Supplemental Materials:

Comparison of phage-derived recombinases in genetic manipulation

of Pseudomonas species

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- **Table S1.** Rif^R mutations in *Pseudomonas*.
- Table S2. Overview of select recombineering efforts to date in Pseudomonas spp.
- Figure S1. Effect of wash buffer on transformation efficiency.
- **Figure S2.** Structure and location of Rif^R binding pocket.
- Figure S3. Growth defect of *rpoB* Q518L mutants in *P. protegens* CHA0.
- Figure S4. Effect of phosphorothiolation of oligonucleotide.
- Figure S5. Multiple sequence alignment of *Pseudomonas* Ssb amino acid sequences.

Strain	Position	Nt change	Mutation	# mutants
P. protegens Pf5	1561	$G \rightarrow A$	D521N	1
P. protegens CHA0	1562	$A \rightarrow G$	D521G	1
P. protegens Pf5	1562	$A \rightarrow G$	D521G	7
P. putida KT2440	1562	$A \rightarrow G$	D521G	10
P. protegens CHA0	1567	$A \rightarrow G$	N524D	1
P. protegens CHA0	1580	$C \rightarrow T$	S527F	1
P. protegens CHA0	1599	$C \rightarrow T$	Silent	1
P. protegens CHA0	1650	$C \rightarrow T$	Silent	1
P. protegens Pf5	1706	$C \rightarrow T$	P569L	1
P. protegens Pf5	1736	$C \rightarrow T$	S579F	1

 Table S1. Rif^R mutations in *Pseudomonas*

Strain	System reported with highest efficiency	Substrate	DNA Load† (approx.)	Genetic Target	Efficiency*	Reference
<i>P. aeruginosa</i> PA14	pUCP-RedS (Arabinose- induced λ Red Beta, Exo, Gam)	PCR product with 100 bp homology arms	7 x 10 ¹² copies	∆ <i>pqsC</i> ::kan (800 bp insertion)	3.4 x 10 ⁻⁹	(28)
P. syringae pv. tomato DC3000	Constitutively expressed RecT/E from <i>P. syringae</i> pv. syringae B728a	ssDNA with 40 bp homology arms	1.2 x 10 ¹⁴ copies	4 bp mutation K43R in <i>rpsL</i>	2.4 x 10 ⁻⁴	(21)
		PCR product with 91 bp homology arms	5 x 10 ¹² copies		4.5 x 10 ⁻⁹	(21)
<i>P. putida</i> KT2440	M-toluic acid inducible λ Red Beta, Exo, Gam	PCR product with 500 bp homology	5.4 x 10 ¹¹ copies	∆pp_0589::kan (800 bp insertion)	2.7 x 10 ⁻⁷	(35)
	M-toluic acid inducible RecT/E from <i>E. coli</i> MG1655	PCR product with 100 bp homology	2.6 x 10 ¹² copies	Δpvd::tetA (1.1 kB insertion)	5 x 10 ⁻⁸	(36)
	M-toluic acid inducible λ Red Beta	ssDNA with 50 bp homology	1.2 x 10 ¹⁴ copies	1 bp mutation (kan knock-in)	2 x 10 ⁻³	(12)
<i>P. putida</i> KT2440	Constitutively expressed λ Red Beta	ssDNA with 45 bp homology	3 x 10 ¹⁴ copies	4 bp Q518L change in <i>rpoB</i>	2 x 10⁻⁵	This study
<i>P. putida</i> EM42 (KT2440 derivative)	3MB inducible Ssr T1E_1405 from <i>P.</i> <i>putida</i> DOT-T1E	ssDNA with 45- 50 bp homology	2.9 x 10 ¹³ copies	100 bp deletion in <i>pyrF</i>	5 x 10 ⁻⁴	(15)
P. protegens Pf-5	Rhamnose inducible RecT/E from <i>P. syringae</i> pv. syringae B728a and λ Red Gam	PCR product with 100 bp homology	1 x 10 ¹³	Gentamicin cassette onto plasmid	1.5 x 10 ⁻⁶	(22)
	Constitutively expressed λ Red Beta	ssDNA with 45 bp homology	3 x 10 ¹⁴ copies	15 bp D521P change in <i>rpoB</i>	1 x 10 ⁻⁴	This study

 Table S2. Overview of select recombineering efforts to date in Pseudomonas spp.

