

# **A missense variant in human perilipin 2 (*PLIN2* Ser251Pro) reduces hepatic steatosis in mice**

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## Table of contents

Supplementary materials.....	2
Fig. S1.....	4
Fig. S2.....	6
Fig. S3.....	8
Fig. S4.....	9
Fig. S5.....	10
Fig. S6.....	12
Table S1.....	13
Table S2.....	17
Table S3.....	18
Table S4.....	18
Table S5.....	18
Supplementary references .....	19

## **Supplementary materials:**

### ***Animal experiment***

Experiments were performed according to the protocols approved by the Institutional Animal Care and Use Committee of the University of Pennsylvania. All efforts were made to treat animals with humane care and minimize their discomfort. Mice were kept in barrier facilities (12-hour light/12-hour dark cycle) with *ad libitum* food and water, unless stated. Littermate controls were utilized.

*Adfp*D2–3 mice were provided by Dr. Palczewski. *Adfp*D2–3 mice lack exons 2 and 3 of the *Adfp* gene but do have mRNA expression of an unstable short-form *Adfp* (1). We crossed these mice from Balb/c to C57B6/J mice for more than ten generations. We called these mice *Plin2* KO-mice (1).

### ***Sphingolipid and ceramide analyses***

Liver tissue was homogenized in a RIPA buffer (50 mM Tris (pH 8.0), 150 mM NaCl, 5 mM EDTA, 1% v/v NP-40, 0.5% w/v sodium deoxycholate, 0.1% v/v SDS, 50 mM sodium fluoride). Protein concentrations were determined by BCA assay. Liver homogenate and plasma samples were analyzed by high performance liquid chromatography-tandem mass spectrometry for ceramide content at the Lipidomics Shared Resource at the Medical University of South Carolina. Hepatic ceramide data were normalized to total protein levels.

### ***Lipidomic***

#### ***Tissue preparation for lipids extraction***

Sections of approximately 10 mg of frozen tissues were cut on a tile kept in dry ice with a new blade kept in dry ice. The tissue was added to a microcentrifuge tube with 0.6 mL 80% methanol (MeOH) and 20  $\mu$ L on internal standard mix (1:1, SPLASH® LIPIDOMIX #330707 and Ceramide/Sphingoid Internal Standard Mixture I #LM6002, both from Avanti Polar Lipids, Alabaster, AL) and kept in dry ice. Samples were homogenized by pulse sonication (30 half-second pulse on ice) and incubated on ice for 20 min for metabolites extraction. Each tube was then vortexed 3x 30 seconds each. The tissue homogenates were then moved to a 10 mL glass borosilicate tube with a screw cap. The remaining tissue in the microcentrifuge tubes was rinsed into the glass tube with 0.5 mL MeOH. In each of the tubes, 5 mL methyl tert-butyl ether (MTBE) were added and then tubes were shaken vigorously for 30 min. In each tube, 1.2 mL water was added and then tubes vortexed for 30 sec each. Centrifugation for 10 min @ 1000xg created two phases. The top clear phase was moved to a clean glass Pyrex tube and dried down under

nitrogen. 100  $\mu$ L MTBE/MeOH=1/3 (v/v) was used to re-suspend the residue. The sample was spun down at 10,000  $\times$  g for 10 min at 4°C and only the top 50  $\mu$ L were transferred to a HPLC vial for LC-MS analysis. A pooled sample was created by mixing 10  $\mu$ L of each re-suspended sample. 2  $\mu$ L injections were made.

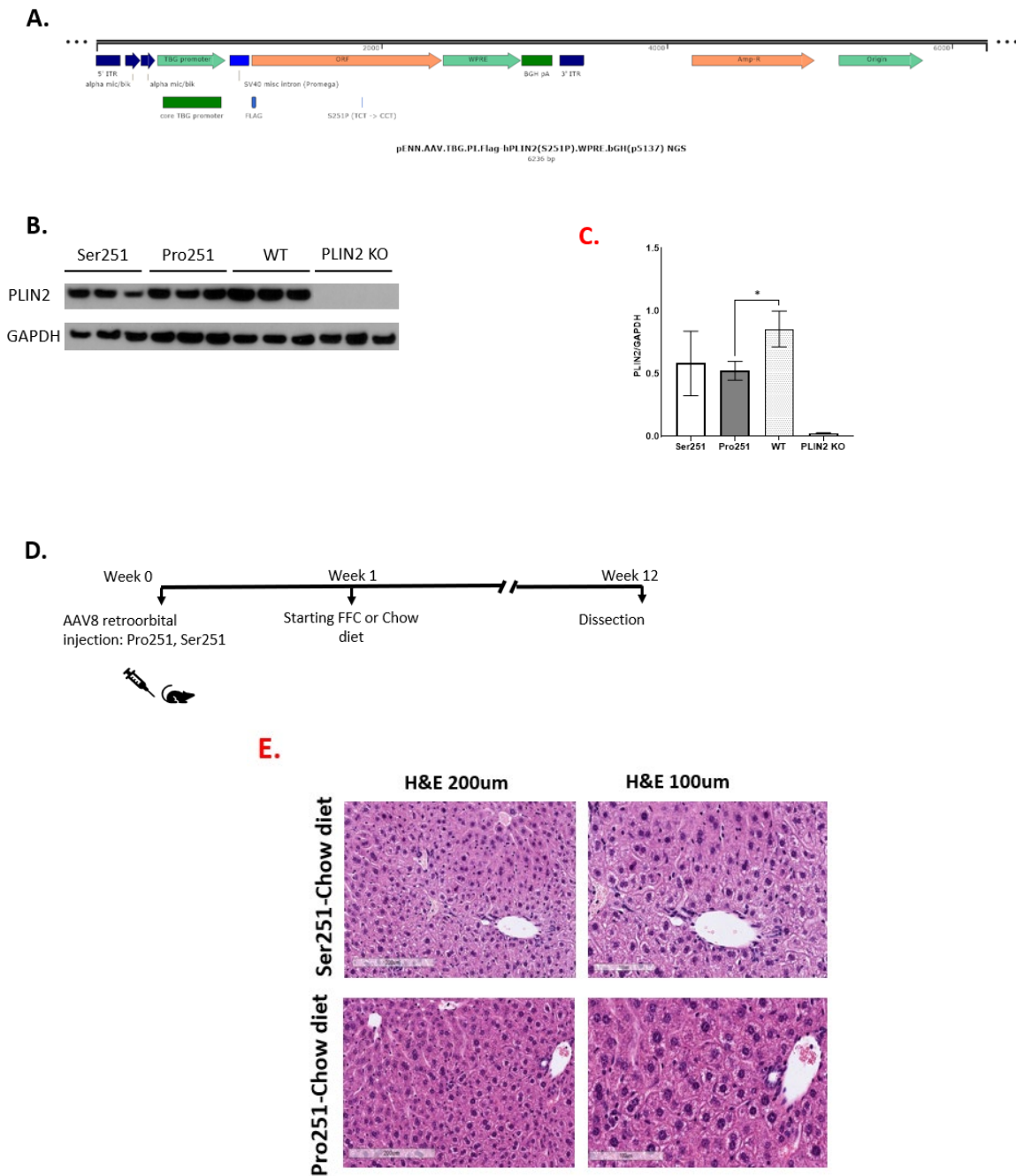
*Liquid chromatography high resolution -mass spectrometry (LC-HRMS) for lipids.*

