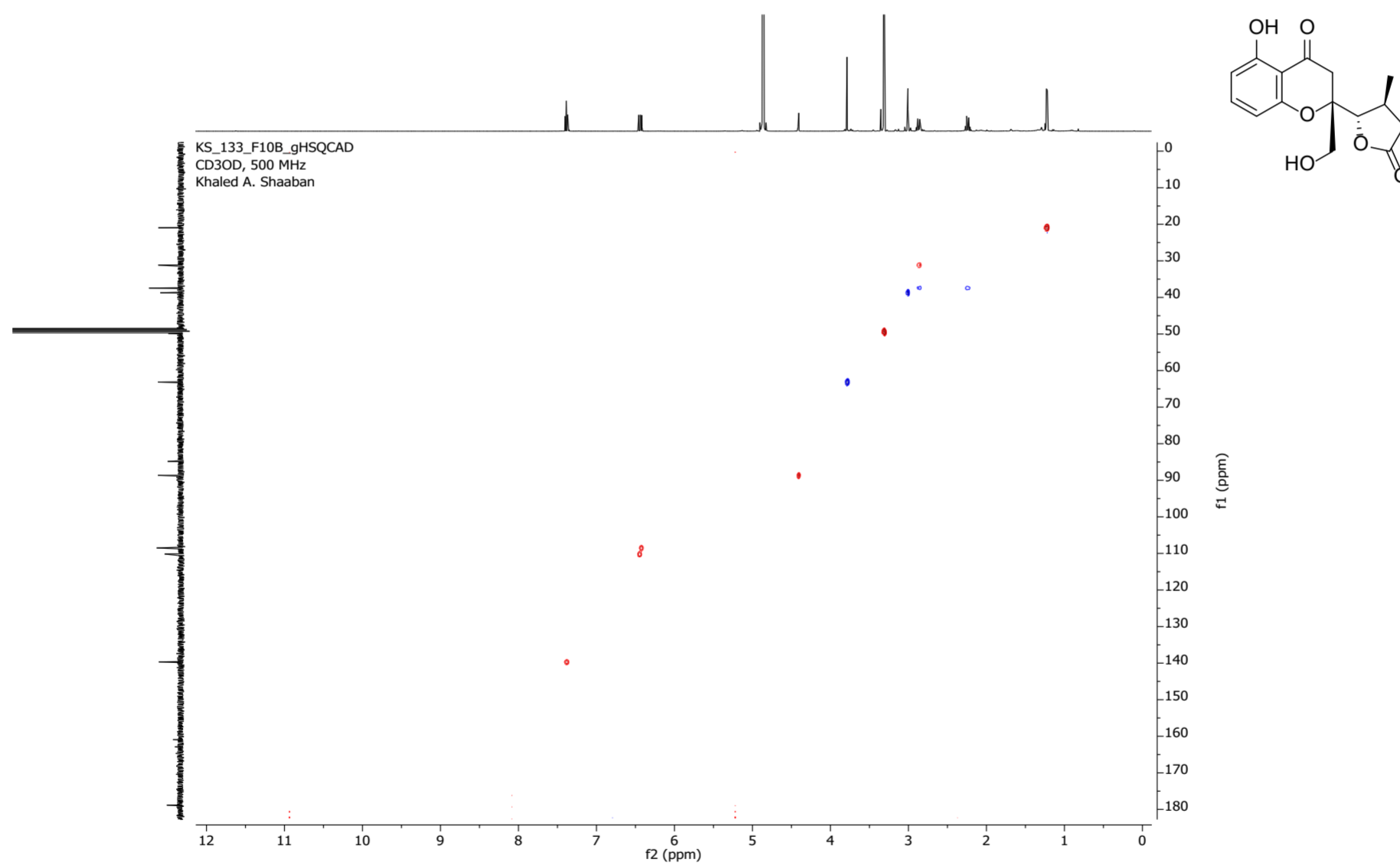
**Fig. 12S** <sup>13</sup>C NMR spectrum (CD<sub>3</sub>OD, 125 MHz) of paecilin Q (1).

## Supplementary material



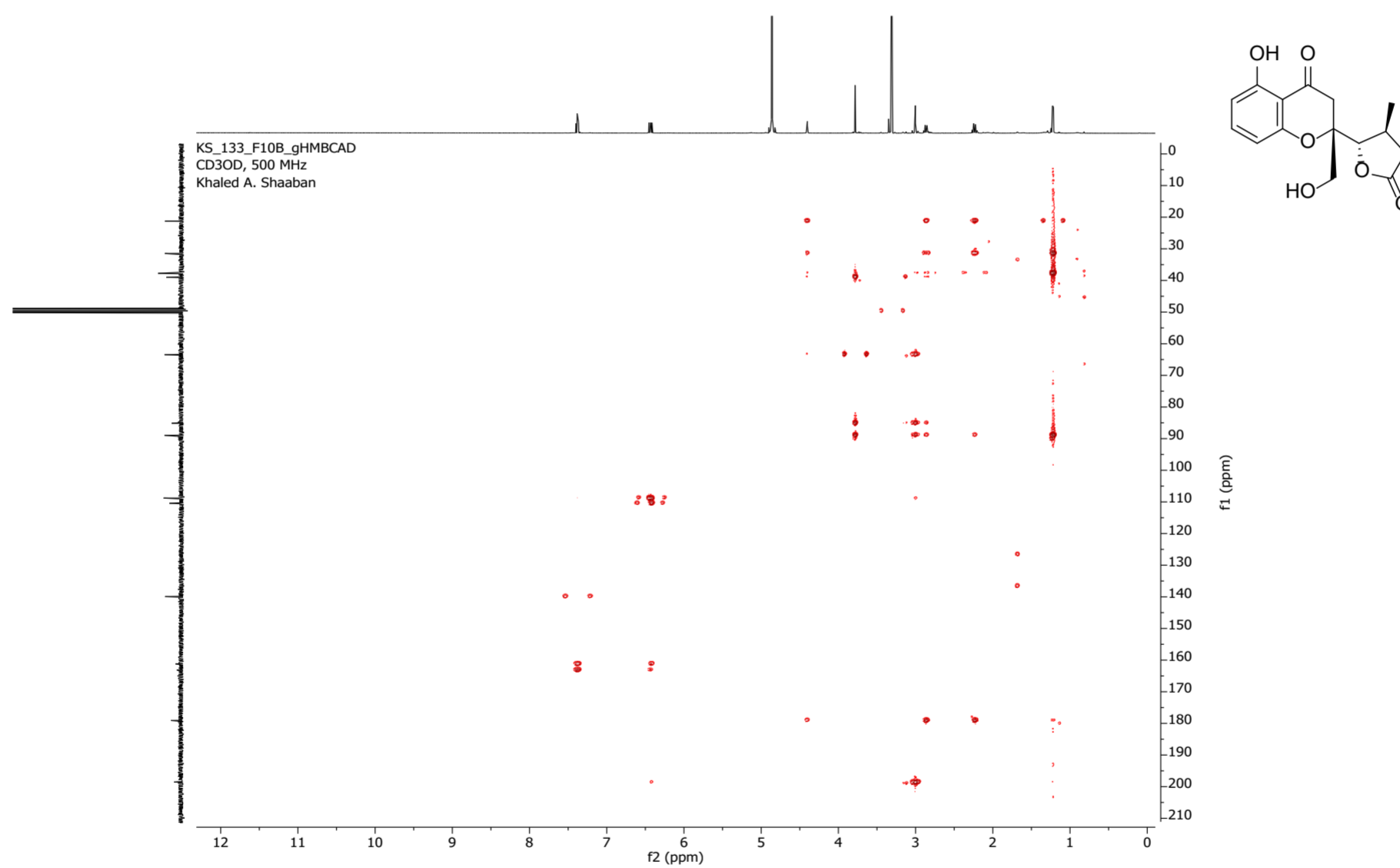
**Fig. 13S**  $^1\text{H}$ ,  $^1\text{H}$ -COSY spectrum ( $\text{CD}_3\text{OD}$ , 500 MHz) of paecilin Q (**1**).

## Supplementary material



**Fig. 14S** HSQC spectrum (CD<sub>3</sub>OD, 500 MHz) of paeclin Q (**1**).

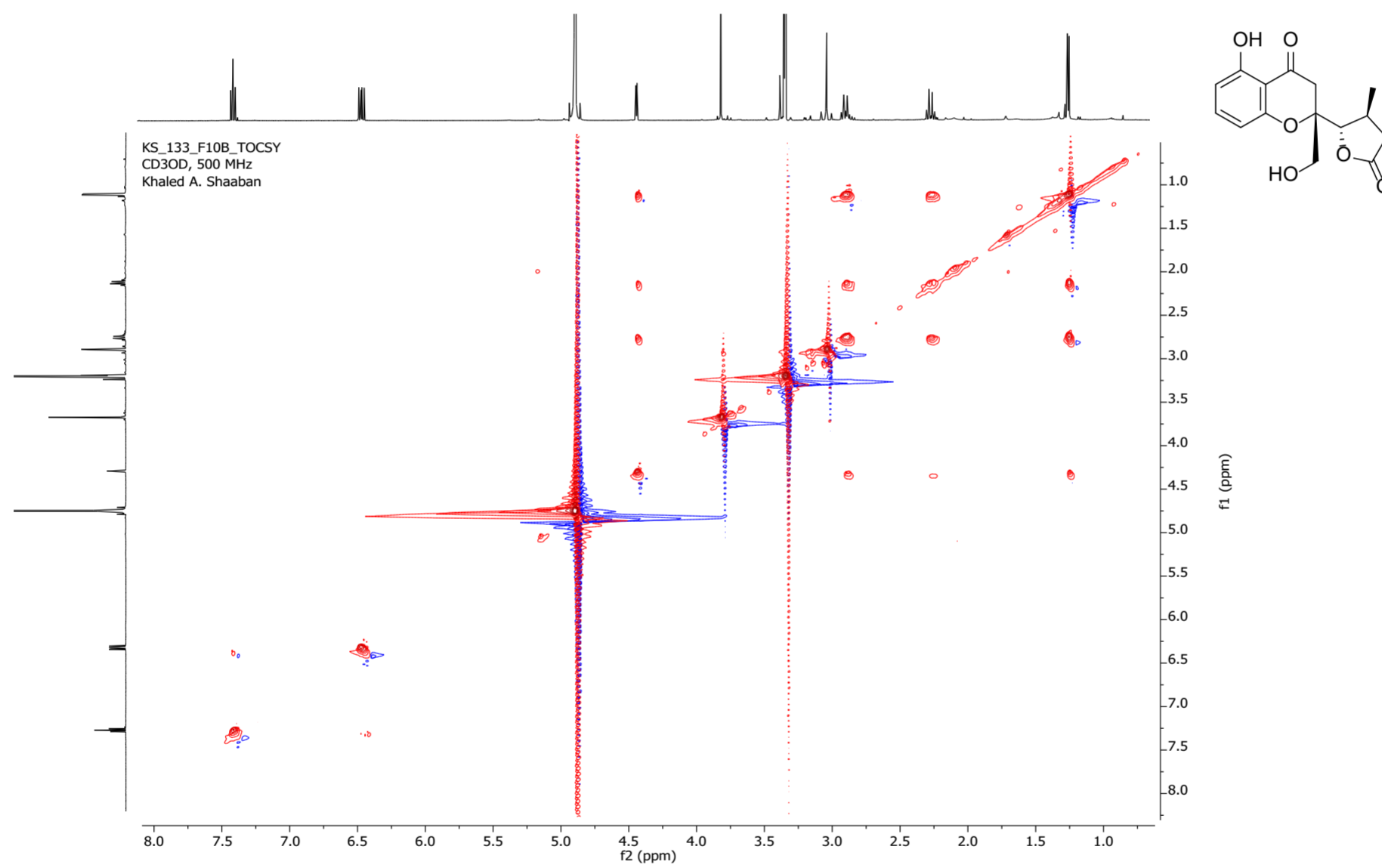
## Supplementary material



**Fig. 15S** HMBC spectrum (CD<sub>3</sub>OD, 500 MHz) of paecilin Q (**1**).

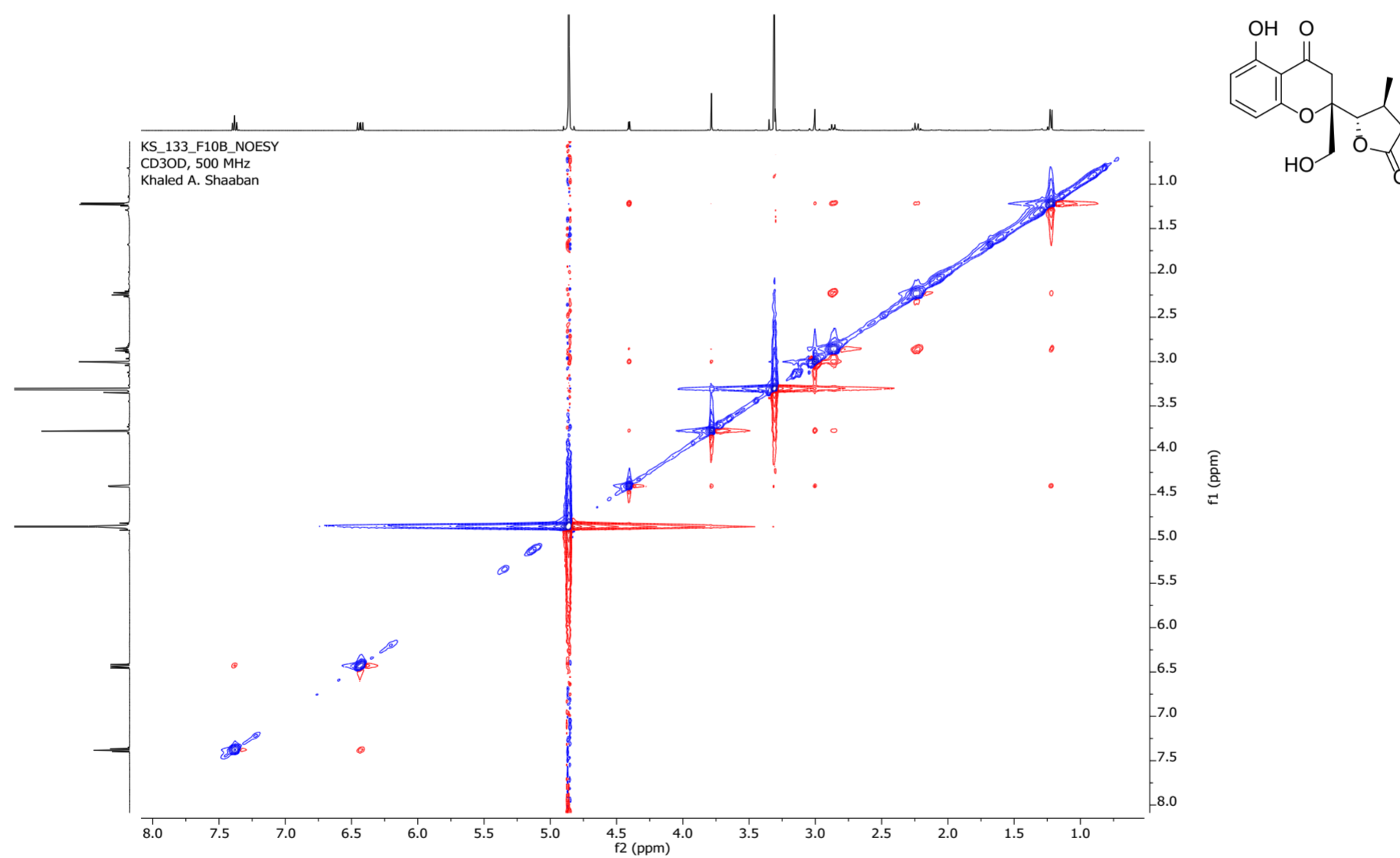


## Supplementary material



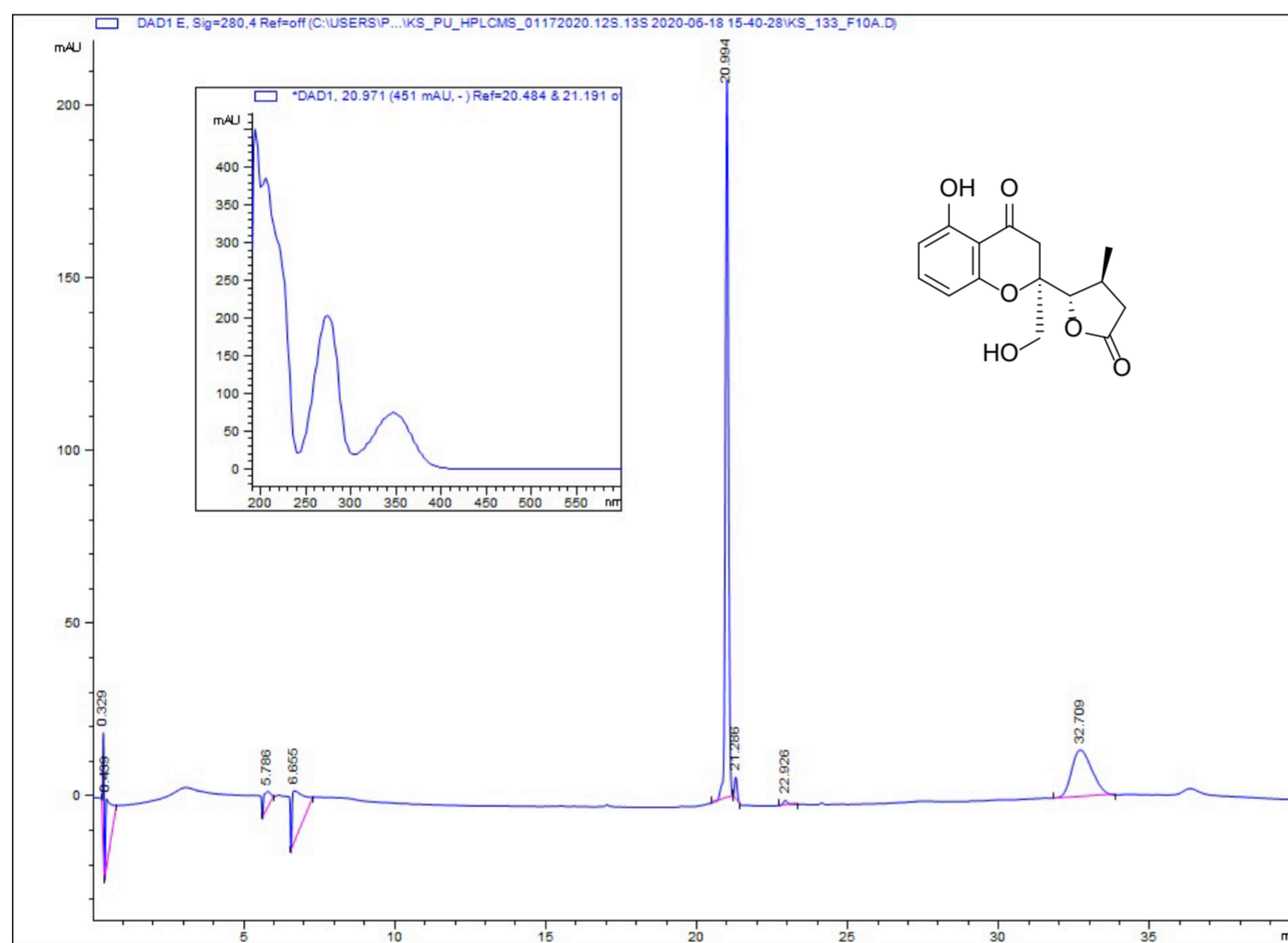
**Fig. 16S** TOCSY spectrum (CD<sub>3</sub>OD, 500 MHz) of paecilin Q (**1**).

## Supplementary material



**Fig. 17S** NOESY spectrum (CD<sub>3</sub>OD, 500 MHz) of paecilin Q (**1**).

## Supplementary material



**Fig. 18S** HPLC analysis of paecilin R (**2**). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 280 nm. UV-vis inset of full wavelength scan (190-600 nm).

## Supplementary material

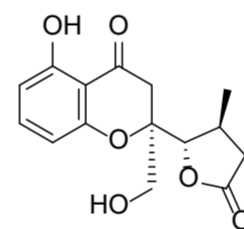
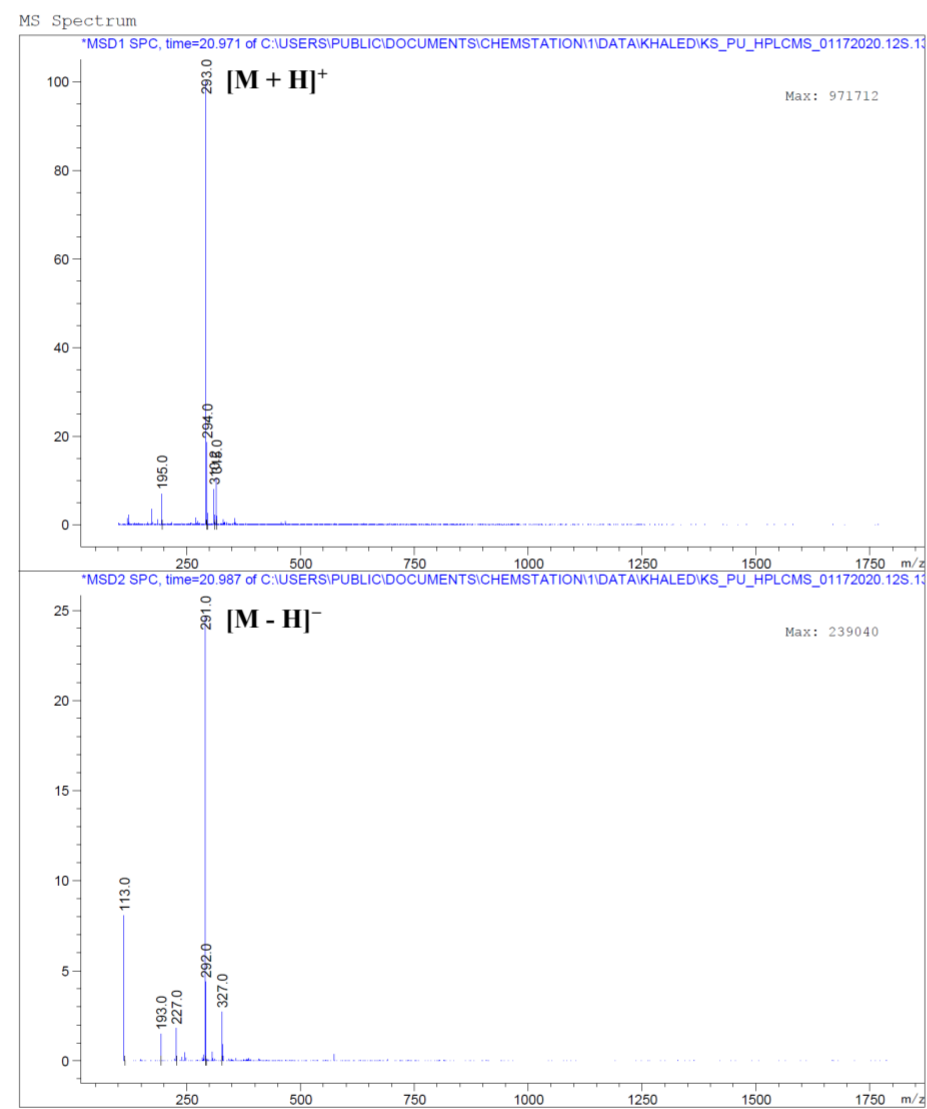
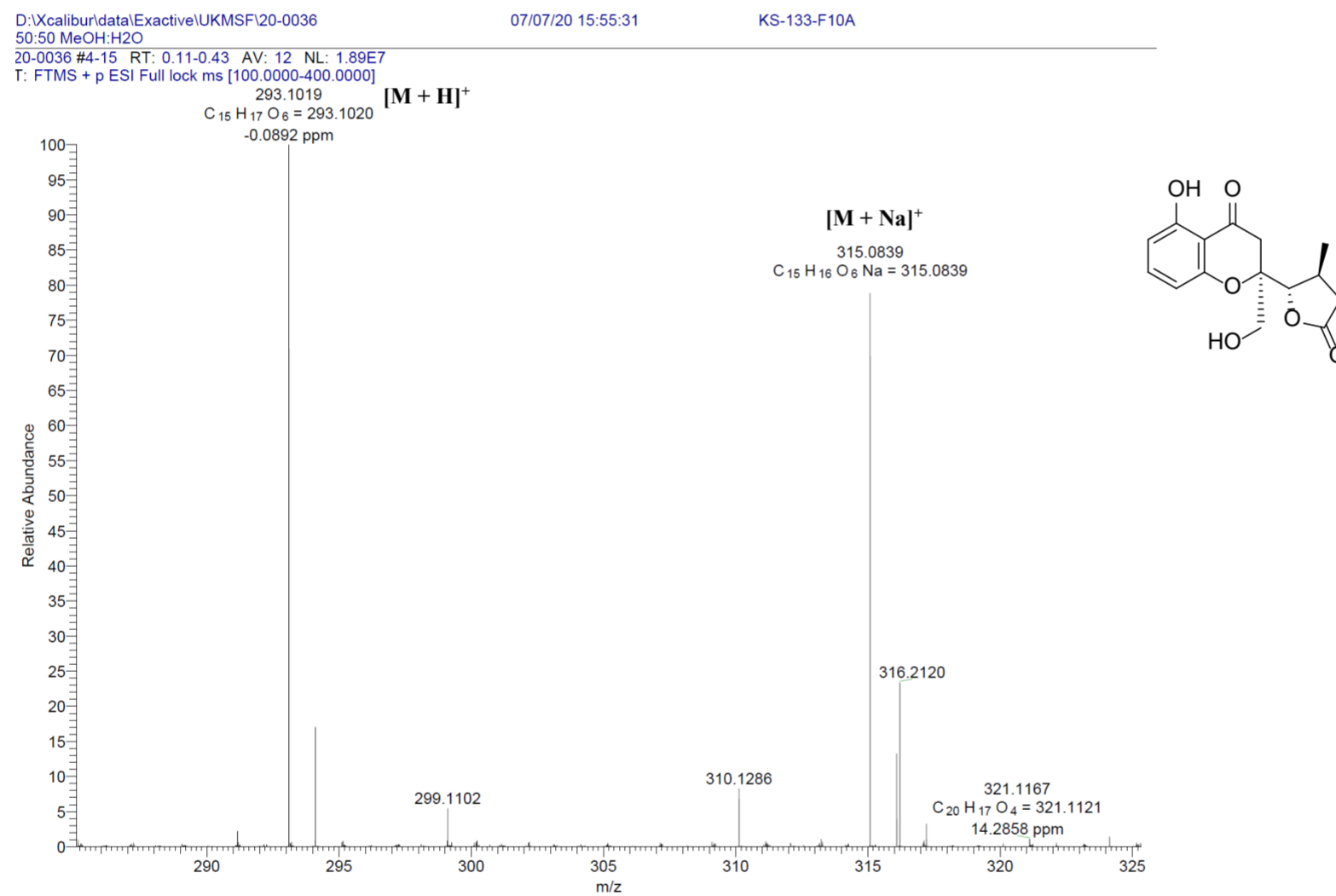


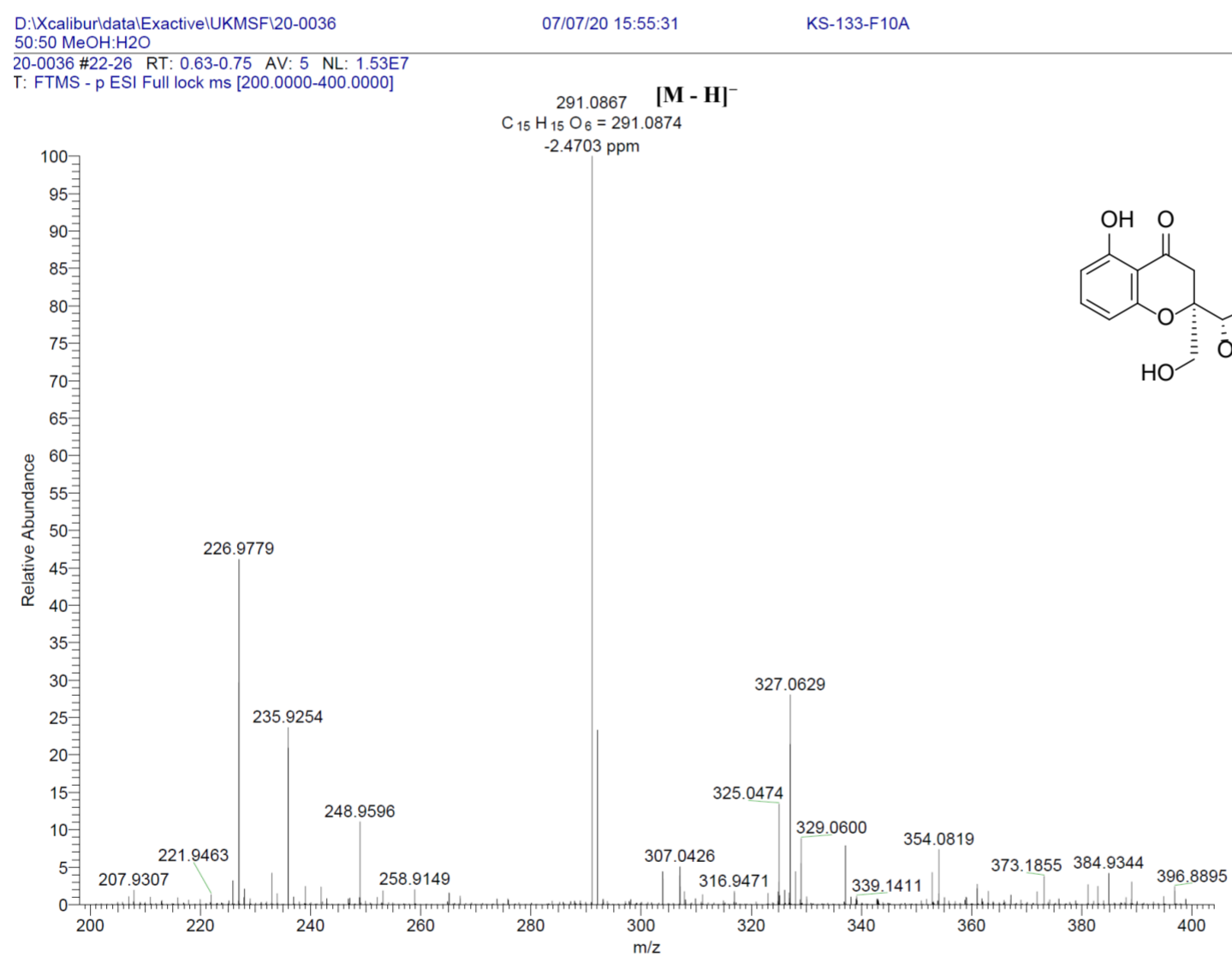
Fig. 19S (+) and (-)-ESI-MS spectrum of paecilin R (2).

## Supplementary material



**Fig. 20S** (+)-HRESI-MS spectrum of paecilin R (**2**).

## Supplementary material



**Fig. 21S** (–)-HRESI-MS spectrum of paecilin R (**2**).

## Supplementary material

KS\_133\_F10A\_1HNMR  
CD3OD, 500 MHz  
Khaled A. Shaaban

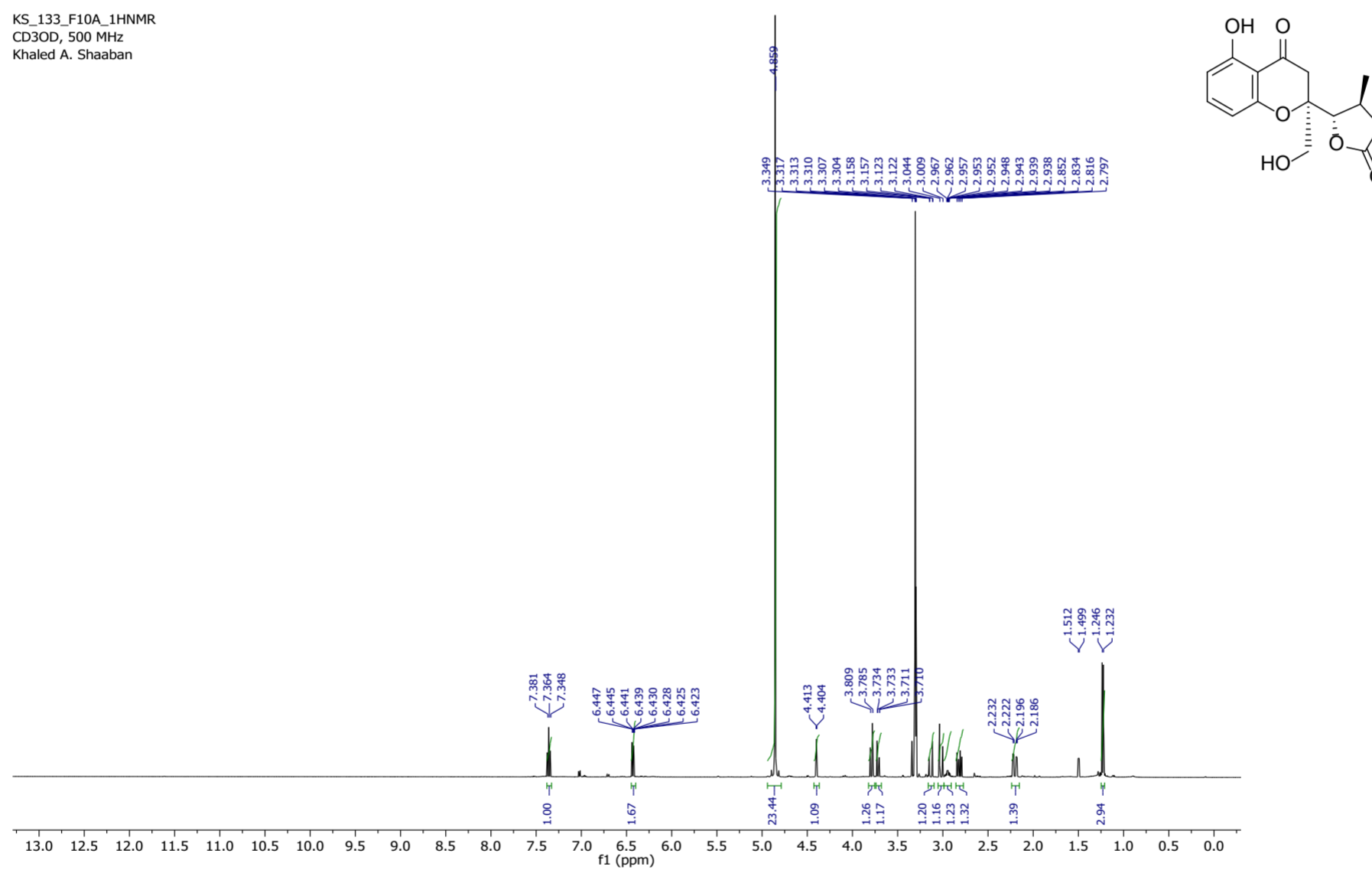
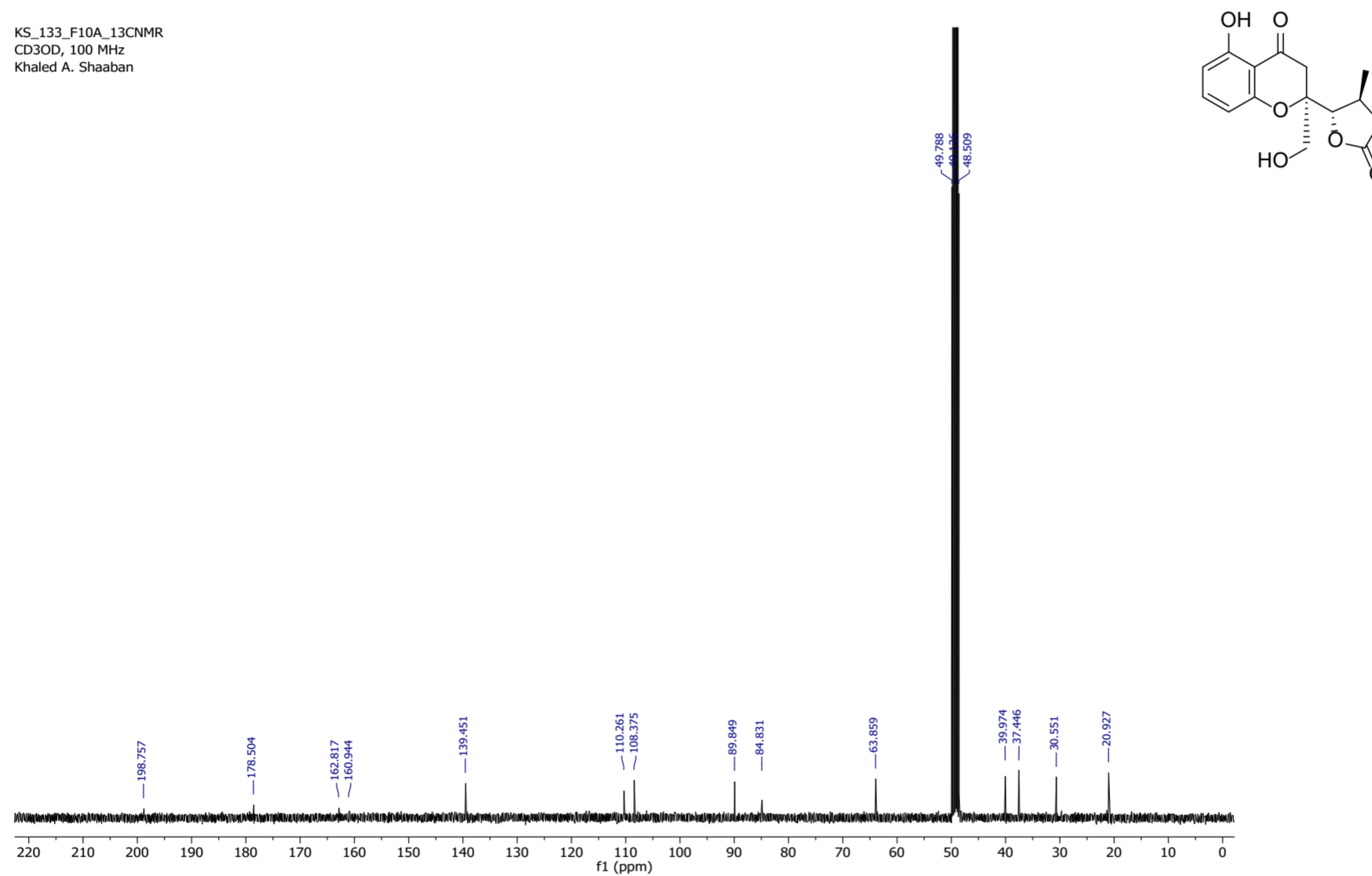


Fig. 22S <sup>1</sup>H NMR spectrum (CD<sub>3</sub>OD, 500 MHz) of paecilin R (2).

## Supplementary material

KS\_133\_F10A\_13CNMR  
CD3OD, 100 MHz  
Khaled A. Shaaban

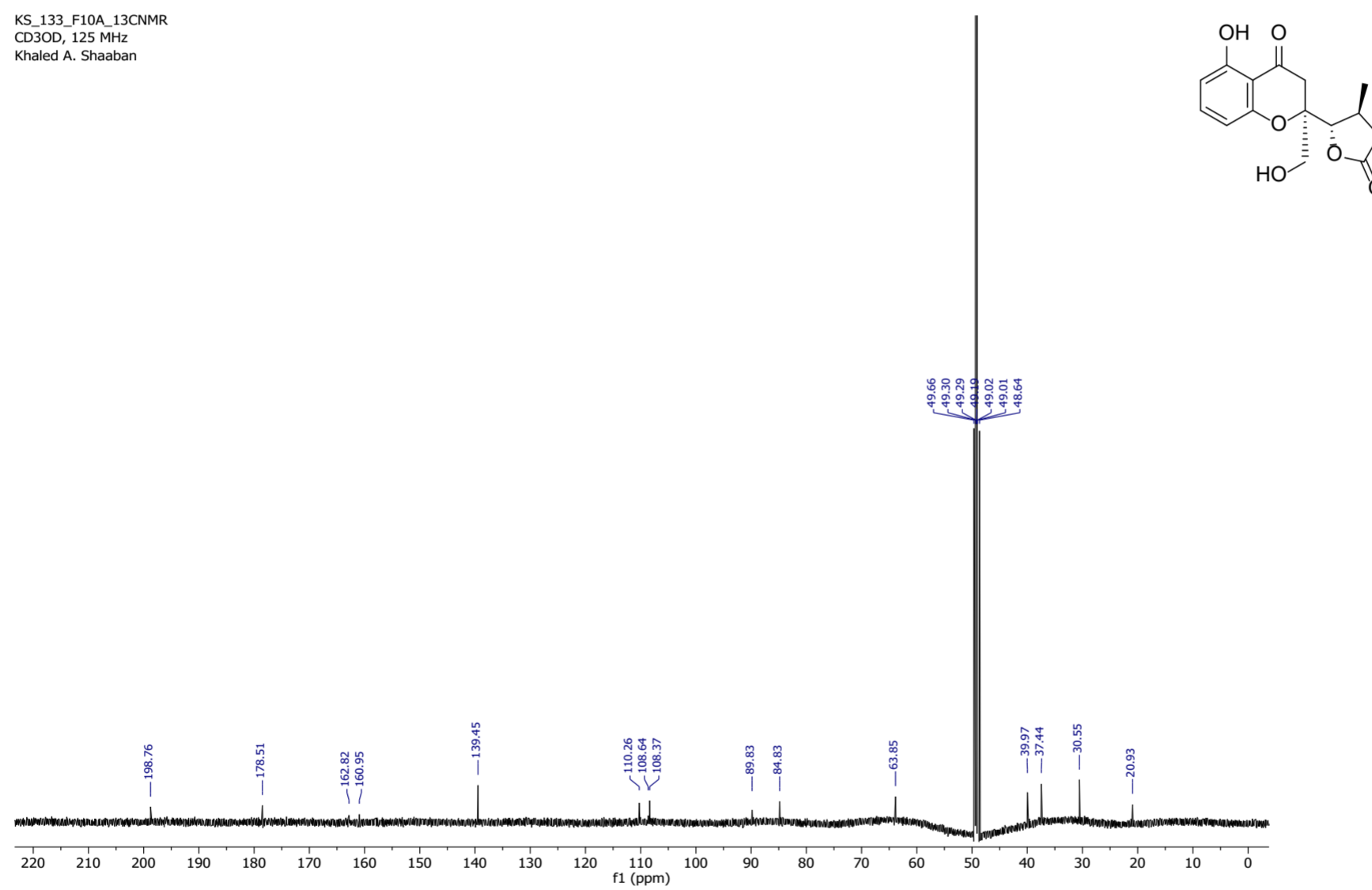


**Fig. 23S**  $^{13}\text{C}$  NMR spectrum ( $\text{CD}_3\text{OD}$ , 100 MHz) of paecilin R (**2**).



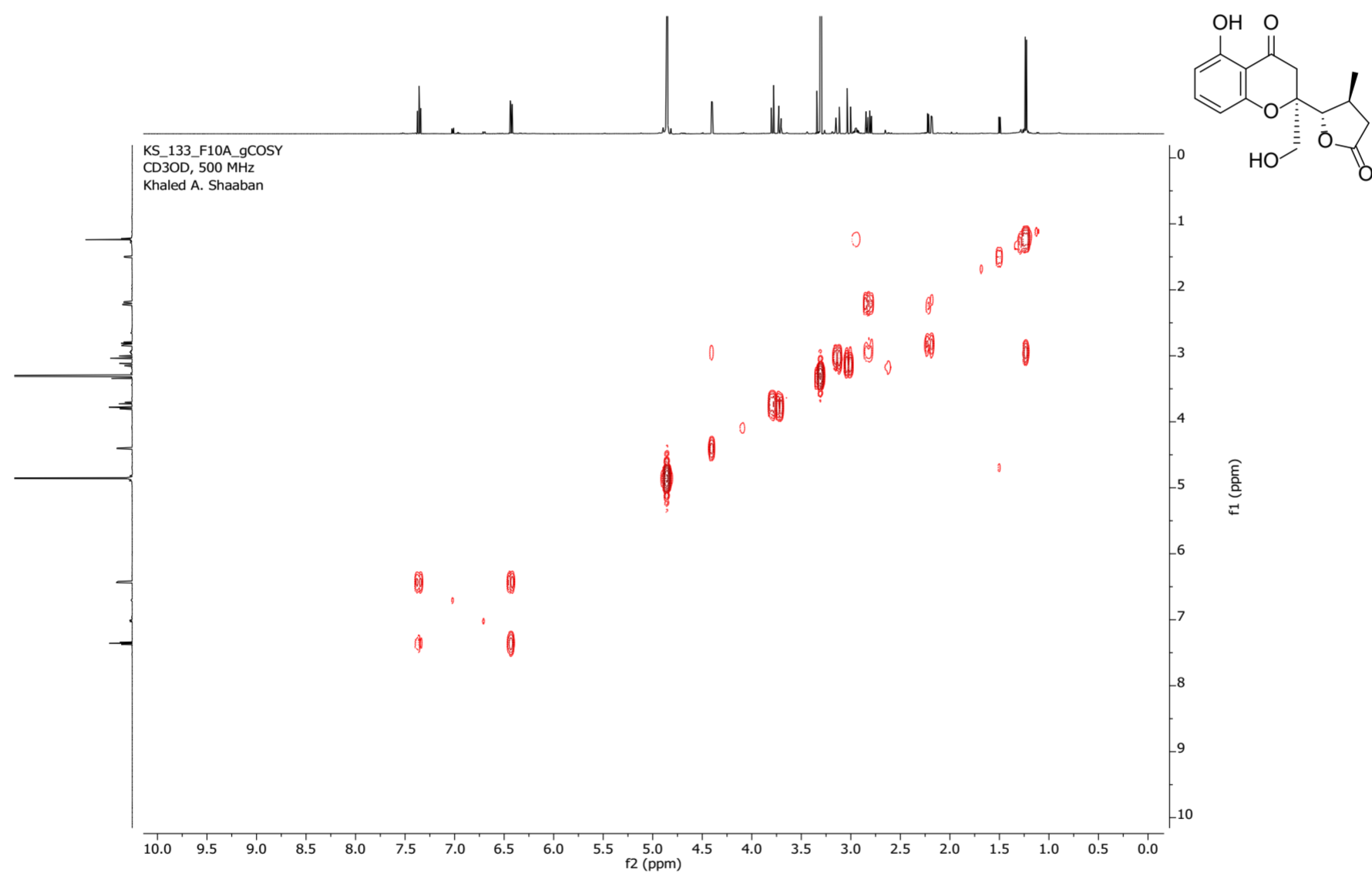
## Supplementary material

KS\_133\_F10A\_13CNMR  
CD3OD, 125 MHz  
Khaled A. Shaaban



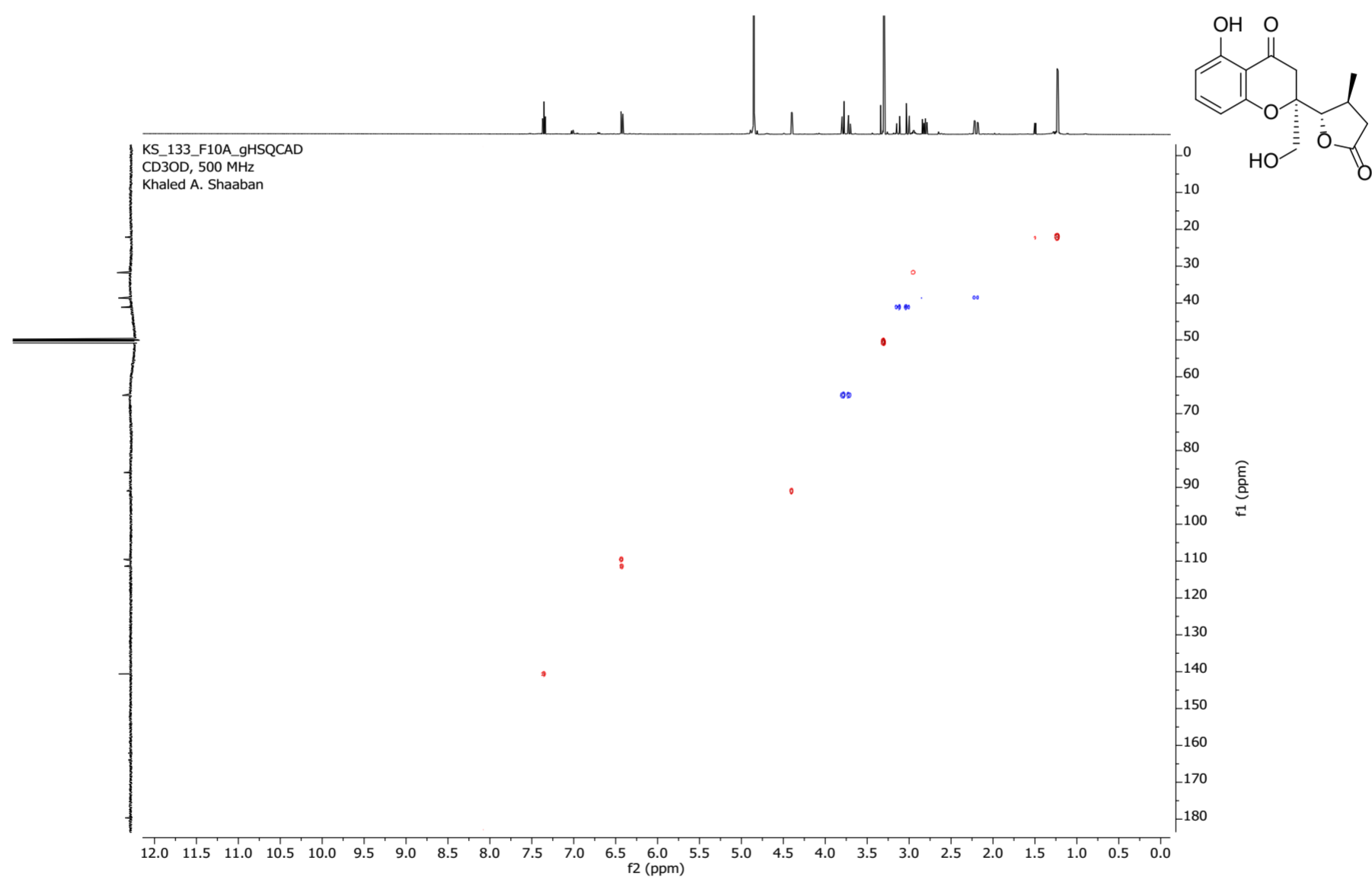
**Fig. 24S** <sup>13</sup>C NMR spectrum (CD<sub>3</sub>OD, 125 MHz) of paecilin R (**2**).

