Supplementary Information

Bioinspired mechanical mineralization of organogels

Jorge Ayarza^{†,1}, Jun Wang^{†,1}, Hojin Kim^{1,2}, Pin-Ruei Huang¹, Britteny Cassaidy¹, Gangbin Yan¹, Chong Liu¹, Heinrich M. Jaeger^{2,3}, Stuart J. Rowan^{1,4,5}, and Aaron P. Esser-Kahn^{*,1}

[†] These authors contributed equally to this work.

*Corresponding author. E-mail: aesserkahn@uchicago.edu

- ¹ Pritzker School of Molecular Engineering, University of Chicago, 5640 South Ellis Avenue, Chicago, Illinois 60637, United States
- ² James Franck Institute, University of Chicago, 929 East 57th Street, Chicago, Illinois 60637, United States
- ³ Department of Physics, University of Chicago, 5720 South Ellis Avenue, Chicago, Illinois 60637, United States
- ⁴ Department of Chemistry, University of Chicago, 5735 South Ellis Avenue, Chicago, Illinois 60637, United States
- ⁵ Chemical and Engineering Sciences Division, Argonne National Laboratory, 9700 Cass Avenue, Lemont, Illinois 60439, United States

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Materials

2-mercapto-5-methyl-1,3,4-thiadiazole 95 % (McMT), tris(2-carboxyethyl)phosphine hydrochloride 98 % (TCEP), iodine \geq 99.8 % (I₂), poly(propylene glycol) tolylene 2,4diisocyanate terminated \geq 99 % (PPG diisocyanate, M_n 2300 Da, 3.60 w % NCO), bisphenol A diglycidyl ether \geq 99 %, triethylenediamine \geq 99 %, methylene diphenyl diisocyanate 98 % (MDI), dibutyltin dilaurate 95 % (DBTDL), tetraethylene glycol 99% (TEG), LiClO₄ 99.9 %, NaN₃ \geq 99.5 % , NaOH 98 %, Na₂SO₄ 99 %, ZnBr₂ \geq 99.9 %, Zn(NO₃)₂·6H₂O 98 %, acetonitrile, dimethyl formamide (DMF), dimethyl sulfoxide (DMSO), methanol, dichloromethane, ethyl acetate, n-hexanes, were purchased from Millipore Sigma and used as received, unless otherwise stated. Zinc oxide nanoparticles (ZnO, 99.95 % purity, 18 nm) were purchased from US Research Nanomaterials Inc. Bis(dibenzocyclooctyne-amine) terminated pentaethylene glycol 95 % (DBCO-PEG₅-DBCO) was purchased from Broadpharm. Deuterated solvents were purchased from Cambridge Isotope Laboratories.

Instruments

Chemical characterization

Nuclear magnetic resonance (NMR) spectra (¹H and ¹³C) were recorded either on a Bruker Avance II+ 500 MHz spectrometer equipped with a Bruker QNP probe (¹H: 500 MHz, ¹³C: 125 MHz) or a Bruker Ascend 9.4 T 400 MHz spectrometer equipped with an Avance III HD console and a BBFO "smart" probe (¹H: 400 MHz, ¹³C: 100 Hz) at 25 °C. Chemical shifts (δ) are reported in parts per million (ppm). ¹H NMR spectra are referenced to TMS, while ¹³C NMR spectra are referenced to either TMS or the residual solvent.

High resolution mass spectroscopy (HRMS) was conducted in an Agilent 6224 TOF LC/MS instrument using electrospray ionization (ESI) and a time of flight (TOF) detector.

Fourier transformed infrared (FTIR) spectra were recorded on a Perkin Elmer Spectrum Two FT-IR spectrometer in the MIR range (4000-400 cm⁻¹) using the ATR mode.

X-ray spectra of the microrods were obtained in a KRATOS AXIS NOVA X-ray Photoelectron Spectrometer (XPS) and a Bruker D8 Powder X-ray Diffractometer (XRD).