Metabolites were separated using a Ascentis Express C18, 2.1  $\times$  150 mm 2.7 $\mu$ m column (Sigma-Aldrich, St. Louis, MO) on an UltiMate 3000 HPLC system. The metabolites were eluted on a 0.4 m/min flow-rate gradient using Solvent A (4:6 v/v water:acetonitrile, 0.1% formic acid, 10 mM ammonium formate) and Solvent B (1:9 v/v acetonitrile:isopropanol, 0.1% formic acid, 10 mM ammonium formate). The gradient was as follows: 10% B at 0 min, 10% B at 1 min, 40% B at 4 min, 75% B at 12 min, 99% B at 21 min, 99% B at 24 min, 10% B at 24.5 min, 10% at 30 min. Separations were performed at 55 °C.

For the HRMS analysis, a recently calibrated QE Exactive-HF mass spectrometer (Thermo Fisher Scientific) was used in positive ion mode with an HESI source. The operating conditions were: spray voltage at 3.5 kV; capillary temperature at 285°C; auxiliary temperature 370°C; tube lens 45. Nitrogen was used as the sheath gas at 45 units, the auxiliary gas at 10 units and sweep gas was 2 units. The same MS conditions were used in negative ionization mode, but with a spray voltage at 3.2 kV. Control extraction blanks were made in the same way using just the solvents instead of the tissue homogenate. The control blanks were used for the exclusion list with a threshold feature intensity set at  $1 \times 10^5$ . Untargeted analysis and targeted peak integration was conducted using LipidsSearch 4.2 (Thermo Fisher Scientific) as described by Wang et al ( DOI: 10.4155/bio-2021-0098).

An external mass calibration was performed using the standard calibration mixture approximately every three days. All samples were analyzed in a randomized order in full scan MS that alternated with MS<sub>2</sub> of top 20, with HCD scans at 30, 45 or 60 eV. Full scan resolution was set to 120,000 in the scan range between m/z 250–1800. The pool sample was run every 15 samples. Lipids quantification was done from the full scan data. The areas were normalized based on the amount of the internal standard added for each class (as found in the Table x1). All amounts were normalized to the original tissue weight.

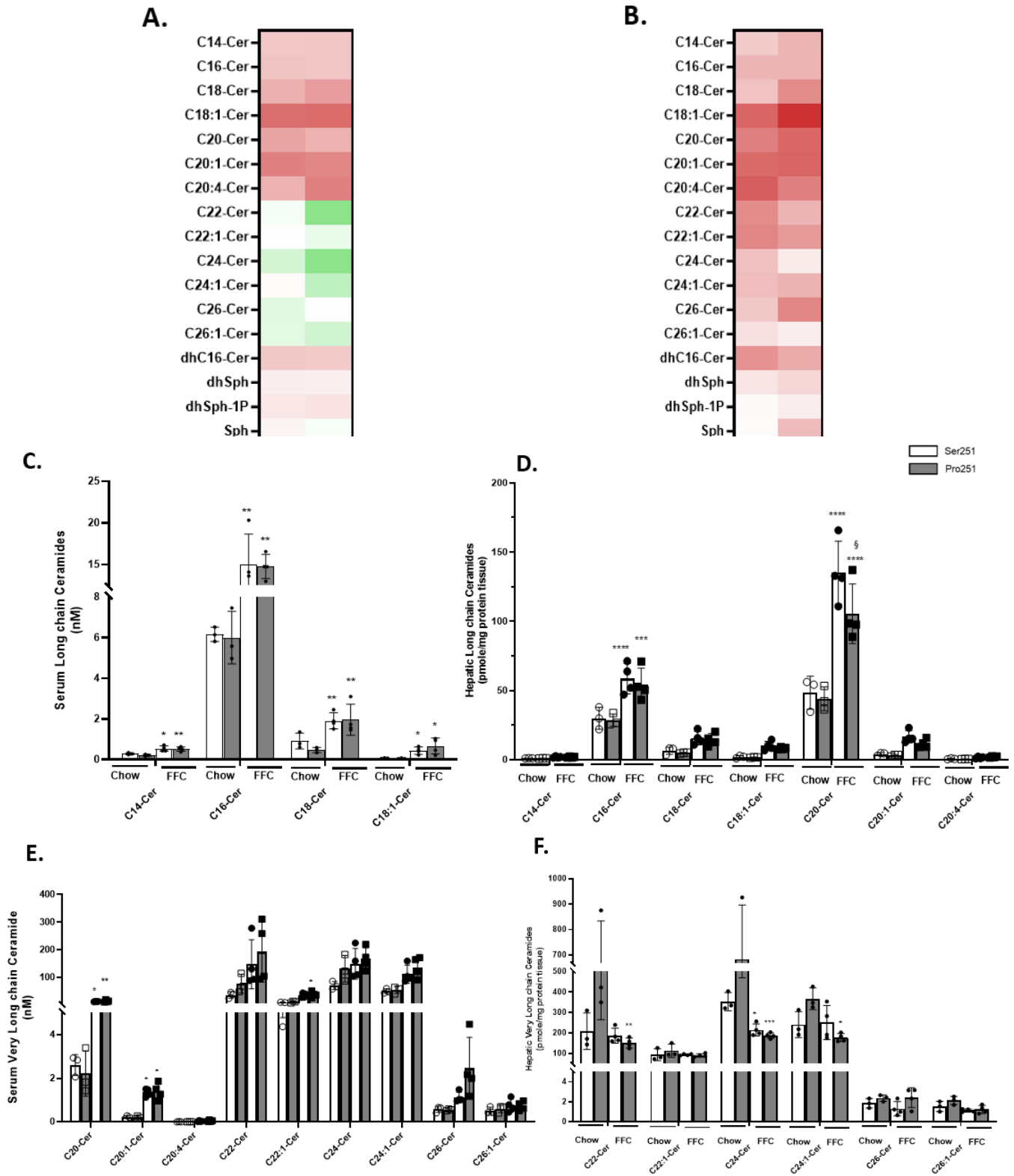
**Fig. S1**



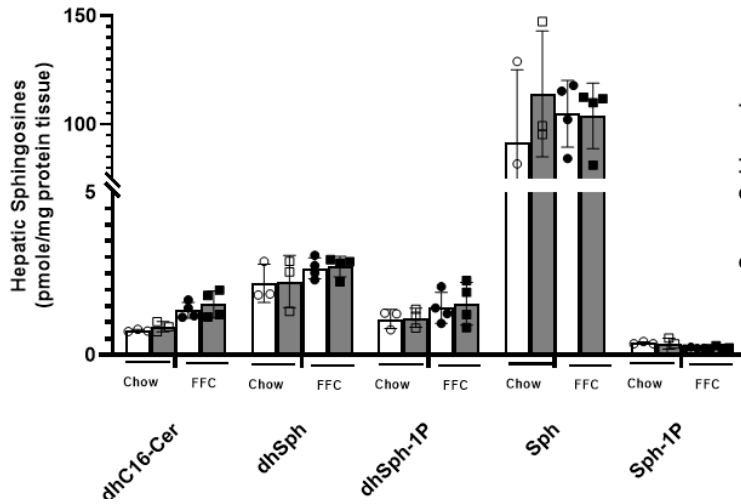
Generation of a mouse model of *Plin2*-Pro251 in hepatocytes and experimental design. Map of the *Plin2*-Pro251 plasmid (A), hepatic PLIN2 protein and mRNA expression (B-C), study design (D). Liver histology for the Pro251 and Ser251 groups on chow diet (E). PLIN2 relative protein