## Supplementary material



**Fig. 25S**  $^1\text{H}, ^1\text{H}$ -COSY spectrum ( $\text{CD}_3\text{OD}$ , 500 MHz) of paecilin R (**2**).

## Supplementary material



**Fig. 26S** HSQC spectrum (CD<sub>3</sub>OD, 500 MHz) of paecilin R (**2**).

## Supplementary material

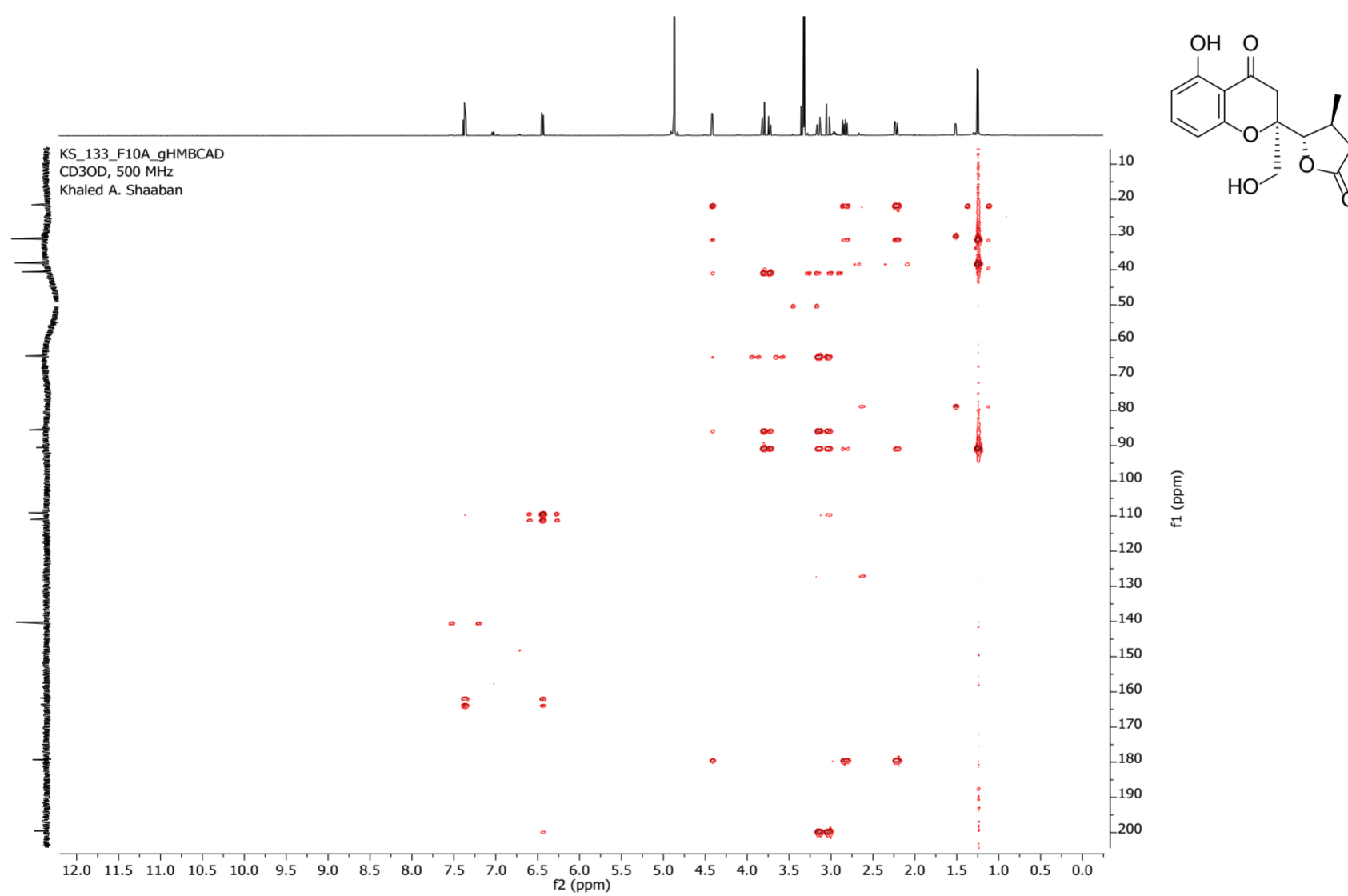
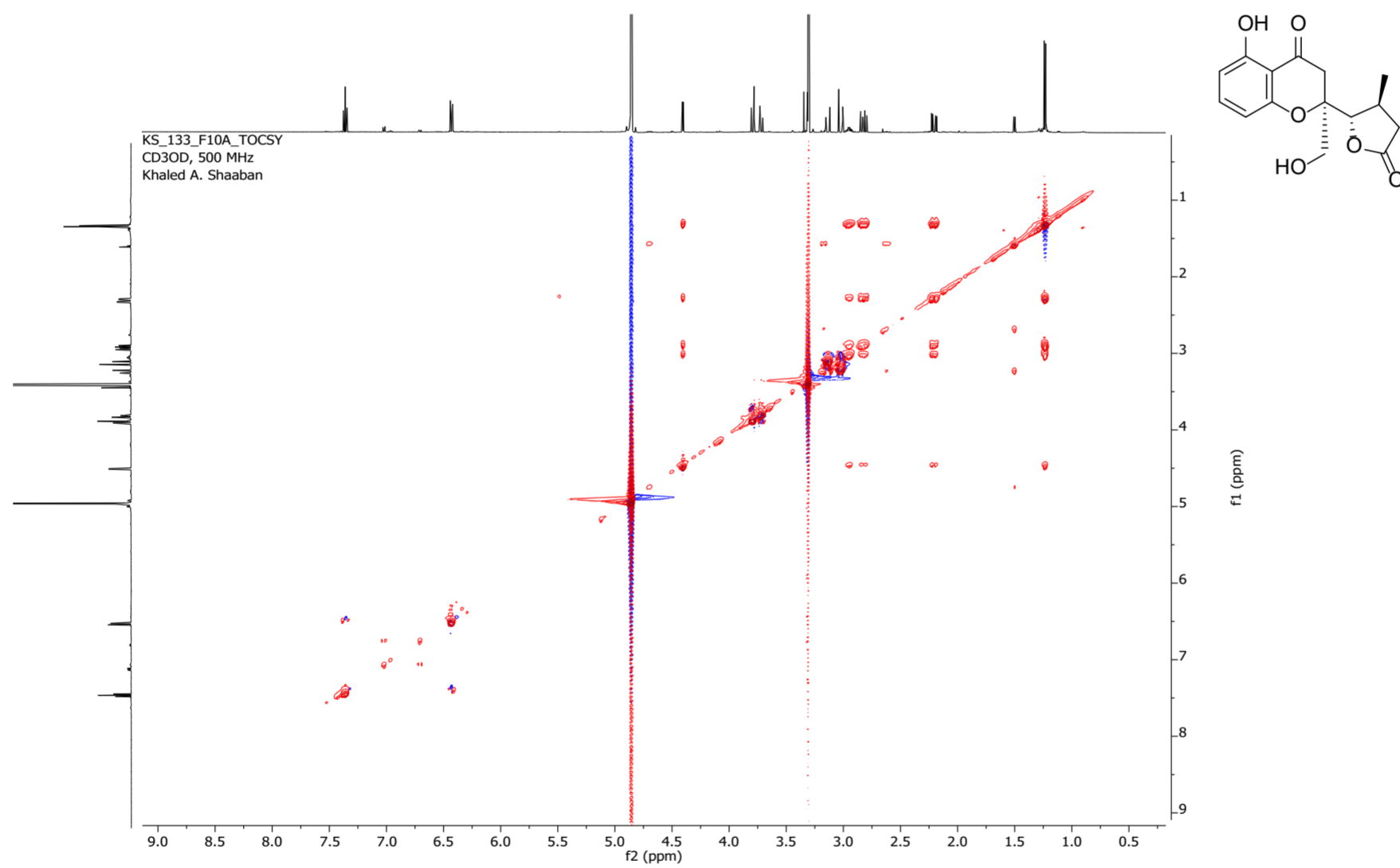


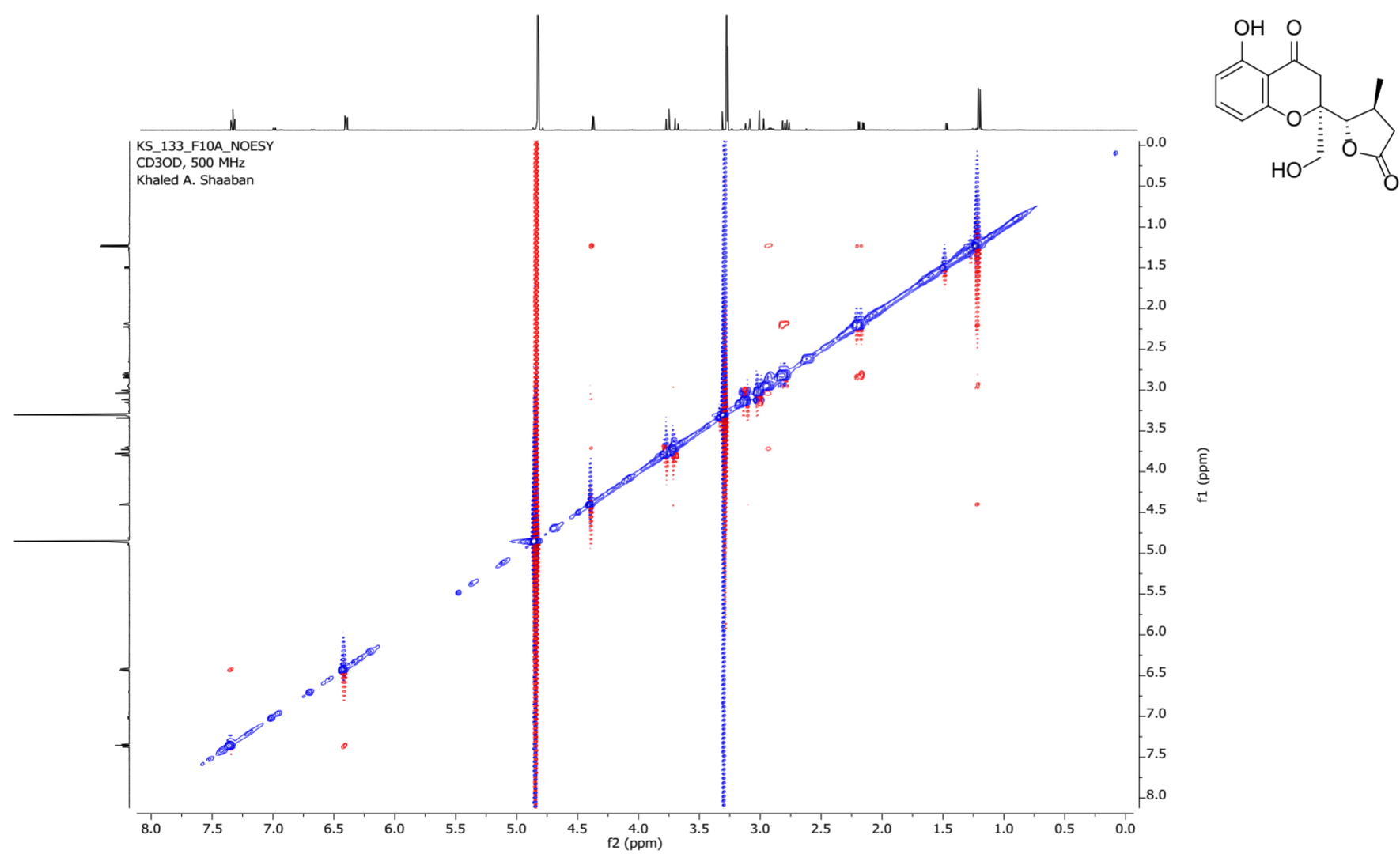
Fig. 27S HMBC spectrum (CD<sub>3</sub>OD, 500 MHz) of paecilin R (2).

## Supplementary material



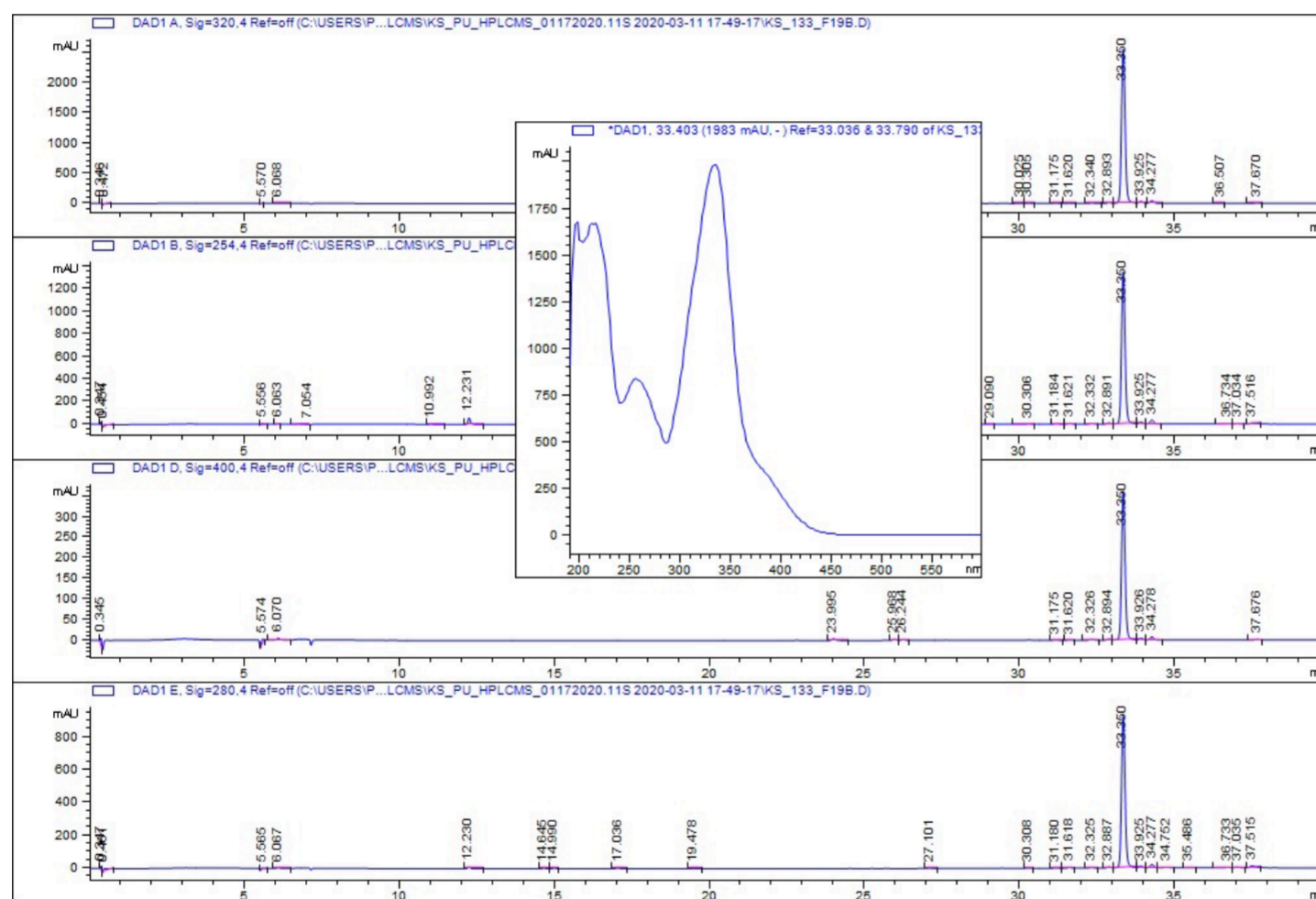
**Fig. 28S** TOCSY spectrum (CD<sub>3</sub>OD, 500 MHz) of paecilin R (**2**).

## Supplementary material



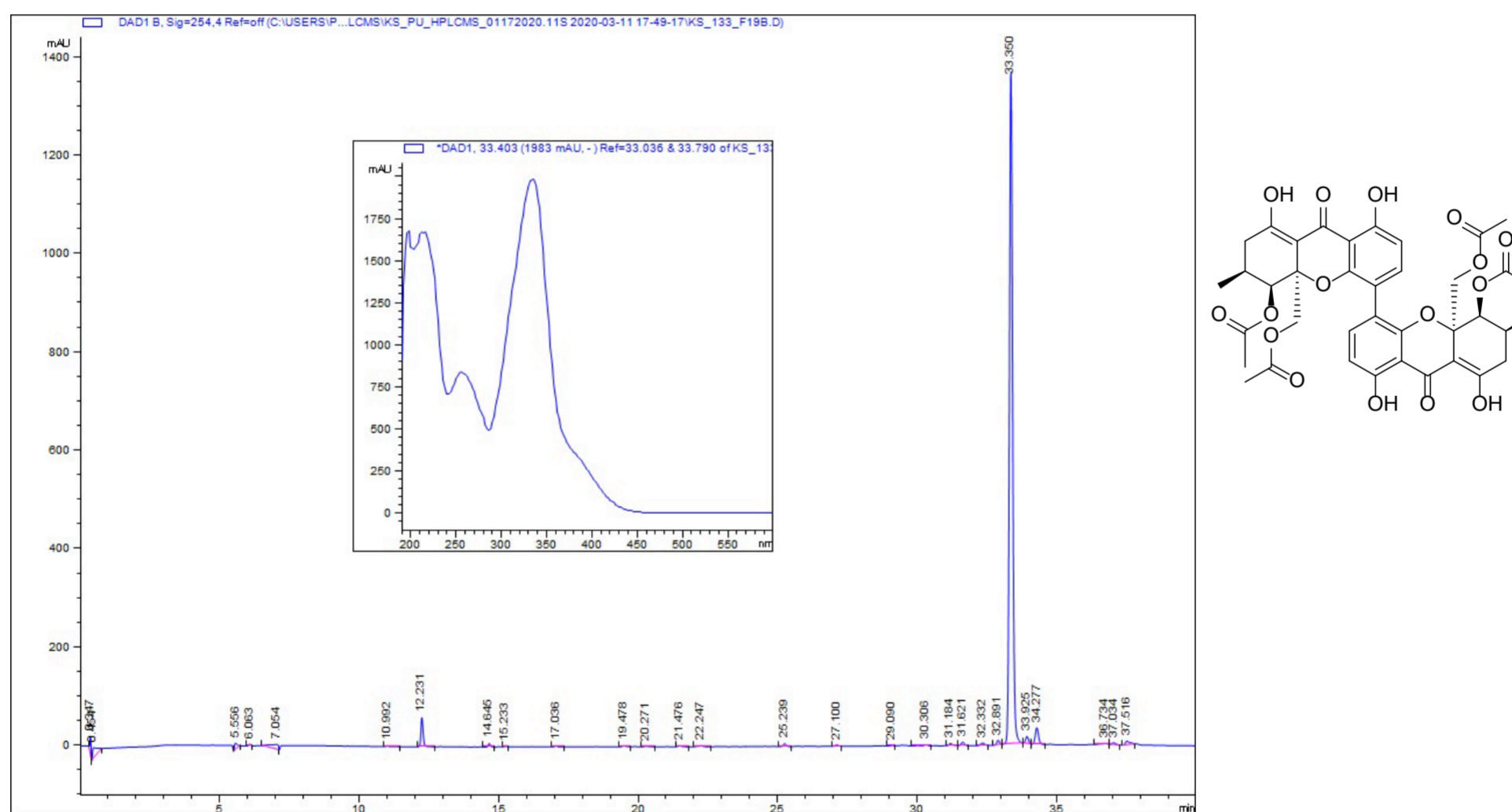
**Fig. 29S** NOESY spectrum (CD<sub>3</sub>OD, 500 MHz) of paecilin R (**2**).

## Supplementary material



**Fig. 30S** HPLC analysis of phomoxanthone A (**3**). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 320, 254, 280, 400 nm. UV-vis inset of full wavelength scan (190-600 nm).

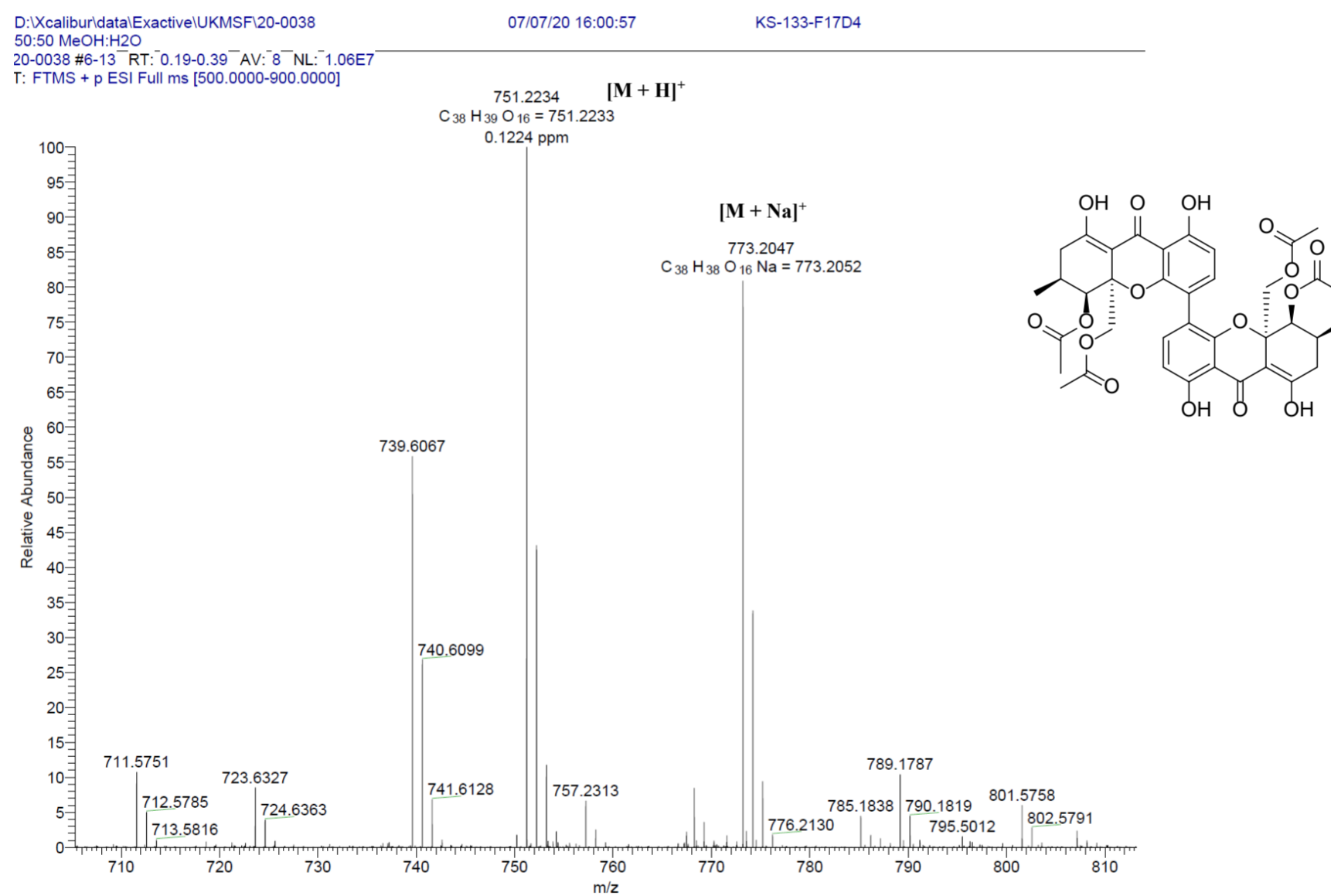
## Supplementary material



**Fig. 31S** HPLC analysis of phomoxanthone A (**3**). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 254 nm. UV-vis inset of full wavelength scan (190-600 nm).

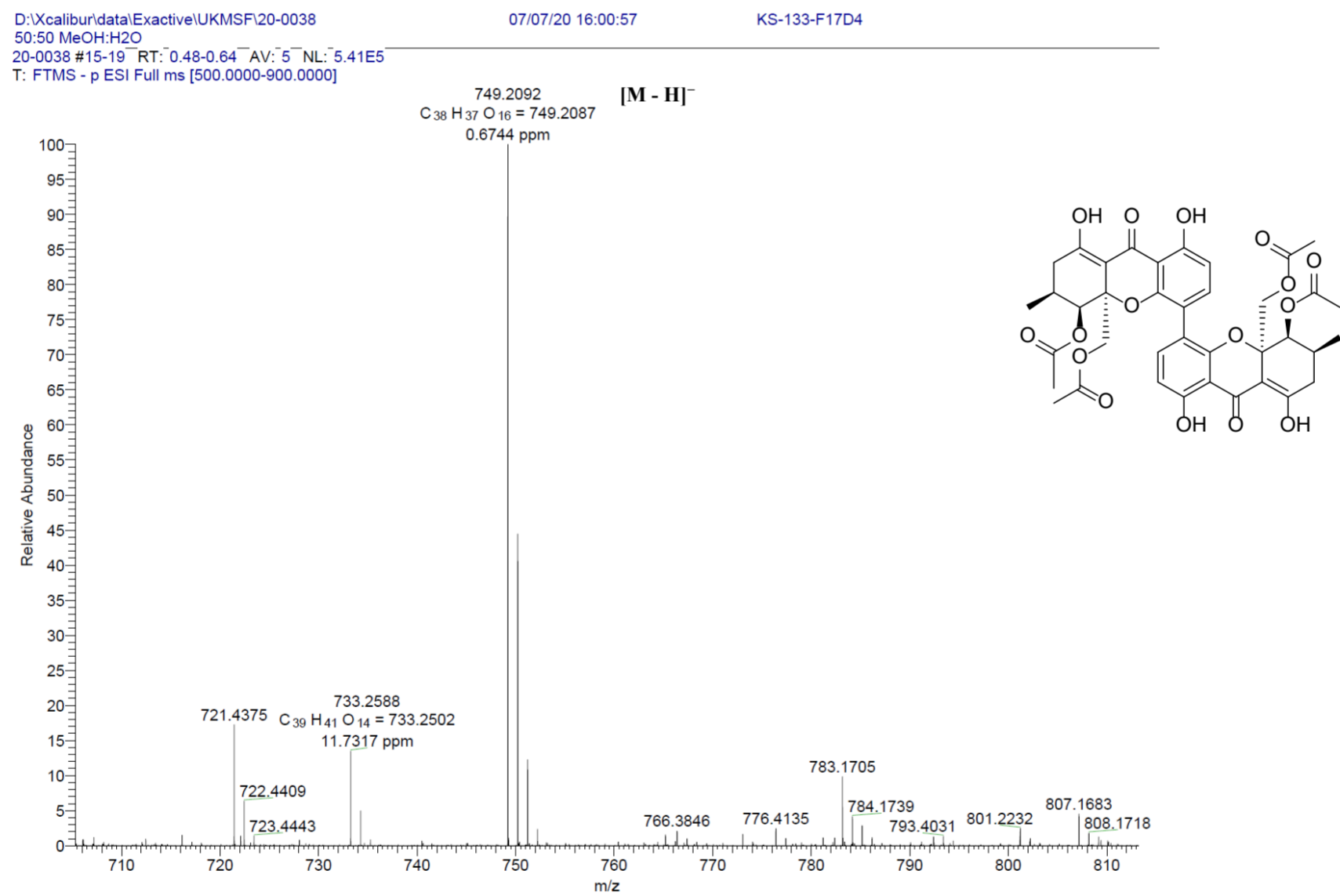


## Supplementary material



**Fig. 32S** (+)-HRESI-MS spectrum of phomoxanthone A (**3**).

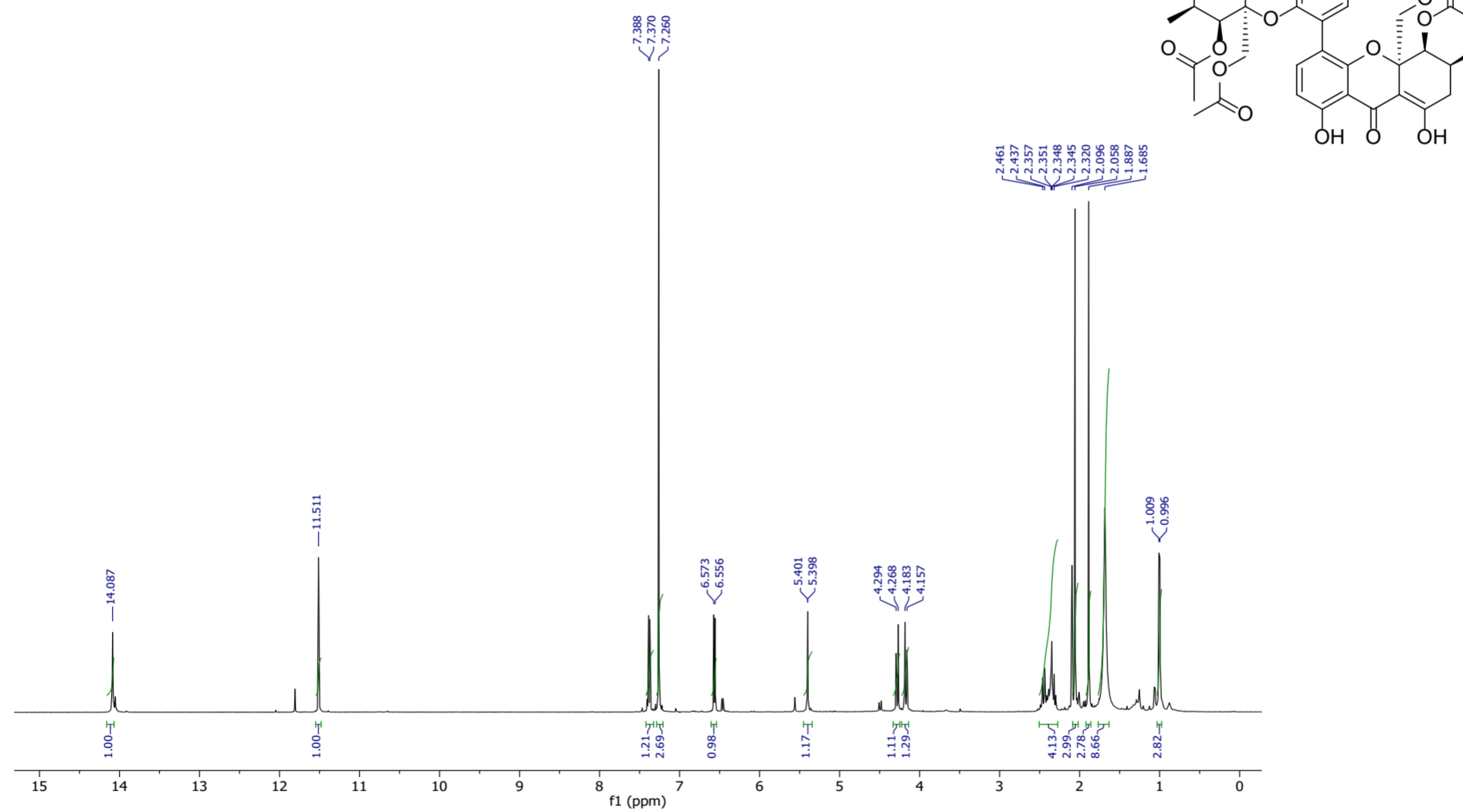
## Supplementary material



**Fig. 33S** (-)-HRESI-MS spectrum of phomoxanthone A (**3**).

## Supplementary material

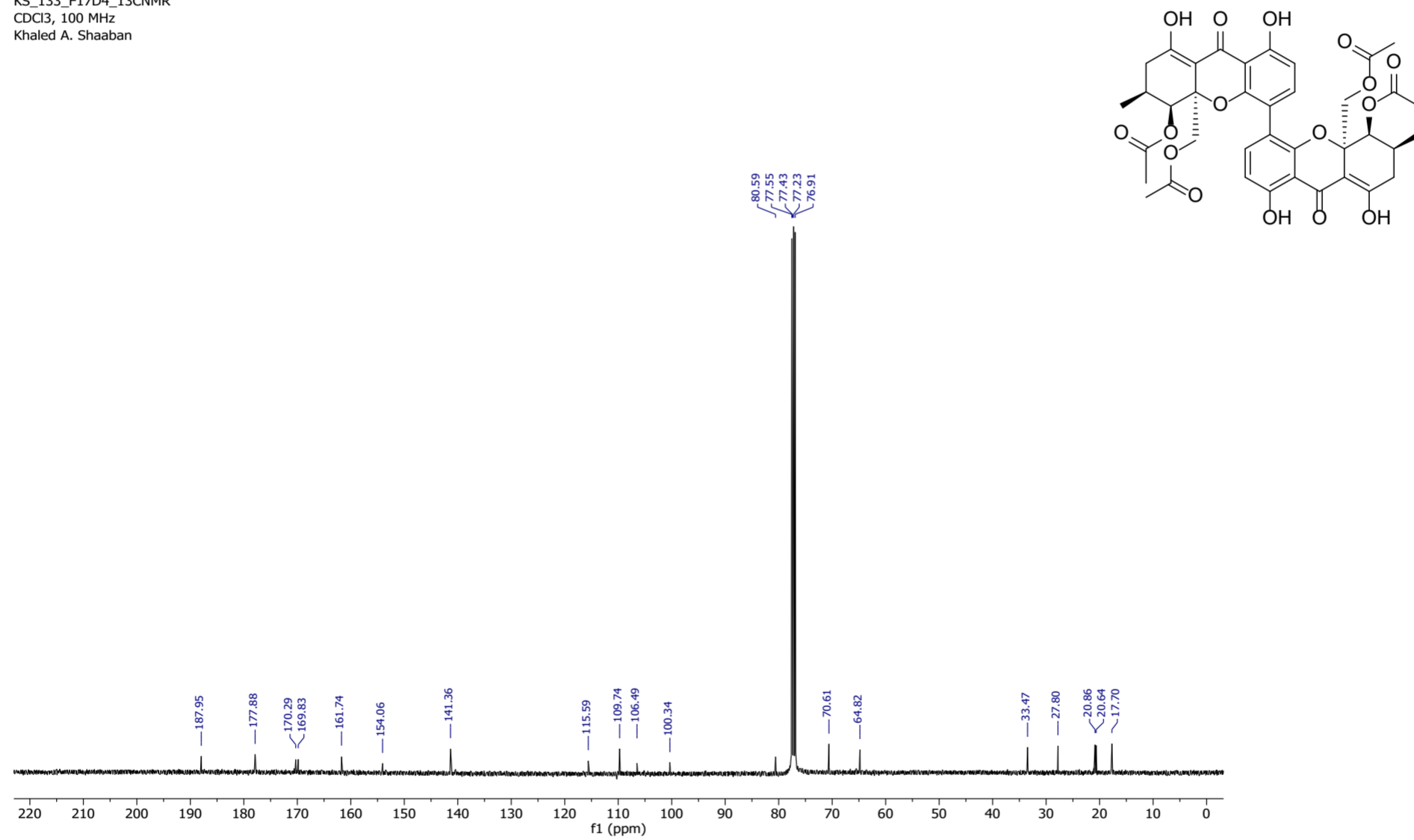
KS\_133\_F17D4\_1HNMR  
CDCl<sub>3</sub>, 500 MHz  
Khaled A. Shaaban



**Fig. 34S** <sup>1</sup>H NMR spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone A (**3**).

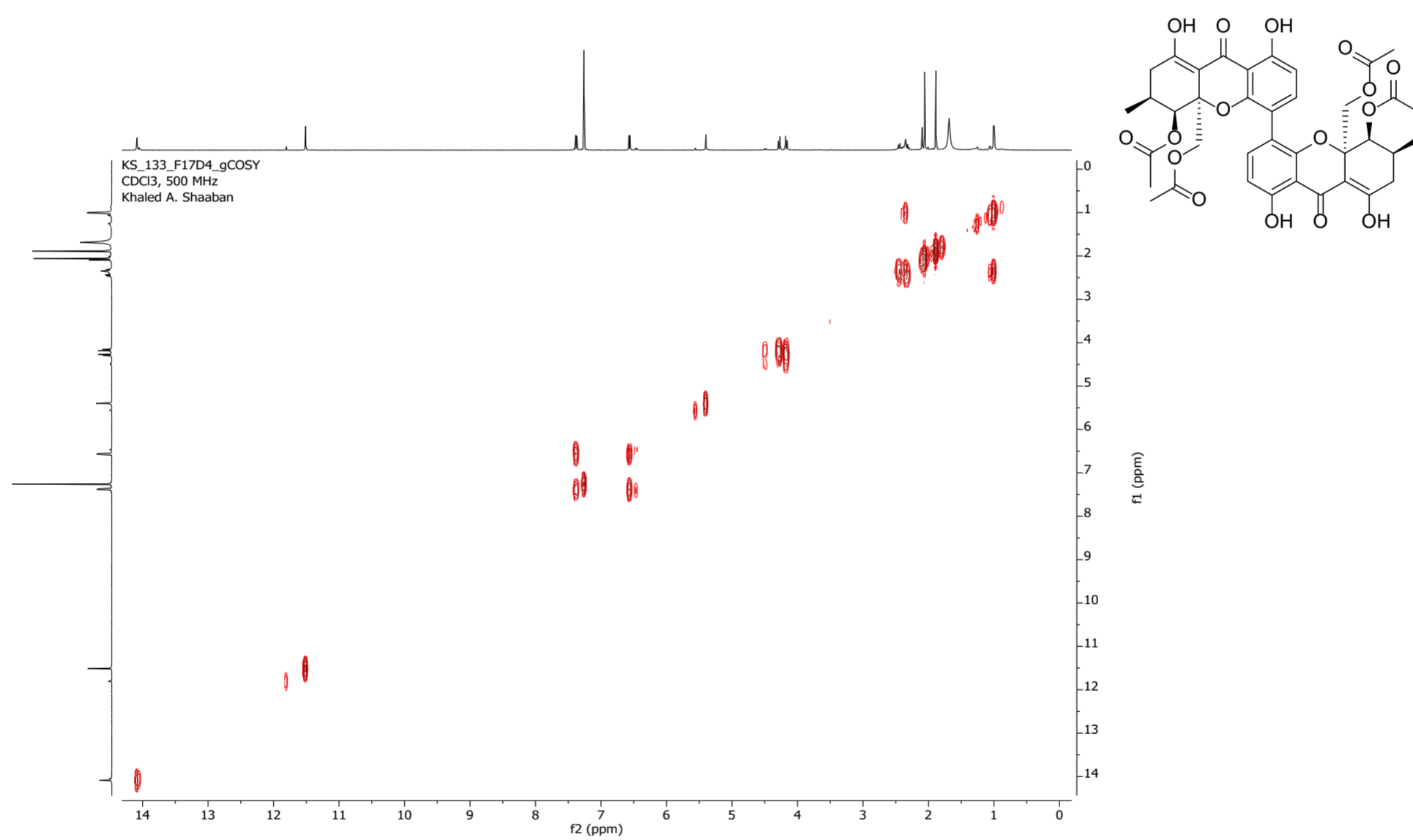
## Supplementary material

KS\_133\_F17D4\_13CNMR  
CDCl<sub>3</sub>, 100 MHz  
Khaled A. Shaaban



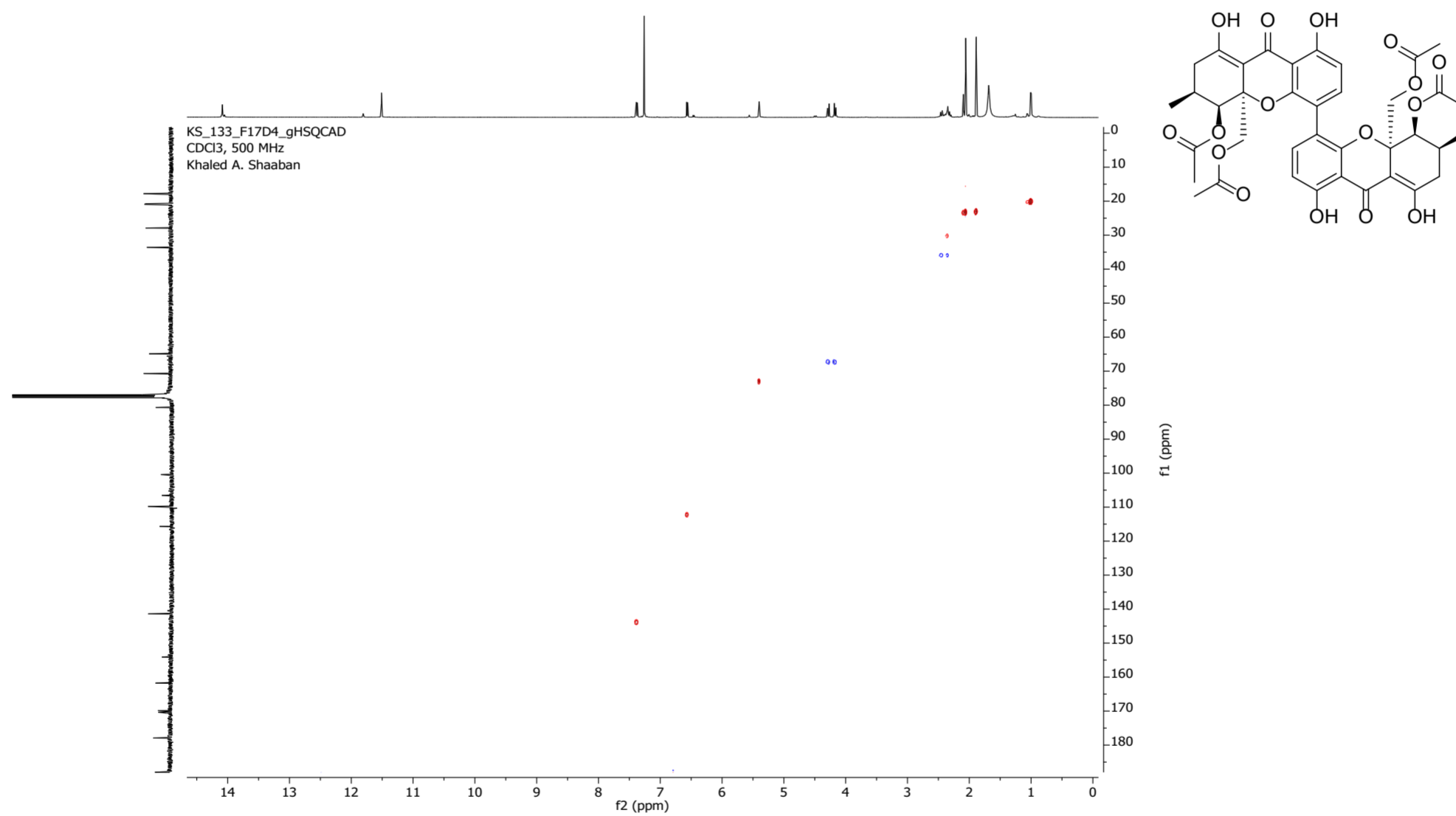
**Fig. 35S** <sup>13</sup>C NMR spectrum (CDCl<sub>3</sub>, 100 MHz) of phomoxanthone A (**3**).