Gel permeation chromatography (GPC) was conducted in a Tosoh EcoSEC instrument, equipped with Tosoh SuperAW3000 + Tosoh SuperAW4000 + guard columns, eluent: DMF 0.01 M LiBr, flow rate: 0.3 mL.min⁻¹, T = 50 °C and refractive index (RI) and ultraviolet (UV) detectors. The system was calibrated with polystyrene (PS) standards.

Imaging techniques

Scanning electron microscopy (SEM) was conducted in a Carl Zeiss Merlin High-Resolution Field Emission Scanning Electron Microscope (FE-SEM). Samples were previously coated with a Pt film (10 nm thickness) using a Cressington 208 HR sputter coater with Pd/Pt target.

Scanning transmission electron microscopy (STEM) images were acquired using JEOL ARM 200F equipped with a cold field emission source operated at 200 kV. Energy dispersive X-ray (EDS) mapping was acquired using an Oxford X-Max 100TLE windowless SDD detector equipped with JEOL ARM 200F.

Light microscopy images were obtained in a Leica DM2700 P Polarization Microscope under brightfield and polarization modes. A few drops of tetrahydrofuran (THF) were used to swell the polymer composite samples prior to imaging.

Transmission electron microscopy (TEM) was conducted in a ThermoScientific Titan Krios G3i instrument. Cross-sections of polymer composite samples were obtained with a Leica EM UC6 Ultramicrotome.

Thermal and mechanical characterization

Differential scanning calorimetry (DSC) was conducted with a DSC 2500 instrument (TA Instruments) using a modulated DSC program in the range -90 to 250 °C.

Rheological measurements were conducted with a rotational rheometer (TA Instruments, ARES G2) equipped with a peltier element and a parallel plate fixture (25 mm). A volume of 100 μ L of sample was used for each measurement, with a measuring gap of 250 μ m, and the temperature set at 25 °C.

Dynamic mechanical analysis (DMA) was conducted with a TA Instruments RSA-G2 DMA using the compression mode parallel disc fixture (25 mm).

1. Synthetic procedures and characterization

Representative synthesis of the microrods

In a cylindrical polypropylene vial, McMT (66 mg, 0.50 mmol) was dissolved in DMF (400 μ L). ZnO nanoparticles (20 mg, 0.25 mmol) were added to the solution and homogenously dispersed via sonication for 20 s. The mixture was sonicated (40 kHz) in the dark for 4 h. The product was diluted with methanol (5 mL) and separated by centrifugation (3000 x g for 10 min). The precipitate was washed twice with methanol (10 mL) and again separated by centrifugation. The product was dried at 50 °C under vacuum overnight. Yield: 71 mg, 86 %.

Synthesis of $Zn(McMT)_n$ complex ¹

A solution of $Zn(NO_3)_2$ ·6H₂O (149 mg, 0.50 mmol) in water (1 mL) was prepared. In a separate vial, McMT (132 mg, 1.00 mmol) was dissolved in a NaOH_(aq) 1 M solution (1 mL). The zinc nitrate solution was added dropwise to the McMT solution under vigorous stirring. A white precipitate formed, and the mixture was stirred for 30 min. The solid was recovered by filtration and washed thoroughly with water. The product was dried at 50 °C under vacuum overnight. Yield: 128 mg, 78 %.

Synthesis of McMT disulfide dimer (2-methyl-5-[(5-methyl-1,3,4-thiadiazol-2-yl)disulfanyl]-1,3,4-thiadiazole)²



McMT (1.32 g, 10 mmol) was dissolved in 20 mL of methanol and K₂CO₃ (1.38 g, 10 mmol) was added. The mixture was placed in an ultrasound bath for 10 min and then stirred for 1 h. Then, iodine (1.27 g, 5 mmol) was added, and the mixture was stirred for 1 h at room temperature. The product was concentrated under vacuum and diluted with 20 mL of dichloromethane. The solution was washed with water (3 x 50 mL), brine (1 x 50 mL), and finally the organic phase was dried with anhydrous Na₂SO₄. The organic phase was concentrated under vacuum, resulting in a white solid. The product was dried under a high vacuum overnight. Yield: 1.15 g, 88 %. ¹H NMR (500 MHz, CDCl₃): $\delta_{\rm H}$ (ppm) 2.78 (s, 6H, CH₃). ¹³C NMR (125 MHz, CDCl₃): $\delta_{\rm C}$ (ppm) 168.9 (SC(N)S), 166.1 (H₃CC(N)S), 16.0 (CH₃).