levels by Western Blotting were normalized to GAPDH (B and C). Statistical analyses were performed using a two-tailed unpaired *t*-test or one-way ANOVA with Tukey's posthoc test. \*Indicates statistical difference between diets, \*P < 0.05; \*\*P < 0.005; \*\*\*P < 0.0005; \*\*\*\*P < 0.0001. §Indicates statistical difference between genotypes, §P < 0.05; §§P < 0.005; §§§P < 0.0005; §§§§P < 0.0001.

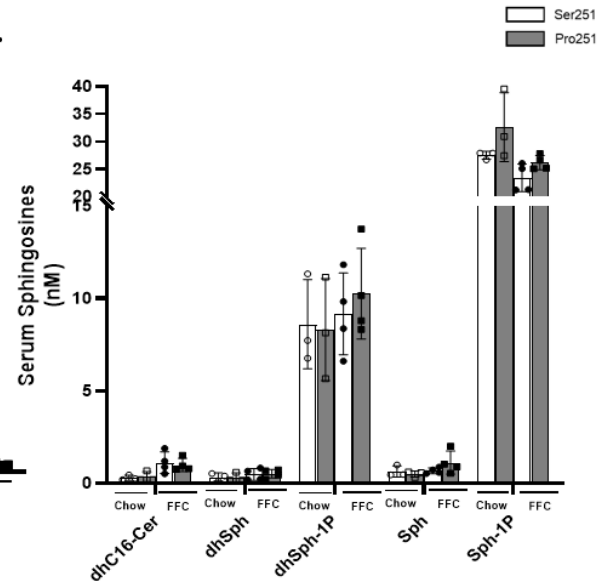
**Fig. S2**



G.



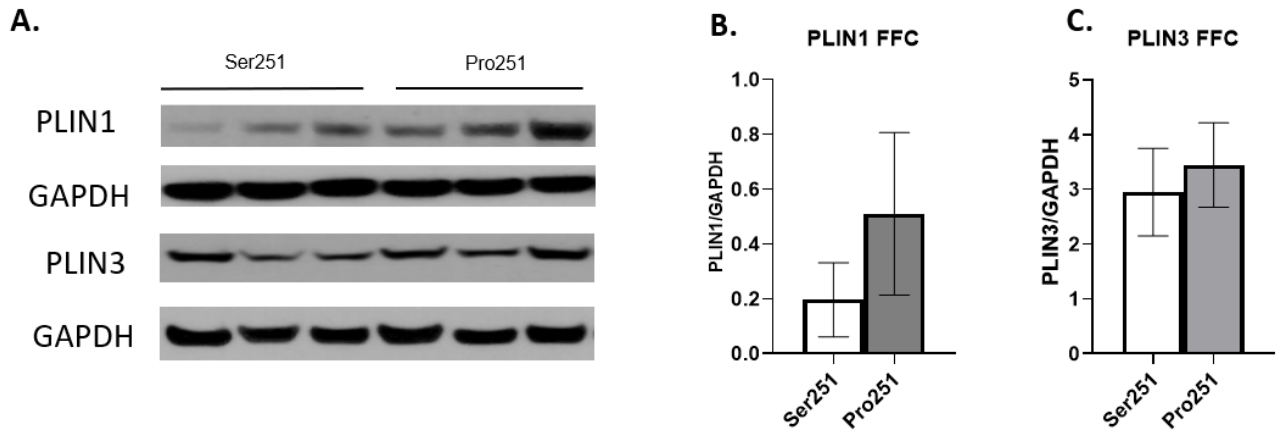
H.



In the heatmap, the color code indicates the log<sub>2</sub> of the ratio between means of the groups for an individual ceramide. A more intense red color indicates a greater increase of absolute concentration of the individual ceramide in the liver (A) and serum (B). Serum long chain ceramides (C), hepatic long chain ceramides (D), serum very long chain ceramides (E), hepatic very long chain ceramides (F), hepatic sphingolipids (G) and serum sphingolipids (H). Statistical analyses were performed using a two-tailed unpaired *t*-test or one-way ANOVA with Tukey's posthoc test. \*Indicates statistical difference between diets, \**P* < 0.05; \*\**P* < 0.005; \*\*\**P* < 0.0005; \*\*\*\**P* < 0.0001. §Indicates statistical difference between genotypes, §*P* < 0.05; §§*P* < 0.005; §§§*P* < 0.0005; §§§§*P* < 0.0001.