## Supplementary material



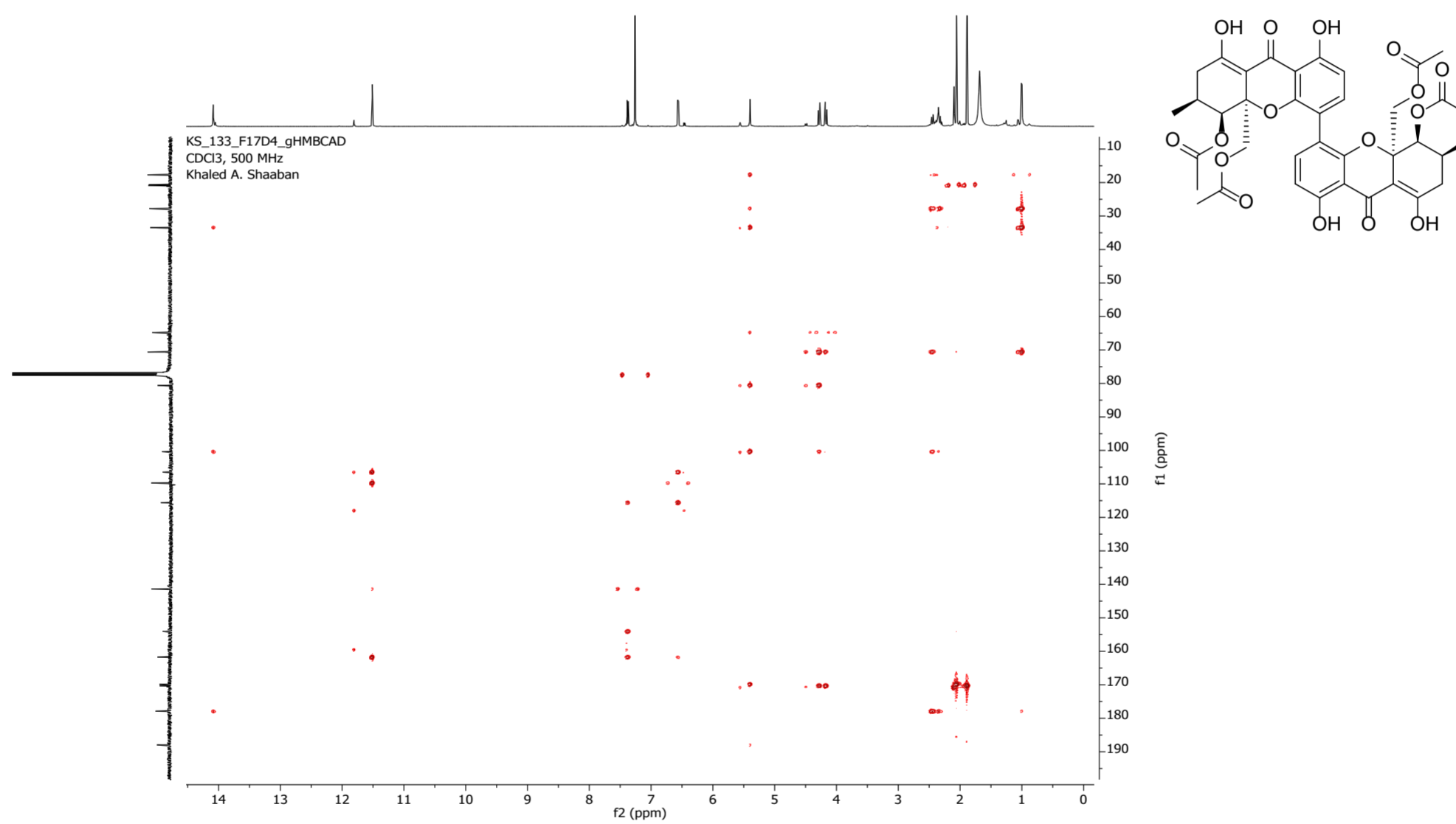
**Fig. 36S** <sup>1</sup>H, <sup>1</sup>H-COSY spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone A (**3**).

## Supplementary material



**Fig. 37S** HSQC spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone A (**3**).

## Supplementary material

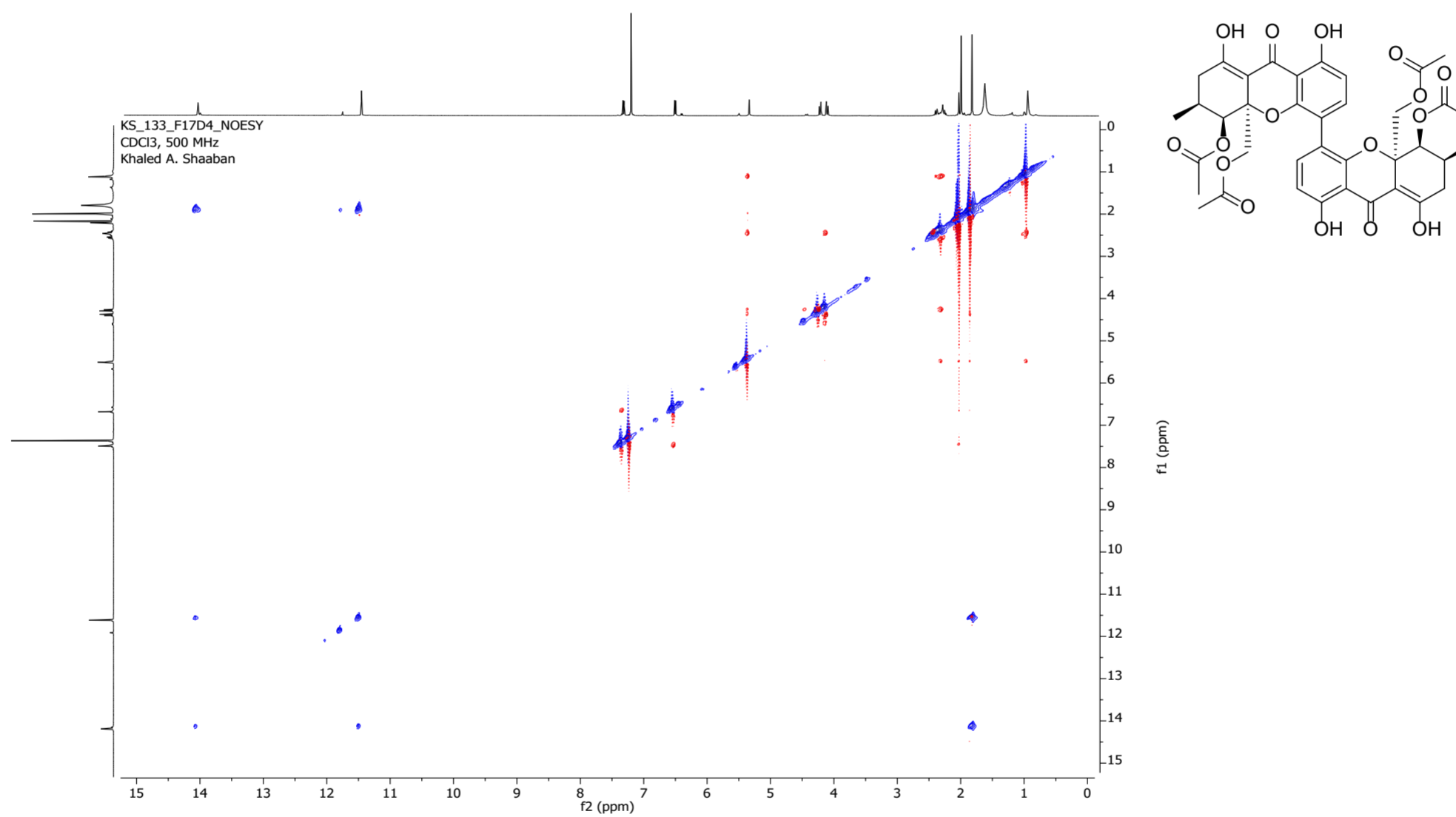


**Fig. 38S** HMBC spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone A (3).



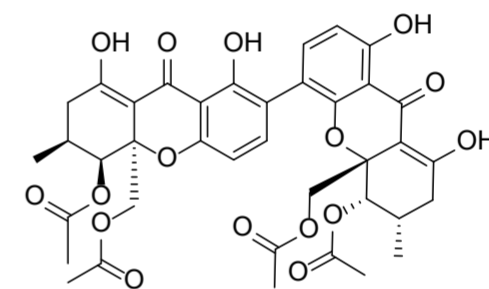
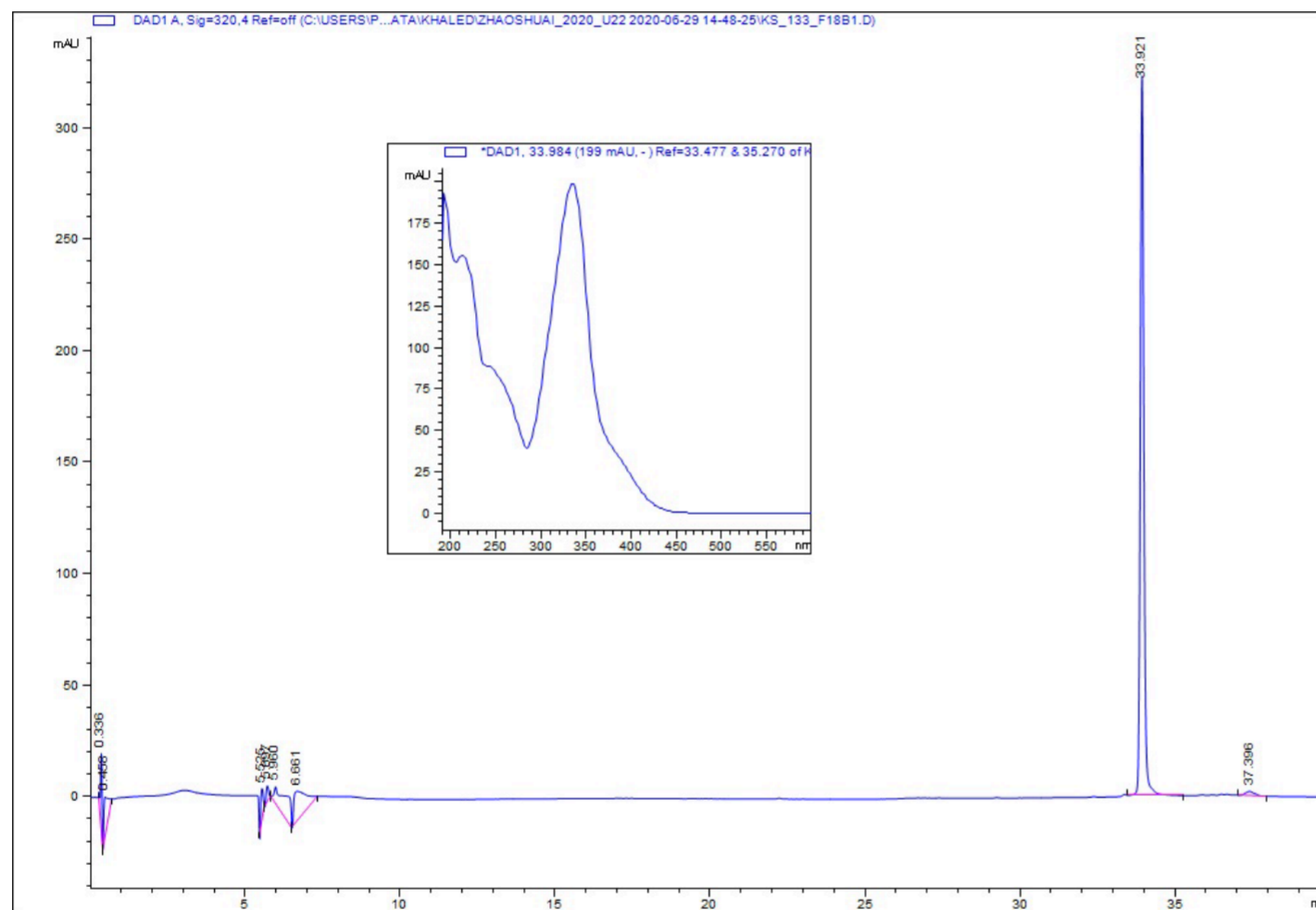


## Supplementary material



**Fig. 40S** NOESY spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone A (**3**).

## Supplementary material



**Fig. 41S** HPLC analysis of phomoxanthone B (**4**). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 320 nm. UV-vis inset of full wavelength scan (190-600 nm).

## Supplementary material

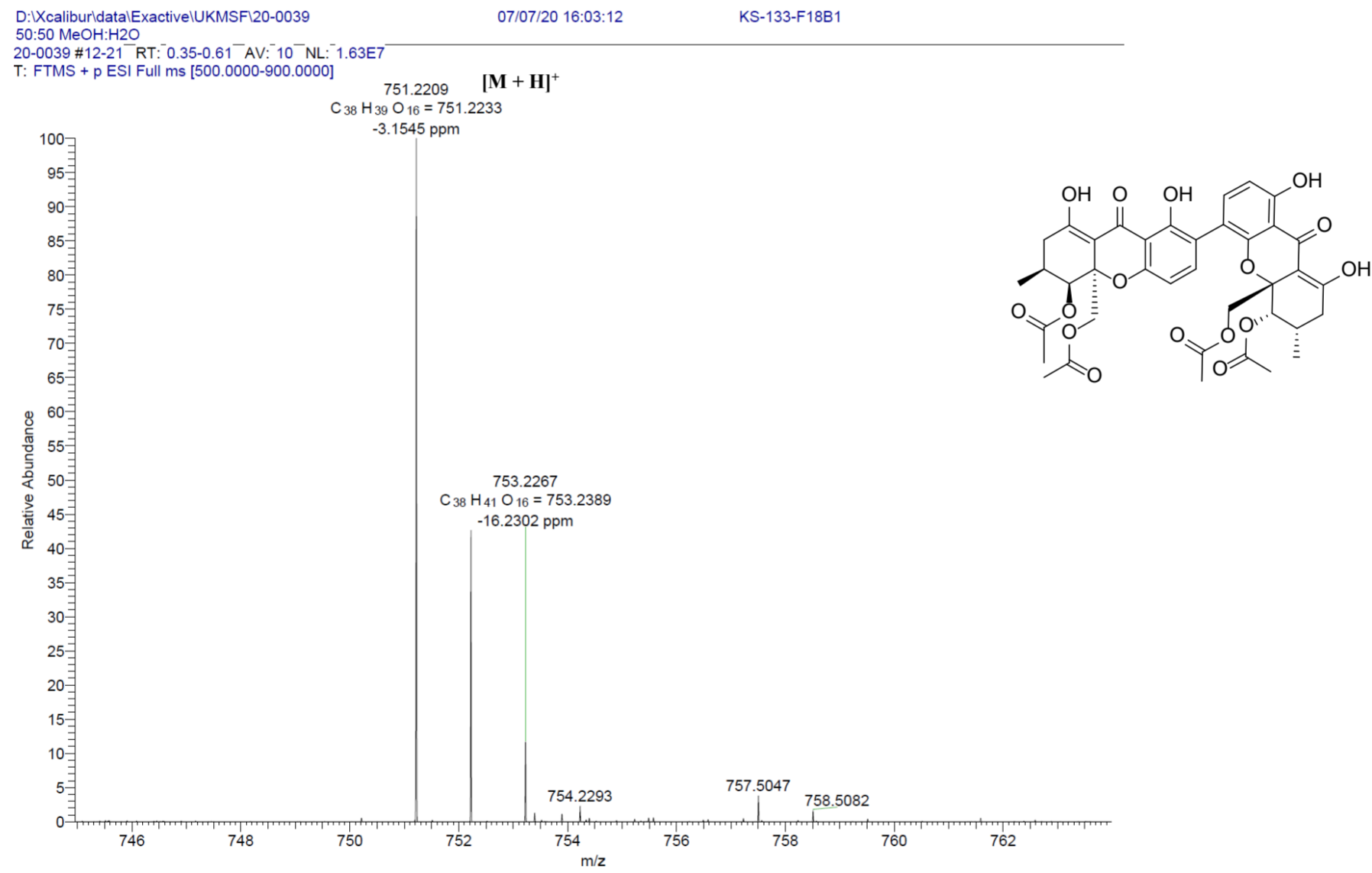


Fig. 42S (+)-HRESI-MS spectrum of phomoxanthone B (4).

## Supplementary material

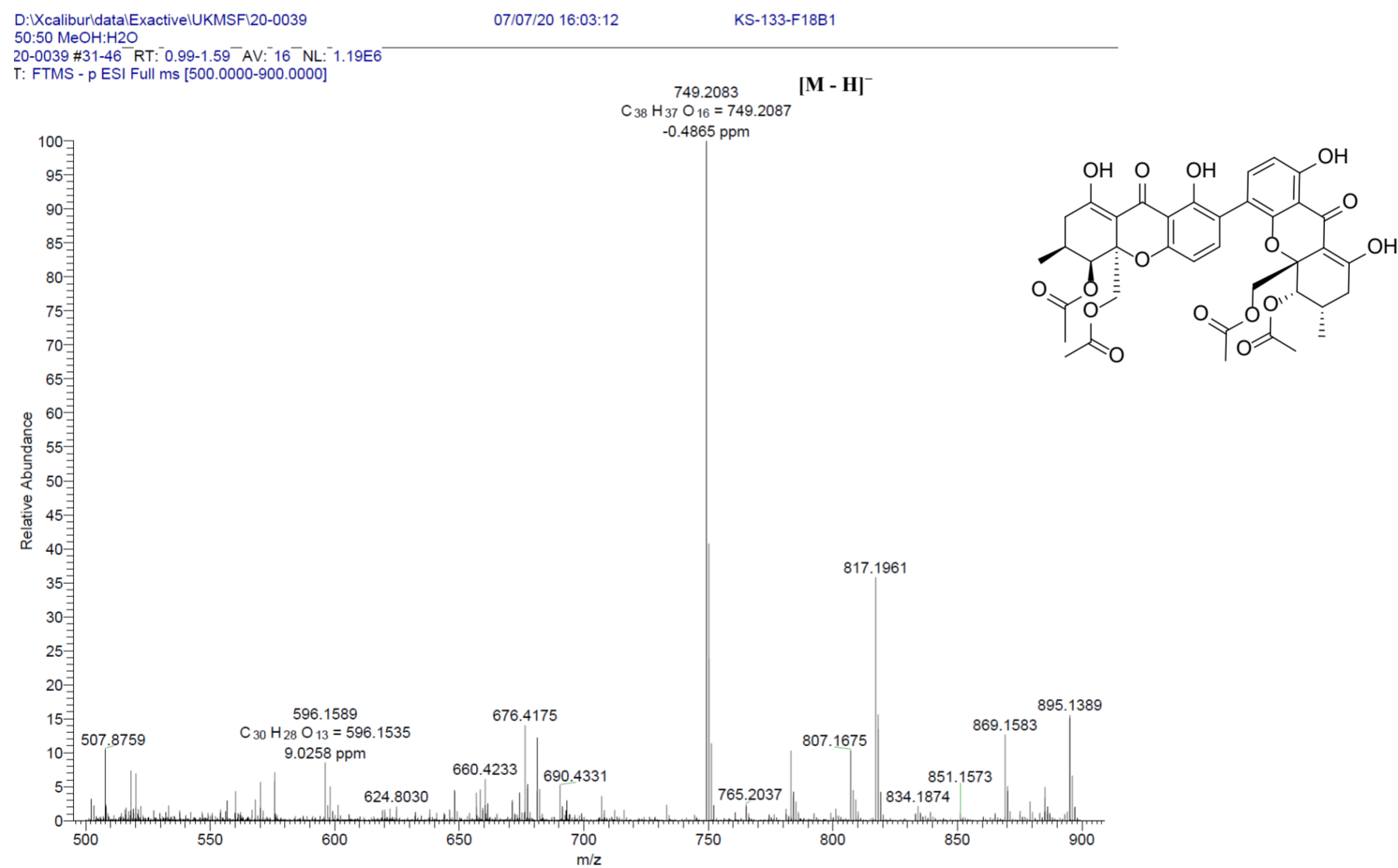


Fig. 43S (-)-HRESI-MS spectrum of phomoxanthone B (4).

## Supplementary material

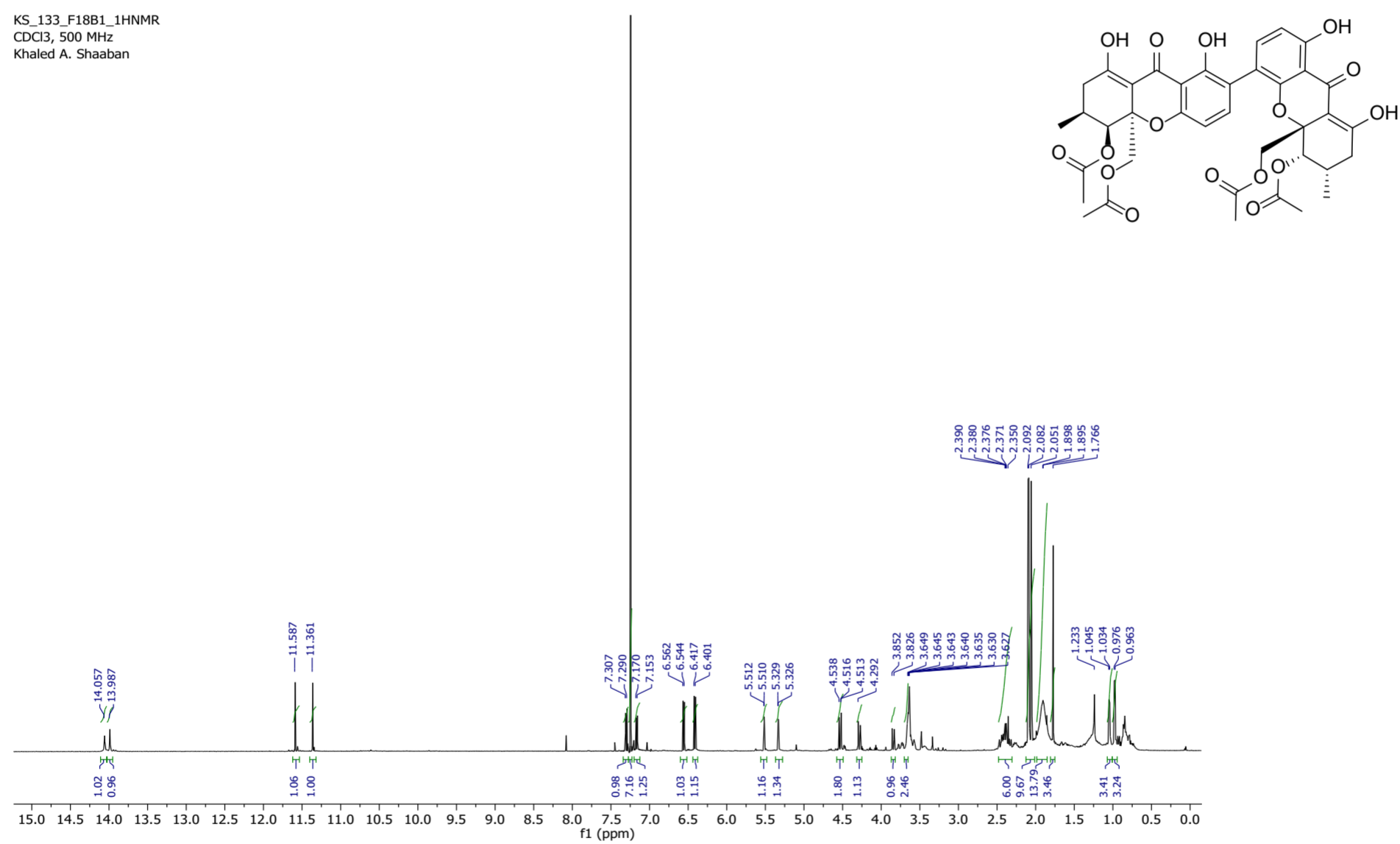
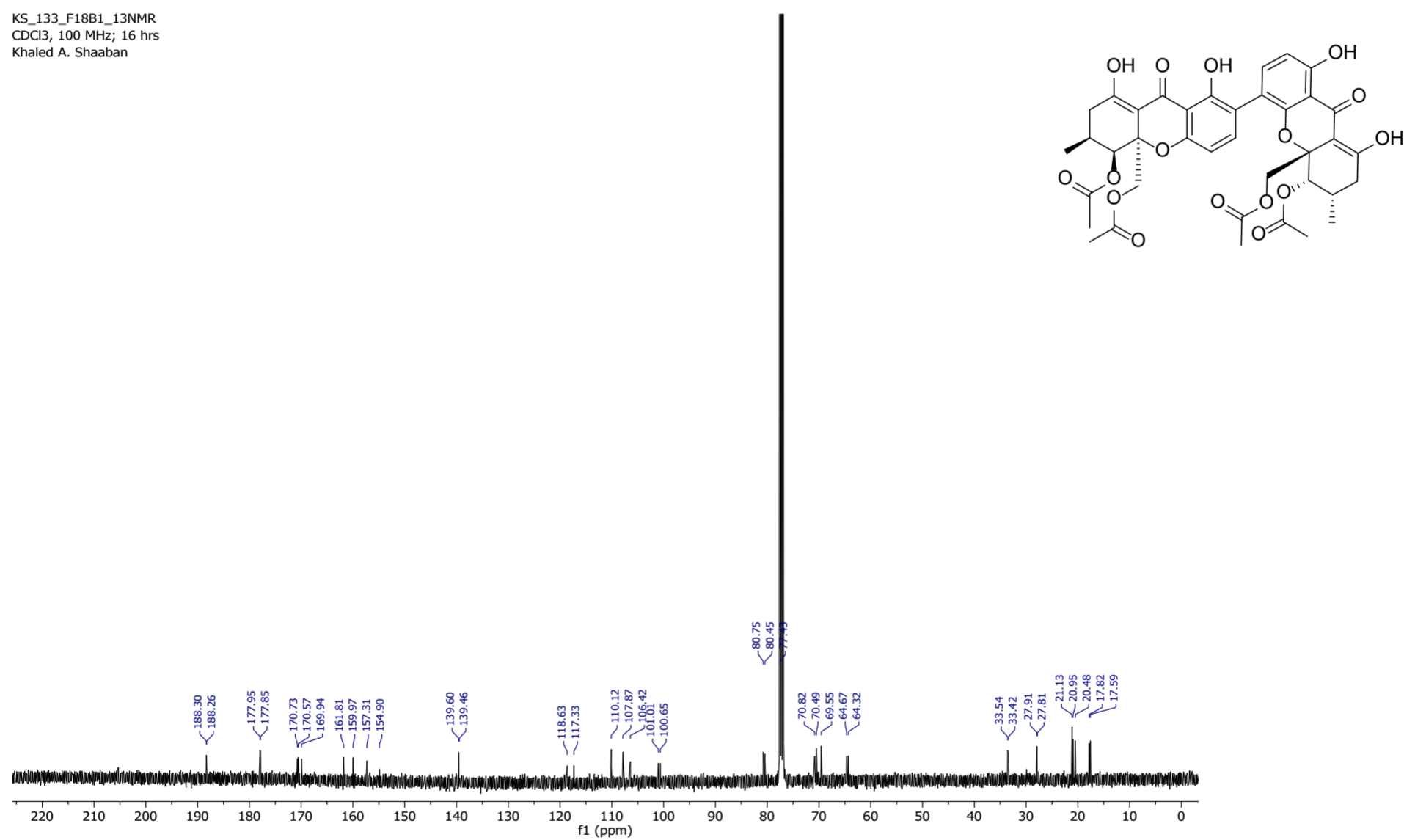


Fig. 44S <sup>1</sup>H, <sup>1</sup>H-COSY spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone B (4).

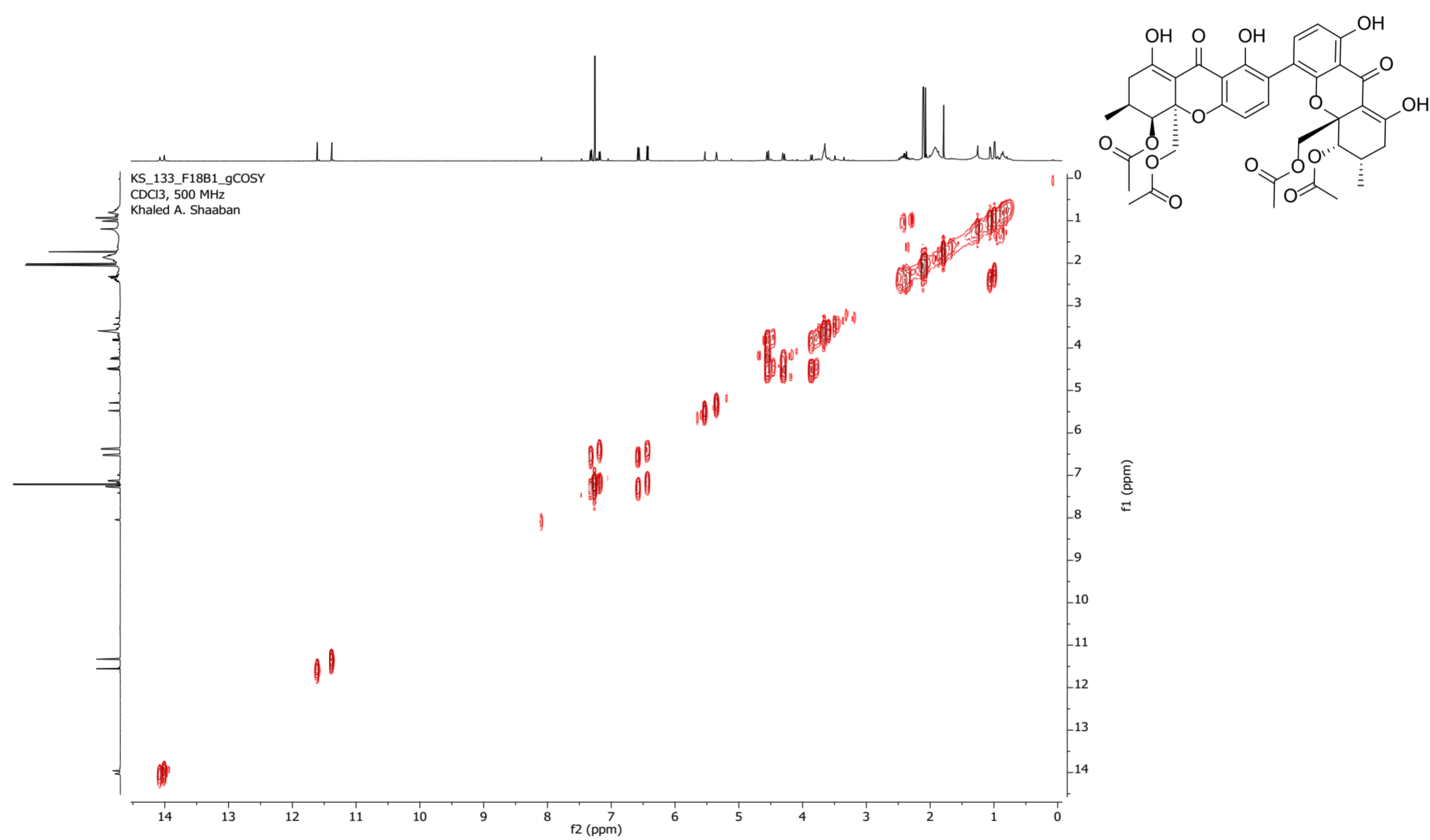
## Supplementary material

KS\_133\_F18B1\_13NMR  
CDCl<sub>3</sub>, 100 MHz; 16 hrs  
Khaled A. Shaaban



**Fig. 45S** <sup>13</sup>C NMR spectrum (CDCl<sub>3</sub>, 100 MHz) of phomoxanthone B (**4**).

## Supplementary material



**Fig. 46S** <sup>1</sup>H,<sup>1</sup>H-COSY spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone B (**4**).

## Supplementary material

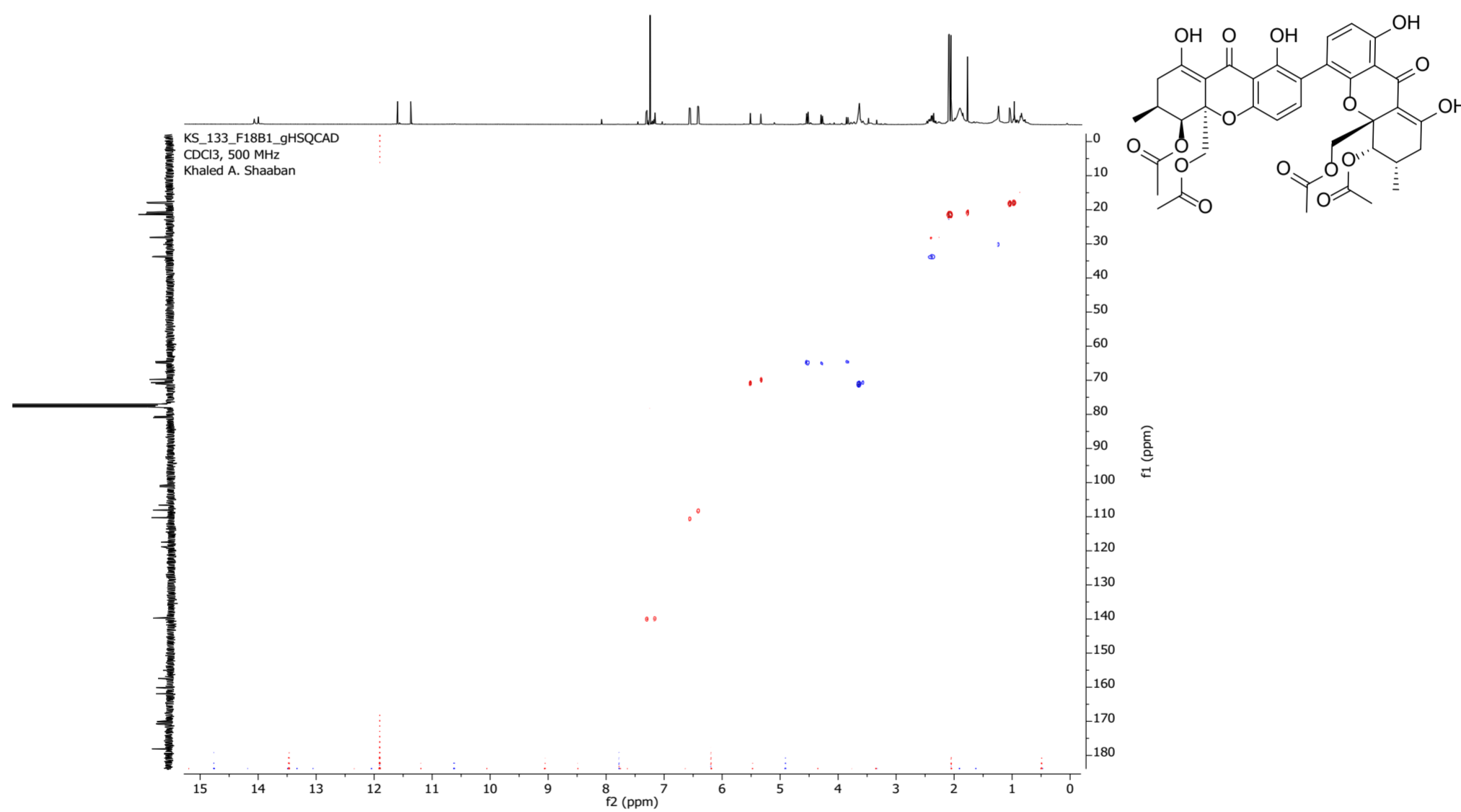
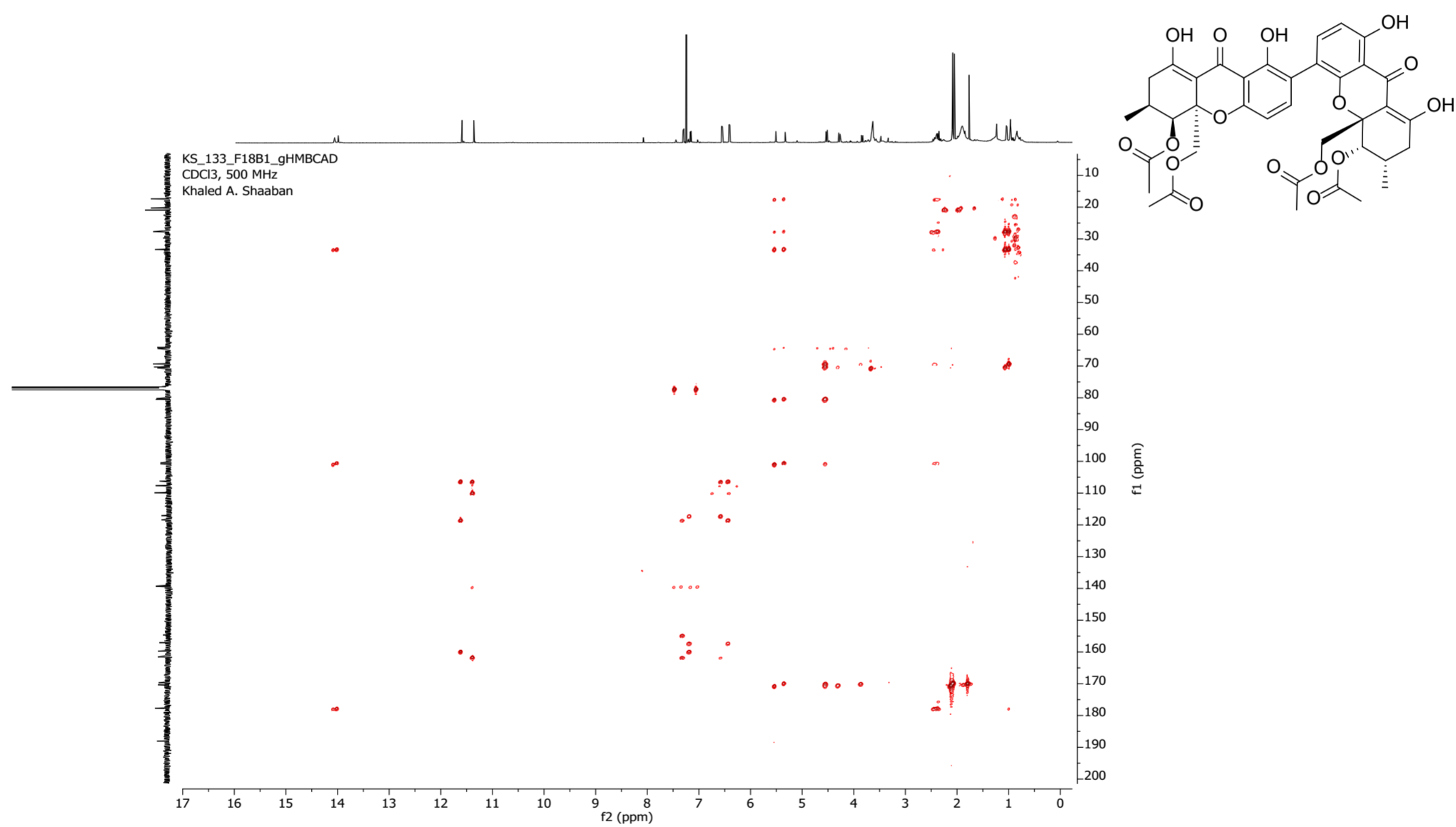


Fig. 47S HSQC spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone B (4).

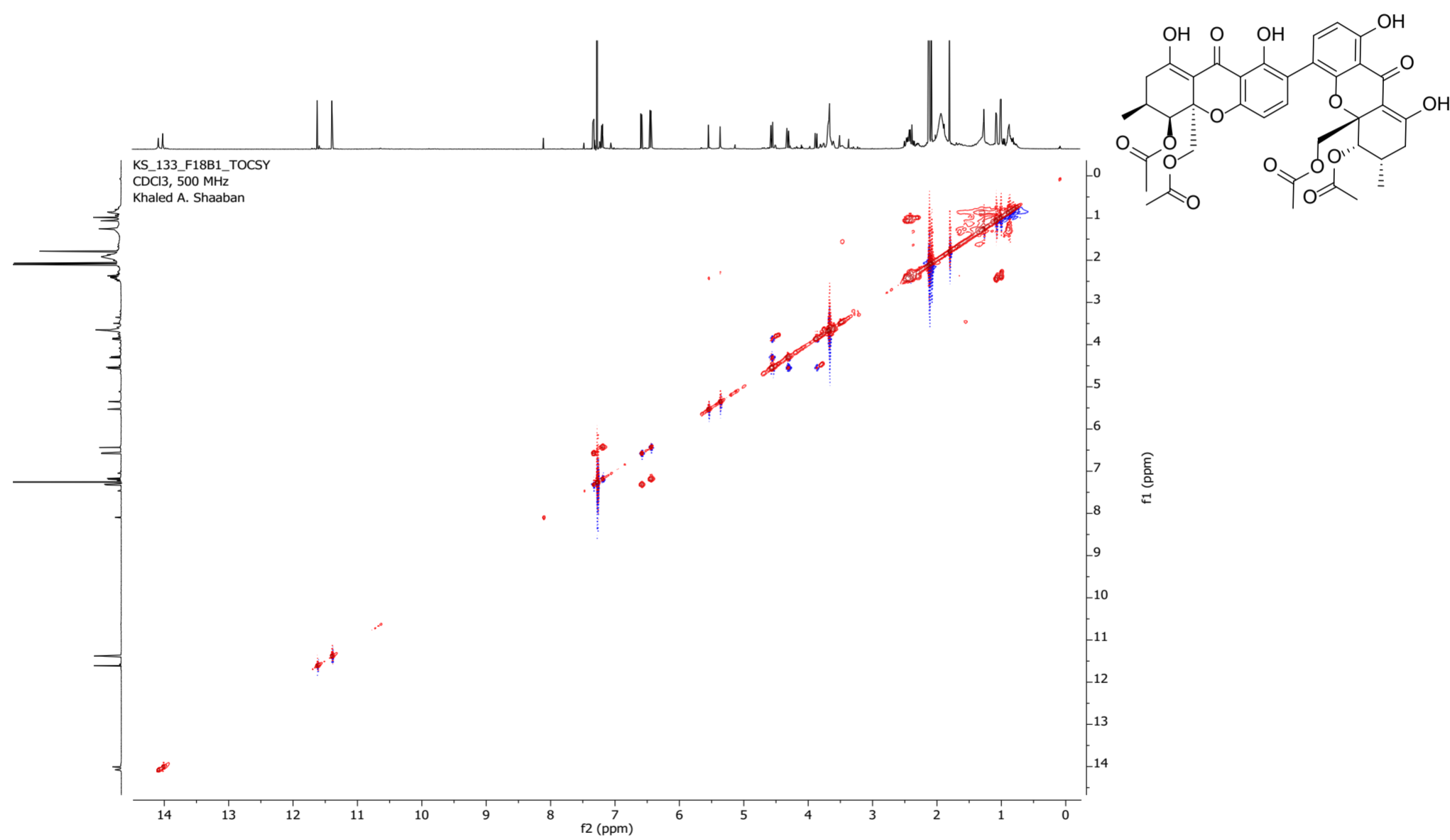


## Supplementary material



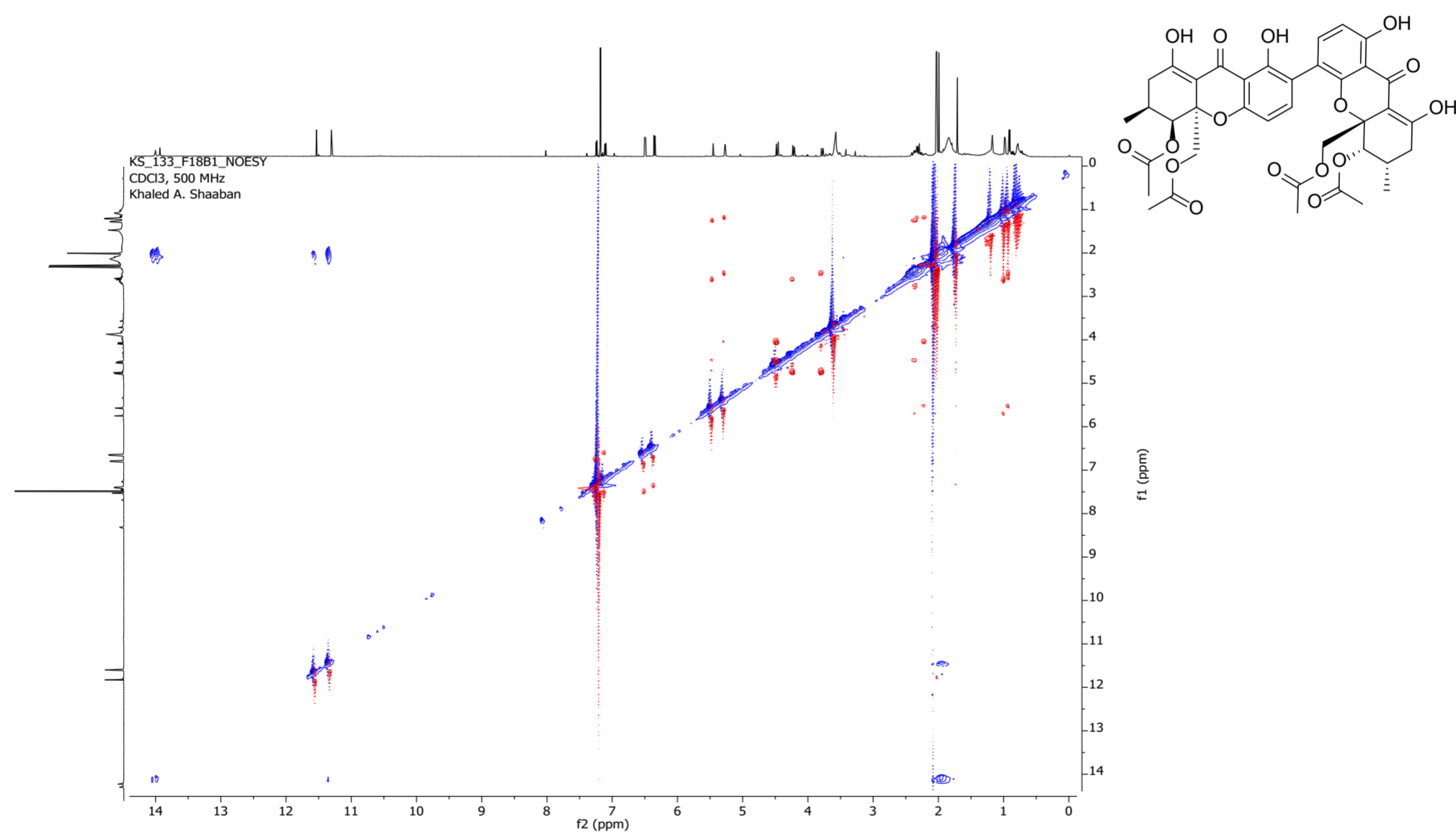
**Fig. 48S** HMBC spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone B (4).

