

Appendix Figures Table of Contents

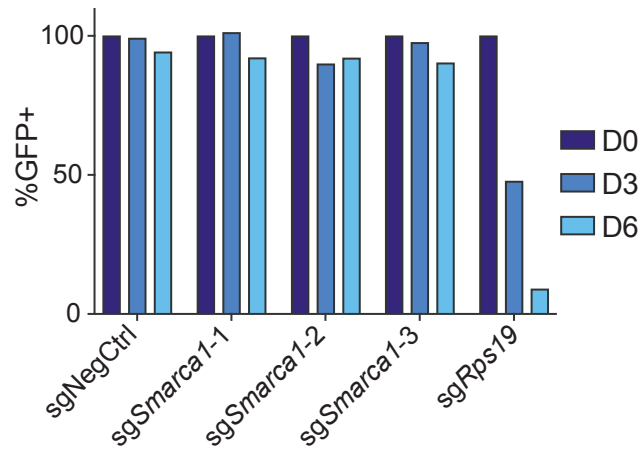
Appendix Figure S1. BPTF and SMARCA5 are essential for maintaining AML in vivo (page 2).

Appendix Figure S2. BPTF and SMARCA5 bind to active promoters in U937 and OCI-AML2 cells (page 3).

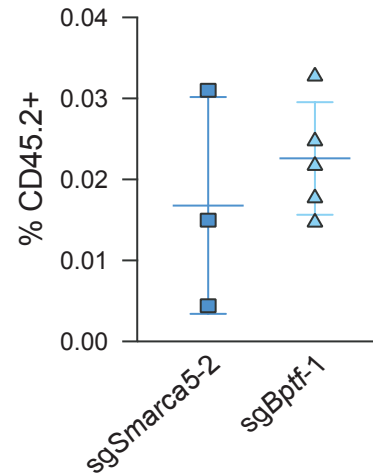
Appendix Figure S3. BPTF and SMARCA5 together remodel insulator regions in AML cells (page 4).

Appendix Figure S1

A



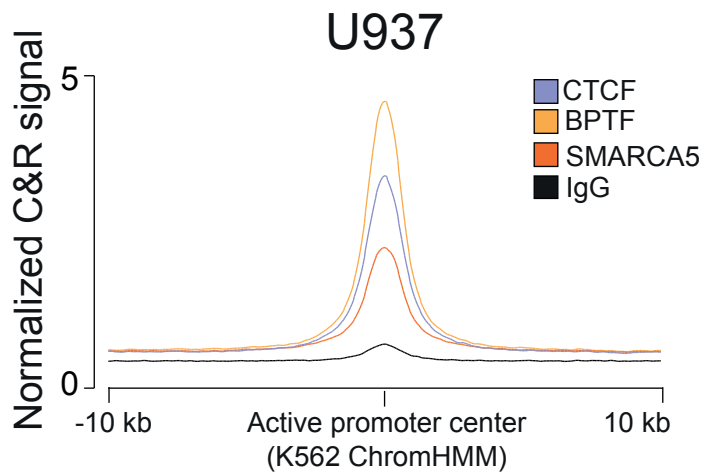
B



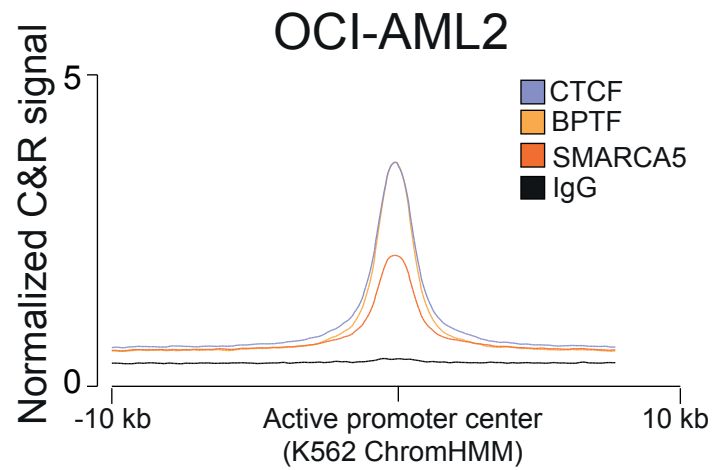
Appendix Figure S1. BPTF and SMARCA5 are essential for maintaining AML in vivo. A) Competition assay after *Smarca1* KO in mouse MLL-AF9 cells with inducible wtCas9 expression. The cells were transduced with the lentiviral cassettes expressing the corresponding sgRNAs and GFP and mixed with untransduced cells. The percentage of GFP-positive cells was measured over time. The data were normalized to Day 0. The results were validated in human AML cells, see Source data. B) Percentages of CD45.2+ (donor) cells present in the blood of the 8 surviving mice before experiment termination. Data are shown as mean \pm SD.