**Fig. S3**

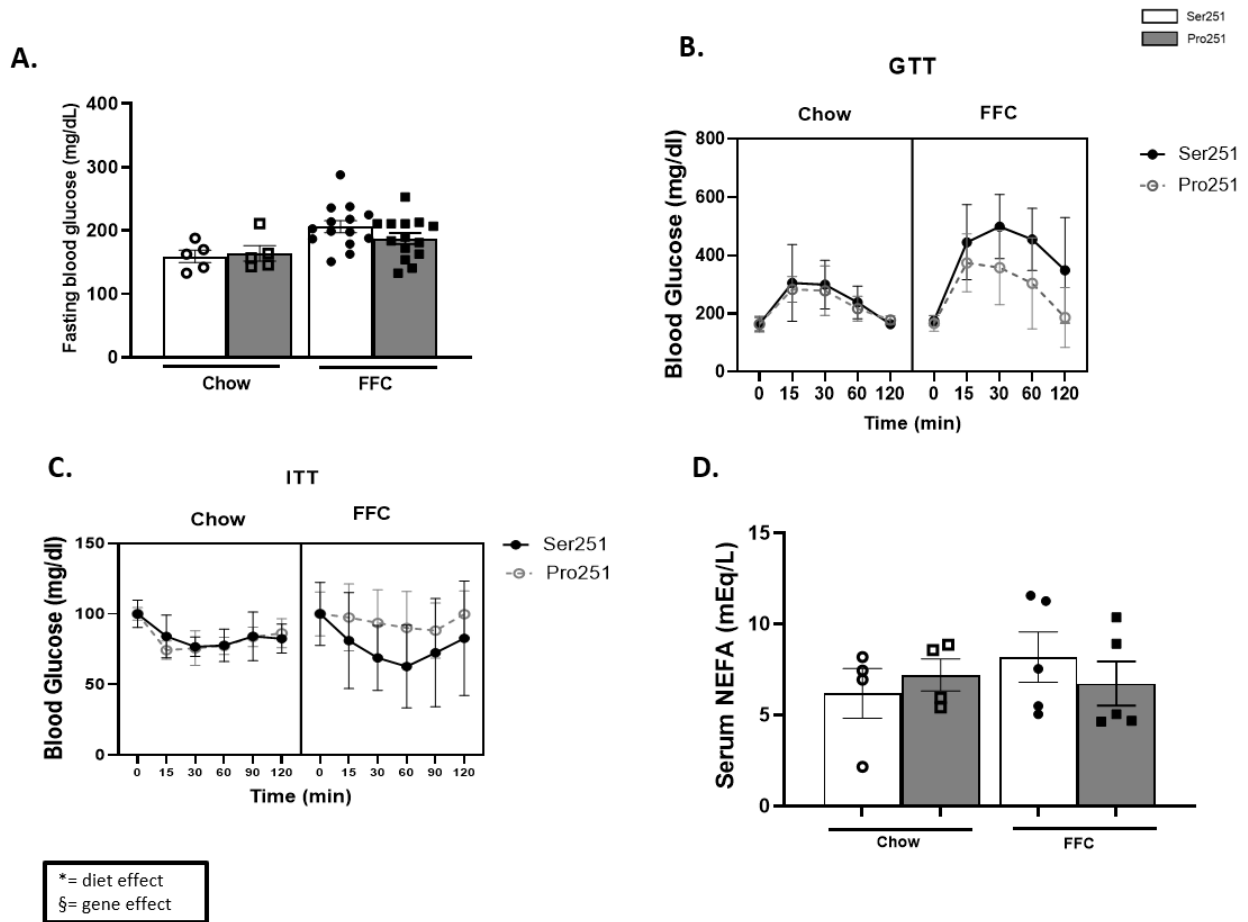
PLIN1, PLIN3 relative protein levels by Western Blotting were normalized to GAPDH (B and C).



Statistical analyses were performed using a two-tailed unpaired *t*-test or one-way ANOVA with Tukey's posthoc test. \*Indicates statistical difference between diets, \* $P < 0.05$ ; \*\* $P < 0.005$ ; \*\*\* $P < 0.0005$ ; \*\*\*\* $P < 0.0001$ . §Indicates statistical difference between genotypes, § $P < 0.05$ ; §§ $P < 0.005$ ; §§§ $P < 0.0005$ ; §§§§ $P < 0.0001$ .



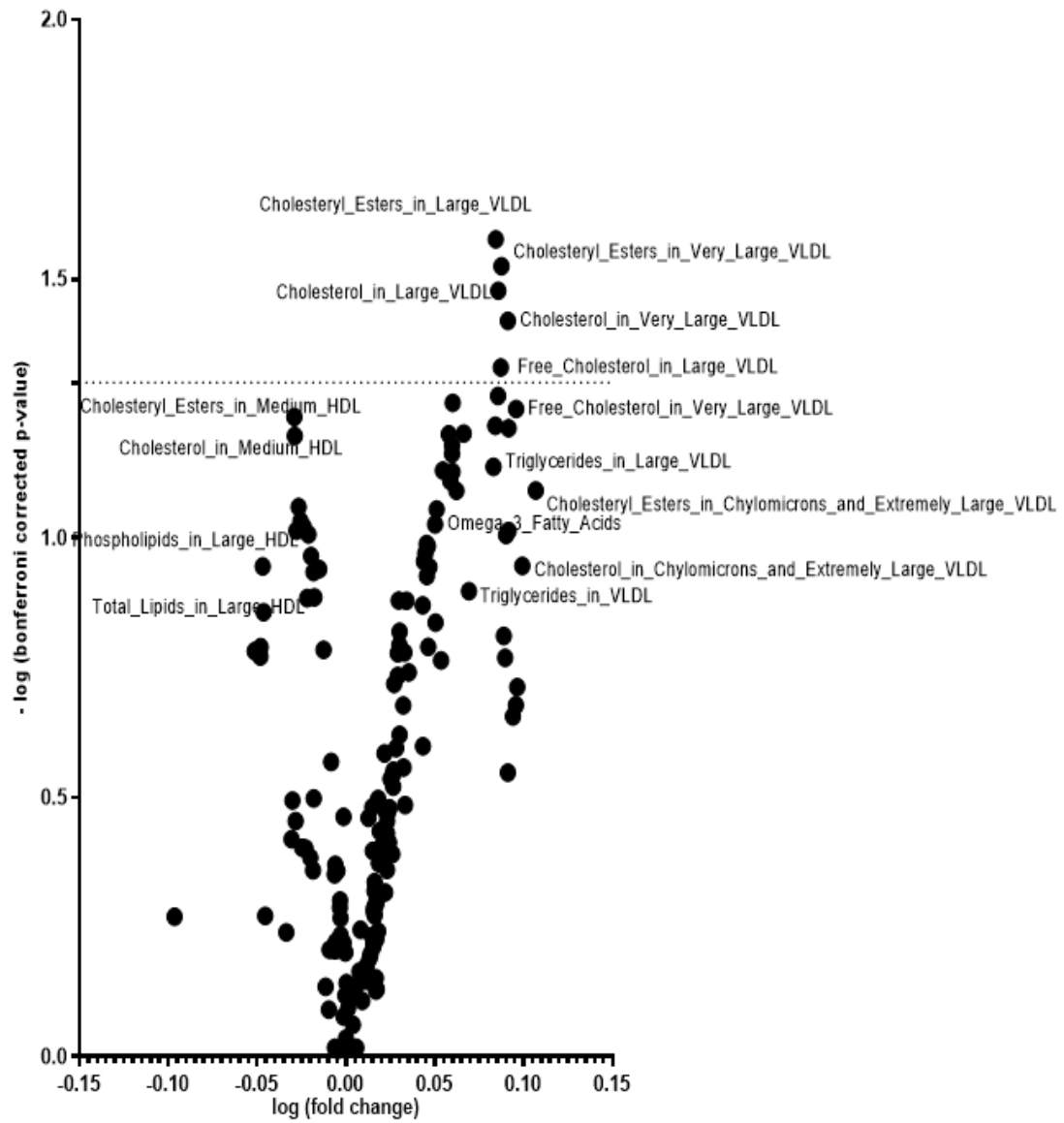
**Fig. S4**



Fasting blood glucose (A), GTT (B), ITT (C), and serum NEFA (D). Statistical analyses were performed using a two-tailed unpaired *t*-test or one-way ANOVA with Tukey's posthoc test. \*Indicates statistical difference between diets, \**P* < 0.05; \*\**P* < 0.005; \*\*\**P* < 0.0005; \*\*\*\**P* < 0.0001. §Indicates statistical difference between genotypes, §*P* < 0.05; §§*P* < 0.005; §§§*P* < 0.0005; §§§§*P* < 0.0001.