## Supplementary material



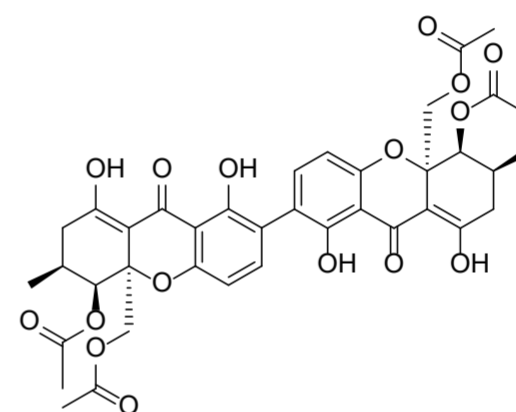
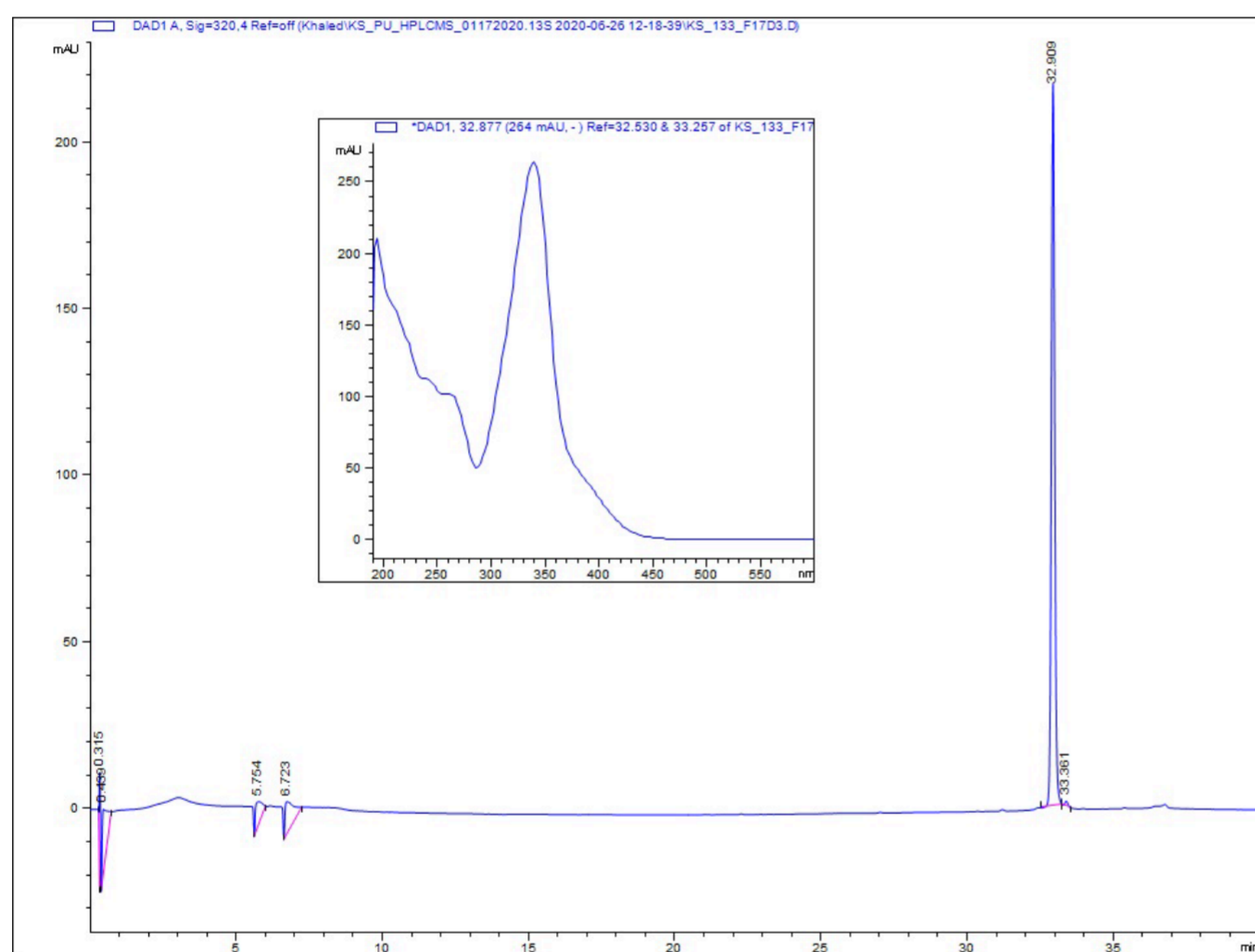
**Fig. 49S** TOCSY spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone B (**4**).

## Supplementary material



**Fig. 50S** NOESY spectrum (CDCl<sub>3</sub>, 500 MHz) of phomoxanthone B (**4**).

## Supplementary material



**Fig. 51S** HPLC analysis of dicerandrol C (**5**). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 320 nm. UV-vis inset of full wavelength scan (190-600 nm).

## Supplementary material

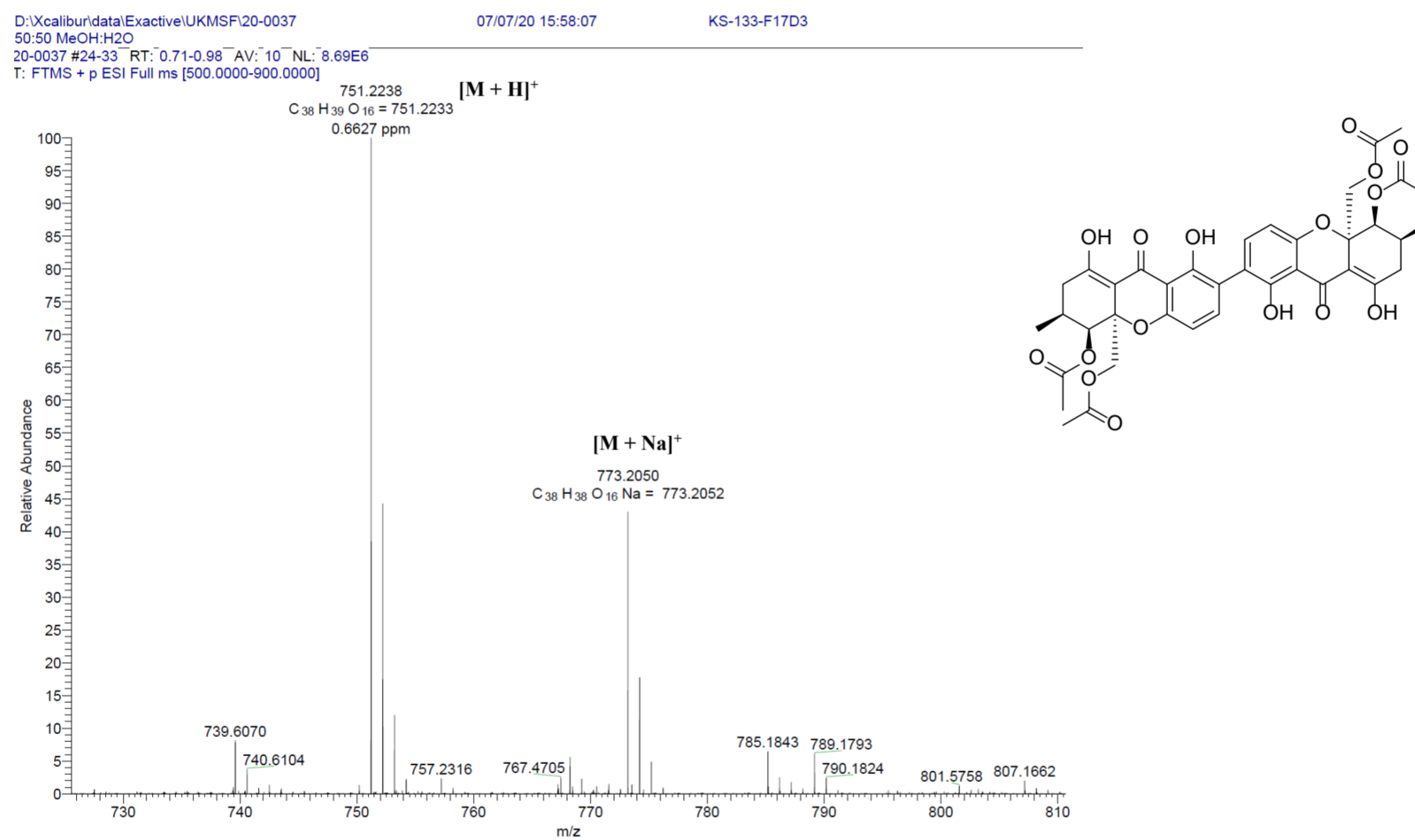


Fig. 52S (+)-HRESI-MS spectrum of dicerandrol C (5).

## Supplementary material

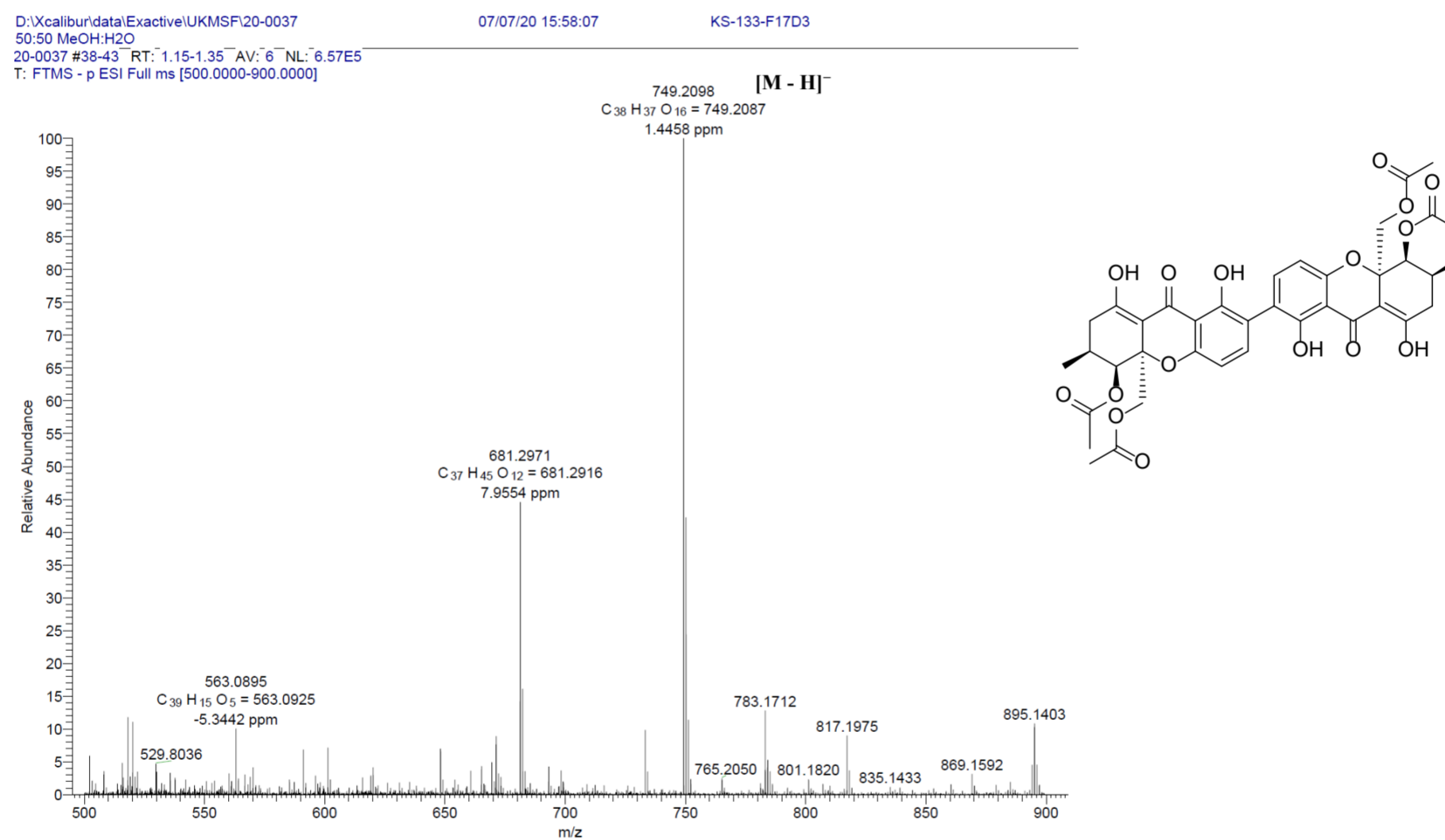
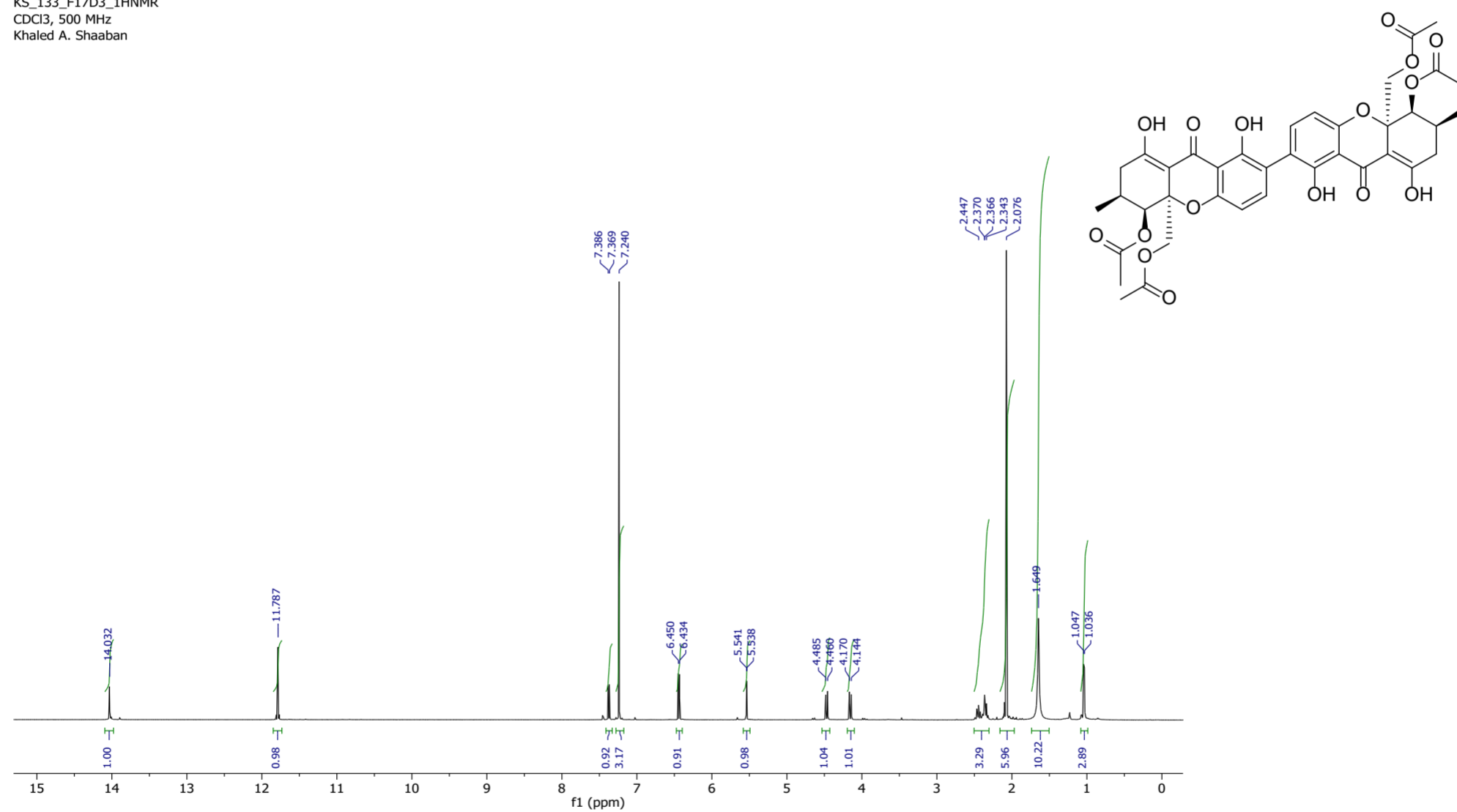


Fig. 53S (-)-HRESI-MS spectrum of dicerandrol C (5).

## Supplementary material

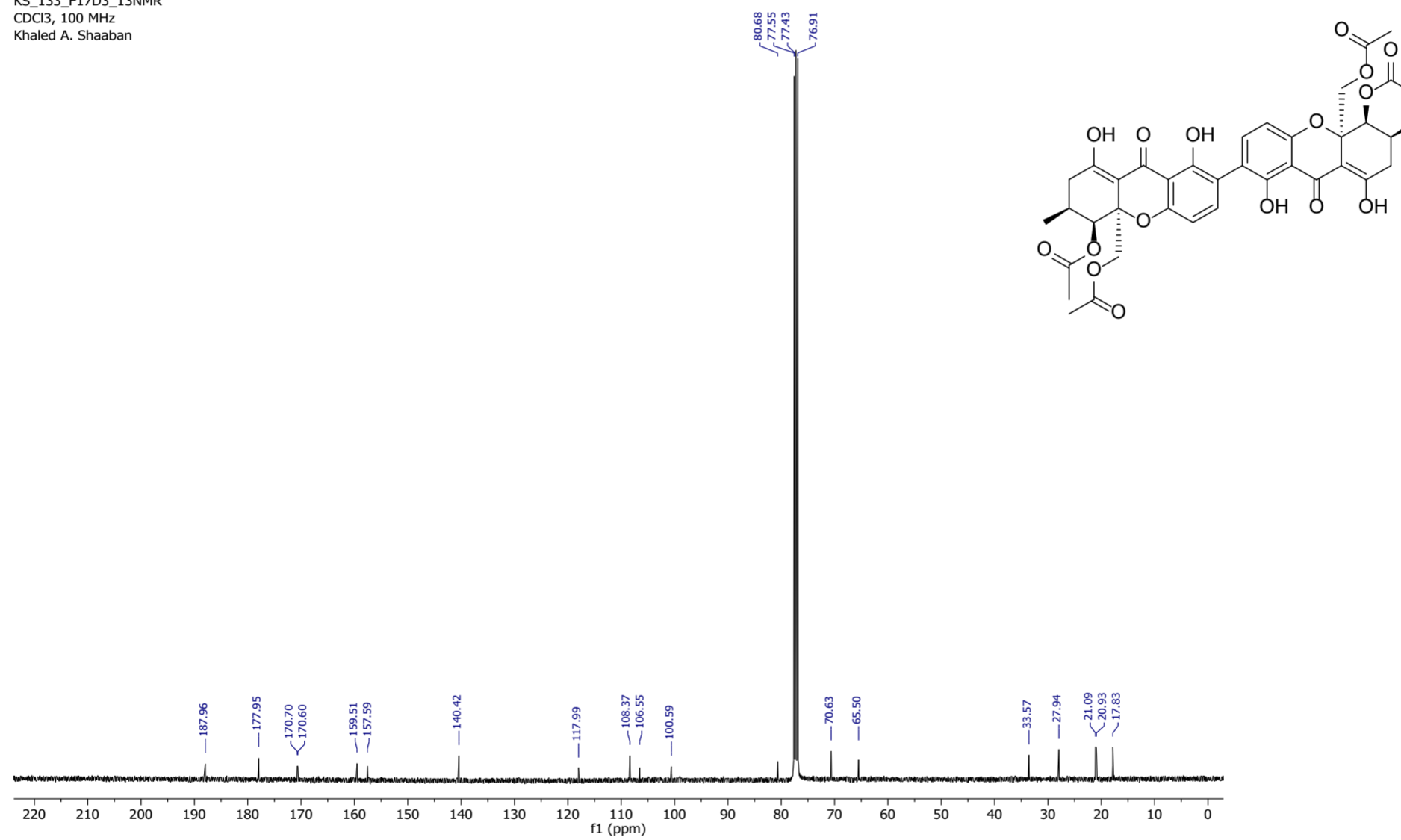
KS\_133\_F17D3\_1HNMR  
CDCl<sub>3</sub>, 500 MHz  
Khaled A. Shaaban



**Fig. 54S** <sup>1</sup>H NMR spectrum (CDCl<sub>3</sub>, 500 MHz) of dicerandrol C (5).

## Supplementary material

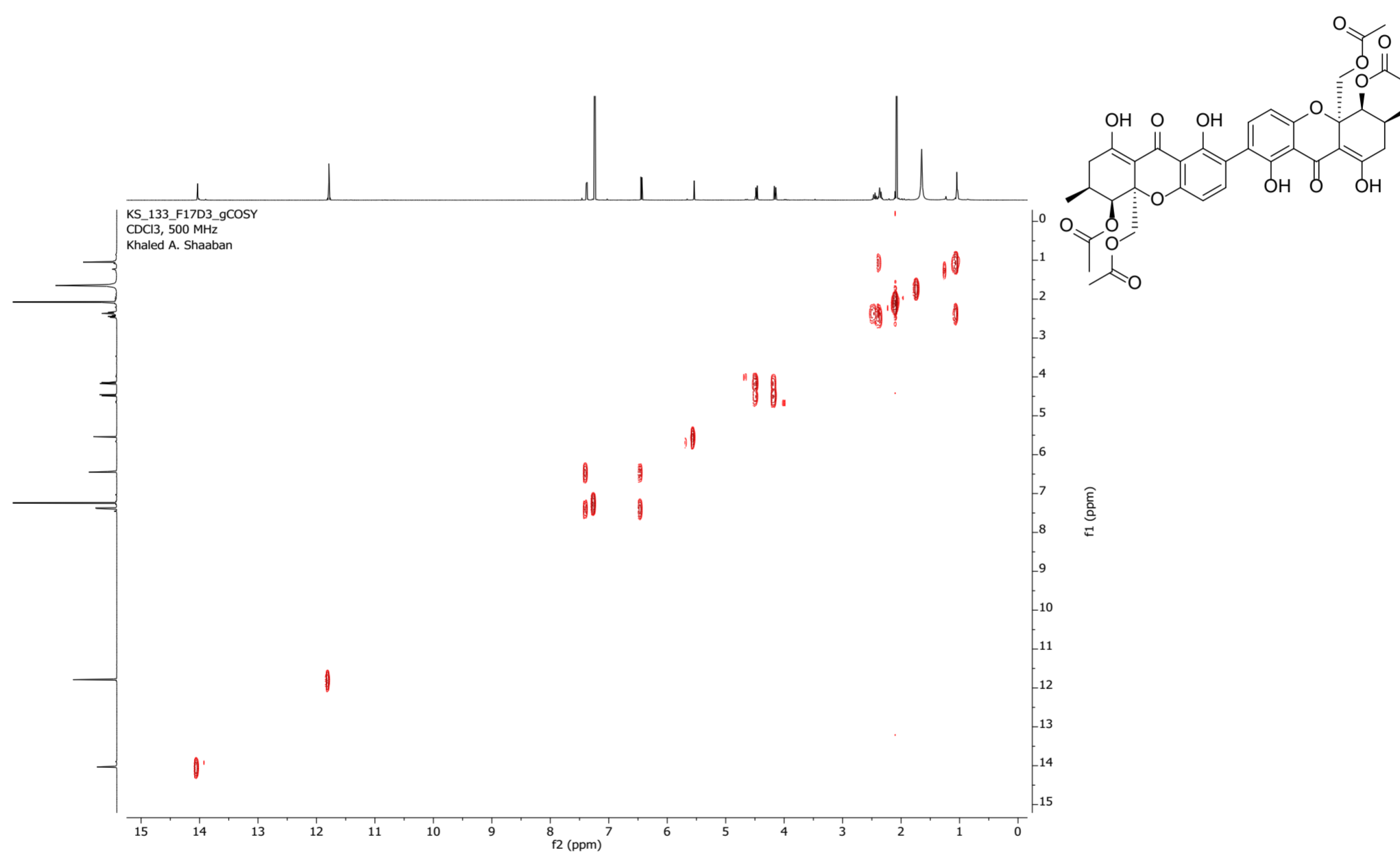
KS\_133\_F17D3\_13NMR  
CDCl<sub>3</sub>, 100 MHz  
Khaled A. Shaaban



**Fig. 55S** <sup>13</sup>C NMR spectrum (CDCl<sub>3</sub>, 100 MHz) of dicerandrol C (5).



## Supplementary material



**Fig. 56S** <sup>1</sup>H,<sup>1</sup>H-COSY spectrum (CDCl<sub>3</sub>, 500 MHz) of dicerandrol C (**5**).

## Supplementary material

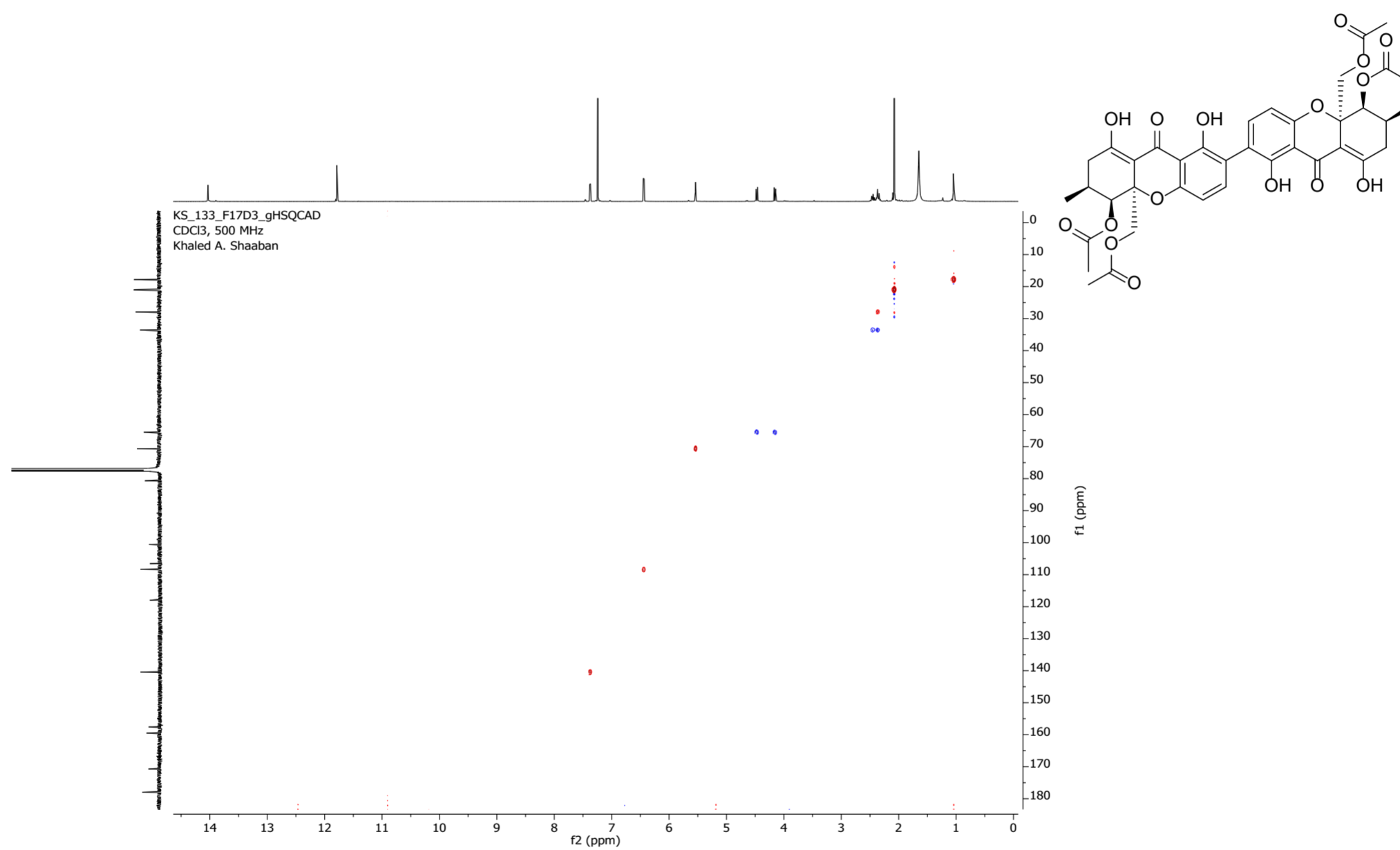


Fig. 57S HSQC spectrum (CDCl<sub>3</sub>, 500 MHz) of dicerandrol C (5).

## Supplementary material

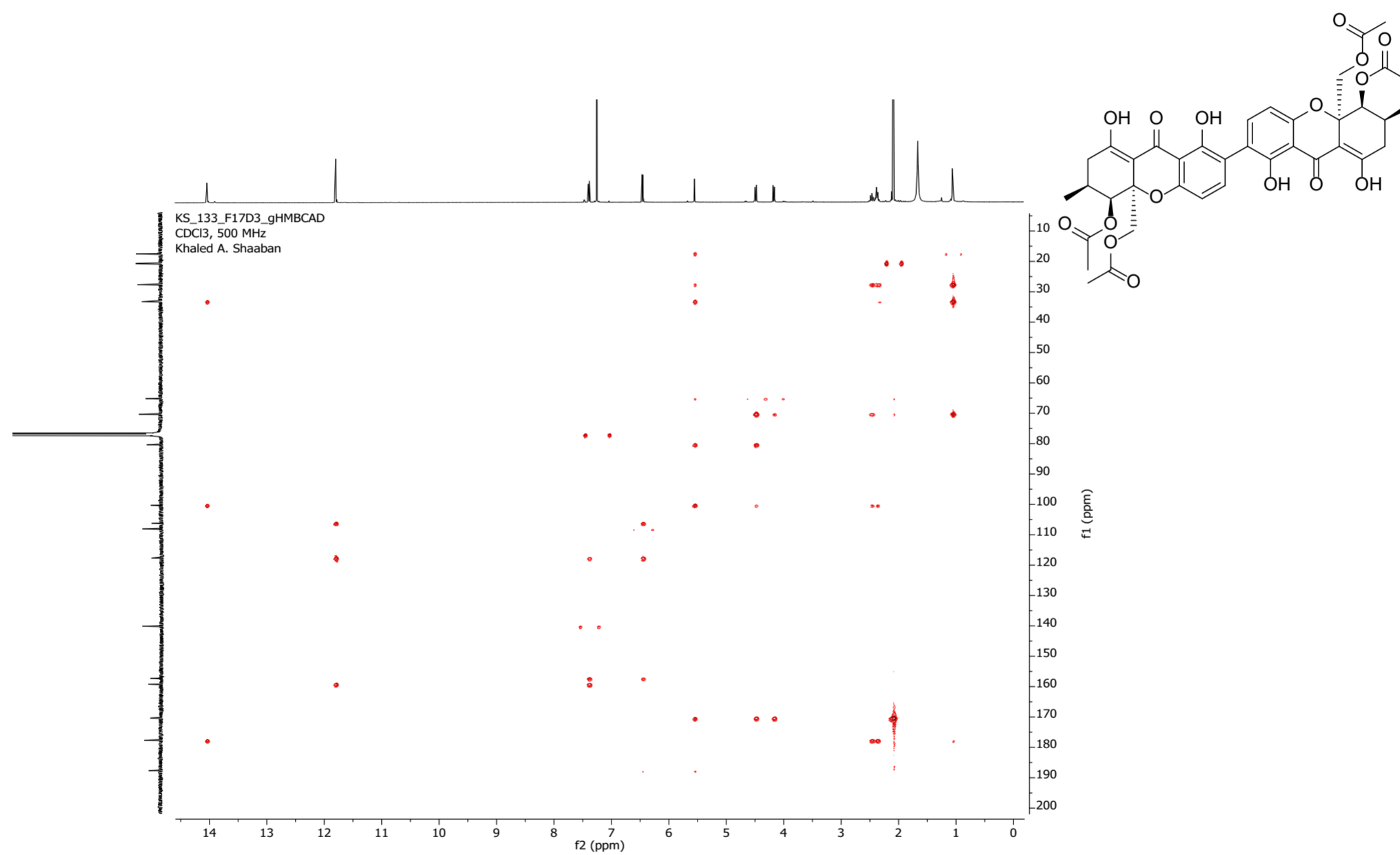
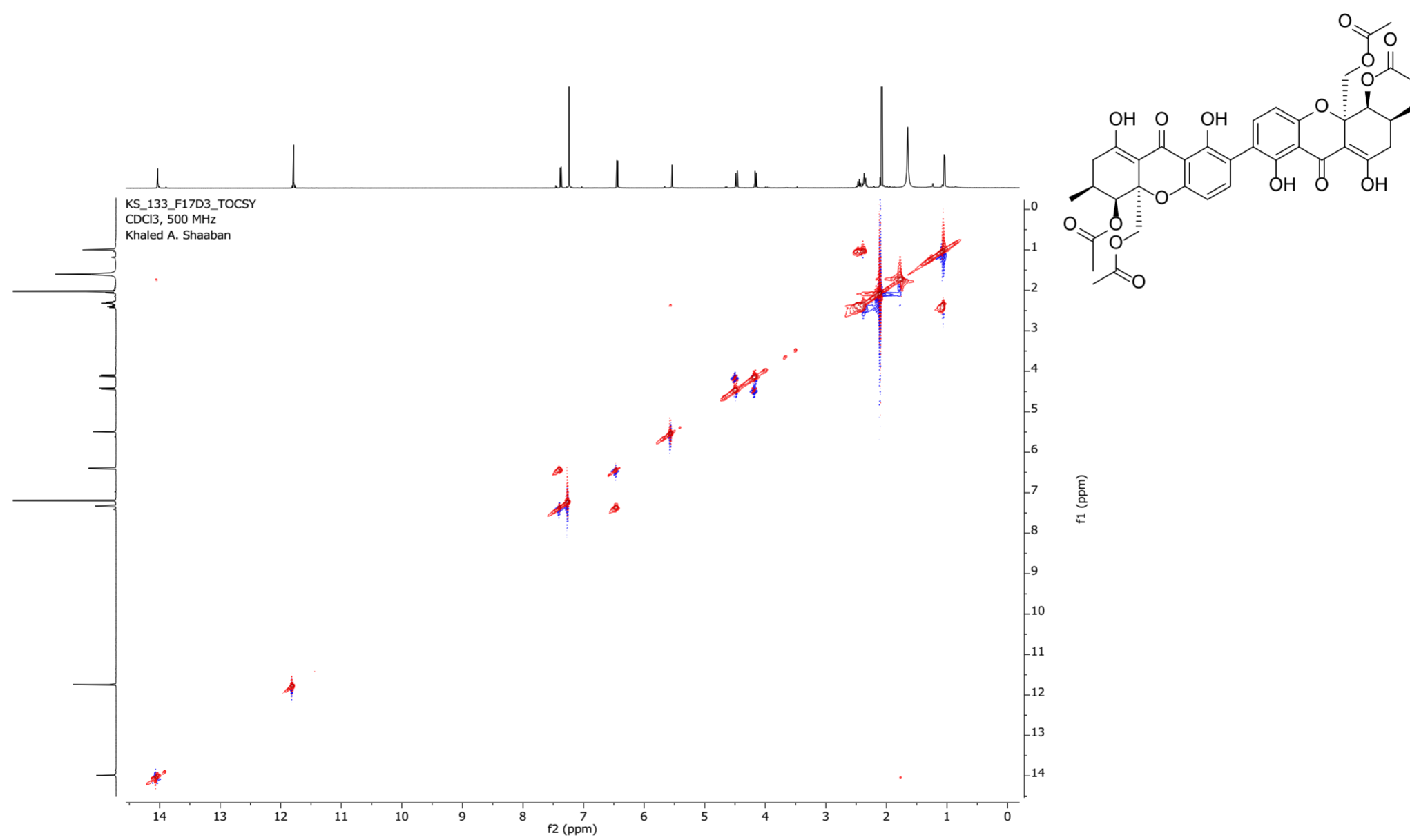


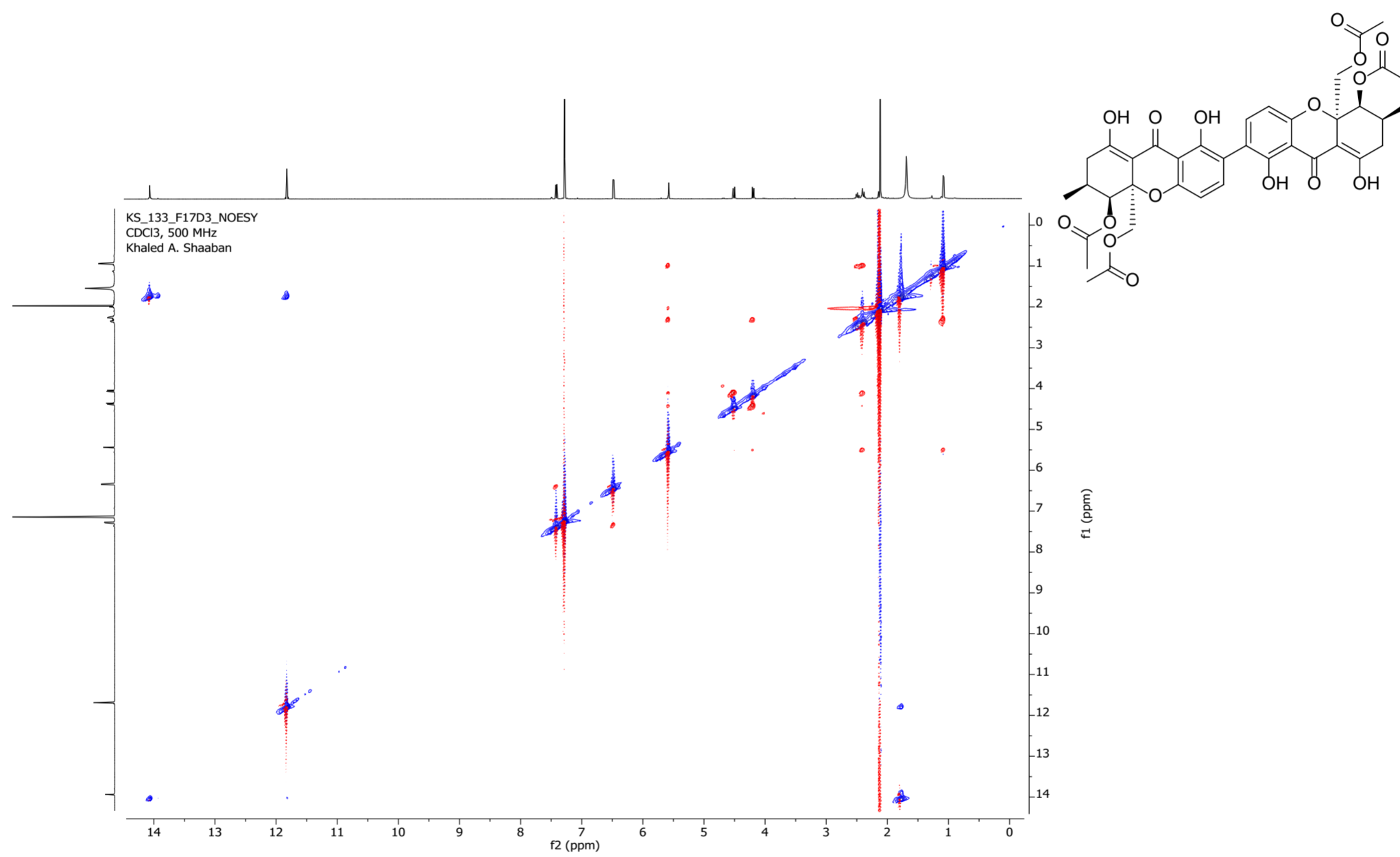
Fig. 58S HMBC spectrum (CDCl<sub>3</sub>, 500 MHz) of dicerandrol C (5).

## Supplementary material



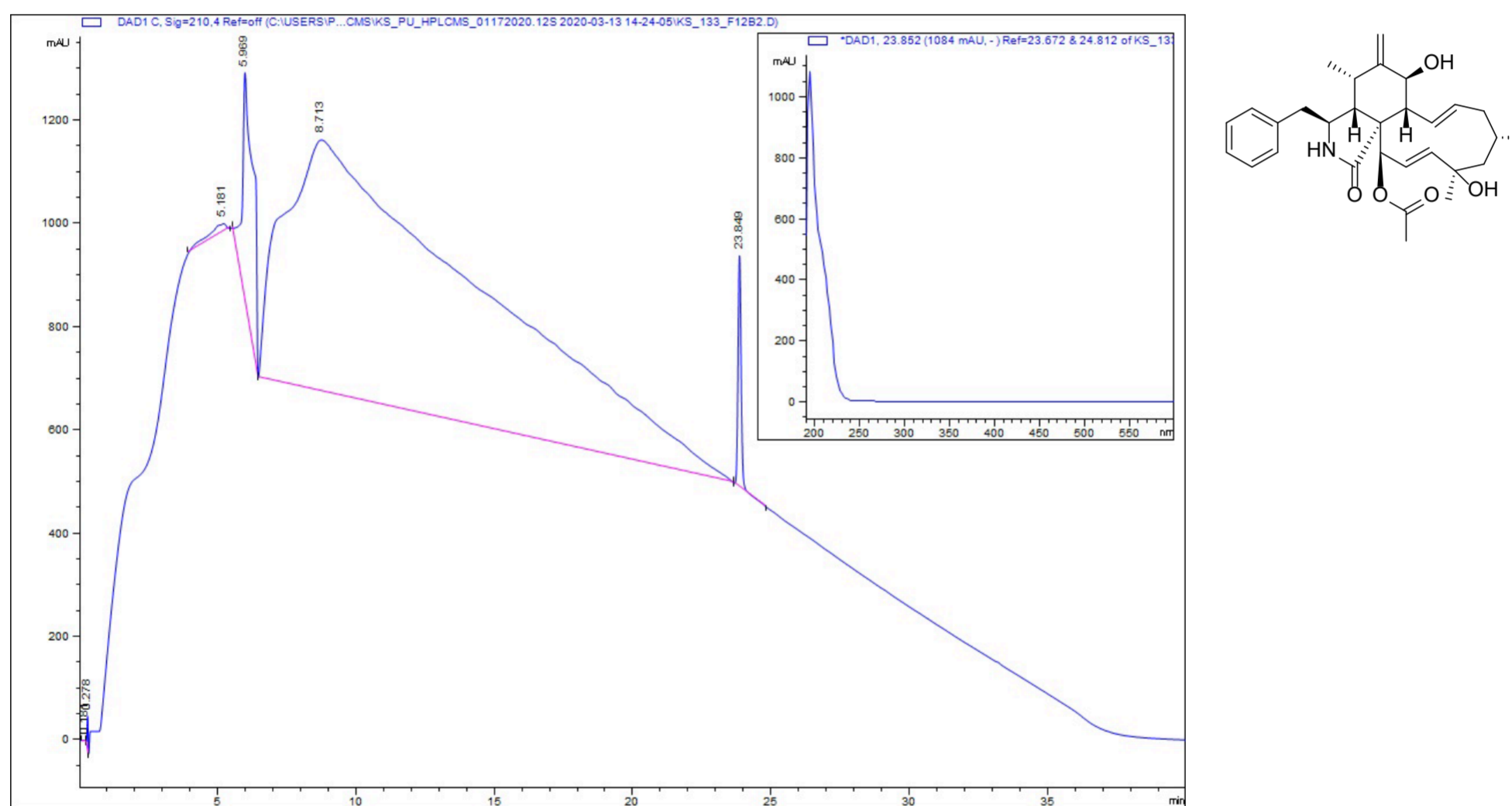
**Fig. 59S** TOCSY spectrum (CDCl<sub>3</sub>, 500 MHz) of dicerandrol C (**5**).

## Supplementary material



**Fig. 60S** NOESY spectrum (CDCl<sub>3</sub>, 500 MHz) of dicerandrol C (**5**).

## Supplementary material



**Fig. 61S** HPLC analysis of cytochalasin H (**6**). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 210 nm. UV-vis inset of full wavelength scan (190-600 nm).

## Supplementary material

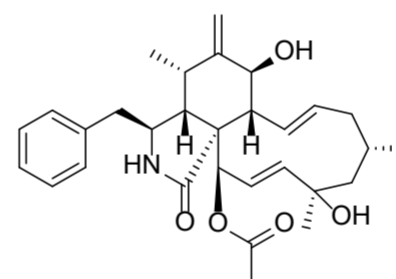
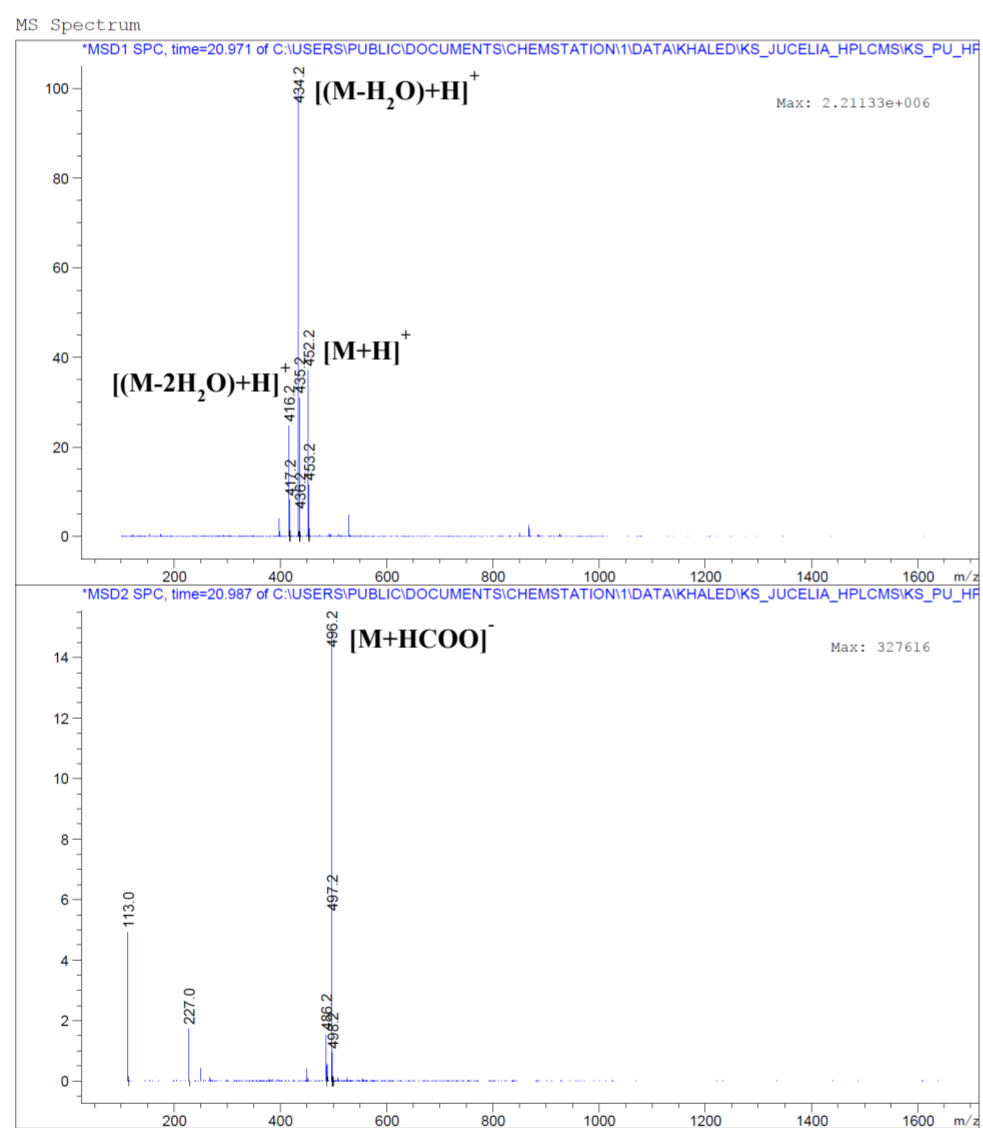
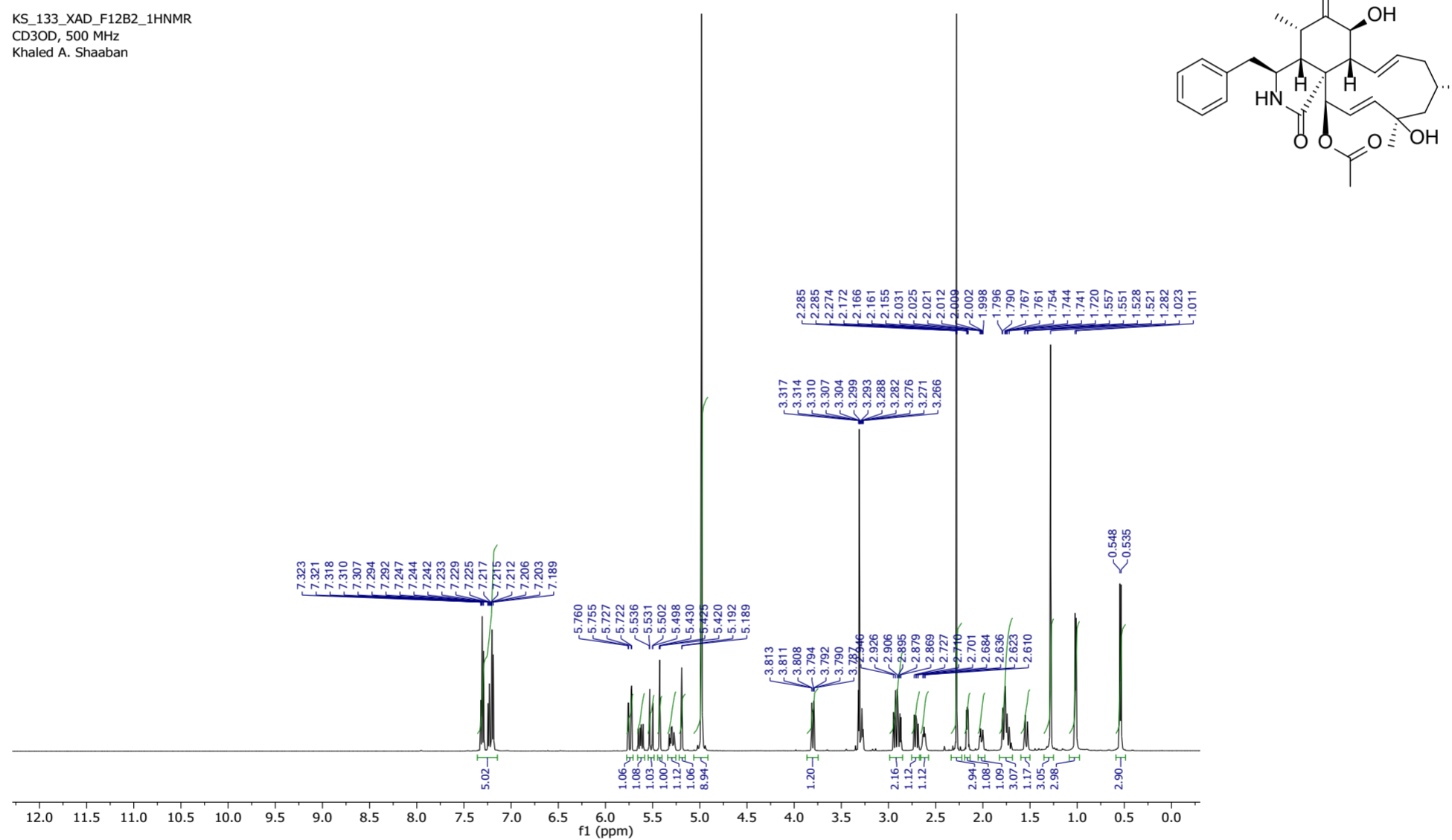


Fig. 62S (+) and (-)-ESI-MS spectrum of cytochalasin H (6).