P. protegens CHA0	Constitutively expressed λ Red Beta	ssDNA with 45 bp homology	3 x 10 ¹⁴ copies	15 bp D521P change in <i>rpoB</i>	5 x 10 ⁻⁴	This study
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*Efficiency selected from highest reported in single study or calculated based upon data provided.

† Calculated based on reported DNA substrate length and amount using following equation

 $Number of \ copies = \frac{Amount \ (ng) \times 6.022 \ \times 10^{23}}{Length \ (bp) \ \times 1 \times 10^9 \ \times Mass \ of \ DNA \ bp} \ , \ \text{where mass of DNA bp is}$

estimated as 660 for dsDNA, and 330 for ssDNA when sequence is not provided.



Figure S1. Effect of wash buffer on transformation efficiency. 4 mLs of log-phase cells were harvested via centrifugation and washed 3 times in 1 mL of wash buffer indicated. 250 ng of pBBR1-MCS2 vector purified from GM2163 was introduced into the final cell suspension and electroporated at 12 kV/cm. Cells were recovered in 1 mL of LB for 2 hours and plated on LB plates with or without kanamycin. Transformants were quantified after 2 days of growth.



Figure S2. Structure and location of Rif^R binding pocket. Cluster I is indicated by cyan, and Cluster II is indicated by tan. The Q518L mutation (pink) and D521P mutation (navy) were mapped onto the existing structure using the Pymol wizard tool. Structure was generated using *P. putida* ATCC 47054 *rpoB* structure (accession no.: Q88QP2).



Figure S3. Growth defect of *rpoB* Q518L mutants in *P. protegens* CHA0. The same plate of *P. protegens* CHA0 pictured 2 days (left) and 4 days (right) after recombineering using λ -Red Beta and a recombinogenic oligo encoding a Q518L point mutation in the *rpoB* gene. All delayed growth colonies (n=19) sequenced were found to carry the Q518L point mutation.