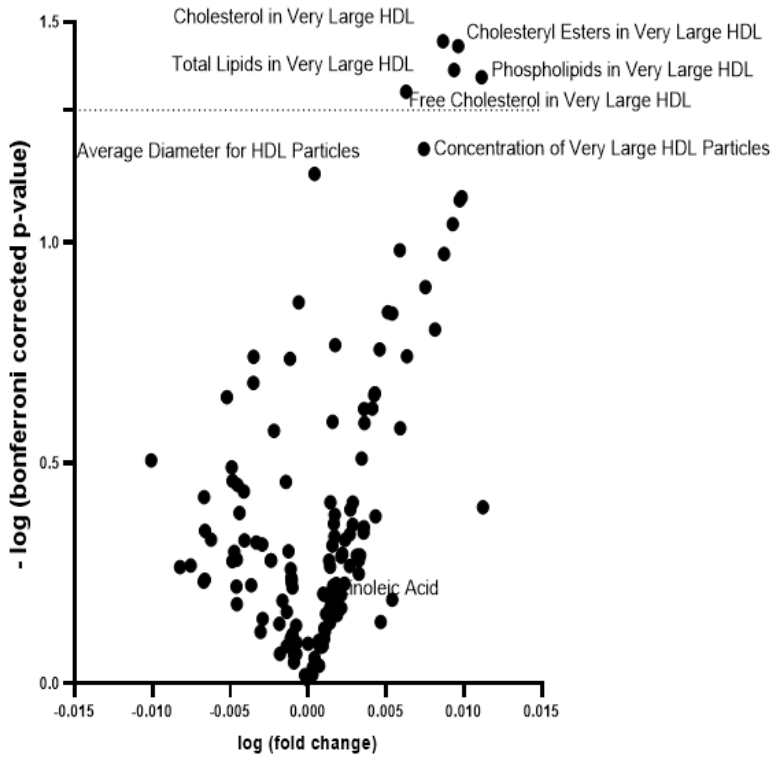
Fig. S5.

A



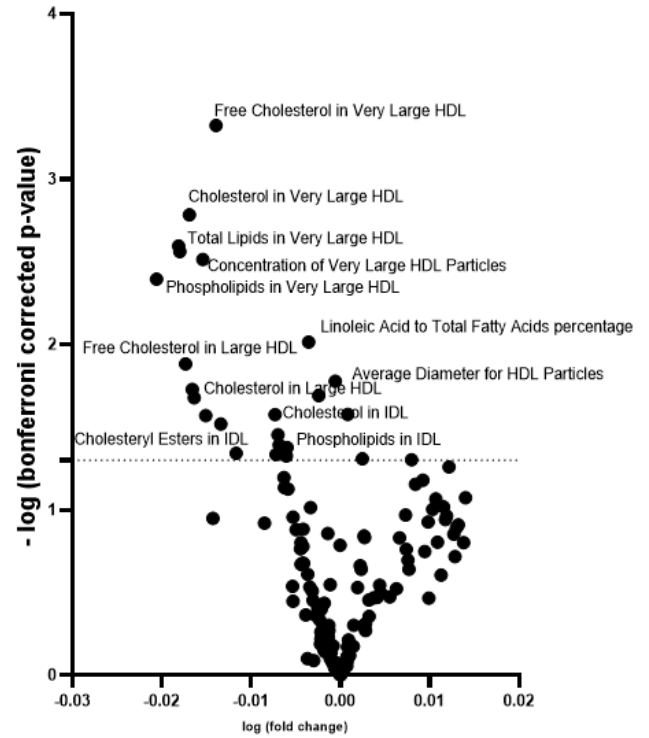
Homozygous vs non carriers

**B.**



**Male homozygous vs non carriers**

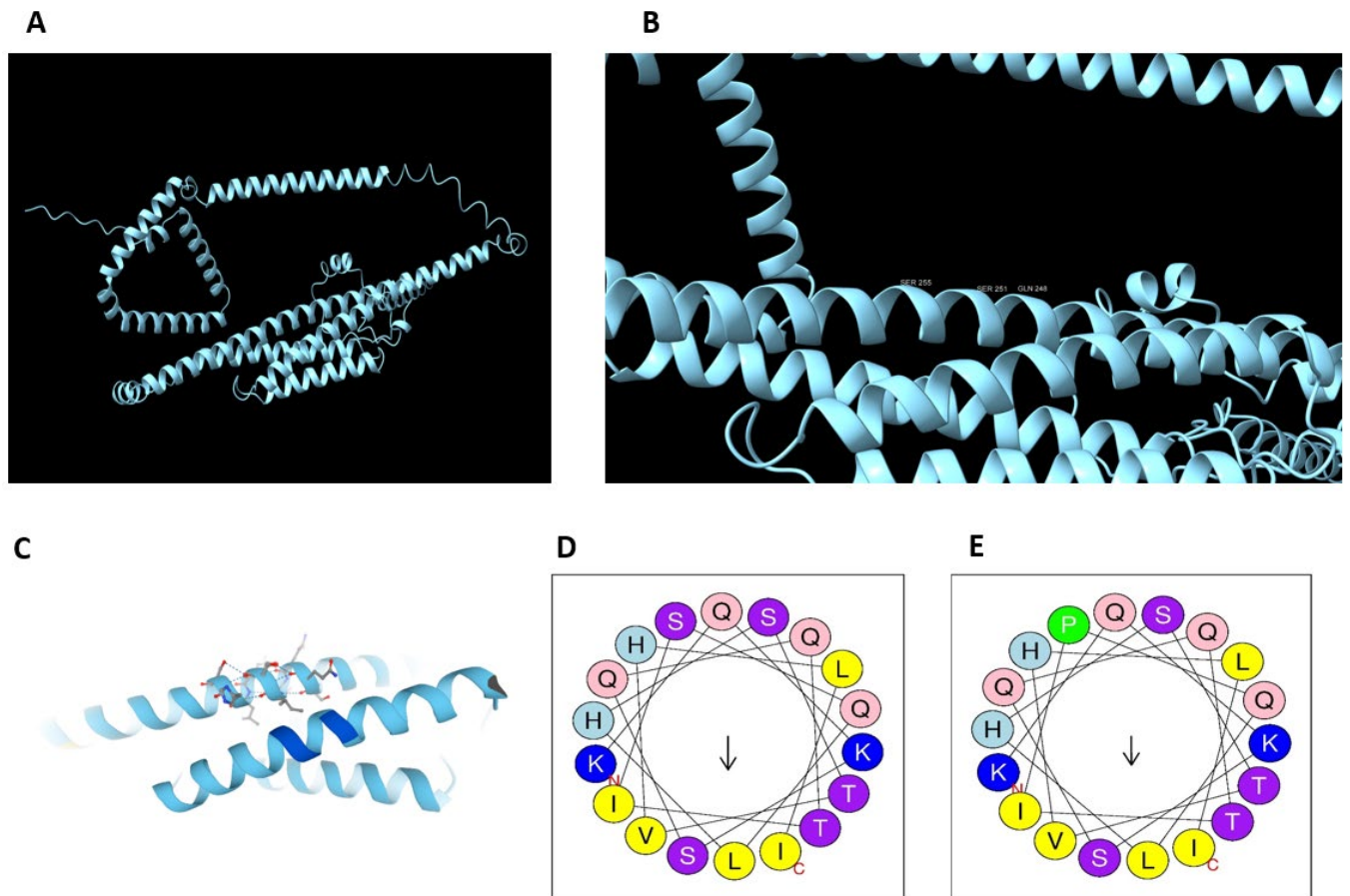
**C.**



**Female homozygous vs non carriers**

Volcano plot for the lipidomic parameters derived from the UKB database (A), volcano plot for the lipidomic parameters in male (B), volcano plot for the lipidomic parameters in female (C). The x-axis displays the log fold change, comparison between Homozygous Pro251 carriers and non-carriers. The dotted line marks the Bonferroni significant.