Control reactions

w/ respect to representative procedure for the synthesis of microrods

- No ZnO: sample without ZnO nanoparticles.
- No US: sample was stirred in a vortex at 100 rpm.
- w/ZnBr₂: instead of ZnO nanoparticles, anhydrous ZnBr₂ (56 mg, 0.25 mmol) was added and dissolved in the reaction mixture.
- w/ McMT disulfide dimer: instead of McMT, the disulfide (66 mg, 0.13 mmol) was added and dissolved in the reaction mixture.

Synthesis of microrods within polymer organic solution

In a cylindrical polypropylene vial, poly(propylene glycol) tolylene 2,4-diisocyanate terminated (PPG diisocyanate, M_n 2300 Da, 3.60 w % NCO, 0.50 g, 0.43 mmol NCO) was weighed. PPG diisocyanate was dissolved in DMF (1 mL) and 21 mg of tetraethylene glycol (0.11 mmol glycerol, 0.22 mmol OH) were added. Dibutyltin dilaurate (3 µL) was added to the mixture and the solution was vortexed for 30 s. Then, McMT (132 mg, 1.00 mmol) was added, and the mixture was vortexed until dissolution. Finally, ZnO nanoparticles (40 mg, 0.50 mmol) were added and dispersed under sonication. The mixture was sonicated in an ultrasound bath (40 kHz) for 6 h.

Synthesis of Bisphenol A di(3-azido-2 hydroxy propan-1-ol) ether (BPA-diazide)³



A solution of bisphenol A diglycidyl ether (16.8 g, 49.5 mmol) and LiClO₄ (16.6 g, 157 mmol) in acetonitrile (150 mL) was prepared. NaN₃ (10.0 g, 154 mmol) was added in portions and the reaction mixture was heated to 90 °C under Ar atmosphere with vigorous stirring for 24 h. Afterwards, the reaction was cooled down to room temperature and water (300 mL) was added. The reaction mixture was extracted with ethyl acetate (3×150 mL), and the combined organic layer was washed with water (3×100 mL) and brine (3×100 mL), dried over anhydrous Na₂SO₄ and concentrated *in vacuo* to yield a light-yellow viscous oil. The product was further purified by flash column chromatography using an ISCO Teledyne instrument with a 330 g RediSep Rf silica column while eluting with a mixture of hexanes and ethyl acetate (gradient of 0 to 40 % ethyl acetate). Fractions were combined and concentrated *in vacuo*, yielding a pale viscous oil of bisphenol A di(3-azido-2 hydroxy propan-1-ol) ether (15.8 g, 37.1 mmol, yield 75 %).

¹**H NMR** (500 MHz, (CD₃)₂SO): $\delta_{\rm H}$ (ppm) 7.10 (d, 4H, ³J_{H-H} = 8.7 Hz, Ar-C_{3,5}-H), 6.83 (d, 4H, ³J_{H-H} = 8.7 Hz, Ar-C_{2,6}-H), 5.51 (d, 2H, ³J_{H-H} = 5 Hz, OH), 3.99 (m, 2H, CH), 3.88 (m, 4H, O-CH₂), 3.35 (m, 4H, CH₂-N₃), 1.57 (s, 6H, CH₃). ¹³C **NMR** (125 MHz, CDCl₃): $\delta_{\rm C}$ (ppm) 156.0 (Ar-C₁),

143.8 (Ar-C₄), 127.8 (Ar-C_{3,5}), 113.9 (Ar-C_{2,6}), 69.3 (CH), 69.0 (O-CH₂), 53.3 (CH₂-N₃), 41.7 (C_{quat}), 30.9 (CH₃). **HRMS** ESI-TOF (m/z): $[M+Cl]^-$ calcd. for C₂₁H₂₆N₆O₄Cl: 461.1704 found: 461.1685.