		cov	pid	1	[80
1	Ecoli	100.0%	100.0%		MAS <mark>R</mark> GVNKVILVCNLGQ DPEVRY MPNGGAVANI <mark>TLATSESWRDKATCEMKEQTEWHR</mark> VVLFC <mark>K</mark> LAEVASEYLRKGSQVYI	
2	putida	95.5%	53.2%		-MA <mark>RGVNKVILVGTCGQDPEVRYLPNGNAVTN</mark> LS <mark>LATSEQWTDK</mark> QSGQKV <mark>ERTEWHRVSLFCKVAEIAGEYLRKGSQ</mark> CYI	
3	aeruginosa	96.1%	52.7%		-MA <mark>RGVNKVILVGTCGQDPEVRYL</mark> PNGNAVTNLSLATSEQWTDKQSGQKVERTEWHRVSLFGKVAEIAGEYLRKGSQCYI	
4	ogarae	93.3%	51.6%		-MA <mark>RCVNKVILVGTCGQDPEVRYLPNGNAVT</mark> NLS <mark>LATSE</mark> QWTDKQTGQKVEKTEWHRVSMFCKVAEIACEYLRKGSQVYI	
5	fluorescens	92.7%	51.6%		-MA <mark>RCVNKVILVCTCCQDPEVRYLENCNAVTN</mark> LS <mark>LATSE</mark> QMTDK <mark>OTC</mark> OKV <mark>EKTEWHRV</mark> SM <mark>PCKVAEIACEYLRKCSQVYI</mark>	
6	chlororaphis	91.6%	52.7%		-MA <mark>RCVNKVILVGTCGQDPEVRYLPNGNAVT</mark> NLSLATSEQWTDKQTGQKVEKTEWHRVSMFCKVAEIACEYLRKGSQVYI	
7	protegens_CHA0	91.0%	50.5%		-MA <mark>RGVNKVILVG</mark> TC <mark>GQDPBVRY</mark> LPNGNAVTNLSLATSEQMTDKQTGQKVBRTEWHRVSMFGKVABIAGBYLRKGSQVYI	
8	protegens_Pf-5	91.0%	50.5%		-MARGVNKVILVGTCCQDPEVRYLPNGNAVTNLSLATSEQWTDKOTCQKVERTEWHRVSMPGKVAEIAGEYLRKGSQVYI	
9	synxantha	92.1%	51.6%		-MARGVNKVILVGTCGQDPBVRYLPNGNAVTNLSLATSEQMTDKQTGQKVBKTEWHRVSMFGKVABIAGBYLRKGSQVYI	
	consensus/100%				.huRGVNKVILVGshCQDPEVRYhPNGsAVsNloLATSEpWpDKtoGphhEpTEWHRVshFGKlAElAuEYLRKGSQSYI	
	consensus/90%				.huRGVNKVILVGshGQDPEVRYhPNGSAVSNLoLATSEDWDDKtoGphhBpTEWHRVshFGKLABLAUEYLRKGSQSYI	
	consensus/80%				.MARGVNKVILVGTCGQDPEVRYLPNGNAVTNLSLATSEQWTDKQGGCKVE+TEWHRVSbPGKVAEIAGEYLRKGSQSYI	
	consensus/70%				.MARGVNKVILVGTCCQDPEVRYLPNGNAVTNLSLATSEQWTDKOTCOKVE+TEWHRVSbFGKVAEIAGEYLRKGSQVYI	
				0.1		160
1	Fcoli	100 0%	100 0%	01		100
2	nutida	95 5%	53 28			
3	aeruginosa	96.1%	52.7%			
4	ogarae	93.3%	51.6%			
5	fluorescens	92.7%	51.6%			
6	chlororaphis	91.6%	52.7%		PGKLOURENEK-DGIKEVOTEIIVDMOGIMOLLGGEPOGDOGOGM-GNSABRPOOSRPOPSOA-P	
7	protegens CHA0	91.0%	50.5%		PGKLOWRENEK-DGINRYWTEIIVDMOGMMOLLGGRPOGDOOGOGGMSNSAPRPOOSRPAP-O-P	
8	protegens Pf-5	91.0%	50.5%		PGKLOAREWEK-DGIKRYTTEIIVDMOGAMOLLGGRPOGDOOGOGGMSNSABRPOOSRPAP-O-P	
9	svnxantha	92.1%	51.6%		PGKLOTREWEK-DGIKEVTTEIVVDMOGAMOLLGGEPOGDOOGOGGM-SNSABRPOOSRPOPAOO-P	
	consensus/100%				PGplpTRcWpc.sg.cRYTTEllVshtGTMOhLGGR.tt.t.	
	consensus/90%				PGpLprRcWpc.sg.cRYTTEllVshtGnMOhLGGR.tt.tt.t	
	consensus/80%				BCKLOTREWEK, DGIKRYTTEIIVDhpGTMOLLGGRPOGtpttttGttuts.PpOpt.t	
	consensus/70%				BCKLOTREWEK.DCIKRYTTE11VDhpCTMOLLGCREOCtpOttGGspuustpoptspt.	
		cov	pid	161] 189	
1	Ecoli	100.0%	100.0%		SRPQQSAPAAPSNBPPMDFDDDIPF	
2	putida	95.5%	53.2%		QA <mark>FQ</mark> RP <mark>AP</mark> QQ <mark>P</mark> AADFD-S <mark>FDDDIPF</mark>	
3	aeruginosa	96.1%	52.7%		QAPQRPAPQQPAPQBAADFD-SFDDDIPF	
4	ogarae	93.3%	51.6%		PQRERPAPQQAAPQPAPDFD-SFDDDIPF	
5	fluorescens	92.7%	51.6%		QQRSAPAPQQAAPQPAPDFD-SFDDDIPF	
6	chlororaphis	91.6%	52.7%		QRESRPAPQQSSPQRAQNFDDSRDDDIPF	
7	protegens_CHA0	91.0%	50.5%		QRESRPAPQQAAPQPAPDFD-SFDDDIPF	
8	protegens_Pf-5	91.0%	50.5%		QRESRPAPQQAAPQPAPDFD-SFDDDIPF	
9	synxantha	92.1%	51.6%		QRESRPAPQQAAQQPAQDFD-SFDDDIPF	
	consensus/100%				.t.ptsAPttsu.pPsslobbIPs	
	consensus/90%				.t.ptsAPttsu.pPsSFDDDIPF	
	consensus/80%				pt.ppPAP00sus0PA.sed.SedDIPe	
	consensus/70%					

Figure S5. Multiple sequence alignment of *Pseudomonas* Ssb amino acid sequences. MSAs were generated using Clustal Omega from EMBL-EBI using standard parameters.