Fig. S6



Structure of human perilipin 2 (437 amino acids) and predicted by AlphaFold (A); Expanded view of PLIN2 structure showing that S251 is situated in the 4-helix bundle domain and in the middle of a long helix that spans residues from T225 to L281 (B); Further expansion of structure to show hydrogen bonding interactions of S251 (C). Helical wheel of residues spanning from K243 (labeled with a red N) to I260 (labeled with a red C) in wild-type human PLIN2 (D); Helical wheel of residues spanning from K243 to I260 in the S251P variant of human PLIN2 (E).

**Table S1**

Change in mice liver triglyceride species in both chow and FFC diet groups. Comparison between *Pro251* and *Ser251*.

Triglycerides	Chow diet		FFC diet	
	Log2 (Fold change)	P value	Log2 (Fold change)	P value
TG(12:0e_6:0_20:4)	-0.4	0.252	-1.62	0.128
TG(12:1e_6:0_22:6)	-0.4	0.302	-1.43	0.155
TG(14:0_18:2_18:3)	-0.9	0.132	-1.95	0.077
TG(14:0_22:6_22:6)	-1.2	0.005	-1.03	0.085
TG(15:0_16:0_18:1)	-1.4	0.094	-1.60	0.077
TG(15:0_16:1_18:1)	-0.5	0.245	-2.14	0.056
TG(15:0_16:1_18:2)	-1.0	0.126	-2.07	0.066
TG(15:0_18:1_18:2)	-1.0	0.063	-1.91	0.109
TG(15:0_18:1_20:5)	-1.6	0.205	-2.14	0.236
TG(15:0_18:1_22:6)	-1.0	0.034	-0.90	0.205
TG(15:0_18:2_18:2)	-0.8	0.047	-1.81	0.116
TG(15:0_18:2_18:3)	-1.0	0.113	-0.89	0.608
TG(15:0_18:2_20:5)	-1.1	0.220	-2.37	0.105
TG(15:0_18:2_20:5)	-0.4	0.499	-1.55	0.222
TG(15:0_18:2_22:6)	-0.8	0.031	-1.19	0.109
TG(15:0_22:6_22:6)	-1.2	0.006	-0.83	0.202
TG(16:0_10:0_18:2)	-1.1	0.046	-1.63	0.184
TG(16:0_12:1_18:1)	-0.5	0.359	-2.29	0.056
TG(16:0_12:1_18:2)	-0.1	0.771	-1.79	0.164
TG(16:0_14:0_16:0)	-1.0	0.337	-1.94	0.066
TG(16:0_14:0_18:1)	-1.3	0.178	-1.99	0.046
TG(16:0_14:0_18:2)	-0.5	0.536	-2.20	0.072
TG(16:0_14:0_22:6)	-1.2	0.028	-1.59	0.050
TG(16:0_14:2_22:6)	-1.5	0.158	-1.99	0.096
TG(16:0_16:0_16:0)	-1.2	0.165	-1.56	0.055
TG(16:0_16:0_17:0)	-1.1	0.141	-0.78	0.213
TG(16:0_16:0_18:1)	-1.3	0.067	-1.90	0.094
TG(16:0_16:0_24:0)	-0.8	0.314	-0.39	0.415
TG(16:0_16:1_16:1)	-0.6	0.469	-2.26	0.067
TG(16:0_16:1_18:1)	-1.2	0.134	-2.11	0.149
TG(16:0_16:1_18:3)	-1.2	0.176	-1.94	0.089
TG(16:0_17:0_18:1)	-1.3	0.060	-1.42	0.120
TG(16:0_17:1_18:1)	-1.5	0.069	-1.60	0.092
TG(16:0_18:1_18:1)	-1.9	0.008	0.78	0.593

TG(16:0_18:1_18:1)	-1.2	0.033	-1.65	0.100
TG(16:0_18:1_18:1)	-0.8	0.386	1.90	0.467
TG(16:0_18:1_18:2)	-0.2	0.581	-2.88	0.301
TG(16:0_18:1_19:0)	-0.9	0.186	-1.49	0.090
TG(16:0_18:1_20:4)	-1.1	0.040	-1.13	0.051
TG(16:0_18:1_21:0)	-1.5	0.173	-1.11	0.096
TG(16:0_18:1_22:0)	-0.9	0.217	-0.83	0.227
TG(16:0_18:1_22:6)	-0.9	0.013	-1.24	0.111
TG(16:0_18:1_23:0)	-0.7	0.340	-0.63	0.287
TG(16:0_18:1_24:0)	-0.7	0.339	-0.72	0.346
TG(16:0_18:2_18:3)	0.1	0.860	-3.25	0.215
TG(16:0_18:3_22:6)	0.1	0.899	-1.91	0.055
TG(16:0_20:5_22:6)	-0.9	0.030	-1.47	0.078
TG(16:0_22:6_22:6)	-1.4	0.026	-1.16	0.103
TG(16:0_6:0_18:1)	-0.5	0.643	-1.36	0.221
TG(16:1_12:1_18:1)	-1.0	0.106	-1.87	0.113
TG(16:1_14:0_18:2)	-1.4	0.081	-2.41	0.053
TG(16:1_14:1_18:2)	-0.3	0.355	-2.20	0.059
TG(16:1_16:1_18:1)	-1.0	0.112	-1.91	0.150
TG(16:1_18:1_18:1)	-0.9	0.030	-1.65	0.065
TG(16:1_18:1_18:2)	-1.1	0.008	-1.32	0.361
TG(16:1_18:2_18:2)	2.4	0.001	-1.11	0.190
TG(16:1_18:2_18:2)	-1.3	0.027	-2.29	0.090
TG(16:1_18:2_18:3)	-0.8	0.060	-1.63	0.111
TG(16:1_18:2_18:3)	0.2	0.780	-1.53	0.225
TG(16:1_18:3_18:3)	-0.9	0.053	-4.11	0.051
TG(17:0_18:1_18:1)	-0.8	0.110	-5.65	0.137
TG(17:0_18:1_20:4)	-1.1	0.017	-1.21	0.062
TG(17:0_18:1_20:5)	-1.1	0.059	-1.62	0.083
TG(17:0_18:1_22:5)	-0.9	0.055	0.21	0.795
TG(17:0_18:1_22:6)	-0.8	0.006	0.11	0.787
TG(17:0_20:5_20:5)	-1.5	0.022	-1.36	0.071
TG(17:0_22:6_22:6)	-0.9	0.006	-2.57	0.089
TG(18:0_16:0_16:0)	-1.4	0.152	-1.32	0.046
TG(18:0_16:0_17:0)	-1.0	0.137	-0.87	0.102
TG(18:0_16:0_18:1)	-1.3	0.055	-1.48	0.076
TG(18:0_18:0_18:0)	-0.8	0.294	-0.69	0.285
TG(18:0_18:0_20:4)	-2.0	0.015	-1.09	0.059
TG(18:0_18:1_18:1)	-1.5	0.015	-1.63	0.098
TG(18:0_18:1_22:6)	-1.2	0.001	-1.13	0.173
TG(18:0_20:4_22:6)	-0.8	0.008	-0.26	0.692
TG(18:0_22:6_22:6)	-0.8	0.070	-1.46	0.004

