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Last updated by author(s): Oct 8, 2023

Reporting Summary

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Statistics

For	all st	atistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Cor	nfirmed
	X	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	X	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	X	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
X		A description of all covariates tested
X		A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	X	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
		For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.
X		For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
X		For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
X		Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated
		Our web collection on statistics for biologists contains articles on many of the points above.

Software and code

Policy information about availability of computer code

iPGCT transplantation was performed using CellTram 4r Oil (Eppendorf) instruments, the transplanting needle was made using Flaming/ Data collection Brown type micropipette puller (P-100, Sutter) and the needle was polished using Micro Grinder (EG-400, NARISHIGE). Testis and early embryos were imaged using a fluorescence stereomicroscope (Axio Zoom.V16, Zeiss), while sections and primitive gonads were imaged using Confocal microscope (SP8, Leica). The migration of iPGCs was tracked using a fluorescence microscope (CTR6500, Leica). The schematic drawing were drawn using Microsoft Office PowerPoint.

Data analysis Fiji was used for image analysis. Graphs were made via GraphPad v8.

For manuscripts utilizing custom algorithms or software that are central to the research but not vet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

The authors declare that the data supporting the findings of this study can be found in the article and Supplementary Information files. All other relevant data supporting the findings of this study are available upon request from the corresponding author. This article provides the source data.

Field-specific reporting

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Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

X Life sciences

Behavioural & social sciences

Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.						
Sample size	All samples that met the requirements were counted and presented in the manuscript.					
Data exclusions	No data were excluded for analysis.					
Replication	At least three independent repeated experiments were carried out and a representative example of three replicate is shown.					
Randomization	No specific randomization methods was used, the samples were carried out according to the specific experimental methods.					
Blinding	No blinding were used.					

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

Methods

n/a	Involved in the study	n/a	Involved in the study
	X Antibodies	X	ChIP-seq
×	Eukaryotic cell lines	X	Flow cytometry
×	Palaeontology and archaeology	X	MRI-based neuroimaging
	X Animals and other organisms		
X	Clinical data		
×	Dual use research of concern		
×	Plants		

Antibodies

Antibodies used	Anti-Vasa, anti-Sycp3 and anti-PCNA were against the antigens, and purified using antigen-affinity chromatography by our lab. These antibodies were conjugated with Alexa Fluor 488, 568 and 680 on demand using Alexa Fluor® Antibody Labeling Kits according to the manufacturer's manual (Thermo Fisher).
Validation	The antibodies used are all proven by a recent study (Ye et al., 2023).

Animals and other research organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research, and Sex and Gender in Research

Laboratory animals	Both zebrafish and <i>Gobiocypris rarus</i> used in this study were raised in the China zebrafish Resource Center (CZRC). Our experiments were carried out using 3-month-old zebrafish from wild-type stocks with AB genetic background and transgenic line Tg(cmv:GFP) and Tg(cmv:mCherry). iPGC induction was performed in zebrafish embryos at 1-cell stage, and iPGC transplantation was performed when the embryos developed to sphere stage.		
Wild animals Reporting	This study did not involve wild animals		
on sex Field-collected	No sex-specific fish were selected for the experiment. The resulting offspring are all male		
samples Ethics	All animal experiments were conducted according to the standard animal guidelines approved by the Animal Care Committee of		
•			
	the University of Chinese Academy of Sciences and the institute of Hydrobiology, Chinese Academy of Sciences.		
oversight	All animal experiments are supervised by the Animal Care Committee of the University of Chinese Academy of Sciences.		

Note that full information on the approval of the study protocol must also be provided in the manuscript.

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