Synthesis of azido-PU³



In a round-bottom flask, poly(propylene glycol) tolylene 2,4-diisocyanate terminated (PPG diisocyanate, M_n 2300 Da, 3.60 w % NCO, 1.35 g, 0.59 mmol) was mixed with methylene diphenyl diisocyanate (440 mg, 1.76 mmol) and dissolved in ethyl acetate (2 mL). Separately, in a vial, BPA-diazide (1.00 g, 2.35 mmol) was mixed with triethylenediamine (28 mg, 0.25 mmol) and dissolved in ethyl acetate (2 mL). The second solution was added to the first one and the mixture was homogenized under vigorous stirring, followed by heating at 70 °C for 2 h. After cooling the reaction mixture to room temperature, it was diluted with ethyl acetate (4 mL) and precipitated in cold hexanes (100 mL). The polymer was recovered by centrifugation, washed overnight in hexanes (50 mL), and dried under vacuum at 60 °C overnight. Yield: 2.68 g, 96 %. ¹H NMR (400 MHz, (CD₃)₂SO): $\delta_{\rm H}$ (ppm) 9.78 (s), 9.53 (s), 9.04 (s), 7.36 (d), 7.08 (d), 6.83 (d), 5.52 (d), 5.18 (t), 4.86 (m), 4.12 (d), 3.87 (t), 3.79 (s), 3.67-3.71 (m), 3.41-3.48 (m), 2.71 (s), 2.10 (s), 1.56 (s), 1.19 (t), 1.04 (s), 1.02 (s).

Synthesis of polymer organogel with embedded microrods

In a cylindrical polypropylene vial, azido-PU (150 mg) was dissolved in DMF (400 μ L) overnight. Separately, in a vial, DBCO-PEG₅-DBCO (15 mg) was dissolved in DMF (50 μ L). McMT (66 mg) was added to the azido-PU solution and dissolved by vigorous stirring. ZnO nanoparticles (20 mg) were added and homogenously dispersed by vigorous stirring and sonication for 20 s. The mixture was cooled down in a fridge (4 °C) for 30 min. The crosslinker solution was quickly added to the azido-PU mixture and rapidly vortexed. The organogel forms within 1 min. It was then sonicated in an ultrasound bath (40 kHz) for 6 h. The organogel was then carefully removed from the vial and dried in a vacuum oven (50 °C) overnight to remove the DMF.

2. NMR spectra







3. Additional characterization of microrods



Supplementary Fig. 1. Graph comparing the XPS spectra of ZnO and microrods (*top*), with close-up images of the Zn and O peaks (*bottom*).



Supplementary Fig. 2. STEM images of the microrods and EDS elemental mapping.



Supplementary Fig. 3. STEM images of the $Zn(McMT)_n$ complex and EDS elemental mapping.

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Element	Theory	Fo	und	Single	Duplicate 🔀
С	21.85	21.12	21.21	Present:	N S Zn O(Possible)
Н	2.45	1.72	1.70	for:	
N	16.99	16.22	16.40	M.P. ≥200 Celcius	B.P. unknown
S	38.89	37.58	37.49	Temp. 50C Rush Service	Vac. yes ime 30min Rush service guarantees analyses will be completed and results available by 5 PM EST on live day live sample is received by 11 AM.
1				Include Email huang	Address or FAX # Below pr@uchicago.edu
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Supplementary Fig. 4. Elemental analysis of the microrods.

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Element	Theory	Fo	und	Single Duplicate		
С	21.85	19.37	19.34	Present:		
Н	2.45	1.55	1.46	for:		
N	16.99	14.85	14.81	M.P. ≥200 Cetclus B.P. unknown To be dried: Yes ⊠ No		
S	38.89	33.92	33.83	Temp. 50C Vac. Ves Time 30min Rush Service In the revice guarantees analyses will be completed and results available by 5 Mk Est on the day the sample is received by 11 AM.		
				Include Email Address or FAX # Below huangpr@uchicago.edu		
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Supplementary Fig. 5. Elemental analysis of the $Zn(McMT)_n$ complex.



Supplementary Fig. 6. Modulated DSC curve of McMT.



Supplementary Fig. 7. Modulated DSC curve of microrods.



Supplementary Fig. 8. Modulated DSC curve of $Zn(McMT)_n$ complex.