## Supplementary material

KS\_133\_XAD\_F12B2\_1HNMR  
 CD3OD, 500 MHz  
 Khaled A. Shaaban

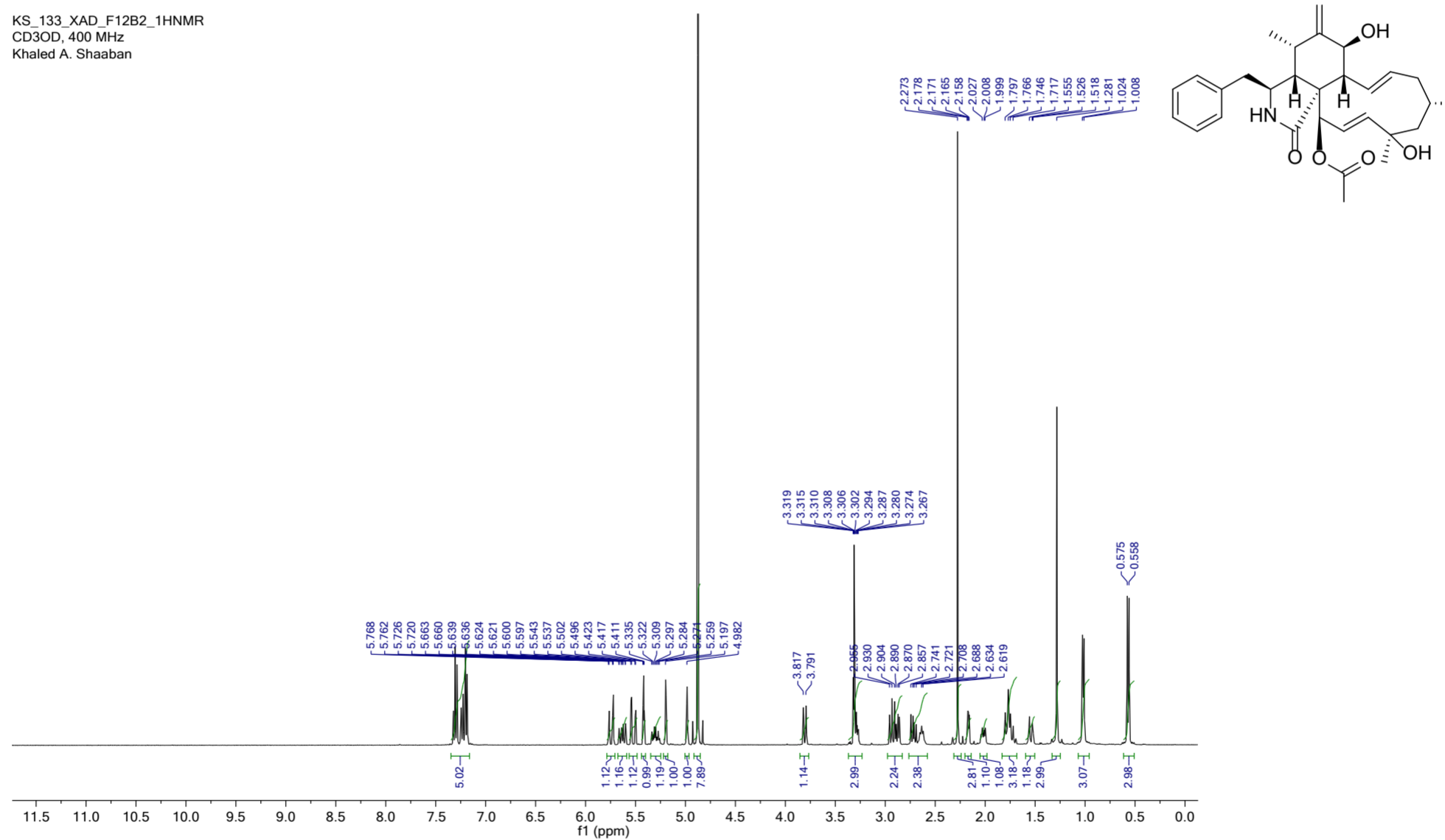


**Fig. 63S**  $^1\text{H}$  NMR spectrum ( $\text{CD}_3\text{OD}$ , 500 MHz) of cytochalasin H (6).



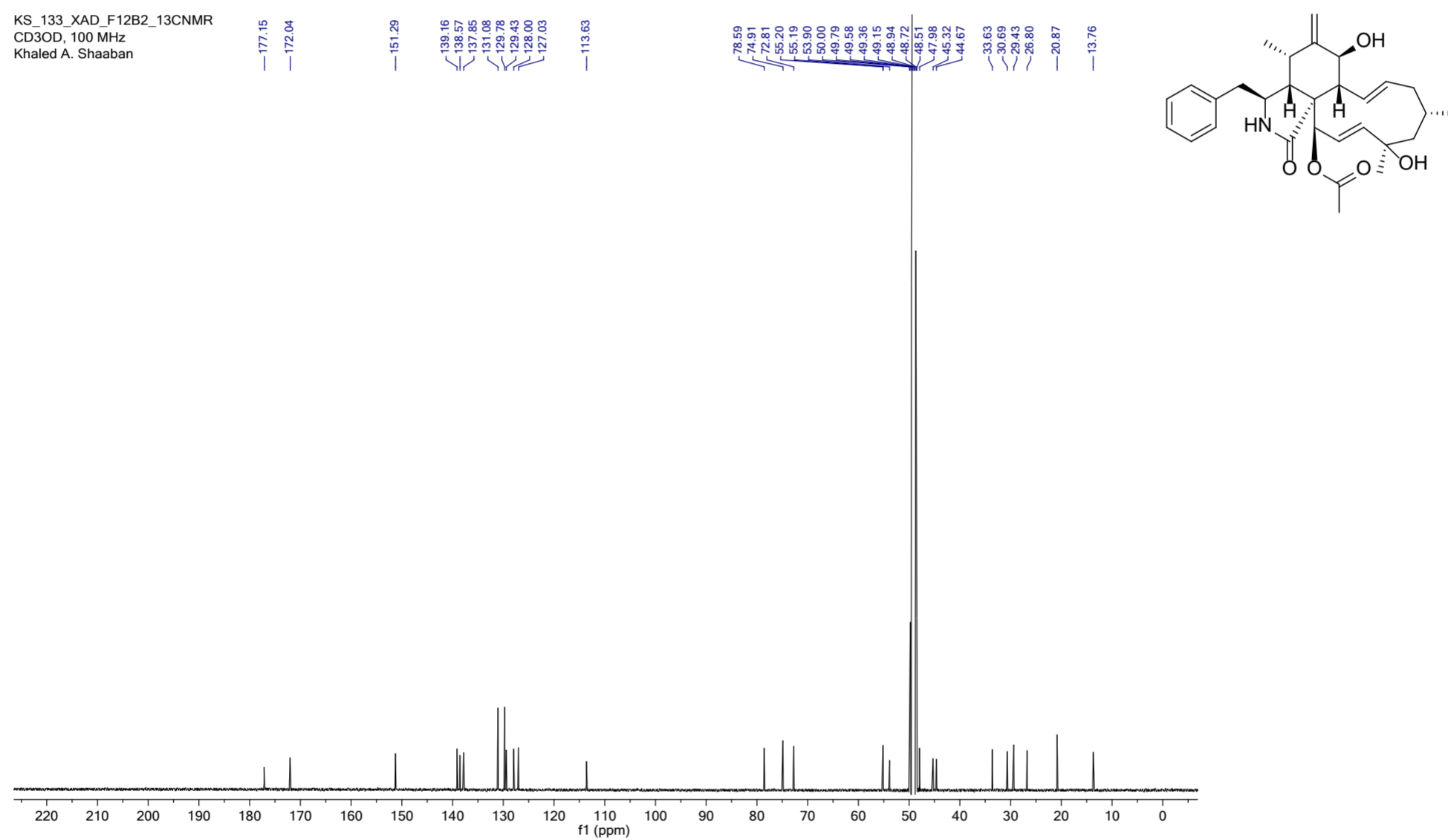
## Supplementary material

KS\_133\_XAD\_F12B2\_1HNMR  
 CD3OD, 400 MHz  
 Khaled A. Shaaban



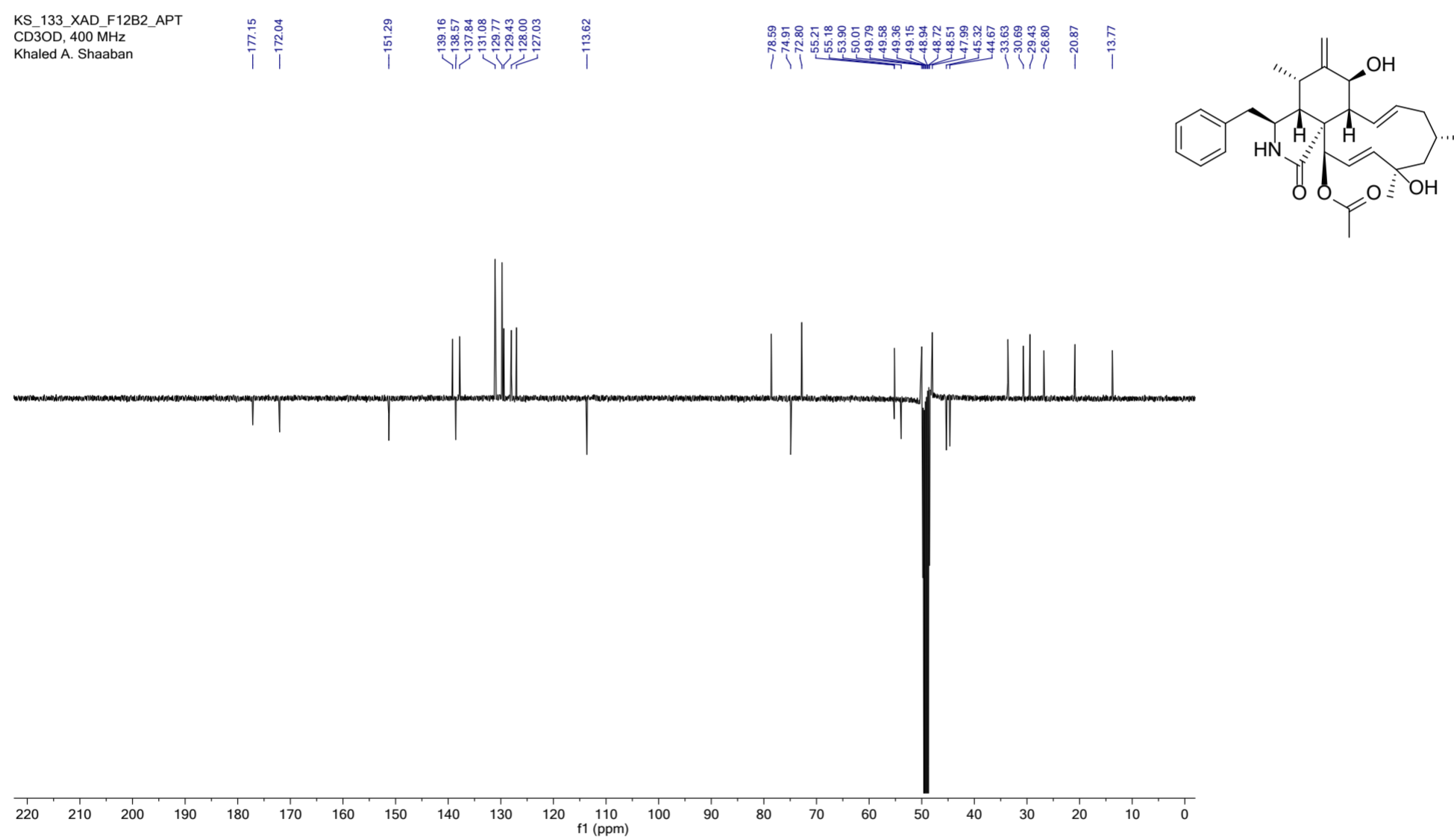
**Fig. 64S** <sup>1</sup>H NMR spectrum (CD<sub>3</sub>OD, 400 MHz) of cytochalasin H (**6**).

## Supplementary material



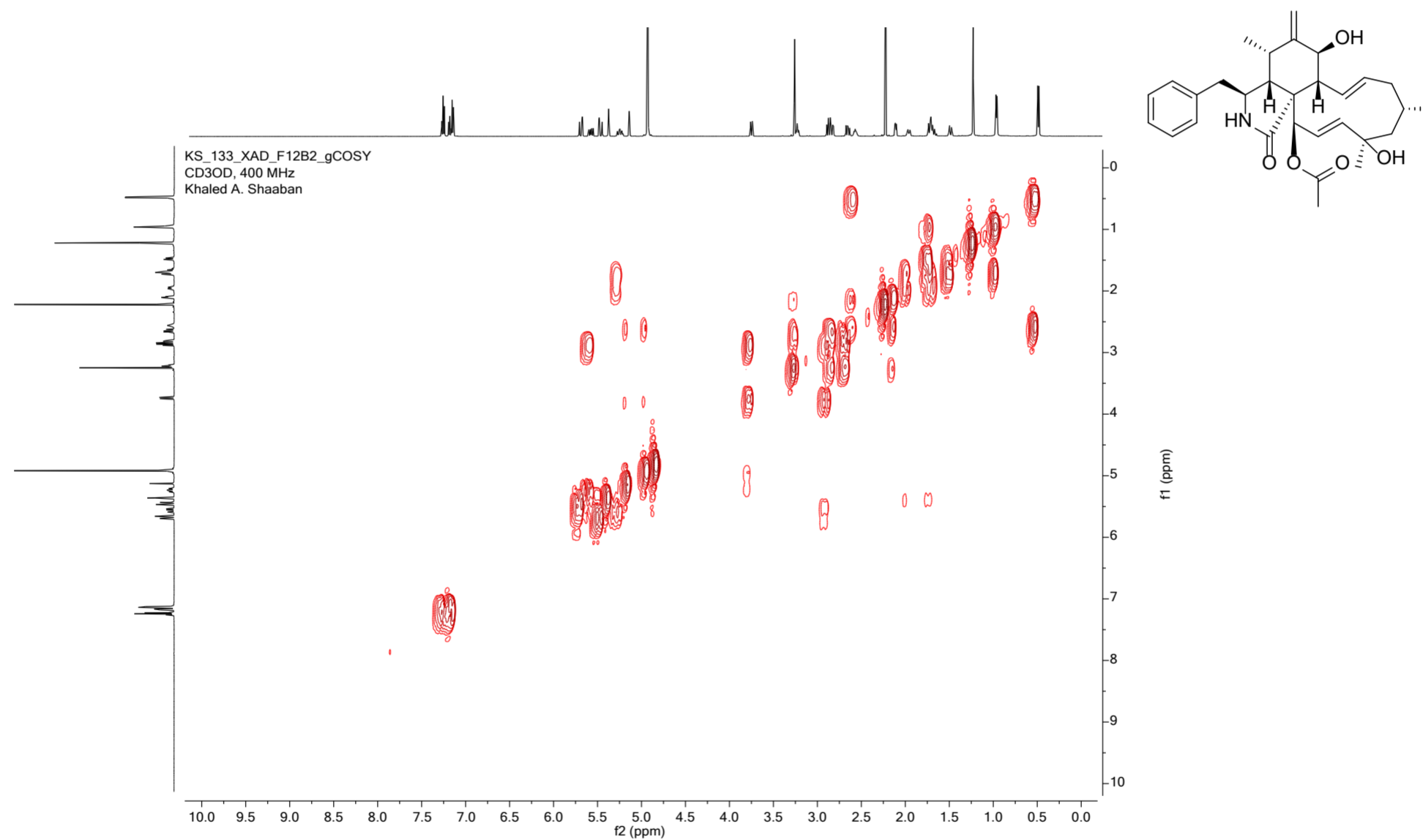
**Fig. 65S**  $^{13}\text{C}$  NMR spectrum ( $\text{CD}_3\text{OD}$ , 100 MHz) of cytochalasin H (**6**).

## Supplementary material



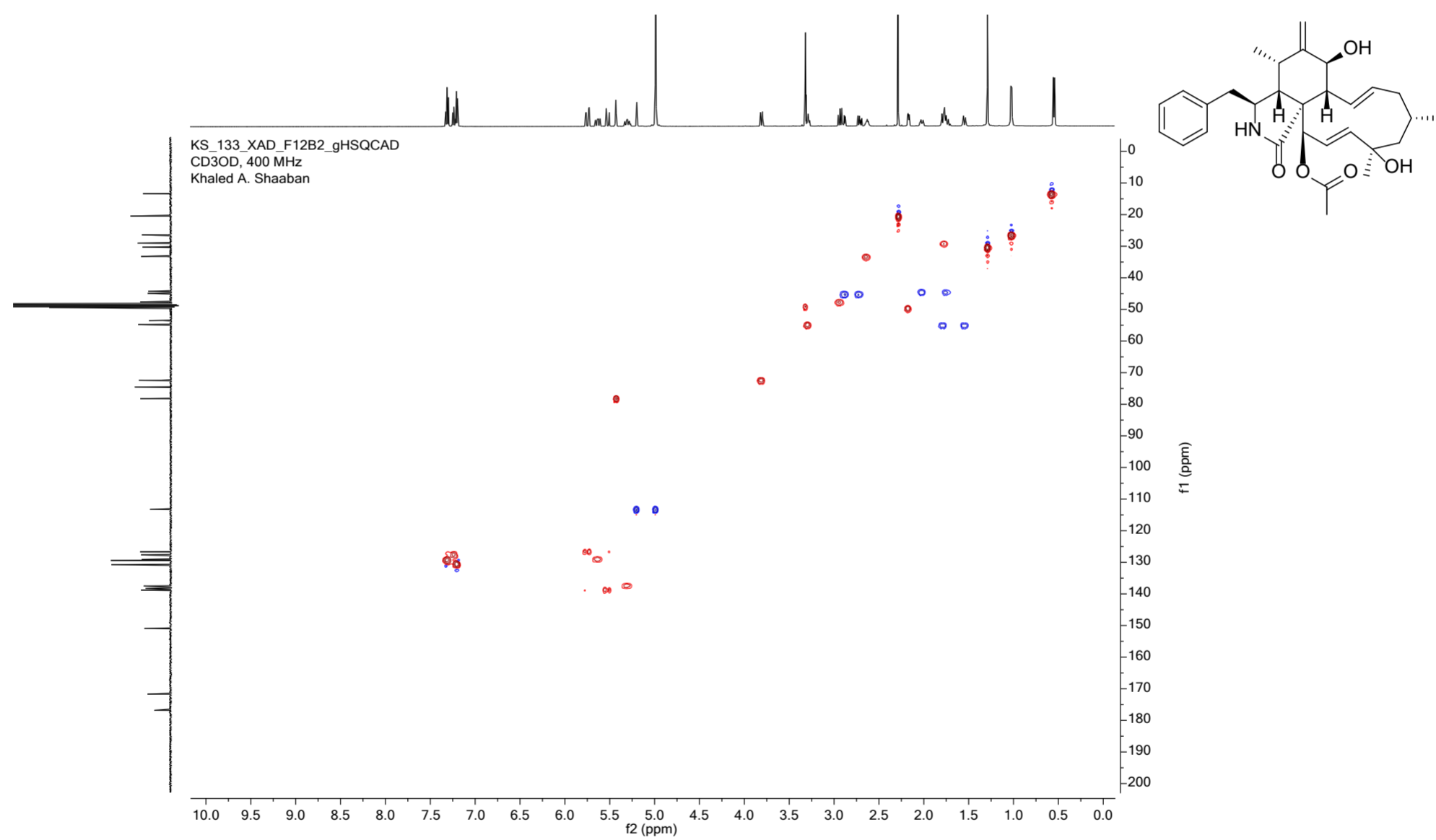
**Fig. 66S** APT NMR spectrum (CD<sub>3</sub>OD, 100 MHz) of cytochalasin H (**6**).

## Supplementary material



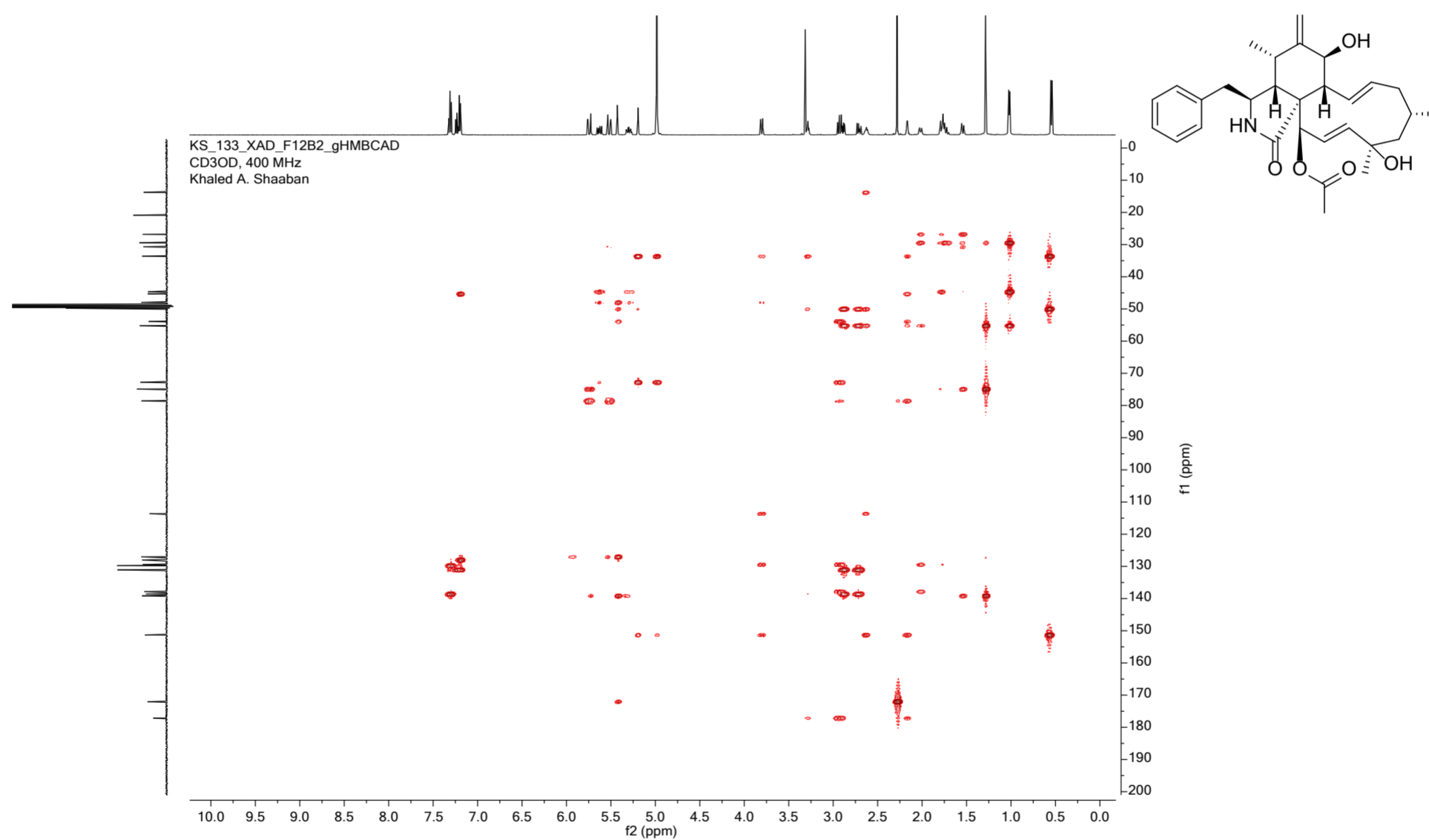
**Fig. 67S**  $^1\text{H}, ^1\text{H}$ -COSY spectrum ( $\text{CD}_3\text{OD}$ , 500 MHz) of cytochalasin H (6).

## Supplementary material



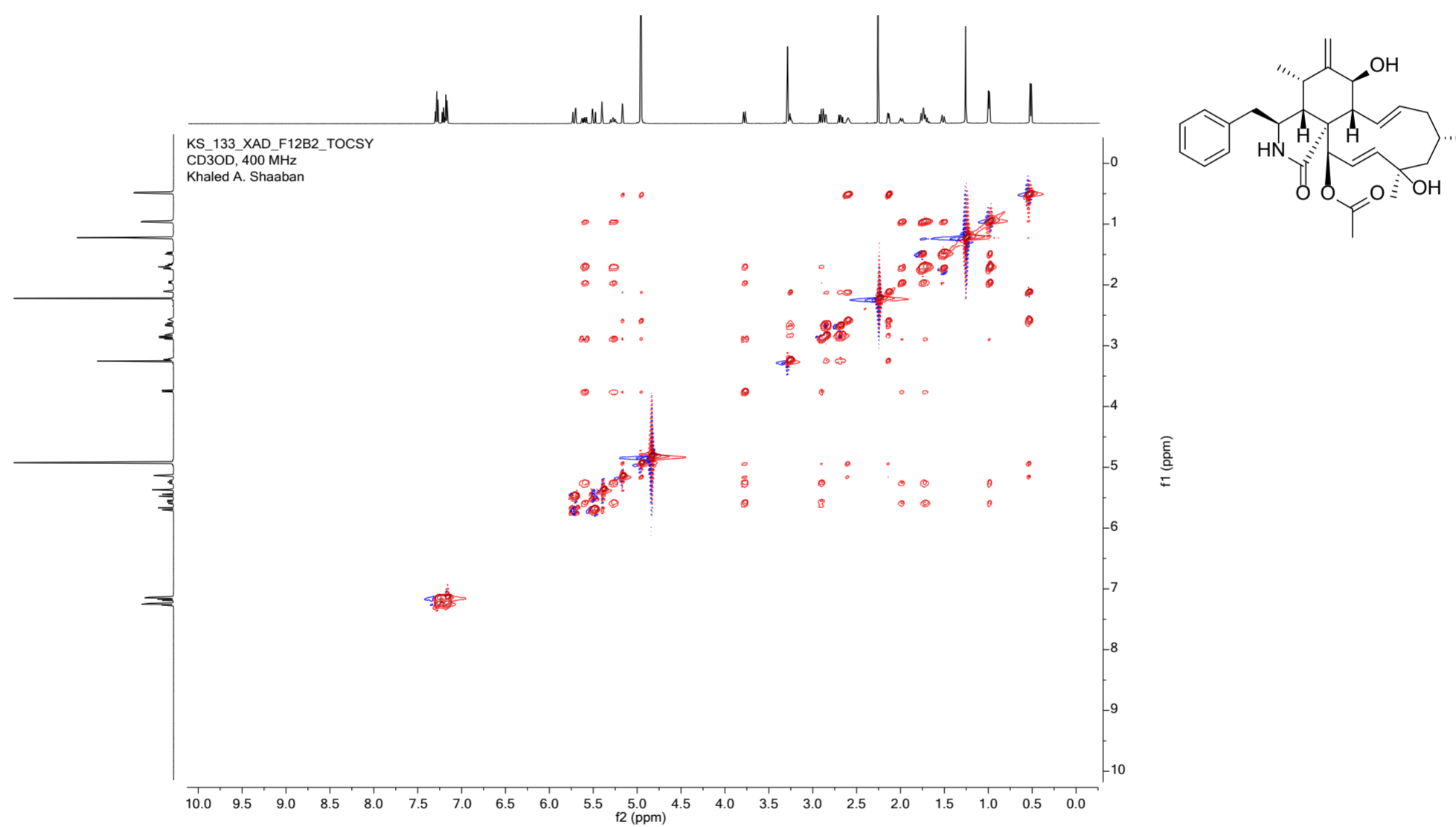
**Fig. 68S** HSQC spectrum (CD<sub>3</sub>OD, 500 MHz) of cytochalasin H (**6**).

## Supplementary material



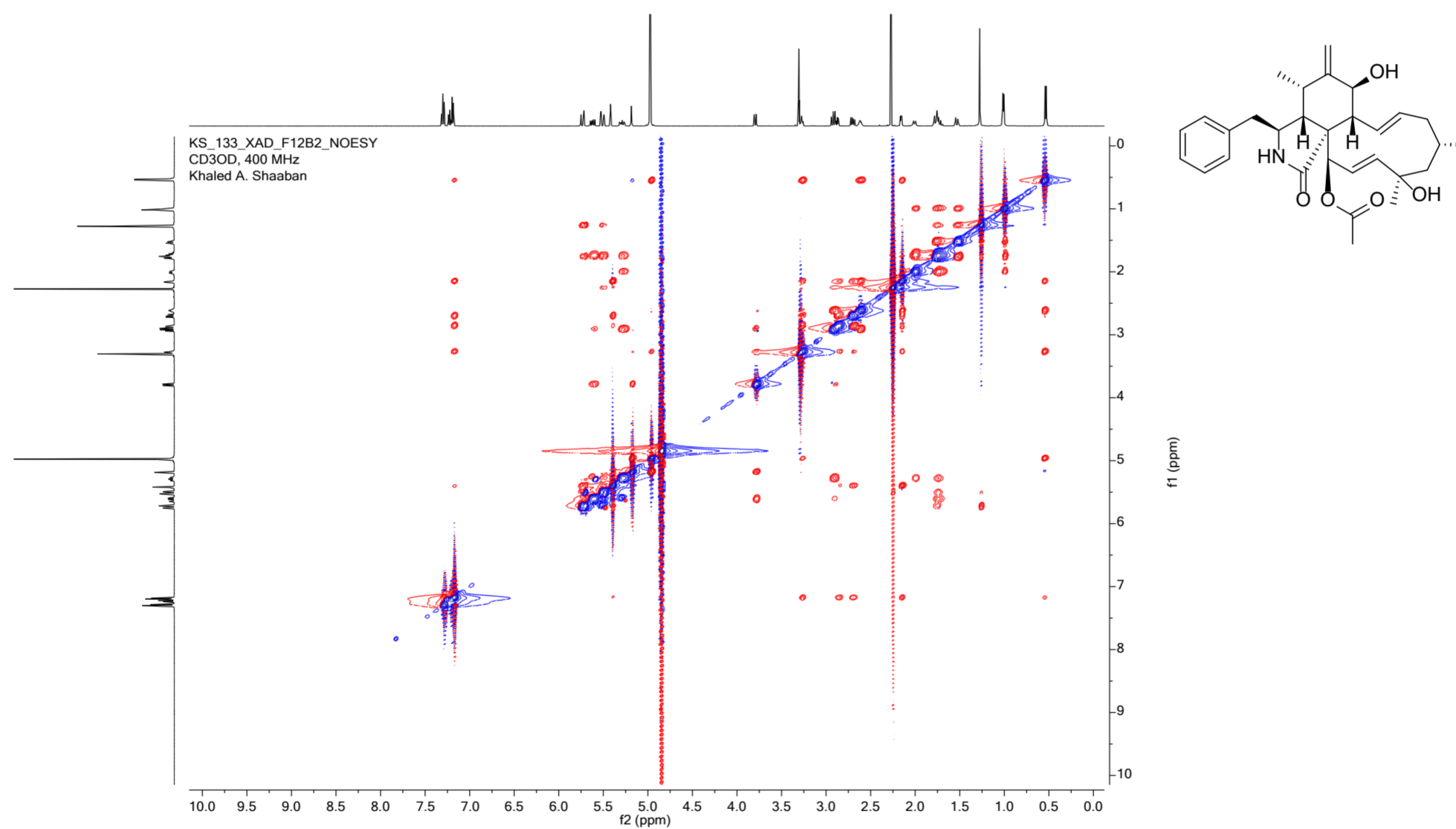
**Fig. 69S** HMBC spectrum (CD<sub>3</sub>OD, 500 MHz) of cytochalasin H (**6**).

## Supplementary material



**Fig. 70S** TOCSY spectrum (CD<sub>3</sub>OD, 500 MHz) of cytochalasin H (**6**).

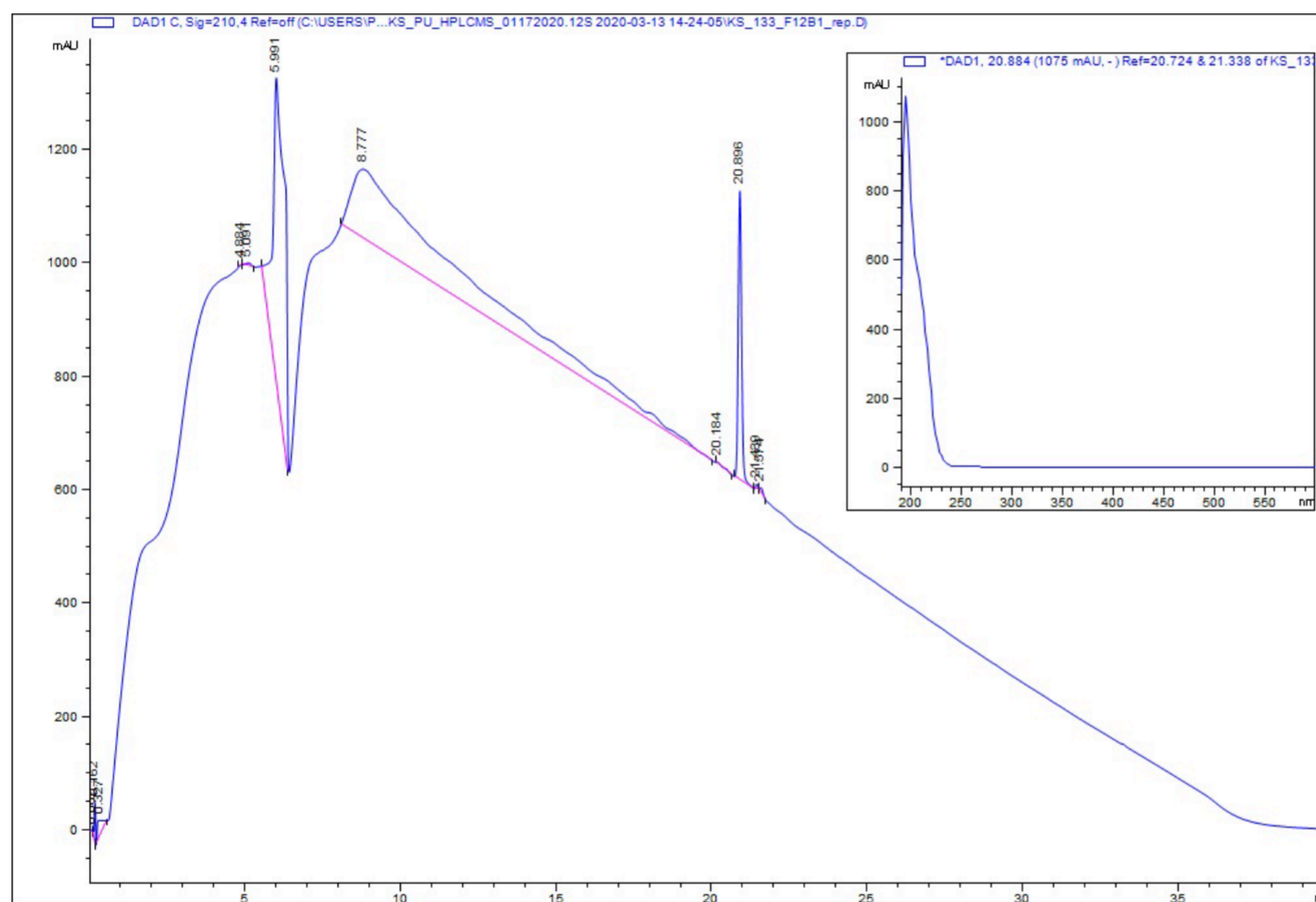
## Supplementary material



**Fig. 71S** NOESY spectrum (CD<sub>3</sub>OD, 500 MHz) of cytochalasin H (**6**).



## Supplementary material



**Fig. 72S** HPLC analysis of cytochalasin J (7). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 210 nm. UV-vis inset of full wavelength scan (190-600 nm).

## Supplementary material

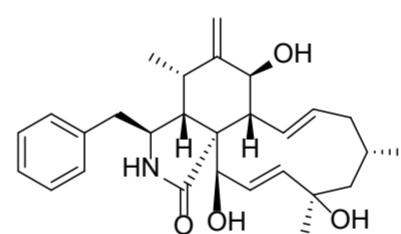
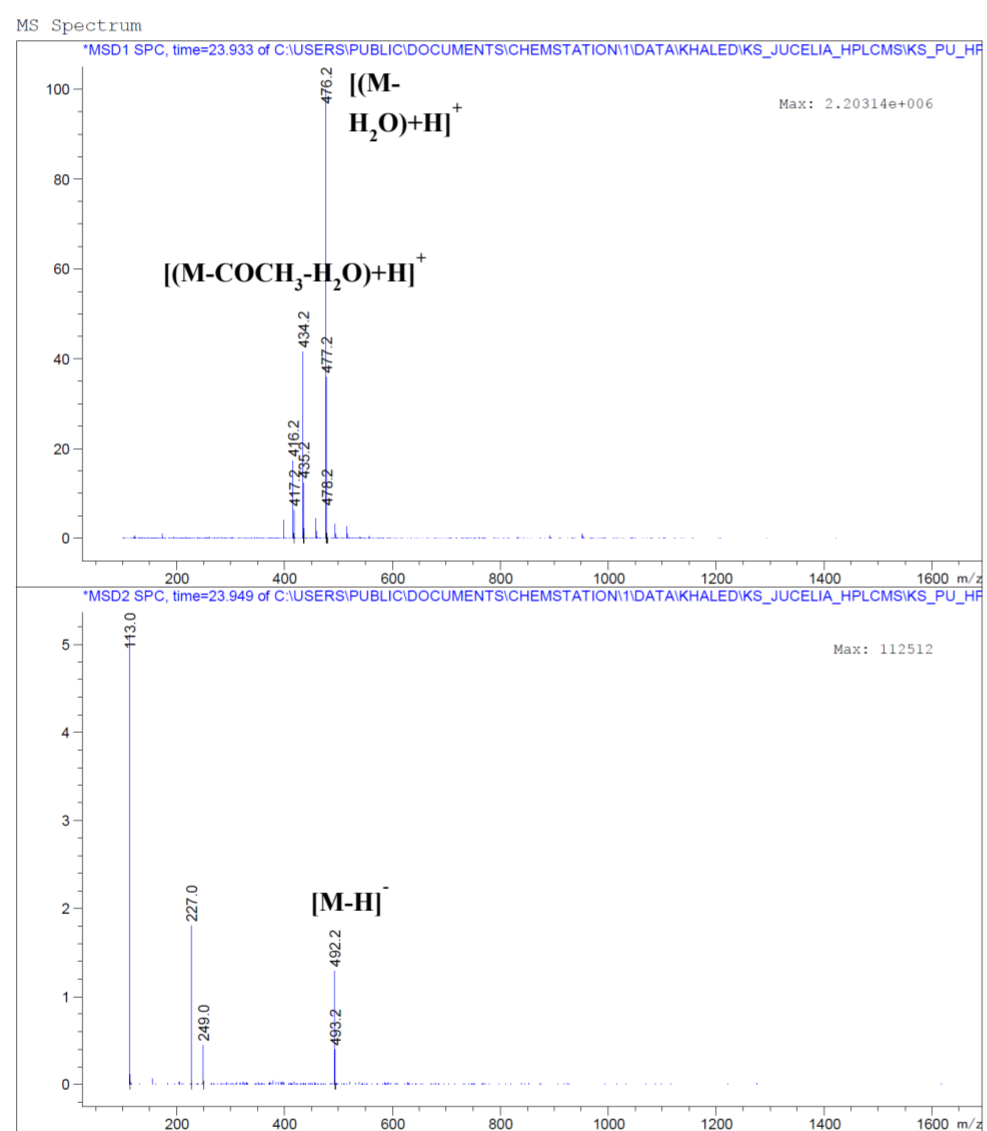


Fig. 73S (+) and (-)-ESI-MS spectrum of cytochalasin J (7).