TG(18:0_22:6_22:6)	-1.4	0.229	-0.92	0.185
TG(18:0e_12:4_12:4)	-0.1	0.763	-1.40	0.310
TG(18:1_14:0_14:0)	-1.1	0.225	-2.19	0.049
TG(18:1_17:1_18:1)	-1.2	0.015	-1.53	0.205
TG(18:1_17:1_18:2)	-1.6	0.057	-1.71	0.126
TG(18:1_17:1_22:5)	-0.5	0.491	-1.45	0.258
TG(18:1_17:1_22:6)	-2.0	0.043	-1.13	0.092
TG(18:1_18:1_18:1)	-1.5	0.008	-1.84	0.087
TG(18:1_18:1_18:1)	0.4	0.728	-2.58	0.358
TG(18:1_18:1_18:2)	-1.0	0.001	-1.62	0.164
TG(18:1_18:1_18:2)	-0.9	0.361	-3.10	0.300
TG(18:1_18:1_18:3)	-0.9	0.003	-1.50	0.229
TG(18:1_18:1_20:3)	-0.9	0.004	-0.77	0.330
TG(18:1_18:1_20:3)	-1.5	0.035	-1.36	0.053
TG(18:1_18:1_20:4)	-1.1	0.025	-1.76	0.071
TG(18:1_18:1_20:4)	-1.9	0.074	-1.11	0.254
TG(18:1_18:1_20:5)	-1.0	0.009	-1.37	0.138
TG(18:1_18:1_21:0)	-1.1	0.124	-0.72	0.347
TG(18:1_18:1_21:1)	-1.2	0.051	-1.71	0.059
TG(18:1_18:1_22:0)	-1.5	0.078	-1.04	0.225
TG(18:1_18:1_22:1)	-1.6	0.148	-1.37	0.117
TG(18:1_18:1_22:3)	0.1	0.785	-1.61	0.001
TG(18:1_18:1_22:4)	-1.2	0.008	-1.34	0.065
TG(18:1_18:1_22:5)	-1.4	0.008	-1.51	0.106
TG(18:1_18:1_22:6)	-1.9	0.036	-1.16	0.221
TG(18:1_18:1_23:0)	-0.8	0.273	-0.44	0.450
TG(18:1_18:1_23:1)	-0.7	0.455	-0.92	0.232
TG(18:1_18:1_24:1)	-1.6	0.050	-1.23	0.217
TG(18:1_18:2_18:3)	-0.5	0.412	-1.04	0.197
TG(18:1_18:2_20:4)	-1.0	0.008	-1.61	0.171
TG(18:1_18:2_20:4)	0.1	0.828	-1.30	0.118
TG(18:1_18:2_20:5)	-0.4	0.431	-1.13	0.191
TG(18:1_18:2_21:1)	-1.3	0.012	-1.59	0.111
TG(18:1_18:2_22:1)	-1.6	0.024	-1.72	0.141
TG(18:1_18:2_22:5)	-1.5	0.000	-1.04	0.130
TG(18:1_18:2_22:6)	-1.0	0.001	-1.26	0.156
TG(18:1_18:2_24:1)	-1.2	0.058	-1.07	0.249
TG(18:1_20:3_22:6)	-1.2	0.019	-1.22	0.079
TG(18:1_20:4_22:6)	-1.3	0.002	-0.78	0.171
TG(18:1_20:5_22:6)	-1.1	0.013	-0.85	0.205
TG(18:1_22:0_22:6)	-0.8	0.157	0.80	0.359
TG(18:1_22:1_22:6)	-1.4	0.024	0.90	0.464

TG(18:1_22:5_22:5)	-0.7	0.363	-0.92	0.060
TG(18:1_22:5_22:6)	-1.4	0.006	-0.31	0.629
TG(18:1_22:6_22:6)	-1.2	0.001	-0.78	0.223
TG(18:2_17:1_18:2)	-0.9	0.033	-1.64	0.151
TG(18:2_17:1_18:2)	0.9	0.327	-1.66	0.016
TG(18:2_17:1_20:4)	-0.9	0.072	-1.88	0.110
TG(18:2_17:1_22:6)	-0.9	0.033	-1.22	0.119
TG(18:2_18:2_18:2)	-1.1	0.000	-1.81	0.219
TG(18:2_18:2_20:4)	-0.8	0.007	-1.08	0.172
TG(18:2_18:2_22:6)	-1.0	0.002	-1.19	0.067
TG(18:2_18:2_22:6)	-0.9	0.007	-1.05	0.256
TG(18:2_22:6_22:6)	-1.0	0.014	-0.71	0.347
TG(18:2_22:6_22:6)	-0.8	0.143	-0.67	0.536
TG(18:2+5O_16:0_16:0)	-0.7	0.008	-1.98	0.103
TG(18:3_17:1_18:2)	-0.9	0.154	-1.48	0.094
TG(18:3_18:2_18:2)	-1.2	0.161	-3.45	0.324
TG(18:3_18:2_18:2)	0.5	0.455	-2.06	0.241
TG(18:3_18:2_18:3)	-0.7	0.036	-1.50	0.276
TG(18:3_18:2_18:3)	-0.3	0.593	-2.34	0.074
TG(18:3_18:2_20:5)	-0.7	0.046	-1.49	0.140
TG(18:3_18:2_22:6)	-0.6	0.042	-1.38	0.192
TG(18:3_18:3_18:3)	-0.5	0.381	-1.47	0.114
TG(18:3_22:6_22:6)	-0.8	0.018	-0.58	0.396
TG(18:3+5O_16:0_16:0)	-0.7	0.011	-1.82	0.203
TG(18:4_16:1_16:1)	-0.9	0.188	-1.97	0.066
TG(18:4_16:1_18:2)	-0.2	0.818	-1.37	0.080
TG(18:4_18:1_18:1)	-1.2	0.006	-1.41	0.096
TG(19:0_18:1_18:1)	-1.3	0.057	-1.29	0.110
TG(19:0_18:1_20:4)	-1.0	0.025	-0.20	0.710
TG(19:1_16:0_18:1)	-1.5	0.037	-1.37	0.102
TG(19:1_18:1_18:1)	-2.4	0.083	-1.55	0.089
TG(19:1_18:1_18:2)	-1.2	0.004	-1.49	0.110
TG(19:1_18:2_18:2)	-1.0	0.185	-1.68	0.113
TG(19:1_18:2_22:6)	-1.4	0.004	-0.82	0.319
TG(19:1_20:5_20:5)	-1.5	0.029	-1.01	0.168
TG(20:0_16:0_16:0)	-1.2	0.196	-0.75	0.140
TG(20:0_16:0_18:1)	-1.2	0.159	-1.29	0.136
TG(20:0_18:1_18:1)	-1.7	0.031	-1.32	0.140
TG(20:0_18:1_22:6)	-1.0	0.021	-0.74	0.344
TG(20:1_18:1_18:1)	-3.6	0.004	-1.70	0.098
TG(20:1_18:1_18:1)	-0.7	0.124	1.57	0.099
TG(20:1_18:1_18:2)	-1.0	0.003	-0.20	0.875