Appendix Figure S2

A

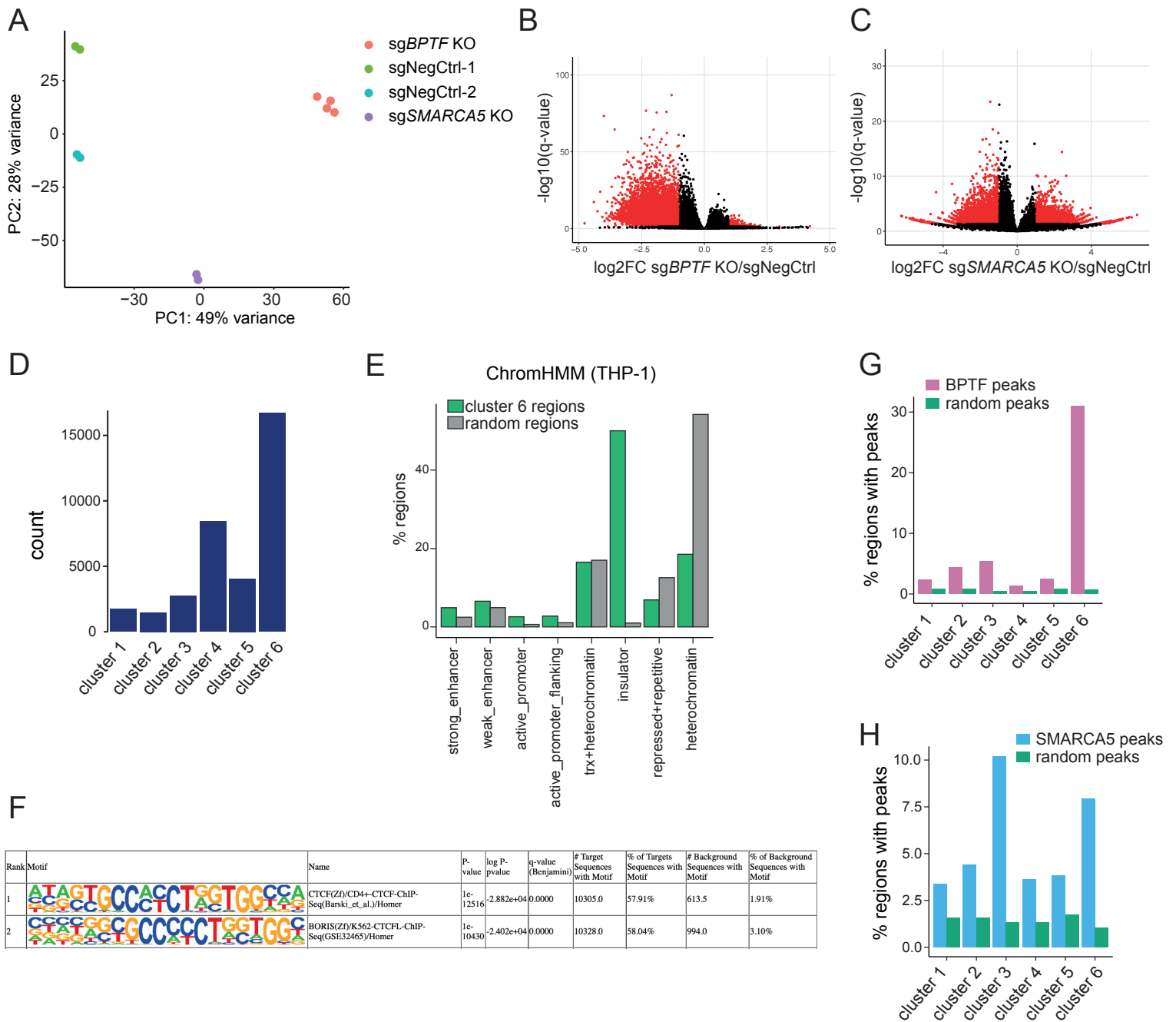


B



Appendix Figure S2. BPTF and SMARCA5 bind active promoter regions in U937 and OCI-AML2 cells. A-B) Average profiles of the CTCF, BPTF, and SMARCA5 Cut&Run signal in U937 (A) and OCI-AML2 (B) cells at the insulator regions predicted by the ChromHMM in K562 cells.

Appendix Figure S3



Appendix Figure S3. BPTF and SMARCA5 together remodel insulator regions in AML cells. A) PCA plot of the ATAC-seq samples. B-C) Volcano plots comparing ATAC-seq changes between the *BPTF* (B) or *SMARCA5* (C) KOs and wild-type cells. The peaks with an absolute log₂ fold change over 1 and a q-value < 0.01 are highlighted in red. D) Number of regions in the differential ATAC-seq clusters described in Figure 4F. E) Bar plot illustrating the percentage of regions in cluster 6 overlapping chromatin domain categories defined by the ChromHMM in THP1 cells. A corresponding set of random regions of the same average length and size was generated and shown in green. Some regions may overlap several categories; hence, the total percentage does not equal 100. ‘trx’ stands for transcription. F) Two top hits of the motif analysis in the differential ATAC-seq cluster 6. G-H) Bar plots illustrating the percentage of regions in differential ATAC-seq clusters overlapping with BPTF (G) or HA-SMARCA5 (H) peaks. A corresponding set of random regions of the same average lengths and sizes was generated and shown in green. All the ATAC-seq experiments were performed in two biological replicates, i.e., independent cell transductions or treatments.