## Supplementary material

KS\_133\_XAD\_F12B1\_1HNMR  
CD3OD, 500 MHz  
Khaled A. Shaaban

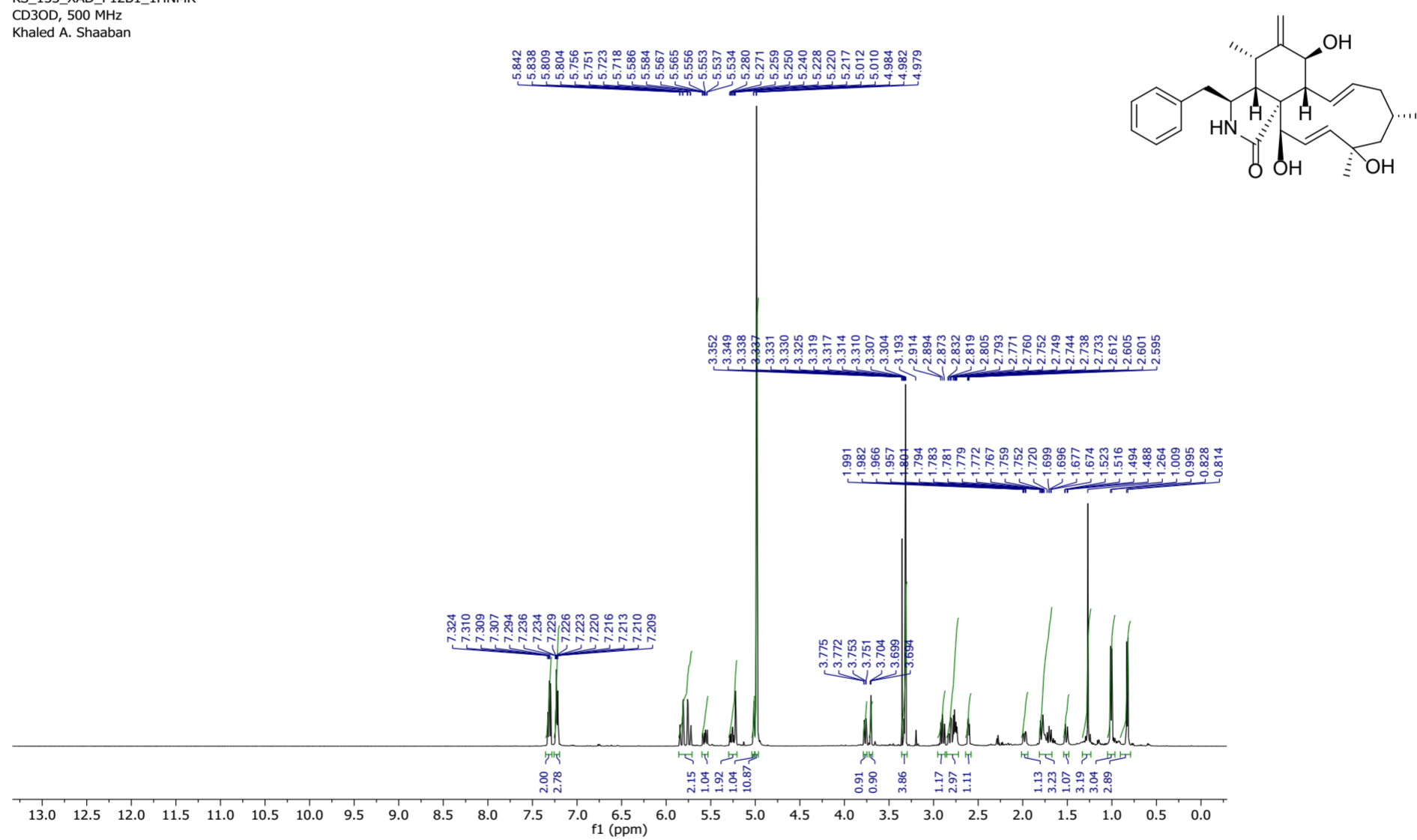
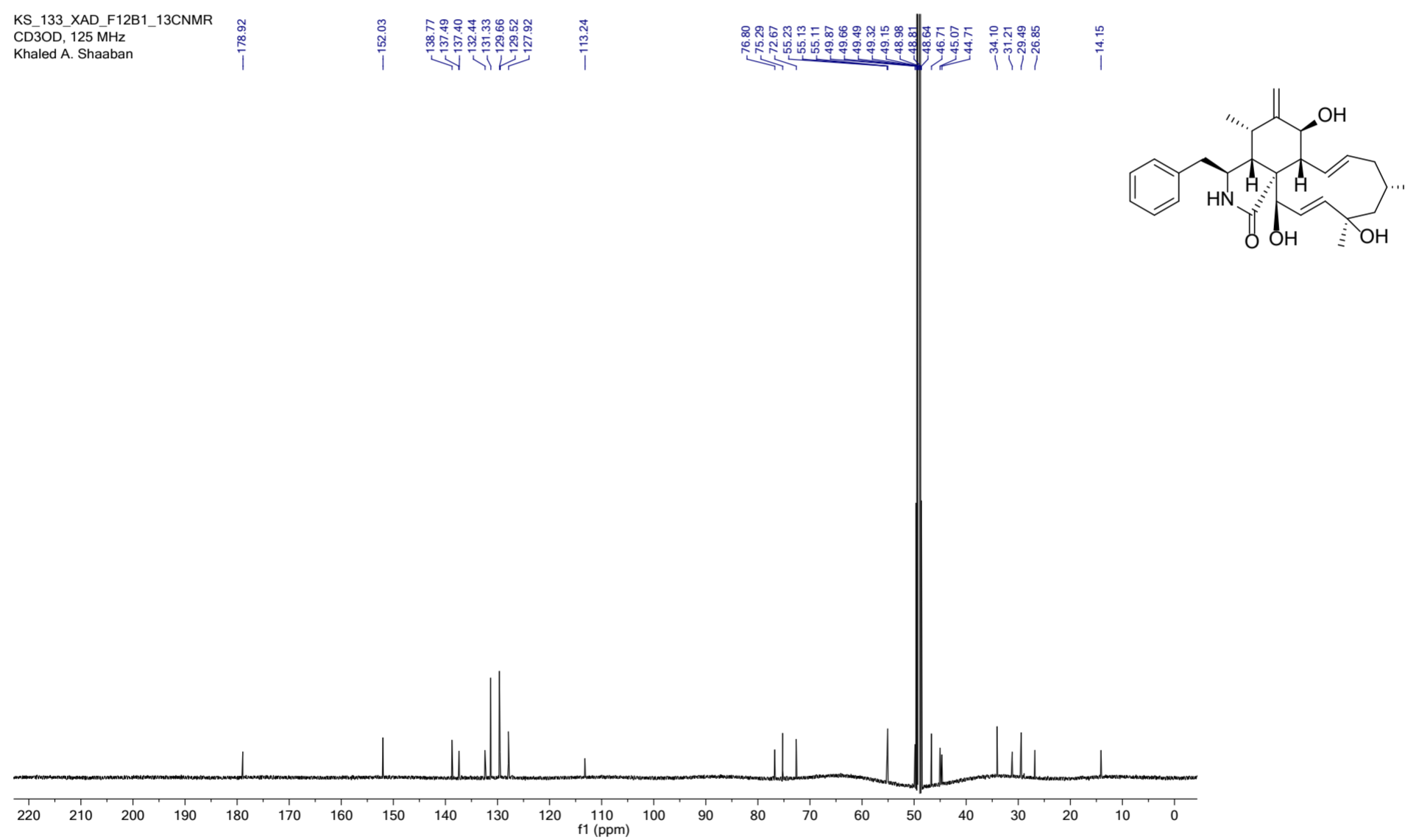


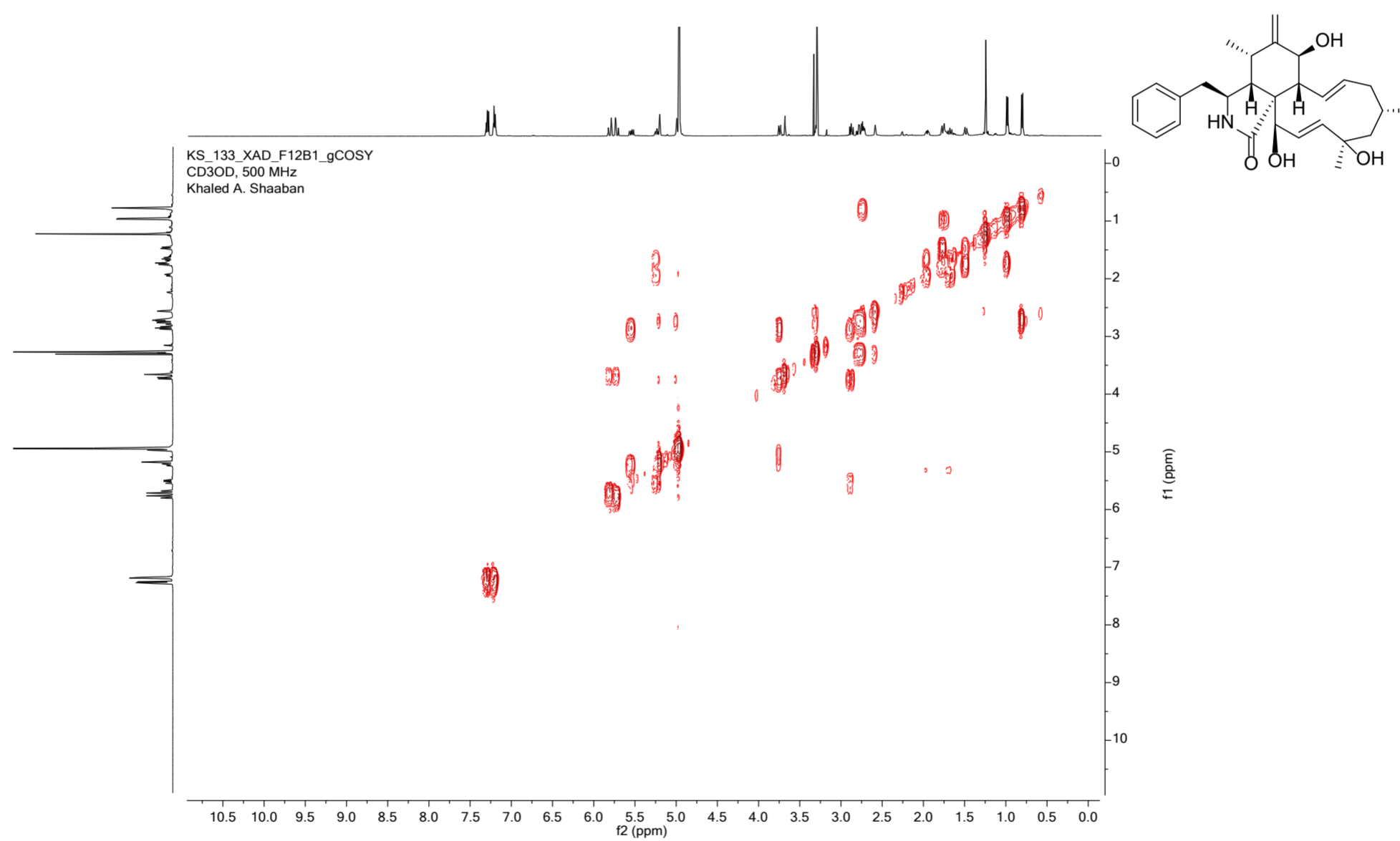
Fig. 74S  $^1\text{H}$  NMR spectrum (CD<sub>3</sub>OD, 500 MHz) of cytochalasin J (7).

## Supplementary material



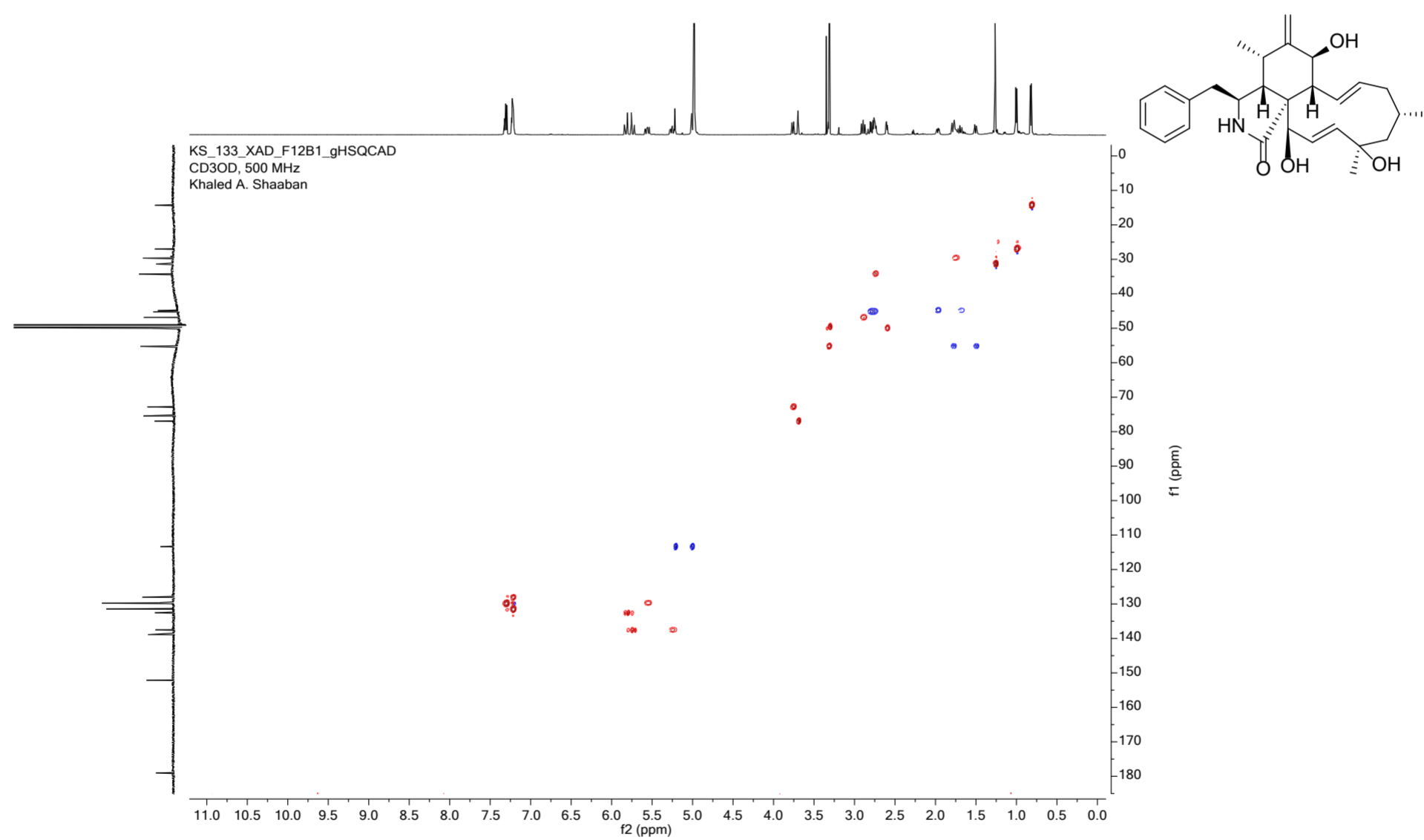
**Fig. 75S**  $^{13}\text{C}$  NMR spectrum ( $\text{CD}_3\text{OD}$ , 100 MHz) of cytochalasin J (**7**).

## Supplementary material



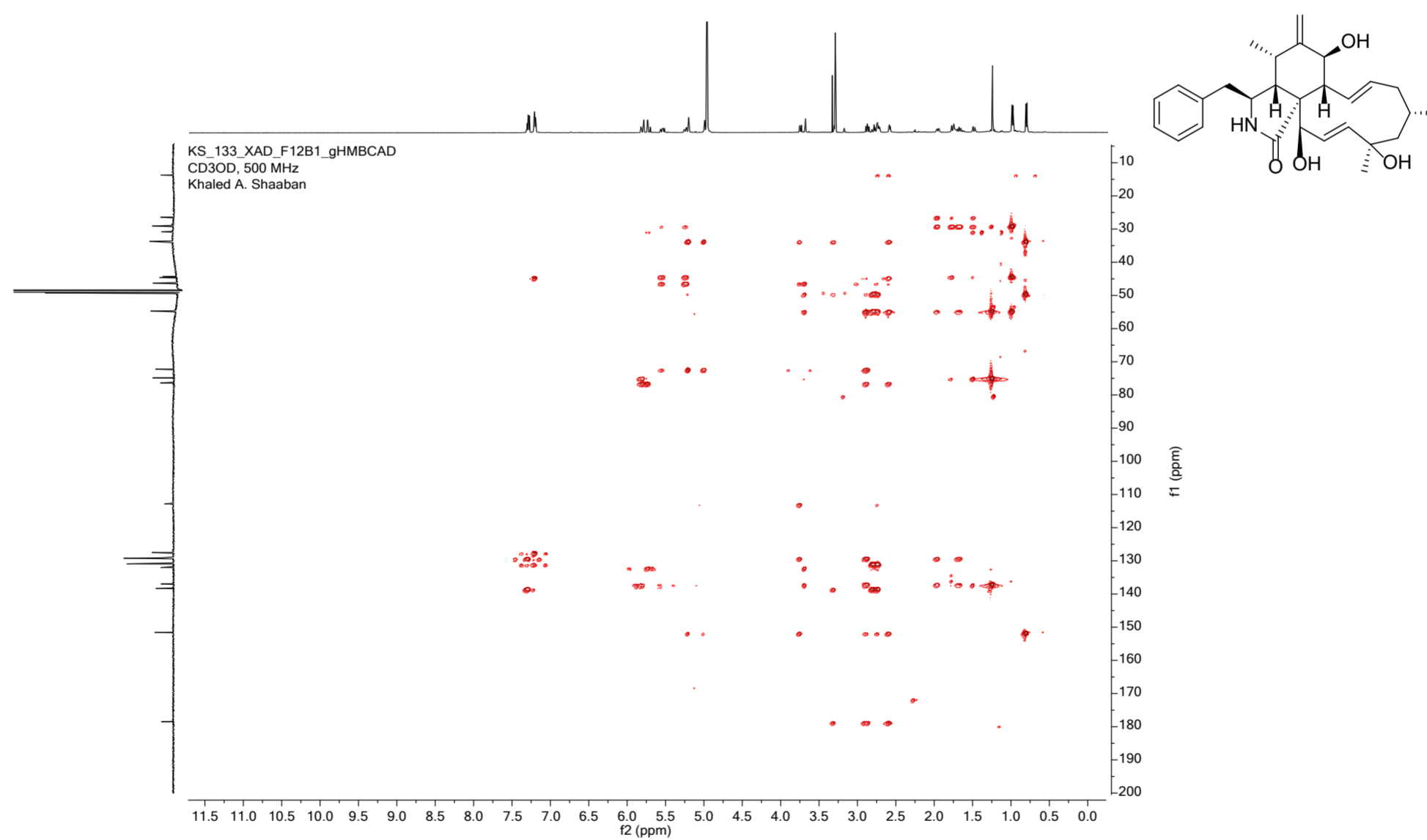
**Fig. 76S**  $^1\text{H},^1\text{H}$ -COSY spectrum ( $\text{CD}_3\text{OD}$ , 500 MHz) of cytochalasin J (7).

## Supplementary material



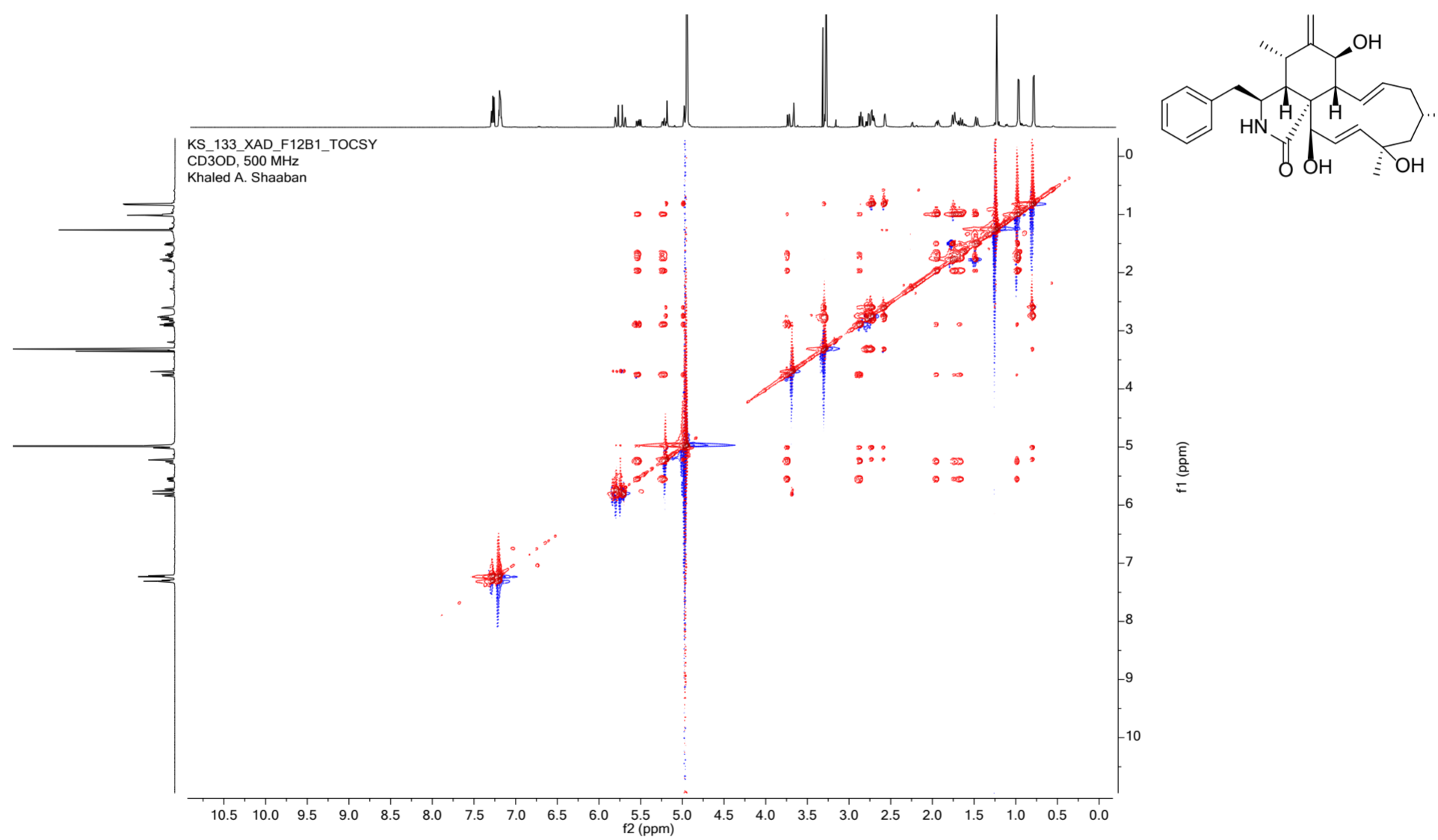
**Fig. 77S** HSQC spectrum (CD<sub>3</sub>OD, 500 MHz) of cytochalasin J (7).

## Supplementary material



**Fig. 78S** HMBC spectrum (CD<sub>3</sub>OD, 500 MHz) of cytochalasin J (7).

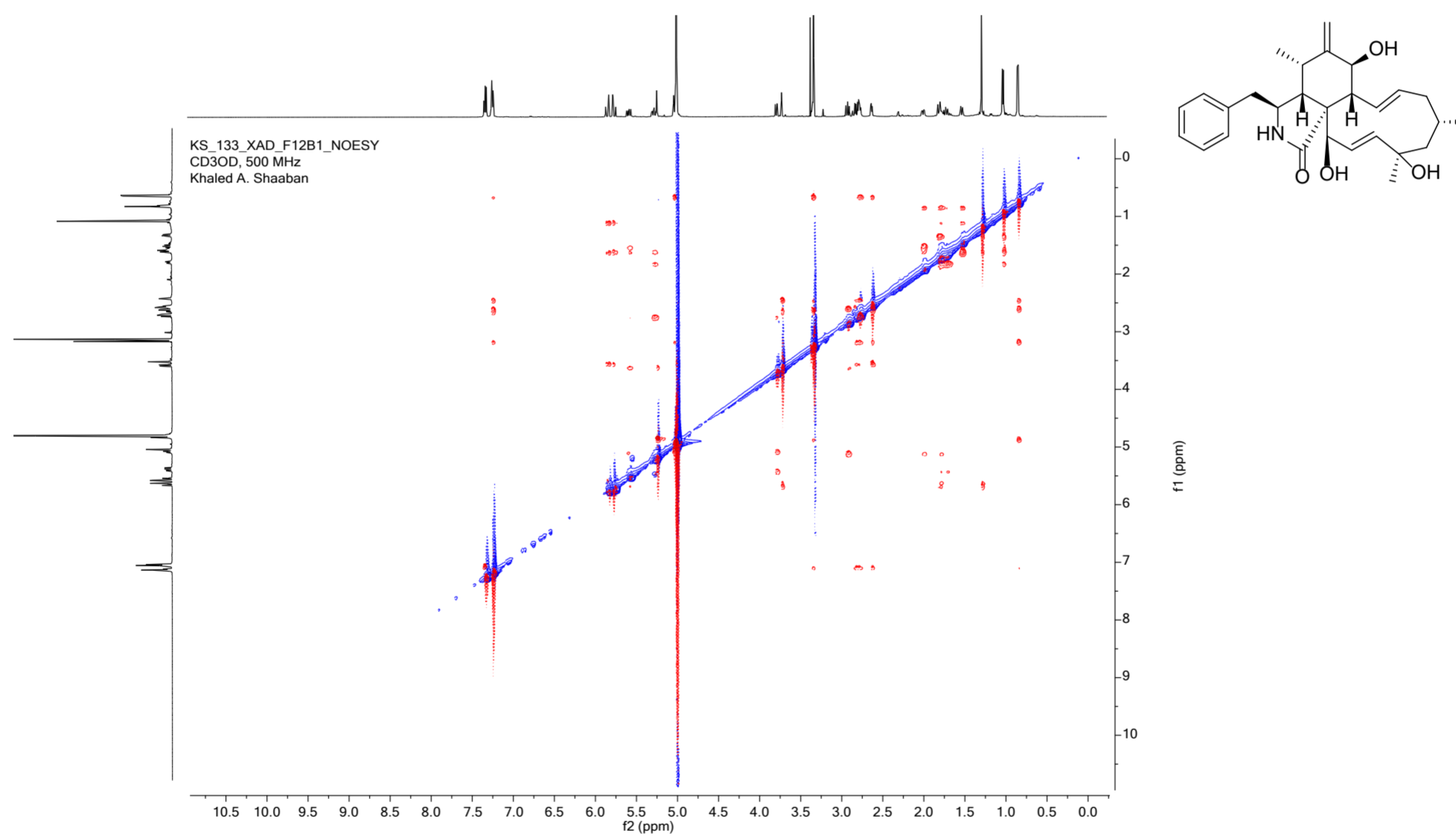
## Supplementary material



**Fig. 79S** TOCSY spectrum (CD<sub>3</sub>OD, 500 MHz) of cytochalasin J (7).

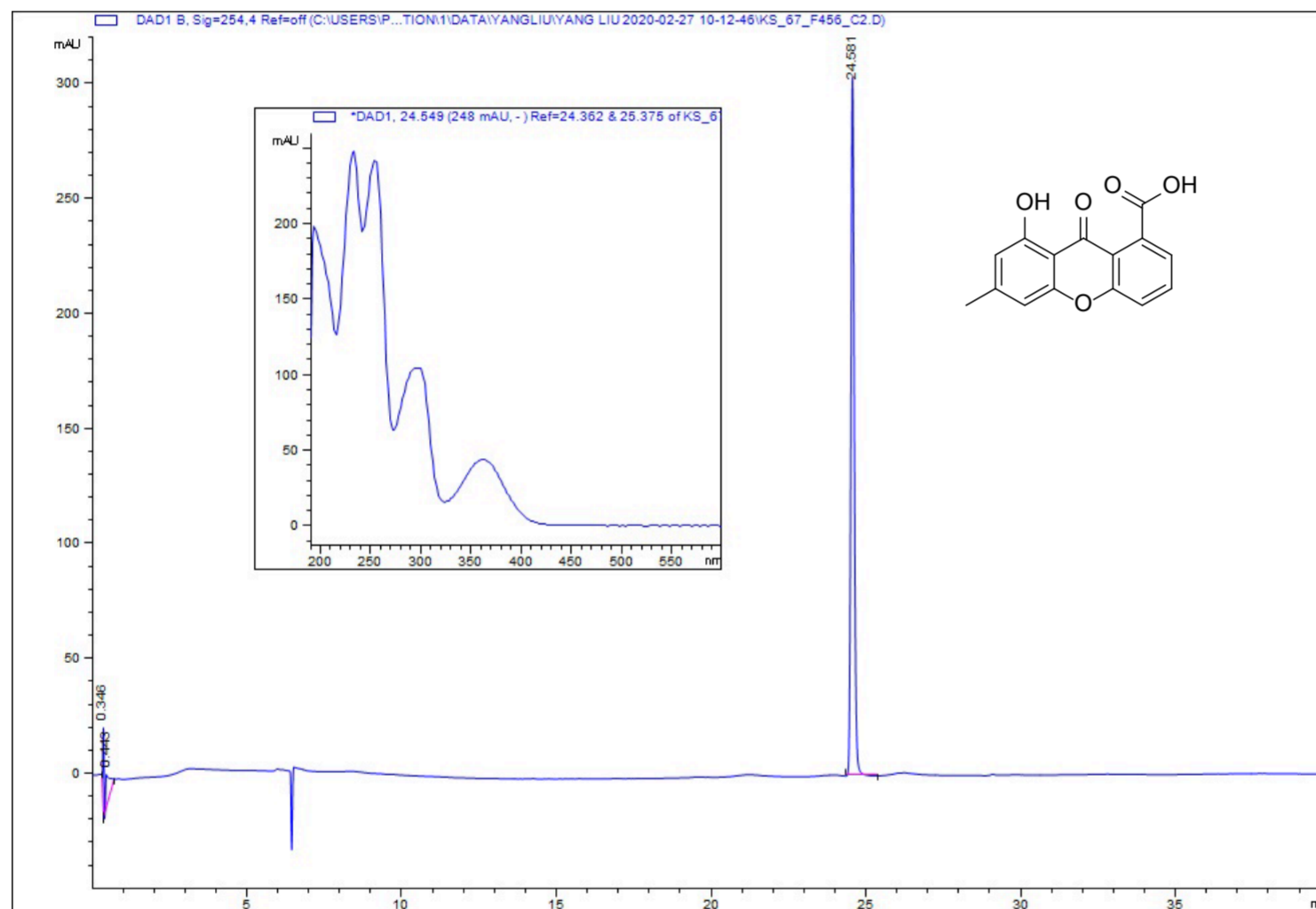


## Supplementary material



**Fig. 80S** NOESY spectrum (CD<sub>3</sub>OD, 500 MHz) of cytochalasin J (7).

## Supplementary material



**Fig. 81S** HPLC analysis of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**)/monodictyxanthone (**8b**). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 254 nm. UV-vis inset of full wavelength scan (190-600 nm).

## Supplementary material

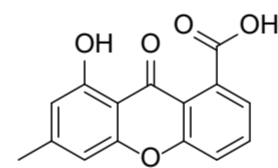
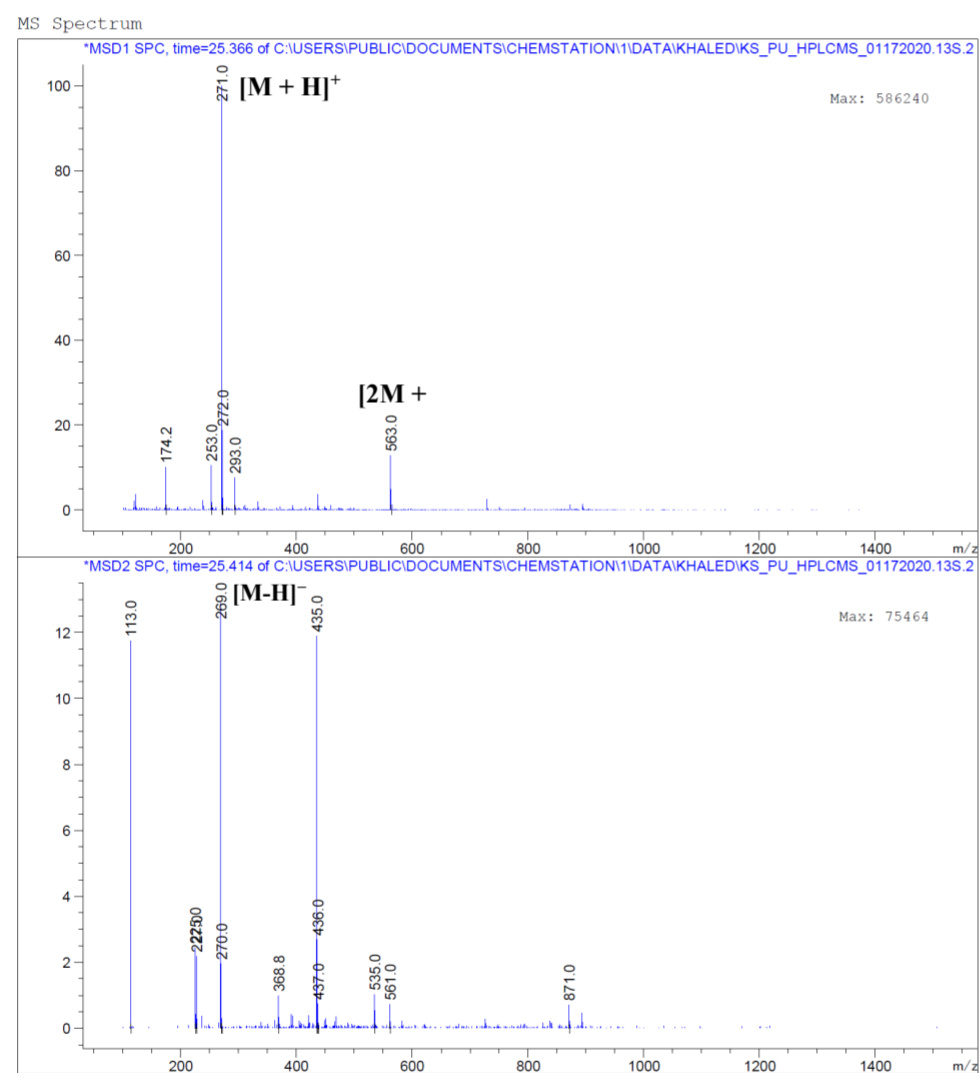
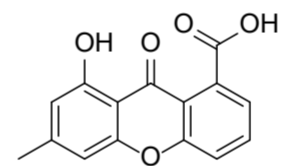
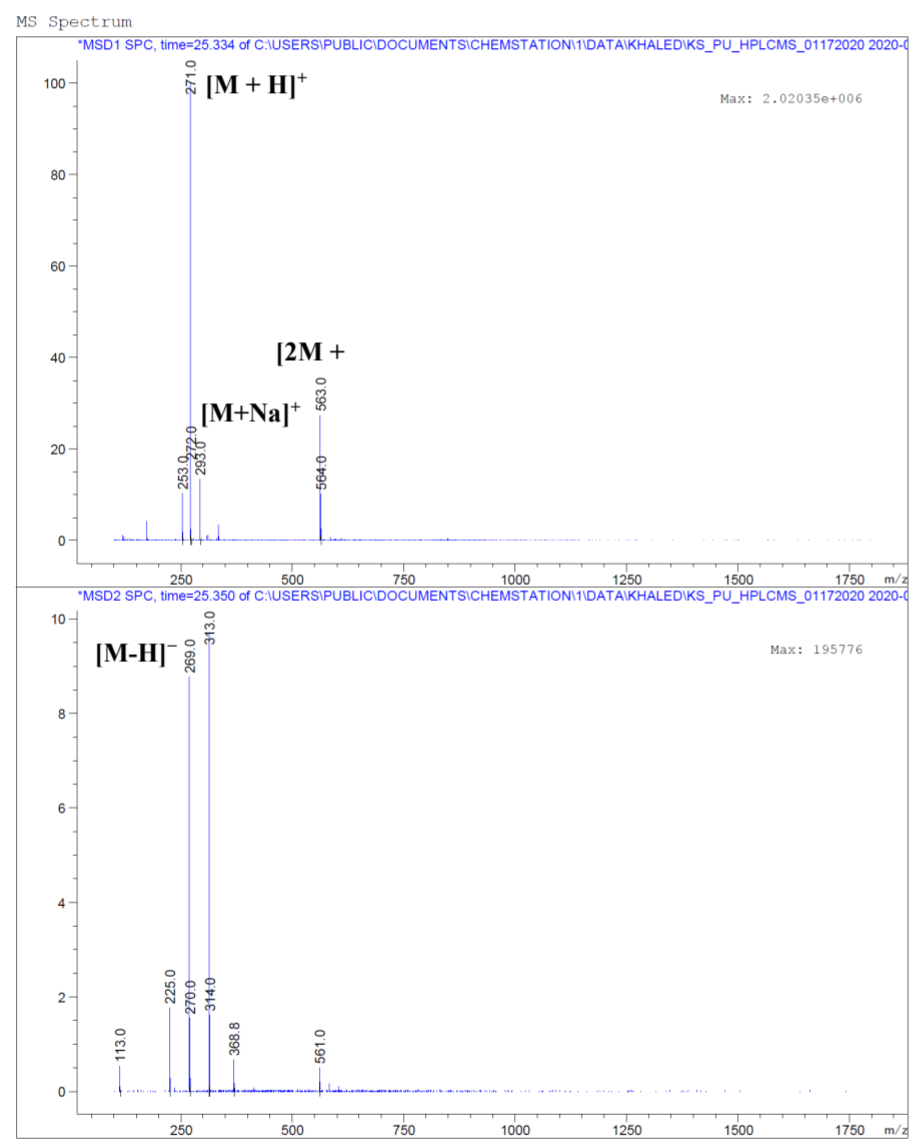


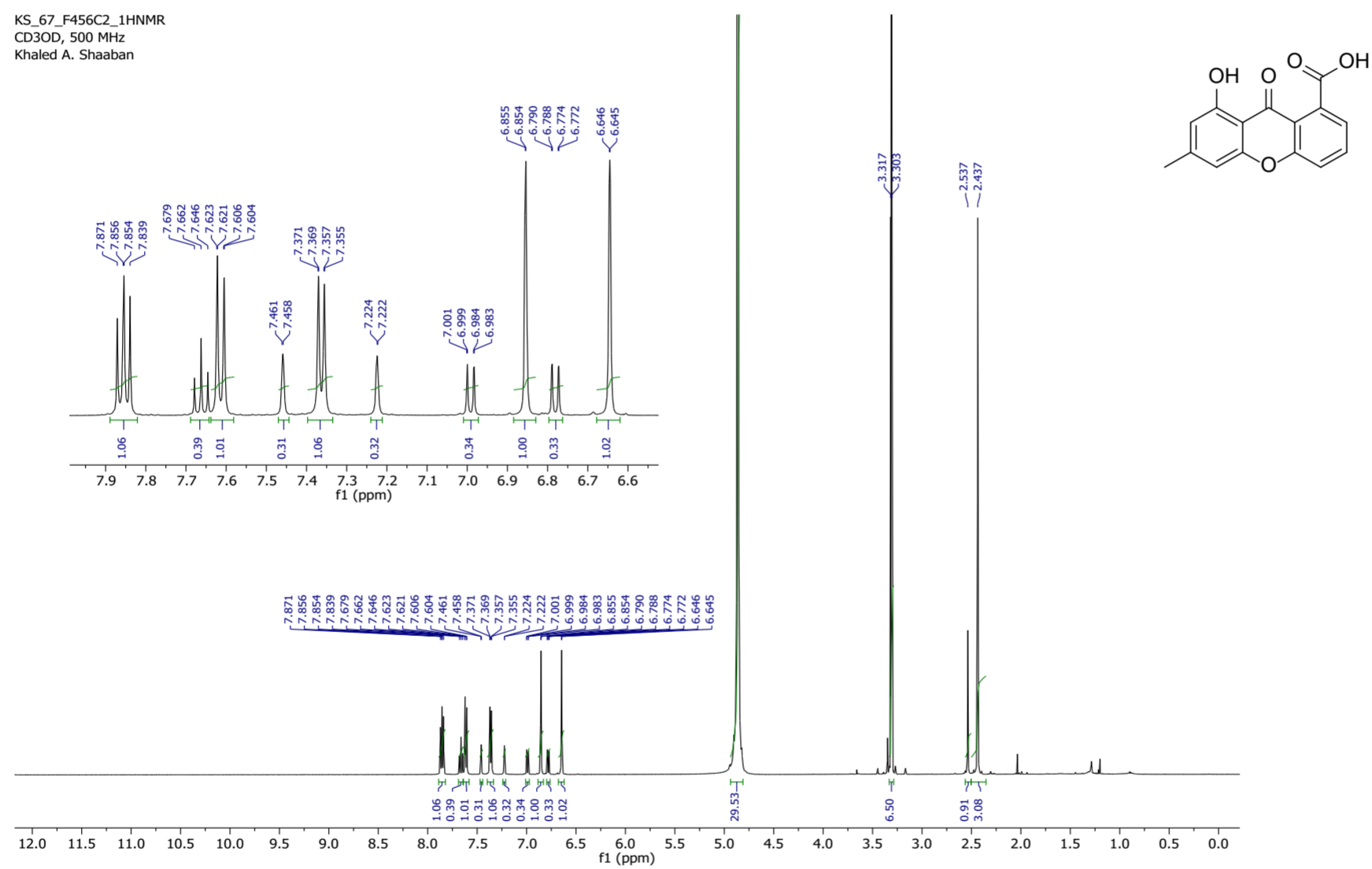
Fig. 82S (+) and (-)-ESI-MS spectrum of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).

## Supplementary material



**Fig. 83S** (+) and (-)-ESI-MS spectrum of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).

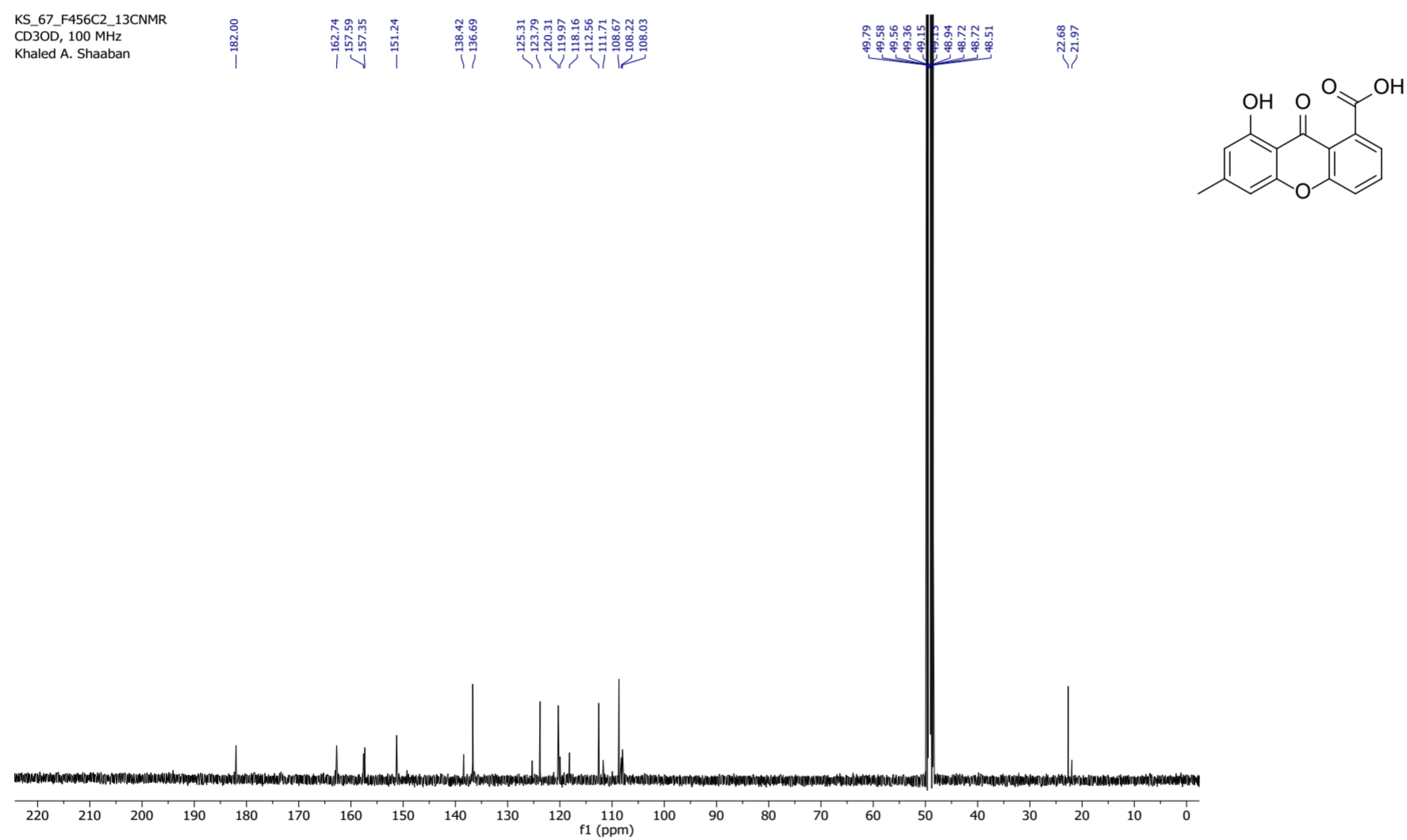
## Supplementary material



**Fig. 84S**  $^1\text{H}$  NMR spectrum (CD<sub>3</sub>OD, 500 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).

Note: this compound contains a trace (~25%) of its isomer monodictyranone (**8b**).

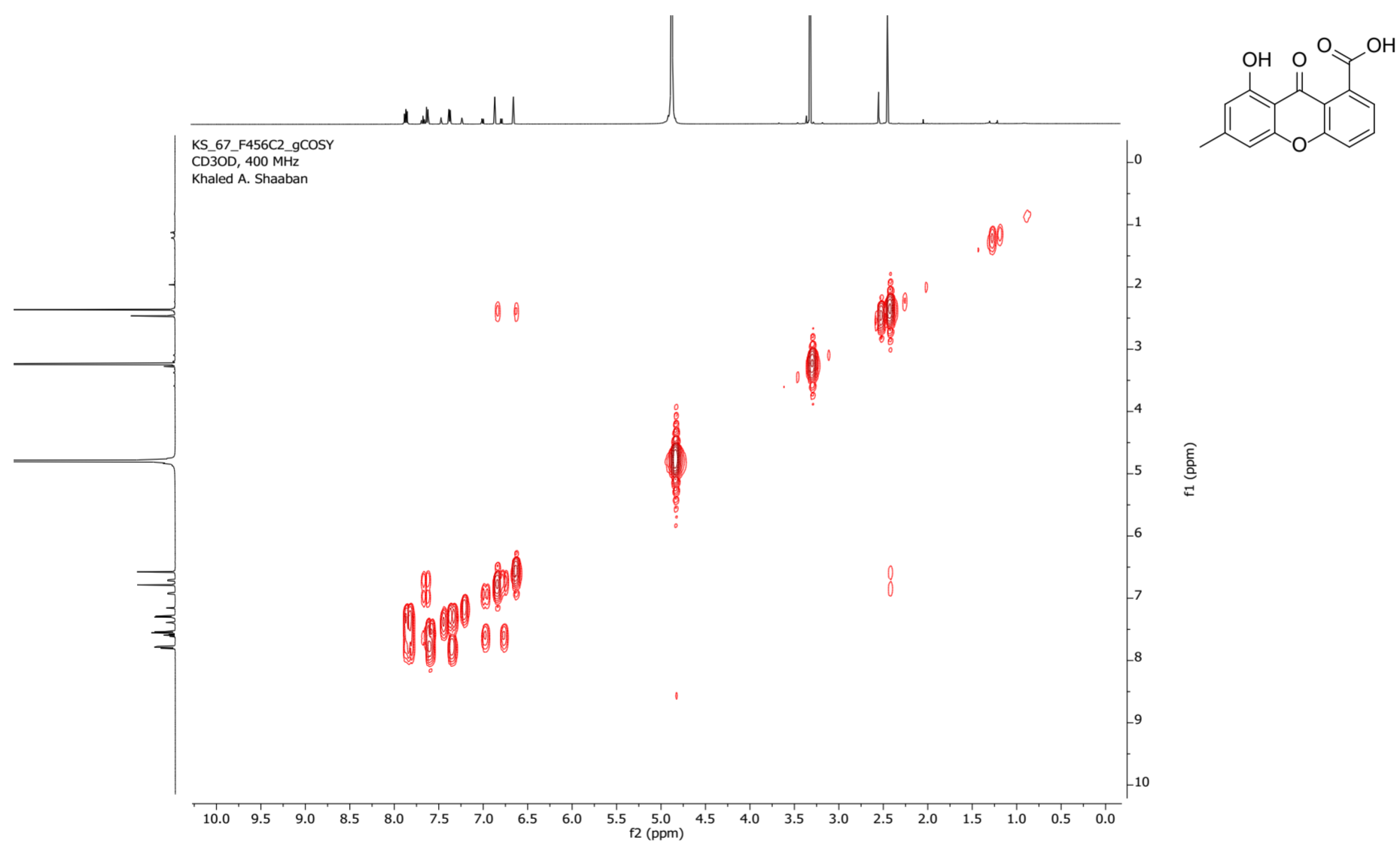
## Supplementary material



**Fig. 85S**  $^{13}\text{C}$  NMR spectrum ( $\text{CD}_3\text{OD}$ , 100 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).

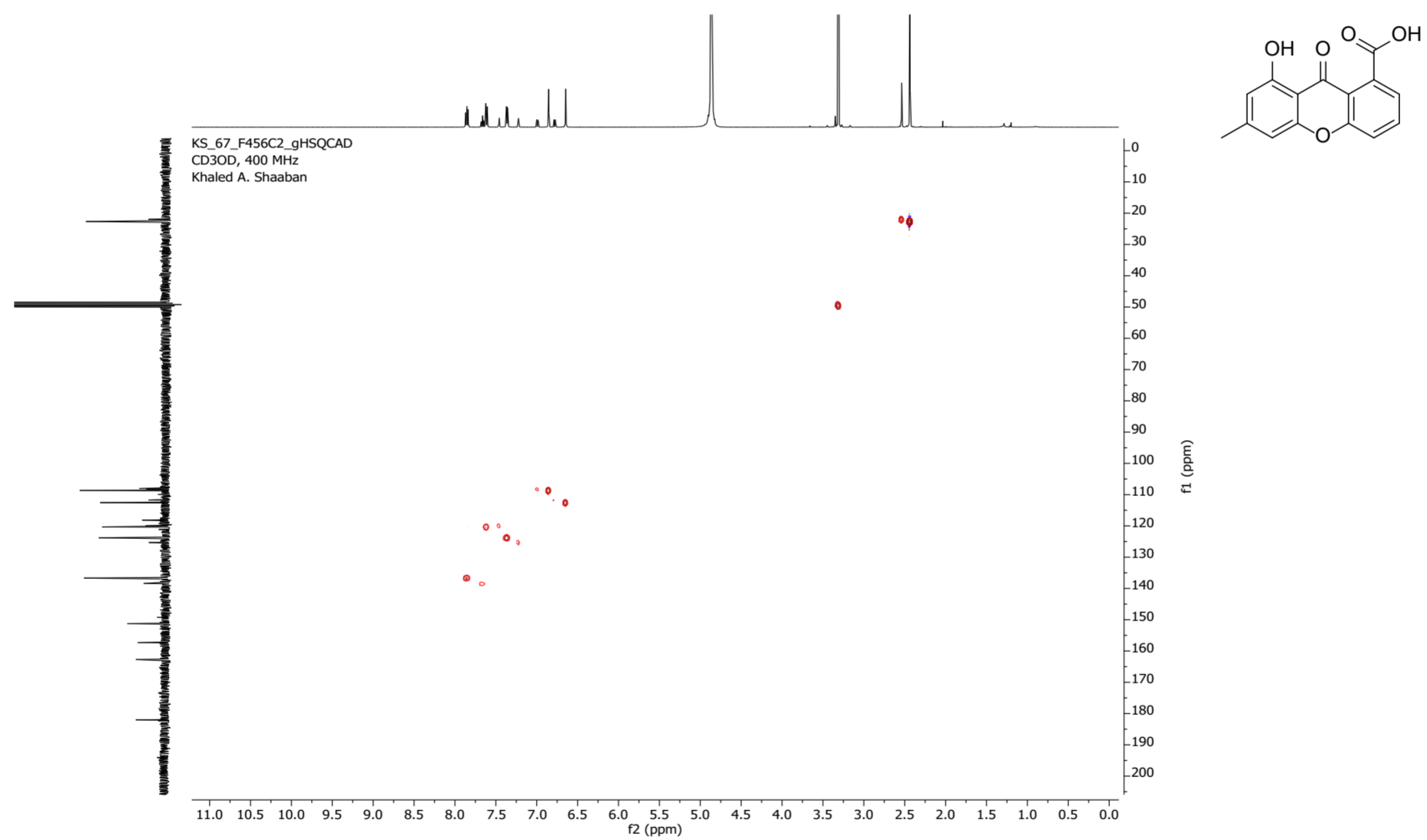
Note: this compound contains a trace ( $\sim 25\%$ ) of its isomer monodictyxanthone (**8b**).