TG(20:1_18:1_18:2)	-1.1	0.004	-1.63	0.137
TG(20:1_18:1_18:2)	-1.0	0.028	-1.36	0.107
TG(20:1_18:1_22:5)	-1.2	0.009	-1.29	0.090
TG(20:1_18:2_22:6)	-1.4	0.061	-1.17	0.067
TG(20:3_18:2_22:6)	-1.4	0.002	-1.18	0.164
TG(20:4e_18:1_18:1)	0.7	0.120	-1.26	0.245
TG(20:5_17:1_18:2)	-2.0	0.017	-1.47	0.007
TG(20:5_18:2_18:2)	-0.9	0.023	-1.49	0.130
TG(20:5_18:2_20:5)	-0.5	0.254	-1.50	0.252
TG(20:5_18:2_22:5)	-0.9	0.014	-1.29	0.206
TG(20:5_18:2_22:6)	-0.5	0.183	-1.25	0.257
TG(22:1_18:2_22:6)	-0.3	0.518	1.58	0.050
TG(22:5_17:1_18:2)	-1.1	0.004	-2.00	0.117
TG(22:5_18:2_18:2)	-1.3	0.001	-0.92	0.184
TG(22:5_18:2_22:6)	-1.2	0.001	-1.01	0.381
TG(22:6_17:1_22:6)	-0.9	0.005	-2.65	0.337
TG(22:6_17:1_22:6)	-0.9	0.073	-0.77	0.486
TG(24:0_18:2_18:2)	-0.3	0.448	-0.35	0.563
TG(28:0_10:4_18:2)	-0.5	0.117	-1.95	0.200
TG(30:0_18:1_18:1)	-1.6	0.184	-0.51	0.521
TG(4:0_16:0_16:0)	-1.1	0.380	-0.68	0.505
TG(4:0_16:0_18:1)	-1.0	0.439	-0.55	0.628
TG(4:0_16:0_18:2)	-0.9	0.441	-0.64	0.684
TG(4:0_18:1_18:2)	-1.2	0.378	-0.88	0.464

Log2 fold change for each triglyceride (TG). Bonferroni corrected/adjusted p value, by dividing the original  $\alpha$ -value (0.05) by the number of analyses on the dependent variable.

**Table S2**

**PLIN2 rs35568725 prevalence in the MVP population (Caucasic, African American, Hispanic)**

Population	Reference	Alternative	MAF	Phenotype	Beta	SE	P
Trans-ethnic	A	G	0.051	ALT	0.005	0.003	0.142
White	A	G	0.055	ALT	0.004	0.003	0.250
Black	A	G	0.013	ALT	0.014	0.013	0.276
Hispanic	A	G	0.028	ALT	0.006	0.015	0.690
Asians	A	G	0.020	ALT	0.031	0.052	0.559
Trans-ethnic	A	G	0.052	NAFLD	0.021	0.015	0.160

White	A	G	0.055	NAFLD	0.031	0.016	0.051
Black	A	G	0.013	NAFLD	-0.068	0.070	0.330
Hispanic	A	G	0.029	NAFLD	-0.089	0.070	0.203
Asians	A	G	0.022	NAFLD	0.120	0.226	0.597

Additive logistic model corrected for age, sex, and the first ten ethnicity-specific principal components.

**Table S3**

**Association between PLIN2-Pro251 and type 2 diabetes, dyslipidemia, hypertension, and NAFLD in the MVP database (before Bonferroni correction)**

Phenotype	Beta	SE	P	OR	95% Low	95% High
NAFLD	0.003	0.003	0.25	1.00	0.99	1.01
T2D	0.06	0.02	0.002	1.07	1.02	1.11
Cardiomyopathy	0.07	0.03	0.016	1.08	1.01	1.15

Multivariable analysis adjusted for age, gender, and 5 PC principal components European Ancestry

**Table S4**

**Hepatic liver fat after adjustment for visceral adipose tissue:**

Phenotype	Beta	SE	P-value
Liver volume	-2.812e-03	3.667e-03	0.443
MRI-PDFF	-0.0973086	0.0732235	0.183
Liver iron	1.505257	0.866653	0.082

Multivariable analyses adjusted for age, sex, BMI, visceral adipose tissue, and PC1-10.

**Table S5**

**Magnetic resonance imaging-derived proton density fat fraction (MRI-PDFF) in BMI categories:**

**BMI: <18.5**

Phenotype	Beta	SE	P-value
MRI-PDFF	0.334	0.186	0.075
Visceral adipose tissue	-0.007	0.160	0.962

Multivariable analyses adjusted for age, sex, and PC1-10.

**BMI: from 18.5 to 25**

Phenotype	Beta	SE	P-value
MRI-PDFF	0.032	0.067	0.630
Visceral adipose tissue	-6.542e-17	2.791e-16	0.814

Multivariable analyses adjusted for age, sex, and PC1-10.

**BMI: from 25 to 30**

Phenotype	Beta	SE	P-value
MRI-PDFF	-0.212	0.118	0.071
Visceral adipose tissue	1.840e-16	1.286e-16	0.152

Multivariable analyses adjusted for age, sex, and PC1-10.

**BMI: >30**

Phenotype	Beta	SE	P-value
MRI-PDFF	-0.203	0.255	0.425
Visceral adipose tissue	-1.494e-16	1.432e-16	0.296

Multivariable analyses adjusted for age, sex, and PC1-10.

### ***Supplementary reference***

1. Carr RM, Peralta G, Yin X, Ahima RS. Absence of perilipin 2 prevents hepatic steatosis, glucose intolerance and ceramide accumulation in alcohol-fed mice. *PLoS one*. 2014;9(5):e97118.