4. Optimization of the stoichiometry of microrod synthesis



Supplementary Fig. 9. SEM image of the microrods obtained via mechanical stirring with a magnetic stir bar (400 rpm). Average length ($3.9 \mu m$) and width ($0.3 \mu m$).

Sample	ZnO (mmol)	McMT (mmol)	Molar ratio (ZnO:McMT)	Yield (mg)
ZnO (10 mg)	0.13	0.50	1:4	33
ZnO (20 mg)	0.25	0.50	1:2	70
ZnO (40 mg)	0.50	0.50	1:1	93*

Supplementary Table 1. Results for experiments varying the concentration of ZnO.

*The increase in yield can be attributed to the unreacted ZnO which is also recovered during the microrod isolation procedure.



Supplementary Fig. 10. Flow viscosity measurements of the microrod suspensions resulting from reactions varying the concentration of ZnO.



Supplementary Fig. 11. SEM image of microrods from sample with ZnO (10 mg).



Supplementary Fig. 12. SEM image of microrods from sample ZnO (20 mg).



Supplementary Fig. 13. SEM image of microrods from sample ZnO (40 mg).

Sample	ZnO (mmol)	McMT (mmol)	Molar ratio (ZnO:McMT)	Yield (mg)
McMT (33 mg)	0.25	0.25	1:1	39
McMT (66 mg)	0.25	0.50	1:2	65
McMT (132mg)	0.25	1.00	1:4	69

Supplementary Table 2. Results for experiments varying the concentration of McMT.



Supplementary Fig. 14. Flow viscosity measurements of the microrod suspensions resulting from experiments varying the concentration of McMT.



Supplementary Fig. 15. SEM image of microrods from sample McMT (33 mg).



Supplementary Fig. 16. SEM image of microrods from sample McMT (66 mg).



Supplementary Fig. 17. SEM image of microrods from sample McMT (132 mg).

Reaction time (h)	Yield (mg)	Particle length (µm)	Particle width (µm)	Aspect ratio
0	-	51.3 x 10 ⁻³	-	1*
1	12	2.1	0.2	11
2	42	3.1	0.2	16
4	71	4.3	0.3	14
6	70	4.1	0.3	14

Supplementary Table 3. Particle size analysis for temporal study of the microrod growth.

* ZnO nanoparticles are spherical.



Supplementary Fig. 18. Higher magnification SEM image of microrod growth at 1 h timepoint.



Supplementary Fig. 19. Higher magnification SEM image of microrod growth at 2 h timepoint.

5. Rheology characterization of microrod/PU polymer solution



Supplementary Fig. 20. Light microscopy images of the *All components* polymer solution containing the microrods under brightfield (*top*) and cross-polarizer filter (*bottom*).



Supplementary Fig. 21. SEM image of the *All components* polymer solution containing the microrods.



Supplementary Fig. 22. Size distribution graphs (length and width) of microrods grown in (A) plain solution and (B) polyurethane polymer solution.



Supplementary Fig. 23. Shear viscosity measurements of microrods grown within polymer organic solution and control reactions (Batch #2).



Supplementary Fig. 24. Shear viscosity measurements of microrods grown within polymer organic solution and control reactions (Batch #3).

6. DMA characterization of microrod/azido-PU composites



Supplementary Fig. 25. Crosslinking reaction of azido-PU with DBCO-PEG₅-DBCO in DMF at room temperature.



Supplementary Fig. 26. Representative compression DMA oscillatory amplitude and frequency measurements of microrod/azido-PU composite and control samples.



Supplementary Fig. 27. Representative DMA oscillatory amplitude measurements for the azido-PU composite samples with different loadings of microrods.



Supplementary Fig. 28. Representative compression DMA oscillatory amplitude and frequency measurements of azido-PU composite samples with different loadings of microrods.

7. Images of cross-sections of the microrods/azido-PU composite



Supplementary Fig. 29. Light microscopy images (bright-field) of the *All components* sample showing the presence of the crystalline microrods in bundles.



Supplementary Fig. 30. TEM images of microrods in azido-PU matrix.

Supplementary References

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