## Supplementary material



**Fig. 86S**  $^1\text{H}$ ,  $^1\text{H}$ -COSY spectrum (CD<sub>3</sub>OD, 400 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).  
Note: this compound contains a trace (~25%) of its isomer monodictyxanthone (**8b**).

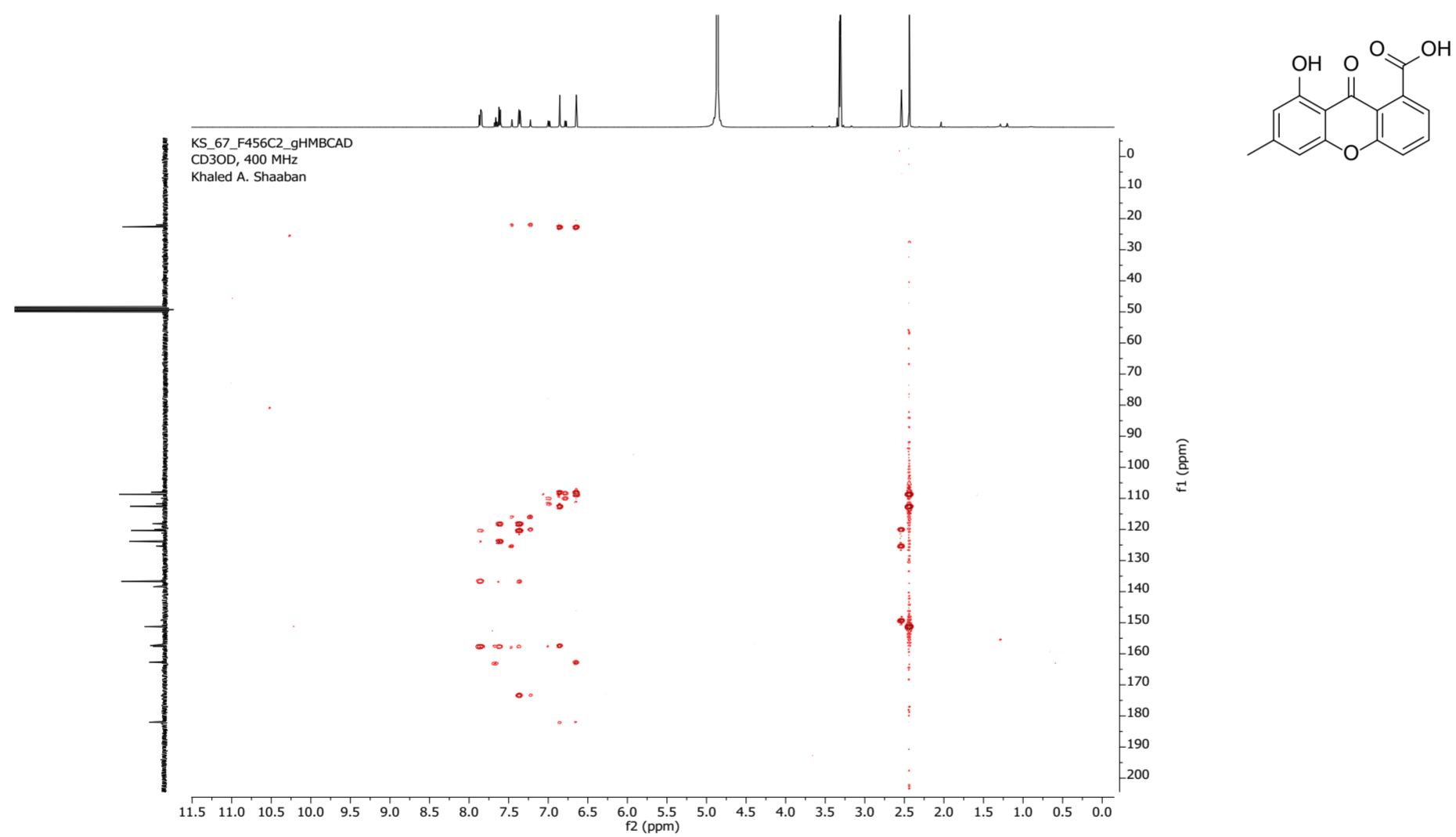
## Supplementary material



**Fig. 87S** HSQC spectrum (CD<sub>3</sub>OD, 400 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthen-1-carboxylic acid (**8a**). Note: this compound contains a trace (~25%) of its isomer monodictyxanthenone (**8b**).

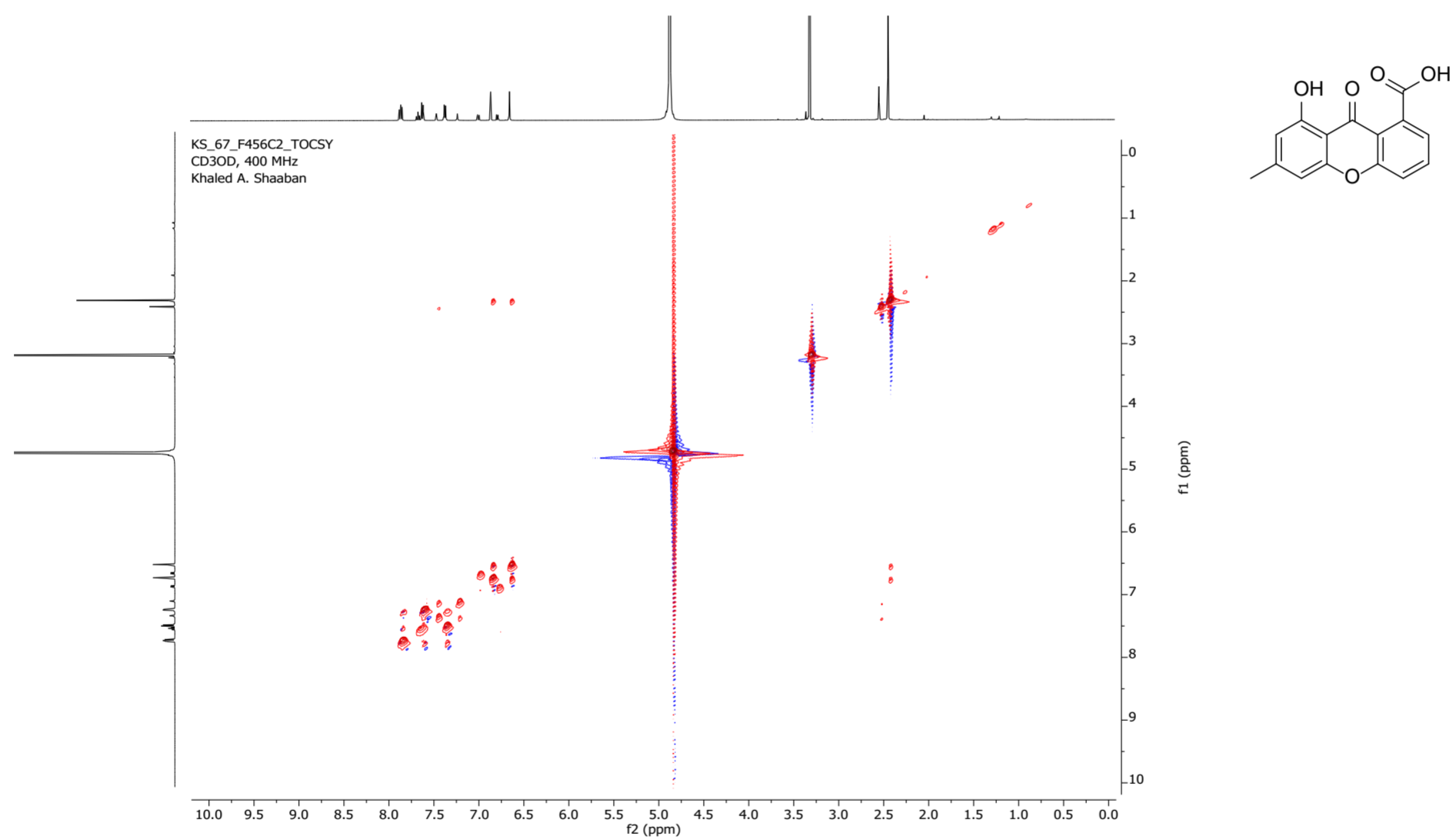


## Supplementary material



**Fig. 88S** HMBC spectrum (CD<sub>3</sub>OD, 400 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).  
Note: this compound contains a trace (~25%) of its isomer monodictyranthone (**8b**).

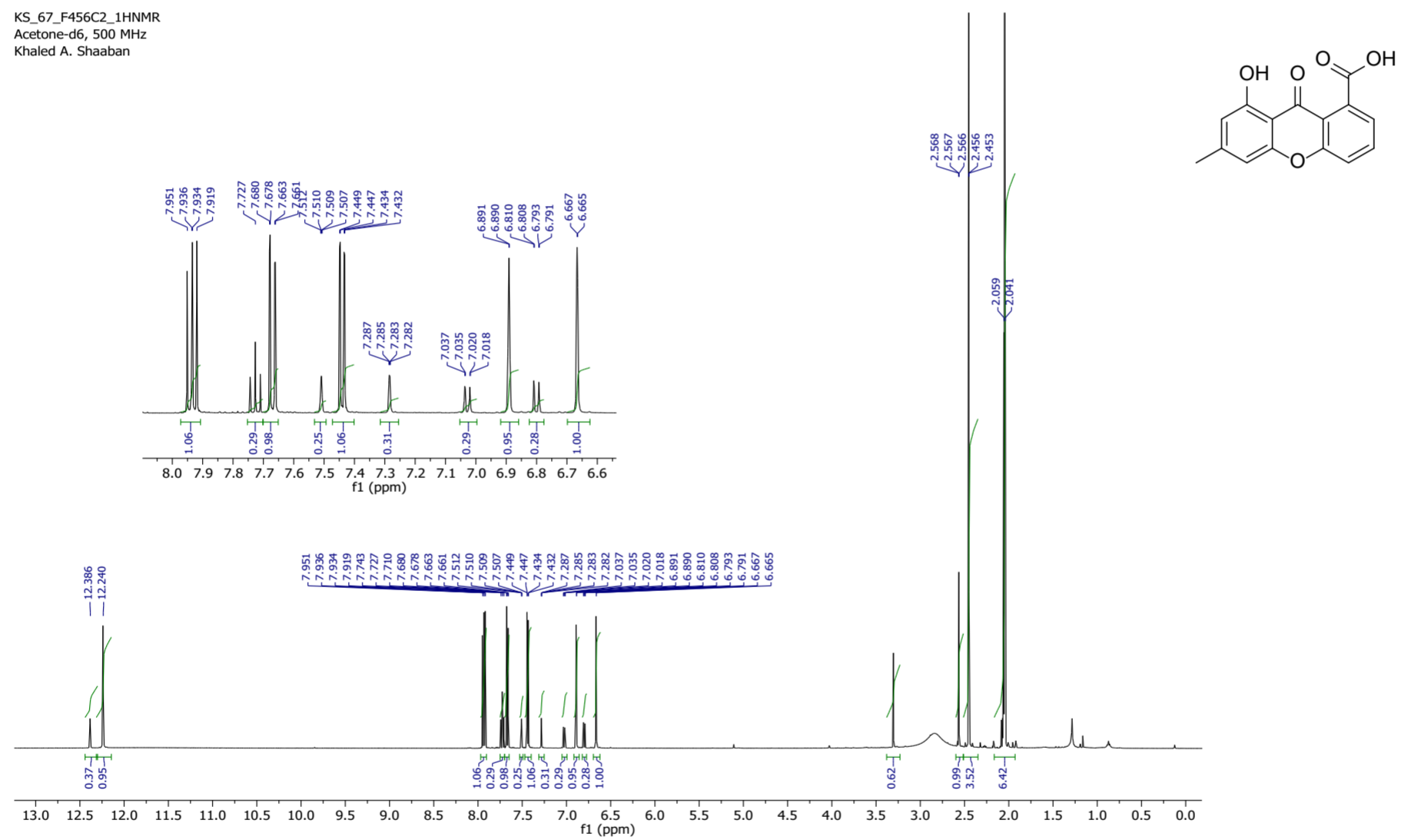
## Supplementary material



**Fig. 89S** TOCSY spectrum (CD<sub>3</sub>OD, 400 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**). Note: this compound contains a trace (~25%) of its isomer monodictyxanthone (**8b**).

## Supplementary material

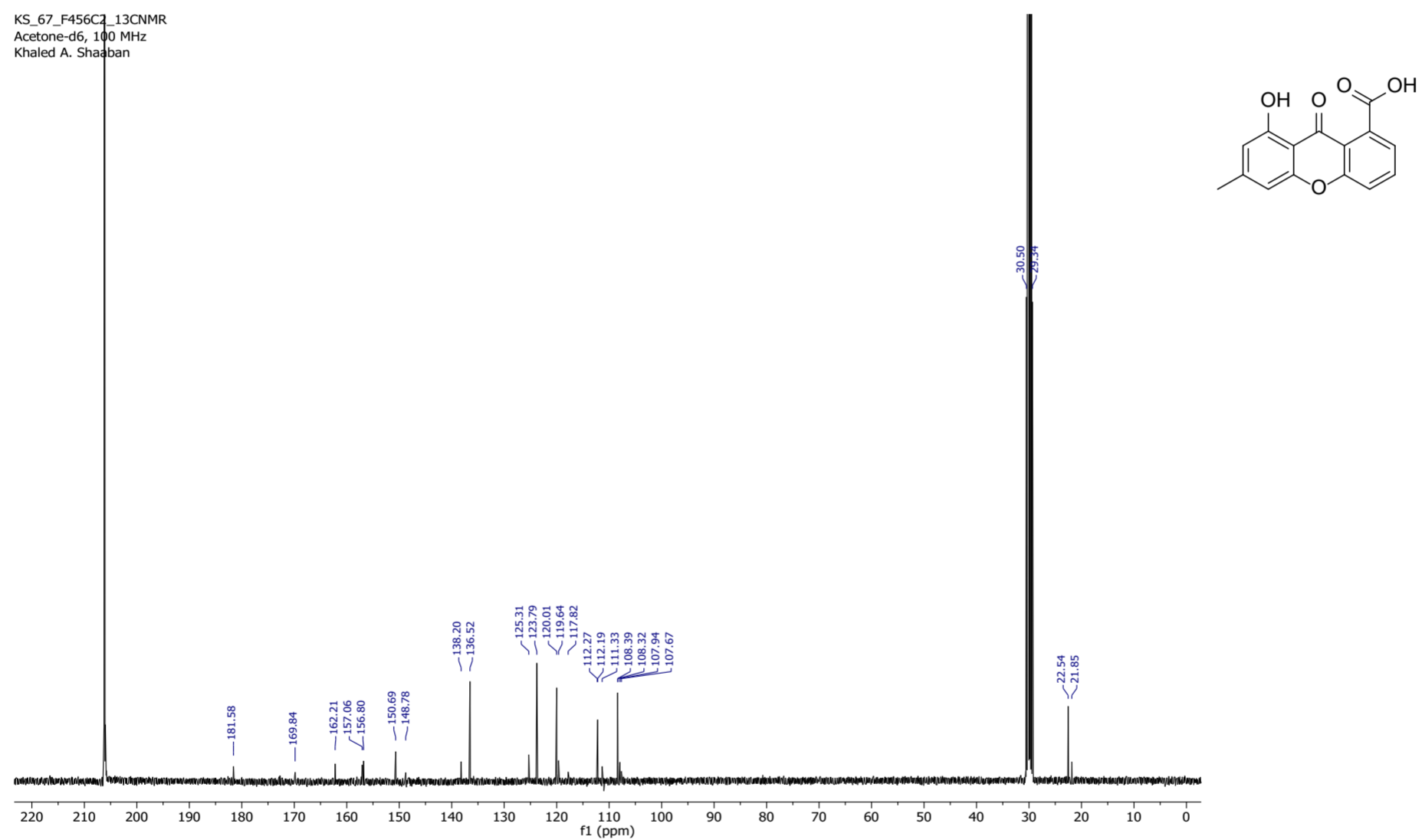
KS\_67\_F456C2\_1HNMR  
Acetone-d<sub>6</sub>, 500 MHz  
Khaled A. Shaaban



**Fig. 90S** <sup>1</sup>H NMR spectrum (acetone-*d*<sub>6</sub>, 500 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).

Note: this compound contains a trace (~25%) of its isomer monodictyxanthone (**8b**).

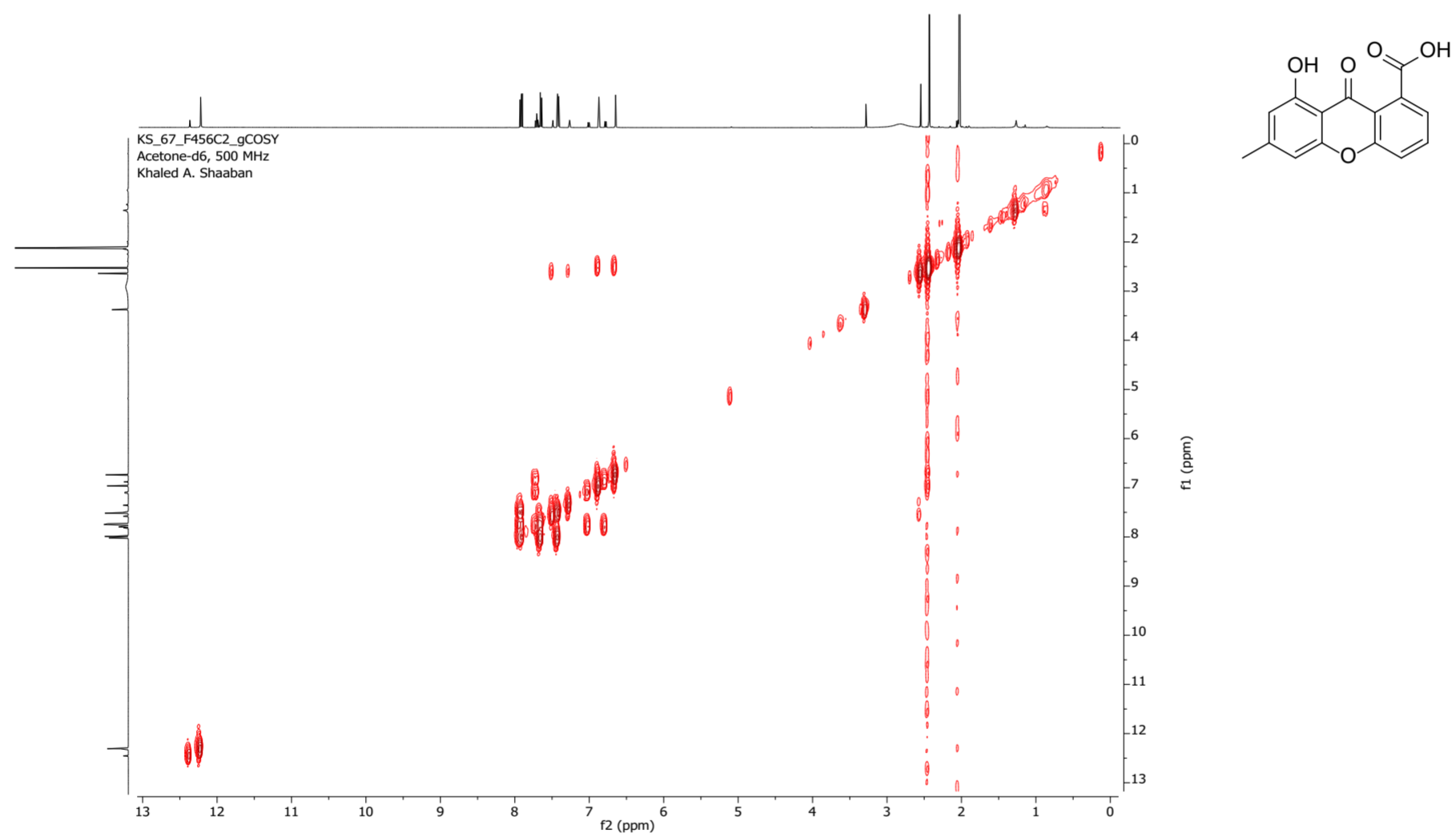
## Supplementary material



**Fig. 91S** <sup>13</sup>C NMR spectrum (acetone-*d*<sub>6</sub>, 100 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).

Note: this compound contains a trace (~25%) of its isomer monodictyranone (**8b**).

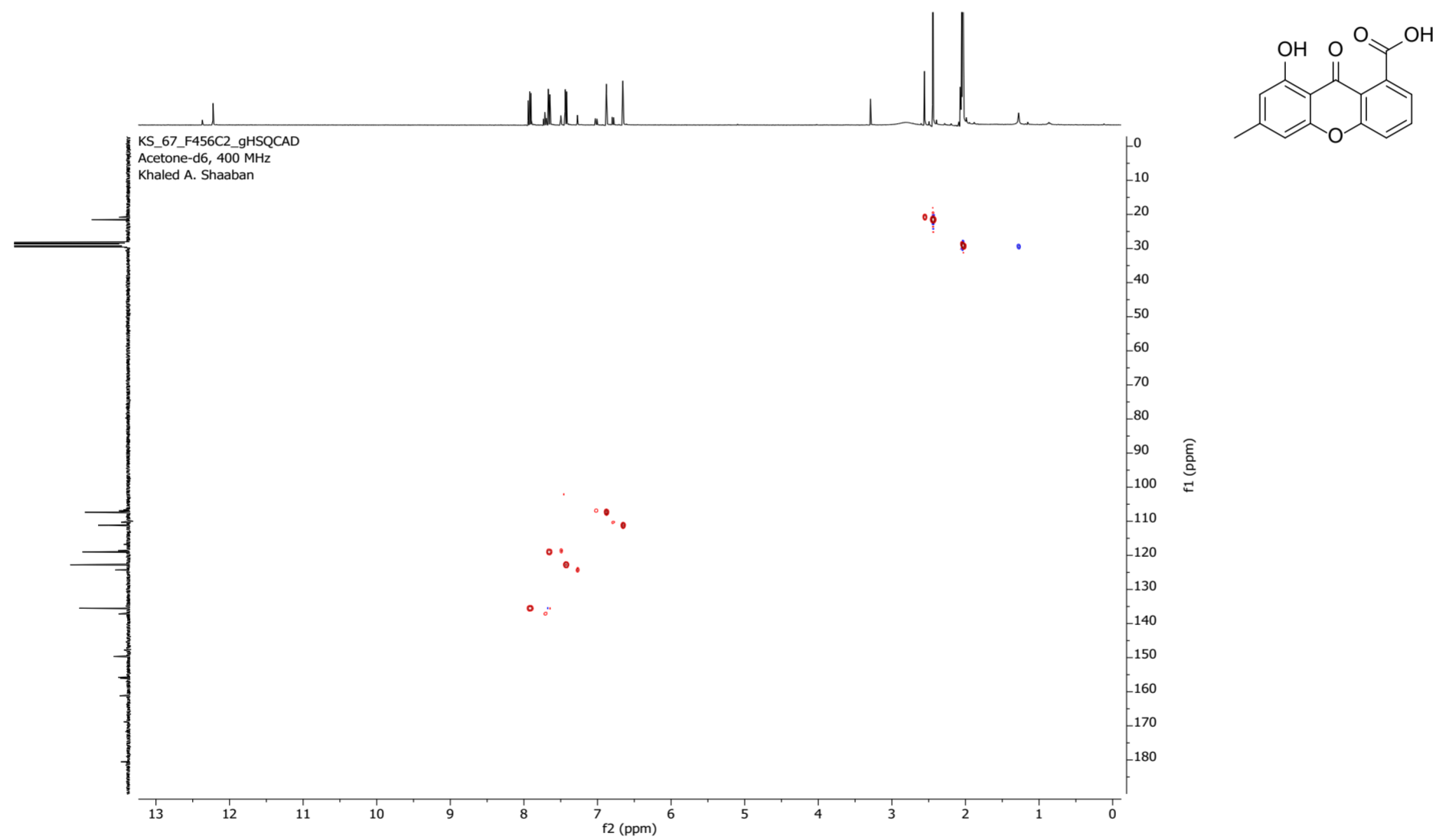
## Supplementary material



**Fig. 92S**  $^1\text{H}, ^1\text{H}$ -COSY spectrum (acetone- $d_6$ , 500 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).

Note: this compound contains a trace (~25%) of its isomer monodictyxanthone (**8b**).

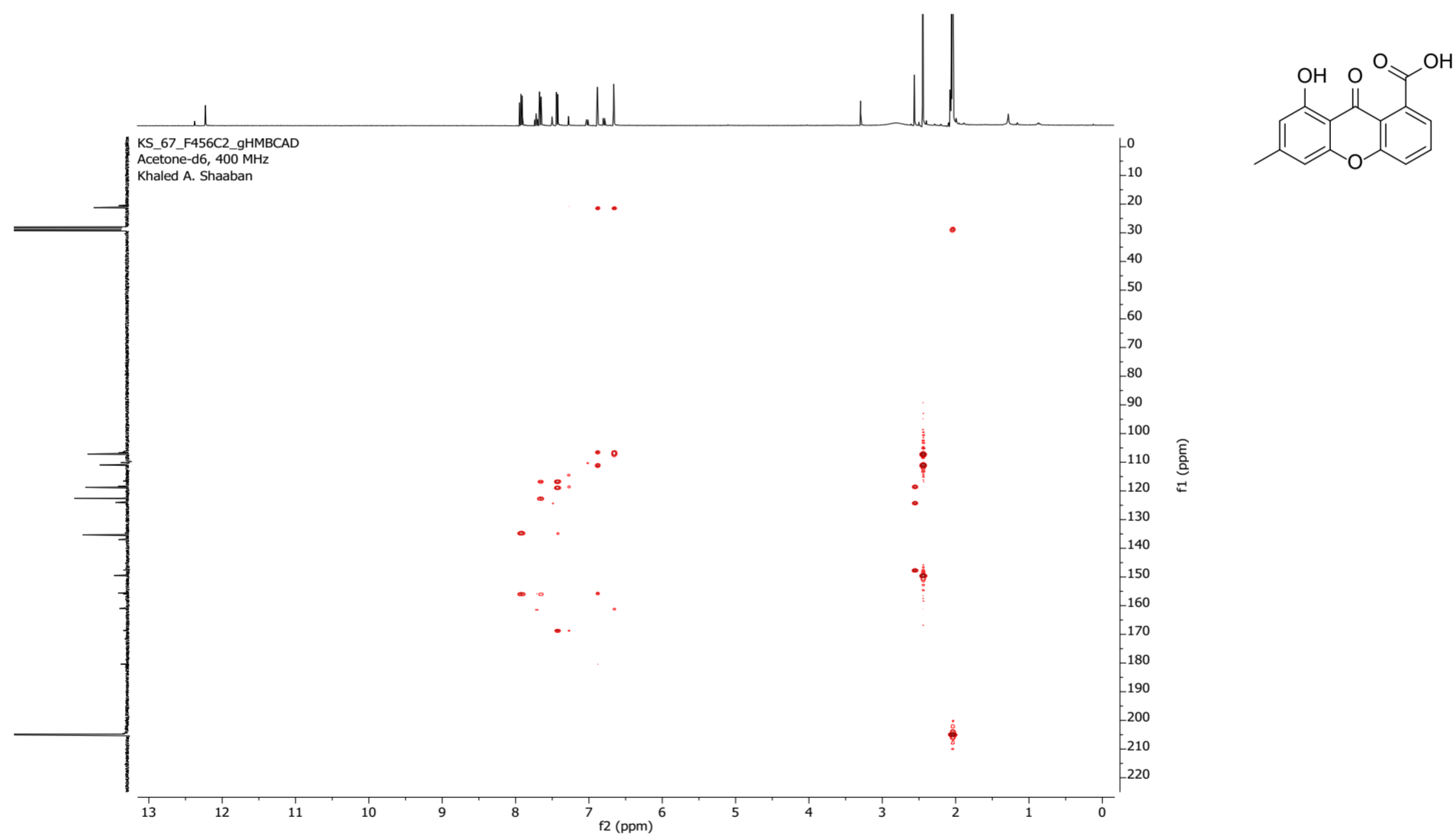
## Supplementary material



**Fig. 93S** HSQC spectrum (acetone- $d_6$ , 400 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).

Note: this compound contains a trace (~25%) of its isomer monodictyranthone (**8b**).

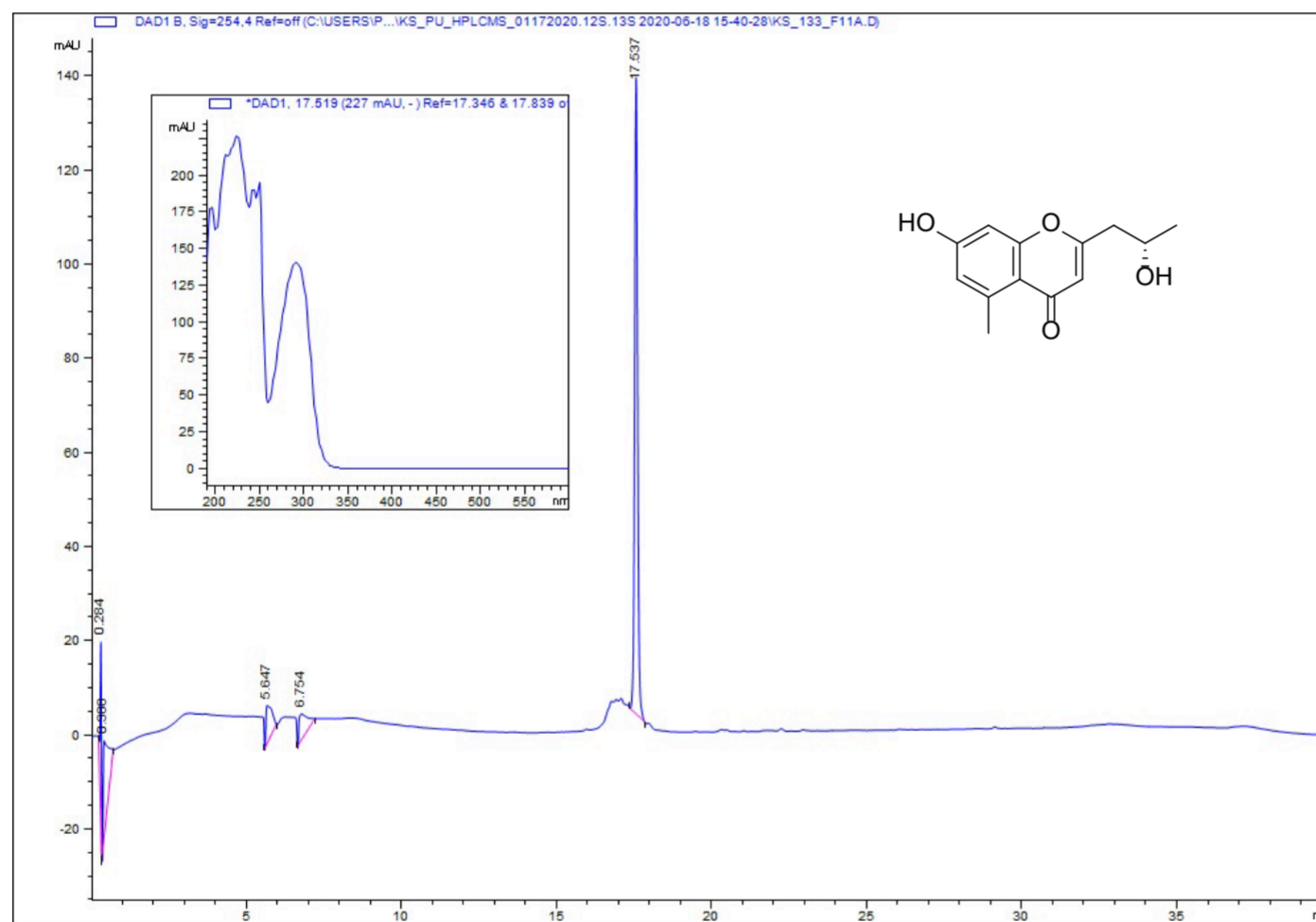
## Supplementary material



**Fig. 94S** HMBC spectrum (acetone-*d*<sub>6</sub>, 400 MHz) of 8-hydroxy-6-methyl-9-oxo-9H-xanthene-1-carboxylic acid (**8a**).

Note: this compound contains a trace (~25%) of its isomer monodictyxanthone (**8b**).

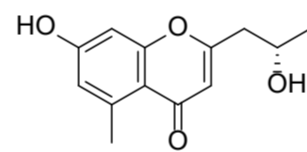
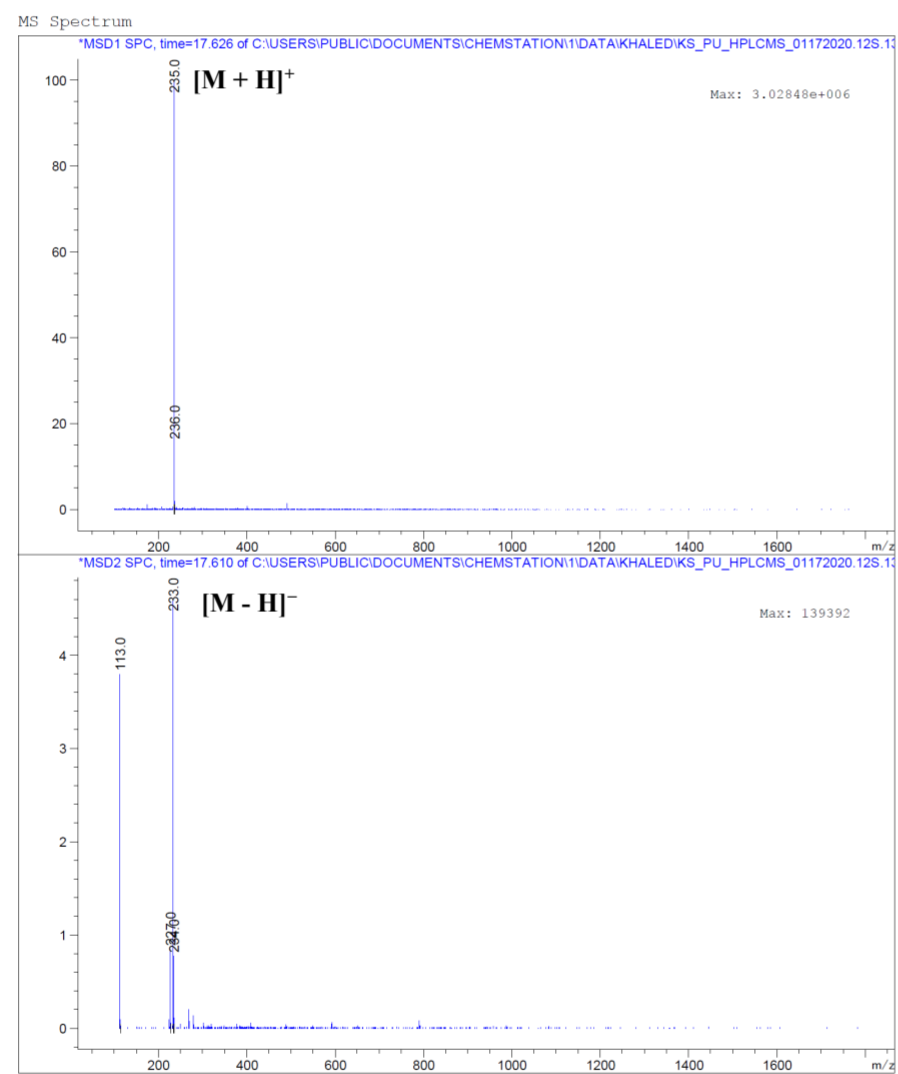
## Supplementary material



**Fig. 95S** HPLC analysis of 5-carbomethoxymethyl-2-heptyl-7-hydroxychromone (**7**). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 254 nm. UV-vis inset of full wavelength scan (190-600 nm).



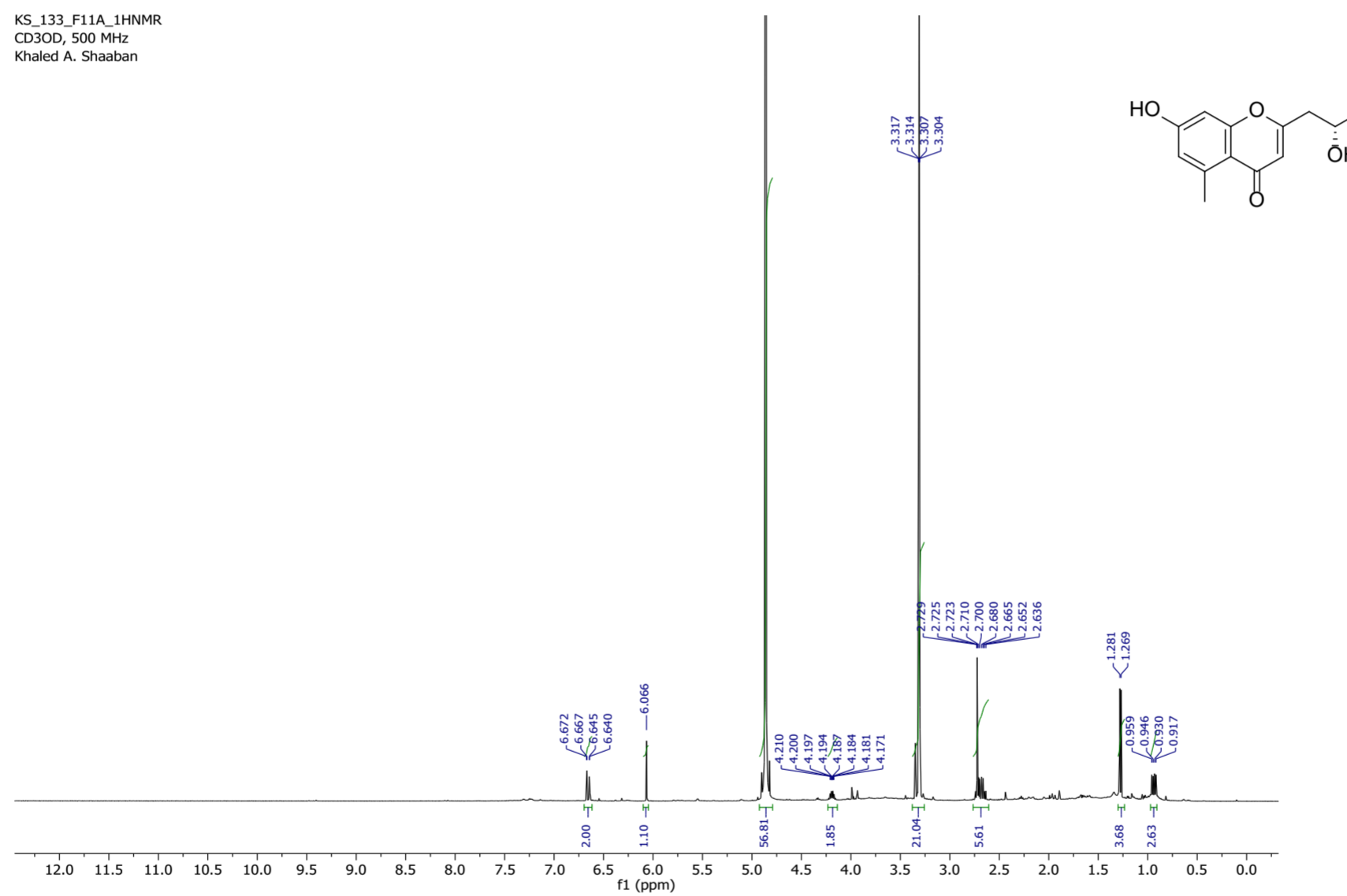
## Supplementary material



**Fig. 96S** (+) and (-)-ESI-MS spectrum of 5-carbomethoxymethyl-2-heptyl-7-hydroxychromone (**9**).

## Supplementary material

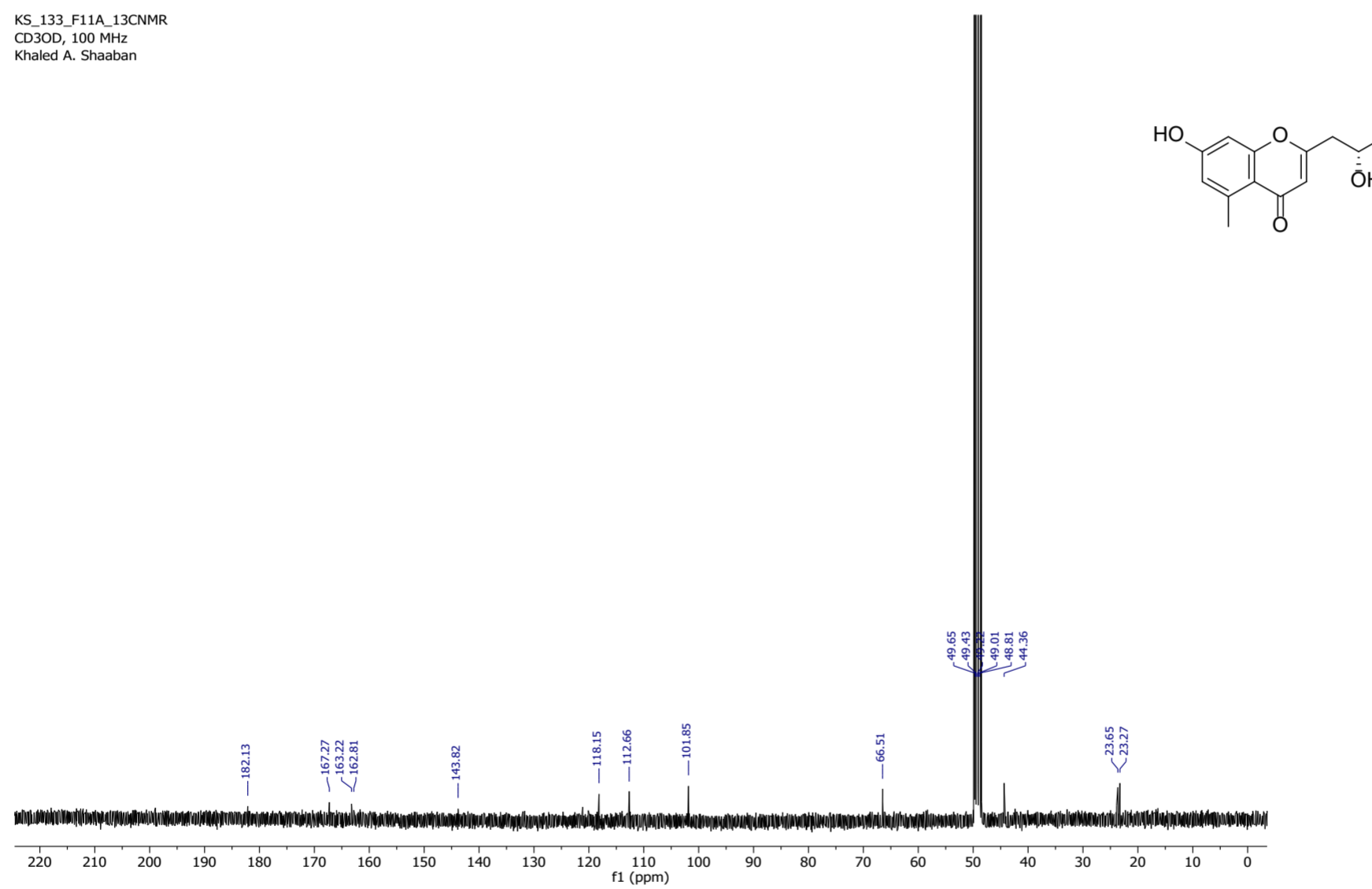
KS\_133\_F11A\_1HNMR  
CD3OD, 500 MHz  
Khaled A. Shaaban



**Fig. 97S** <sup>1</sup>H NMR spectrum (CD<sub>3</sub>OD, 500 MHz) of 5-carbomethoxymethyl-2-heptyl-7-hydroxychromone (9).

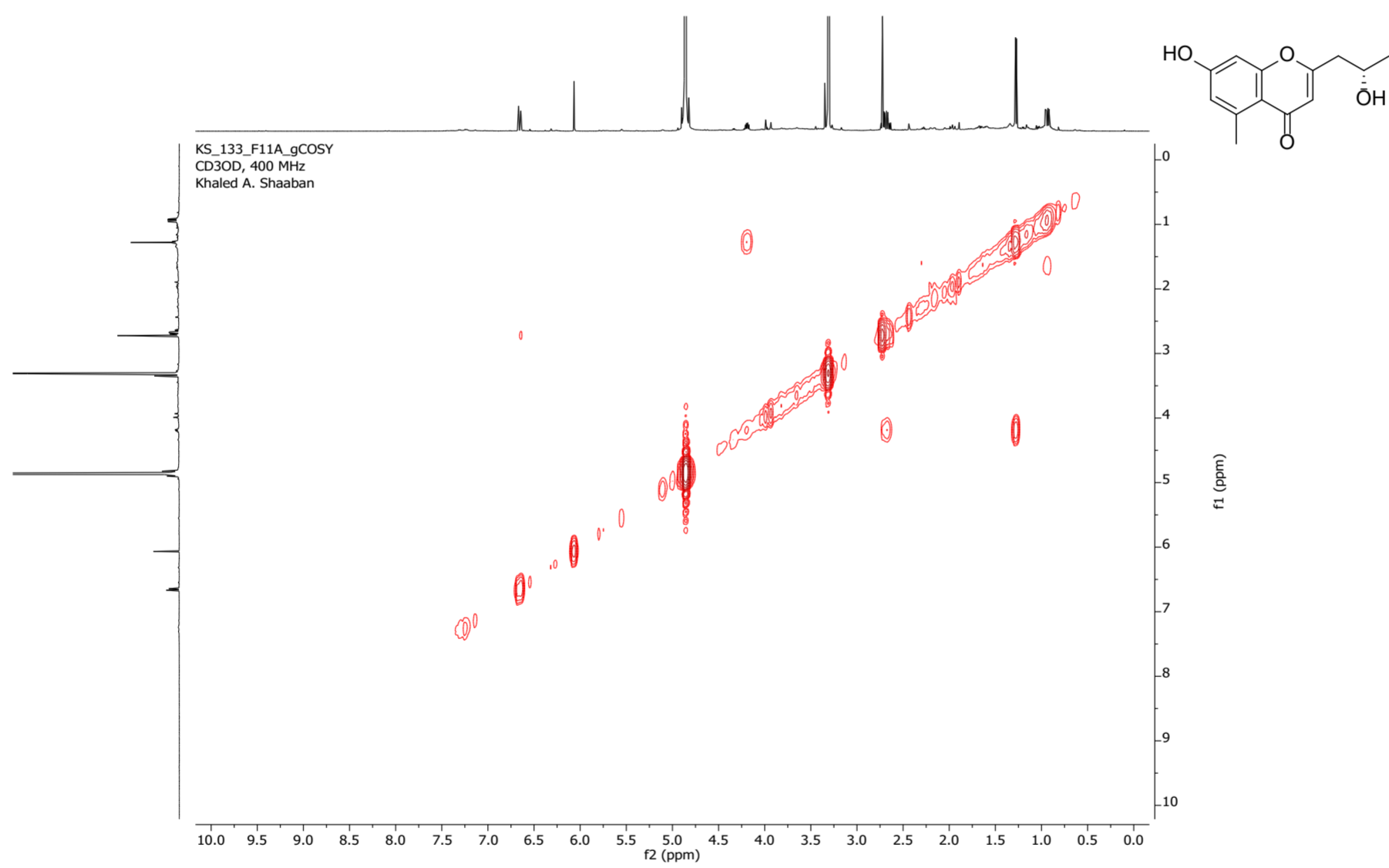
## Supplementary material

KS\_133\_F11A\_13CNMR  
CD3OD, 100 MHz  
Khaled A. Shaaban



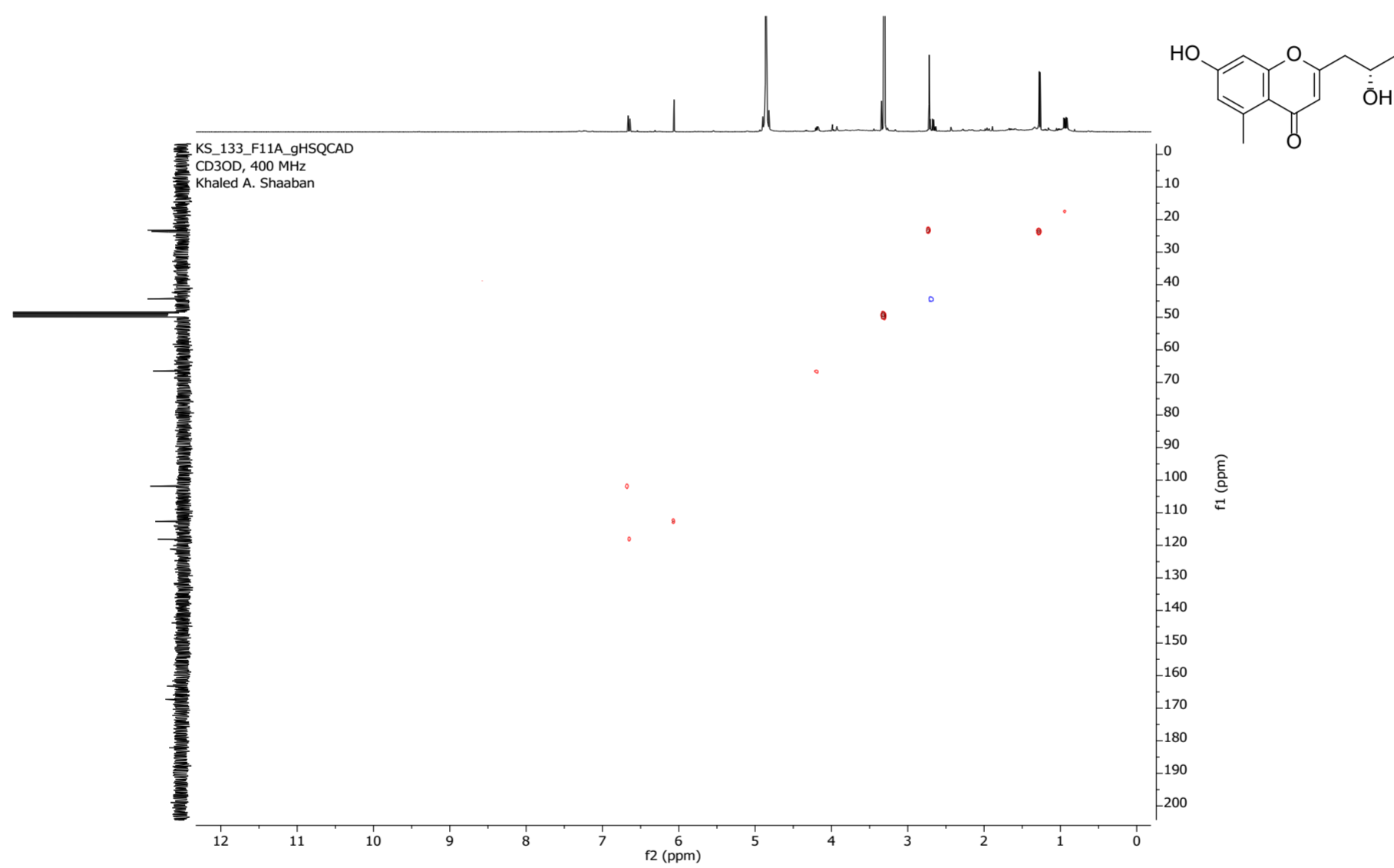
**Fig. 98S** <sup>13</sup>C NMR spectrum (CD<sub>3</sub>OD, 100 MHz) of 5-carbomethoxymethyl-2-heptyl-7-hydroxychromone (**9**).

## Supplementary material



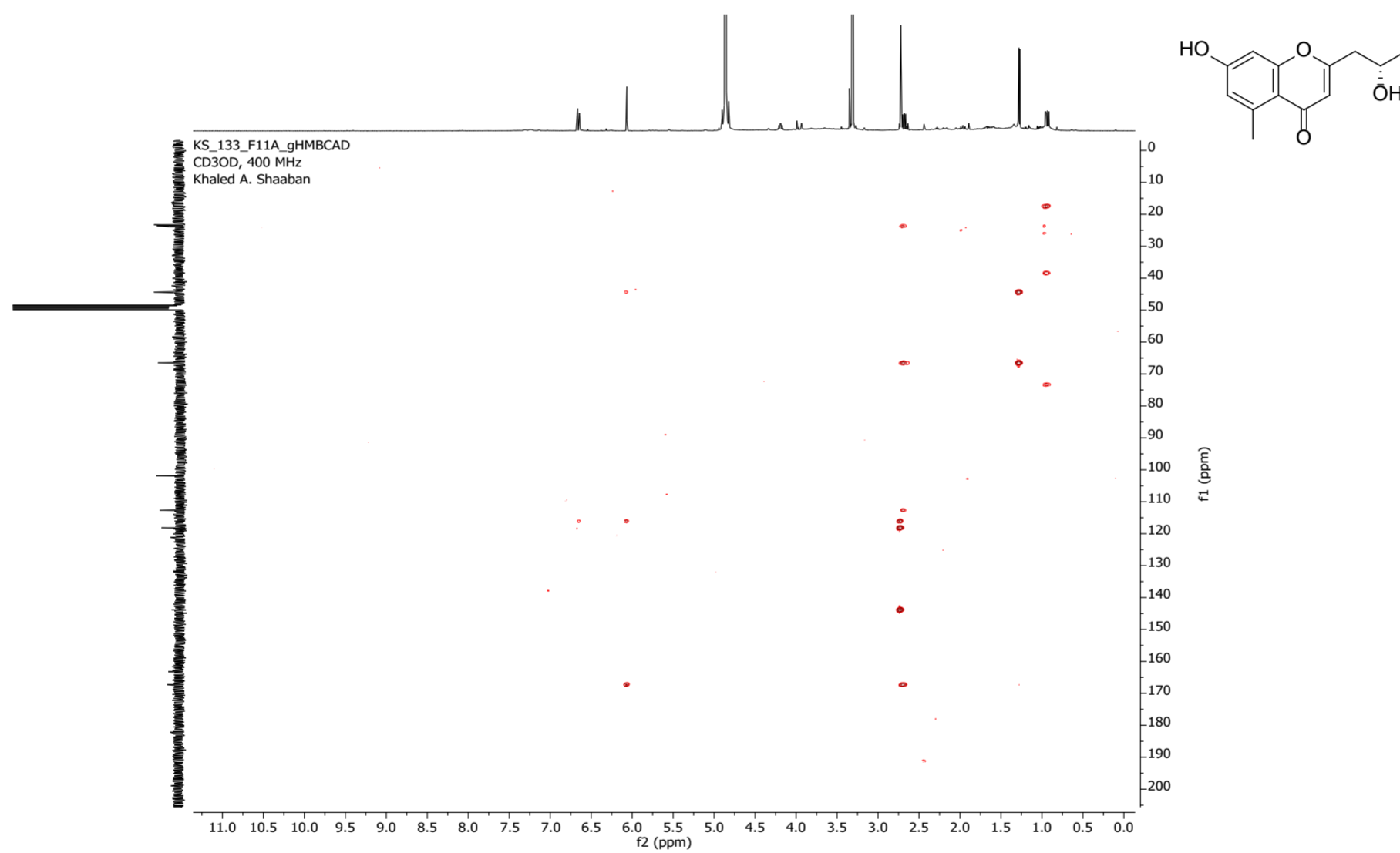
**Fig. 99S**  $^1\text{H}, ^1\text{H}$ -COSY spectrum ( $\text{CD}_3\text{OD}$ , 400 MHz) of 5-carbomethoxymethyl-2-heptyl-7-hydroxychromone (**9**).

## Supplementary material



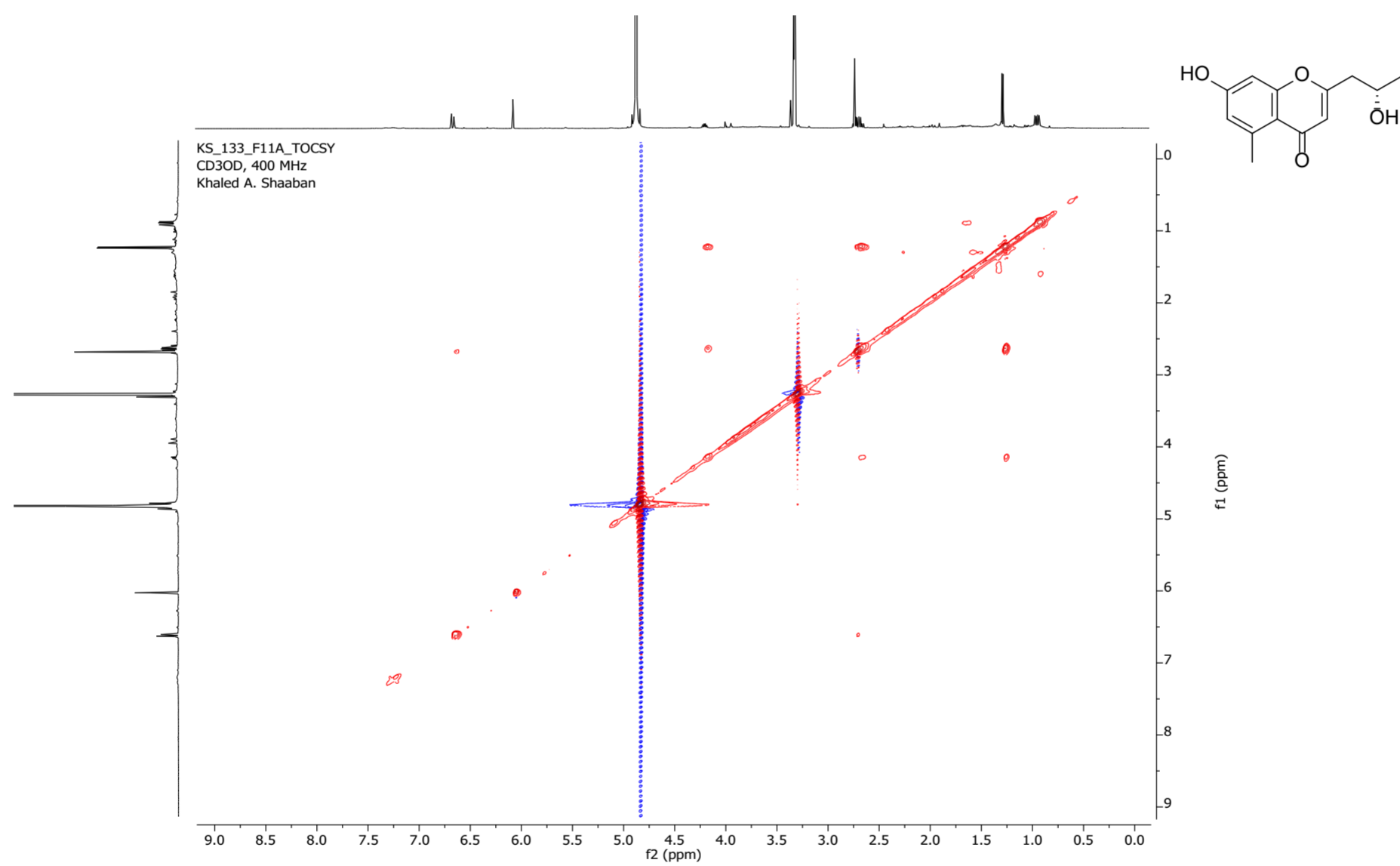
**Fig. 100S** HSQC spectrum (CD<sub>3</sub>OD, 400 MHz) of 5-carbomethoxymethyl-2-heptyl-7-hydroxychromone (**9**).

## Supplementary material



**Fig. 101S** HMBC spectrum (CD<sub>3</sub>OD, 400 MHz) of 5-carbomethoxymethyl-2-heptyl-7-hydroxychromone (**9**).

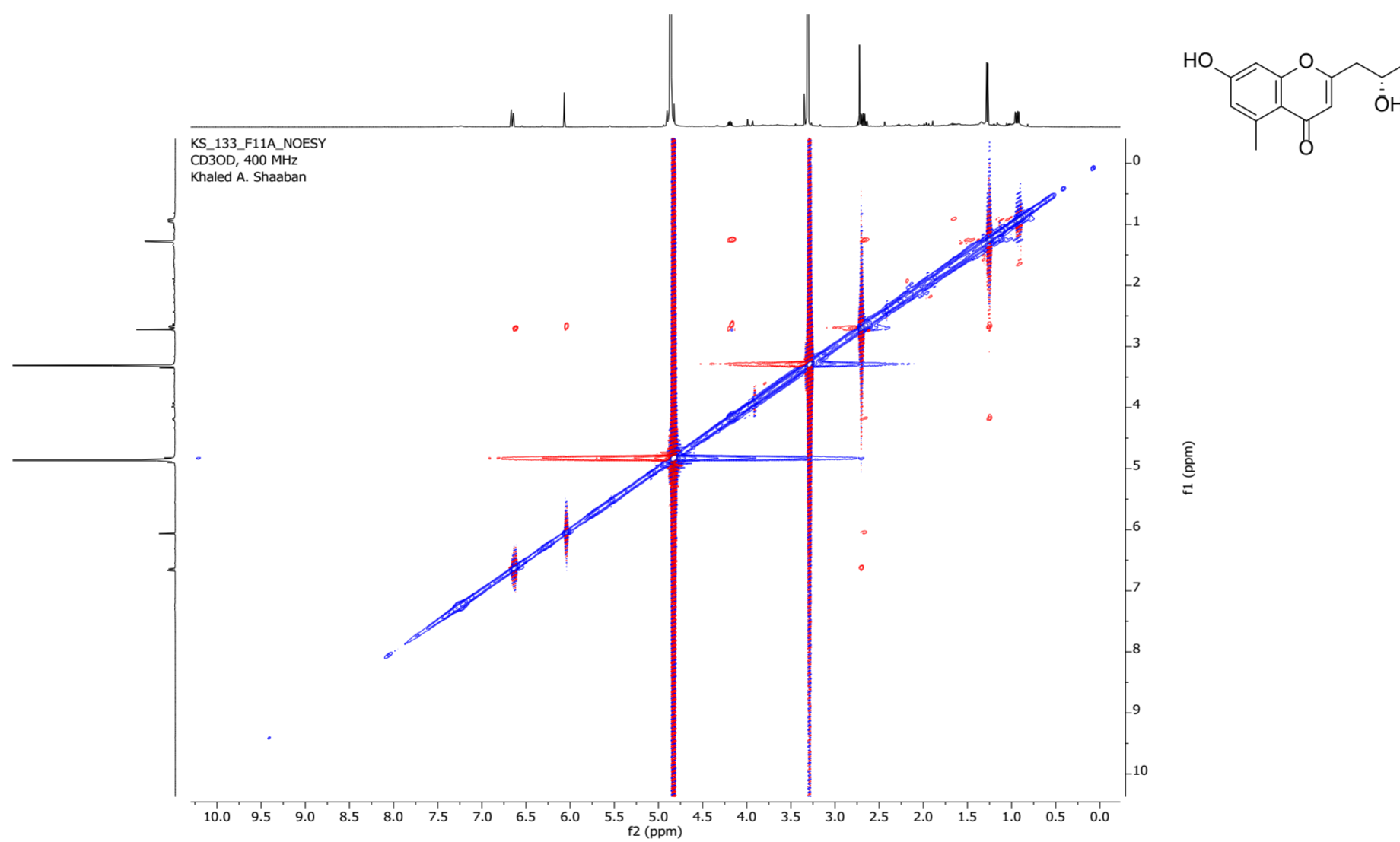
## Supplementary material



**Fig. 102S** TOCSY spectrum (CD<sub>3</sub>OD, 400 MHz) of 5-carbomethoxymethyl-2-heptyl-7-hydroxychromone (**9**).

S111

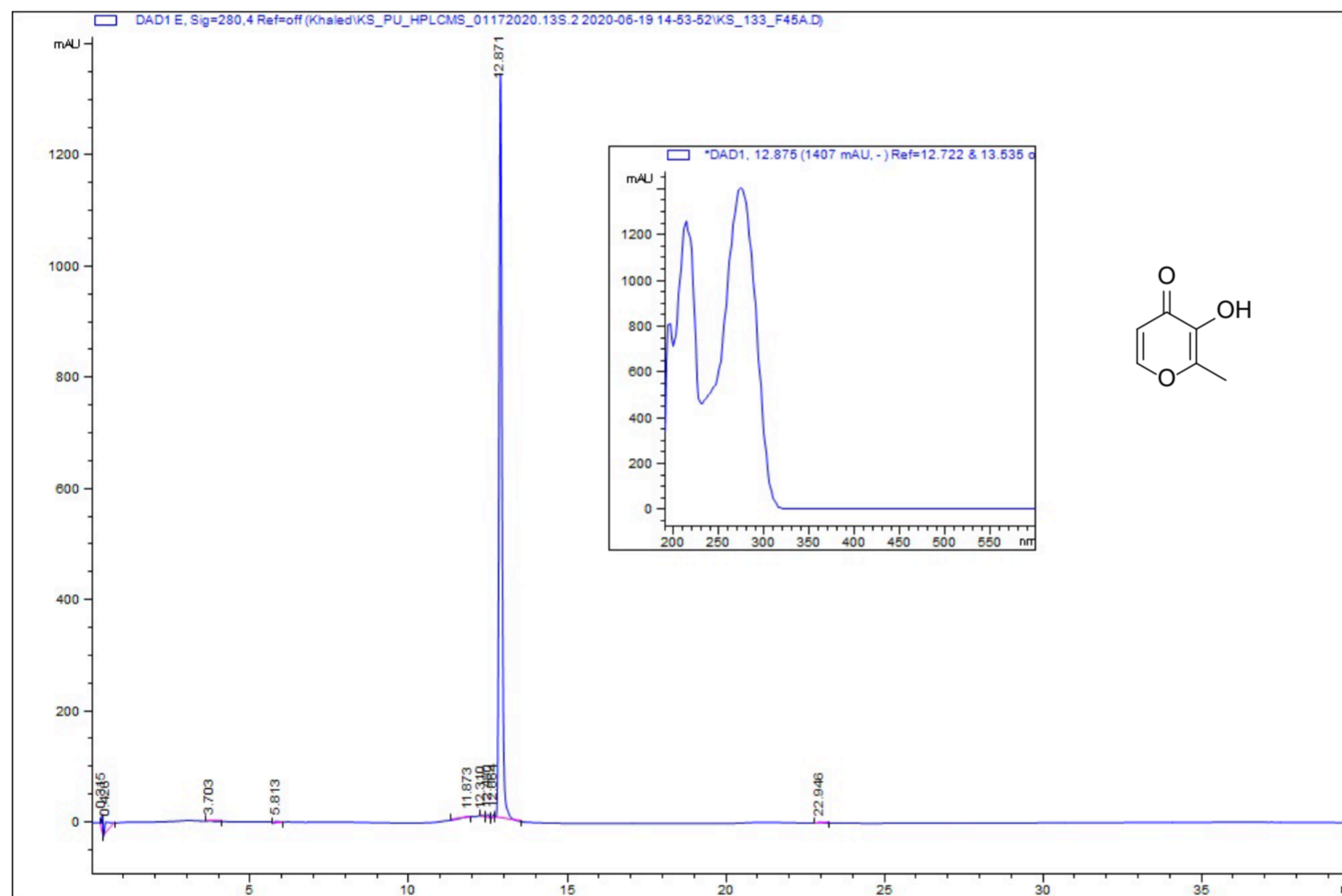
## Supplementary material



**Fig. 103S** NOESY spectrum (CD<sub>3</sub>OD, 400 MHz) of 5-carbomethoxymethyl-2-heptyl-7-hydroxychromone (**9**).

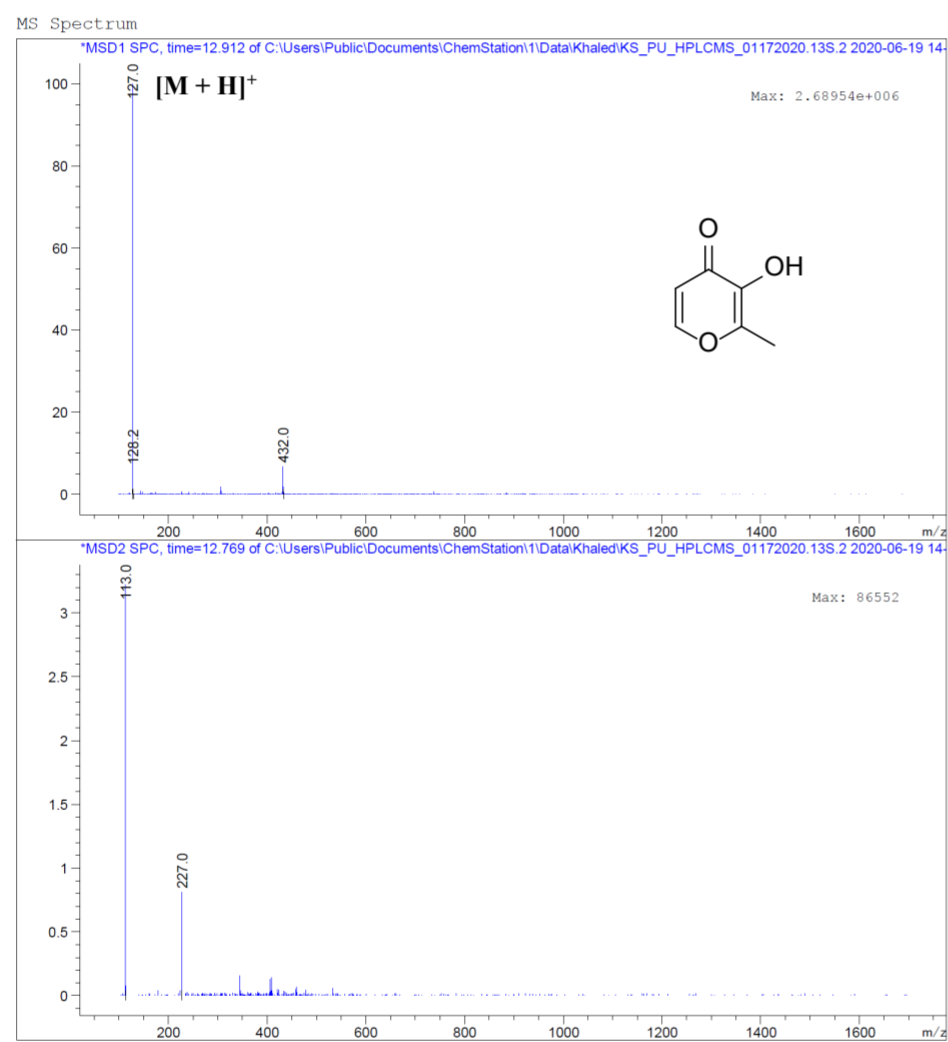


## Supplementary material



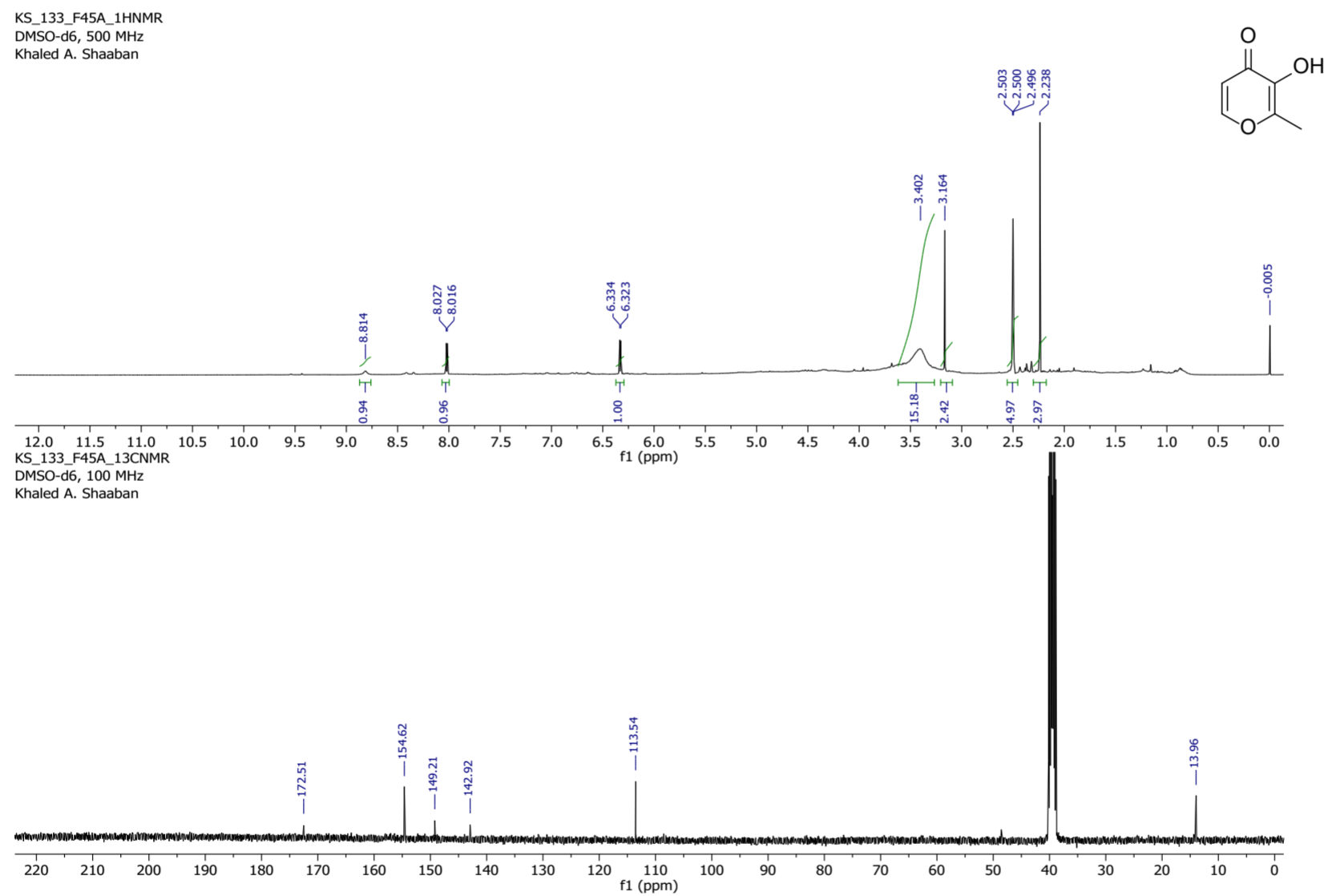
**Fig. 104S** HPLC analysis of maltol (**10**). HPLC conditions: solvent A: H<sub>2</sub>O/0.1% FA; solvent B: CH<sub>3</sub>CN; flow rate: 0.5 mL min<sup>-1</sup>; 0-30 min, 5-100% B; 30-35 min, 100% B; 35-36 min, 100-5% B; 36-40 min, 5% B; Phenomenex NX-C18 column (250 × 4.6 mm, 5 μm); 280 nm. UV-vis inset of full wavelength scan (190-600 nm).

## Supplementary material



**Fig. 105S** (+) and (-)-ESI-MS spectrum of maltol (**10**).

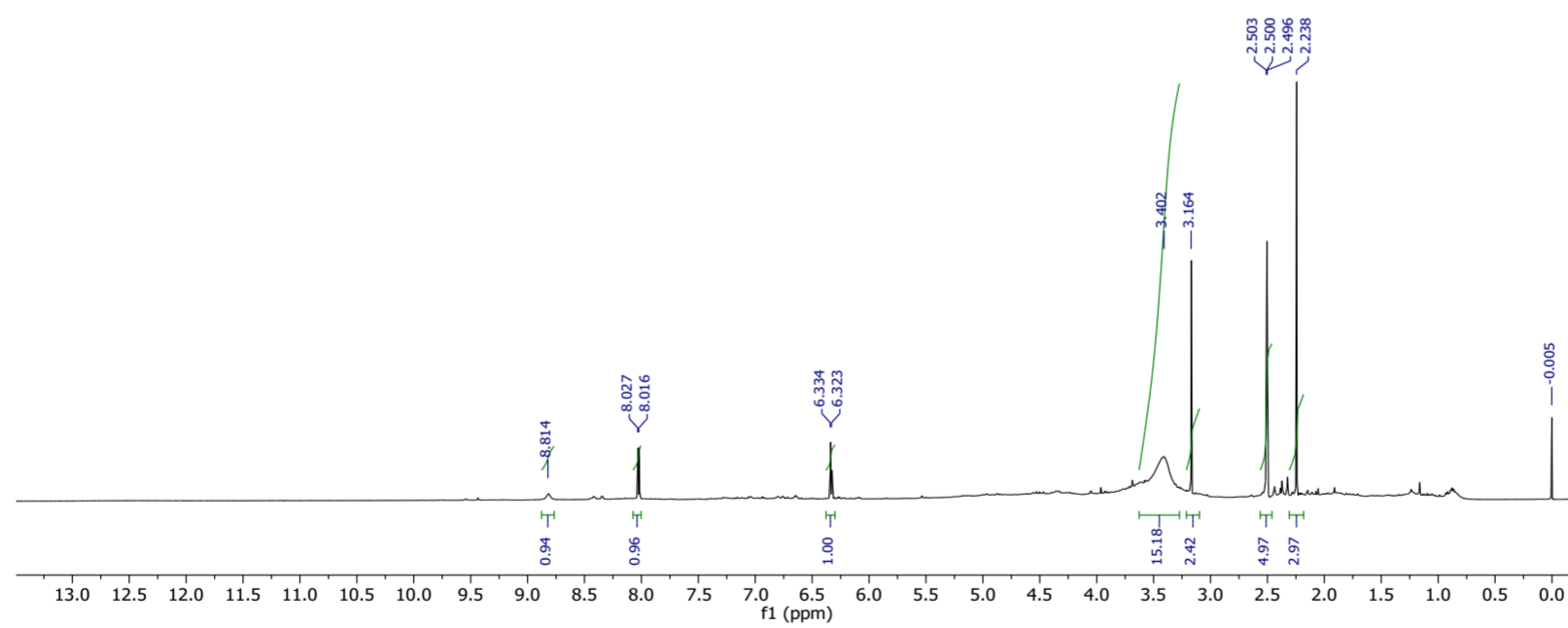
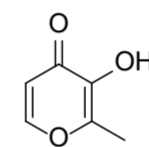
## Supplementary material



**Fig. 106S** <sup>1</sup>H (DMSO-*d*<sub>6</sub>, 500 MHz) and <sup>13</sup>C (DMSO-*d*<sub>6</sub>, 100 MHz) NMR spectrum of maltol (**10**).

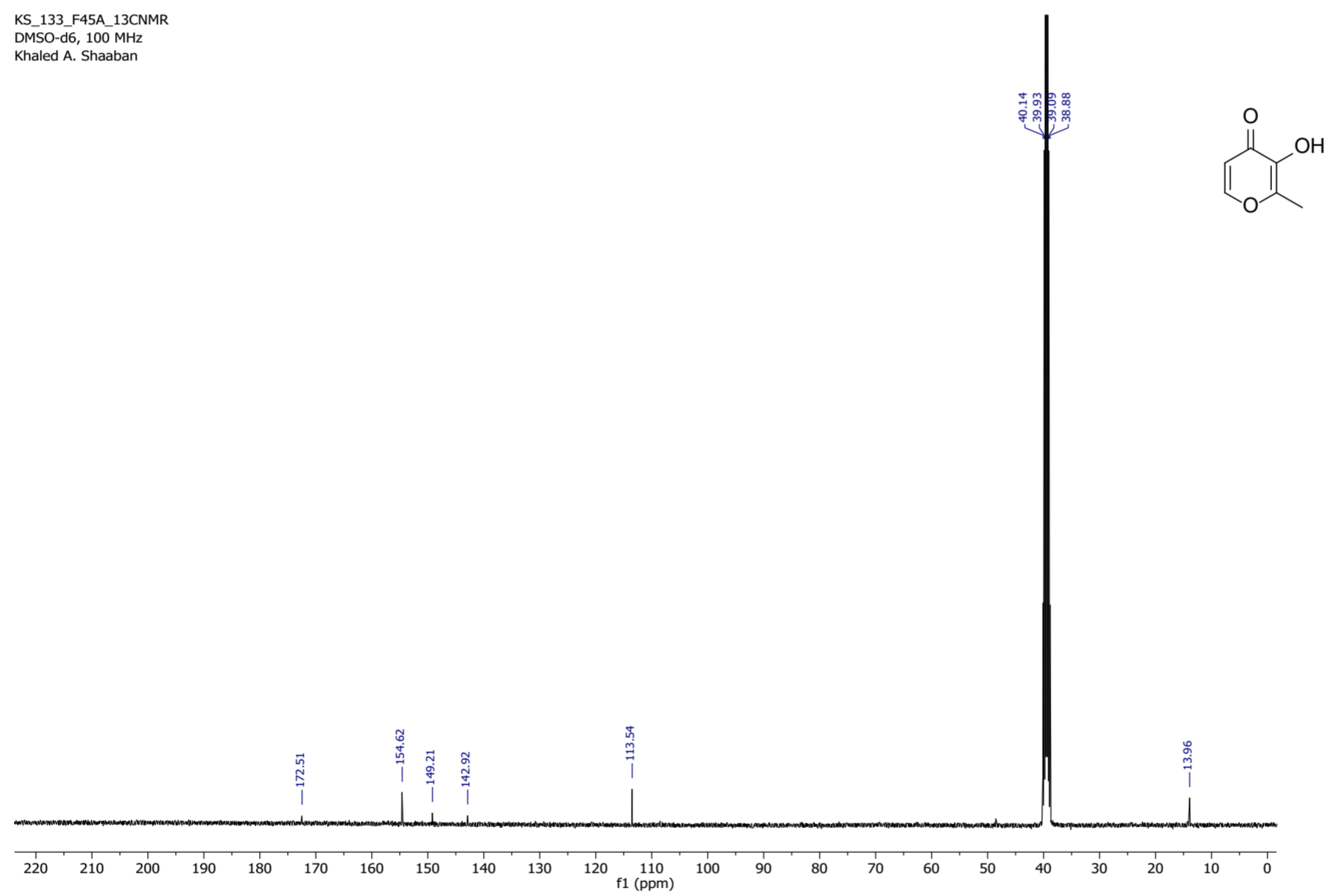
## Supplementary material

KS\_133\_F45A\_1HNMR  
DMSO-d6, 500 MHz  
Khaled A. Shaaban



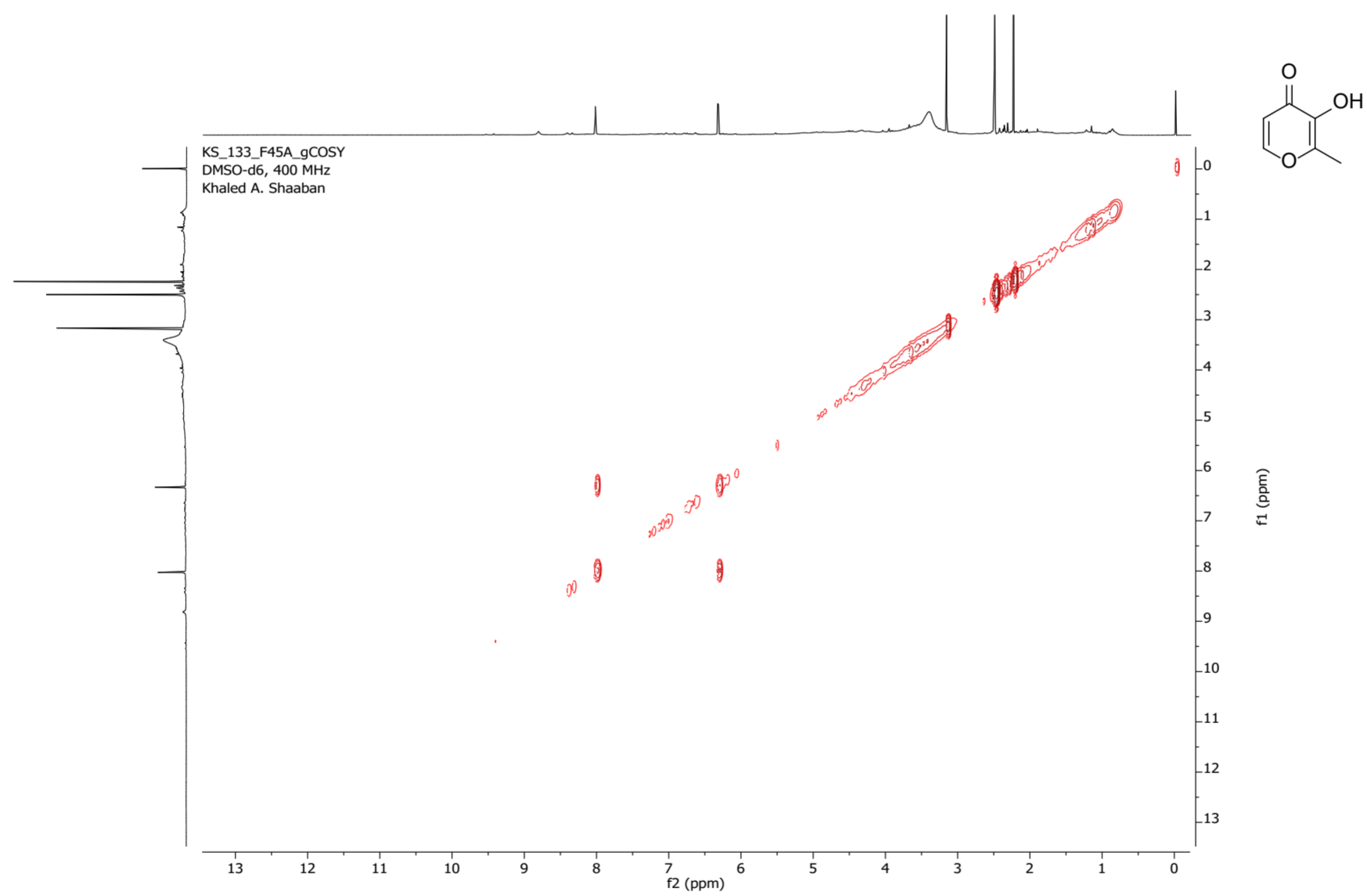
**Fig. 107S**  $^1\text{H}$  NMR spectrum (DMSO- $d_6$ , 500 MHz) of maltol (**10**).

## Supplementary material



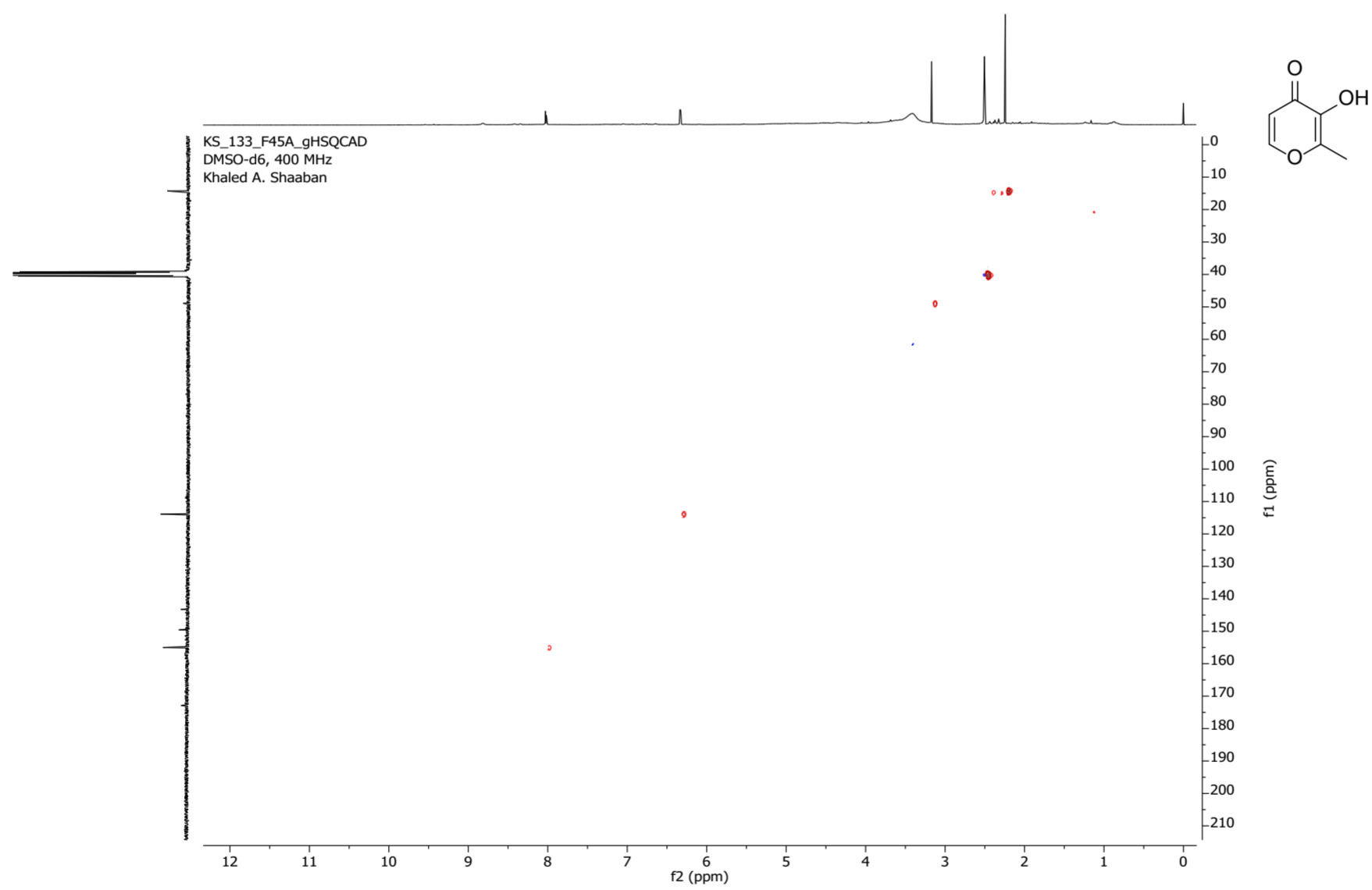
**Fig. 108S**  $^{13}\text{C}$  NMR spectrum (DMSO- $d_6$ , 100 MHz) of maltol (**10**).

## Supplementary material



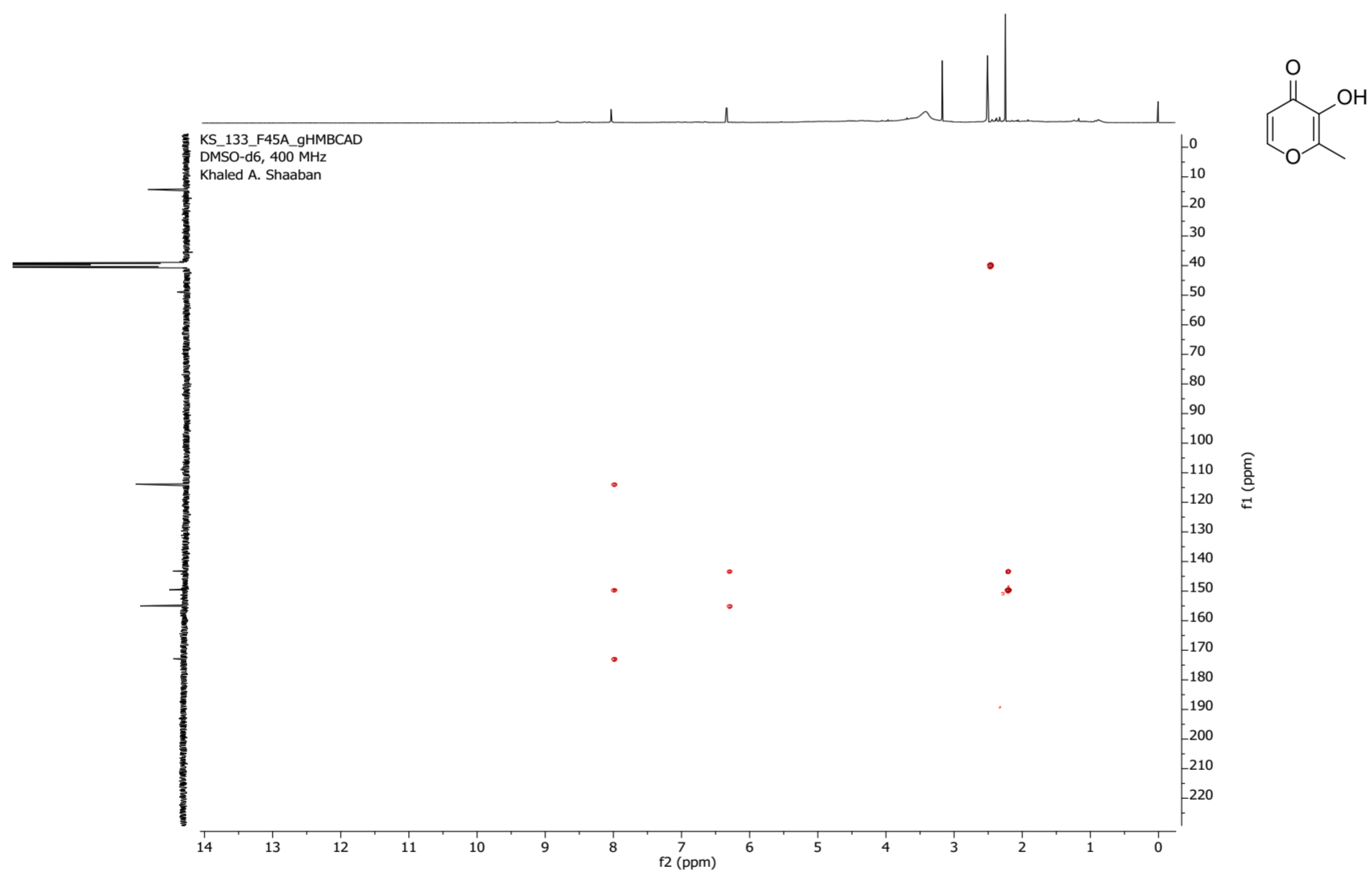
**Fig. 109S**  $^1\text{H},^1\text{H}$ -COSY spectrum (DMSO- $d_6$ , 400 MHz) of maltol (**10**).

## Supplementary material



**Fig. 110S** HSQC spectrum (DMSO-*d*<sub>6</sub>, 400 MHz) of maltol (**10**).

## Supplementary material



**Fig. 111S** HMBC spectrum (DMSO-*d*<sub>6</sub>, 500 MHz) of maltol (**10**).