THE OXYGEN DISSOCIATION CURVE IN ANEMIA OF VARIOUS TYPES [1]

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It is a clinical observation of long standing that the patient who has become anemic gradually, is often well compensated for the lower hemoglobin level he possesses. Thus it is not uncommon for an individual with a hemoglobin level well below 50 per cent, due to pernicious anemia or hypochromic anemia, to carry on his normal activities. It is generally believed that the compensatory adjustment is in part due to increased utilization of oxygen by the tissues and in part to circulatory adaption. It has also been suggested, from studies of the gas transport of red cells in anemia (1–4), that it may in part be due to displacement of the oxygen dissociation curve to the right of normal whereby the amount of oxygen released to the tissues is increased; the number of patients observed in these investigations is small probably because of the difficulties and time-consuming nature of the methods. The fullest report is that of Richards and Strauss (2) who indicated a shift to the right of the oxygen dissociation curve apparent, however, only at the abnormal plasma pH of 7.64. Only the reports of Dill and his coworkers (3) and Isac, Matthes, and Yamanaka (4), demonstrate a shift to the right of the oxygen dissociation curve at the standard plasma pH of 7.4. There are no observations, as far as we are aware, on the oxygen dissociation curve at various stages after recovery from anemia. Many of the patients in these early reports had received whole blood transfusions before the blood gas studies and, as we have shown, this produces an alteration in the oxygen dissociation curve of the anemic recipient (5). Various hypotheses, many of them contradictory, have been offered regarding the cause of the beneficial shift of the oxygen dissociation curve in anemia but none of them affords a satisfactory explanation for the phenomenon.

For these reasons a further and more extensive study of the oxygen dissociation curve in anemia appeared warranted. The present paper describes the oxygen dissociation curves of twenty-nine individuals suffering from various types of anemia. Observations on the position of the oxygen dissociation curves at various times during and after recovery from the anemia are also presented.

MATERIAL AND METHODS

The subjects of this study comprised eleven patients with megaloblastic anemia in relapse and three in therapeutic remission, three patients with nutritional hypochromic anemia, three patients with hypochromic anemia secondary to chronic hemorrhage, two patients with aplastic anemia, four patients with anemia associated with chronic nephritis, acute leukemia, reticulosis, and scurvy respectively, and three patients with hemolytic anemia. Many of the observations were made before specific therapy was administered and subsequent curves were performed at different periods during and after recovery. None of the patients had received blood transfusions before the blood gas studies were performed, with the exception of one patient with aplastic anemia of long standing (Table II, Case 12).

The collection of blood samples, the equilibration in tonometers with the desired gas tensions, the gas analyses, and the plasma pH (pHs) determinations were performed on venous blood samples removed without stasis by the methods described elsewhere (5). The subjects were resting for at least one hour before the removal of the blood samples. The cell pH (pHc) was determined from the Henderson-Hasselbalch equation given below. In the case of the severely anemic subjects the blood samples were first concentrated by removal of plasma under oil to bring the packed cell volume approximately to normal.

$$pHc = pK'c + \log \frac{(BHCO_3) \text{ blood}}{(H_2CO_3) \text{ blood}}$$

where $(BHCO_3)$ blood = the total CO_2 of oxygenated blood at 40 mm. Hg partial pressure of CO_2 and (H_2CO_3) blood = H_2CO_3 content of blood calculated from the formula $(H_2CO_3) = a \, (pCO_2)$ where "a" is the solubility coefficient of carbon dioxide in blood (from data

TABLE I

Normal subjects and megaloblastic anemia [1]

Case no.	Patient (A)	Date (B)	R.B.C. mill./cu. mm. (C)	HbO2 Vol. % (D)	Color index (E)	Packed cell volume (F)	HbO2/P.C.V. (G)	Retics. % (H)	CO Vol. % (I)	(CO2)s Vol. % (J)	(CO2)o Vol. % (K)	Δ(CO2)s-o (L)	Blood pH_s (M)	Blood pH_o (N)	Blood pH_s and pH_o (O)	pH_s = 7.4 (P)	pH_o = 7.22 (Q)	Comments
	6 normal subjects			18–21 19.5					0.25–0.7 0.5	50–62.5 57.5	42–52 48	8–11 9.5	7.33–7.43 7.39	7.17–7.26 7.23	25–28 26.5	25.5–27 26.5	25.5–27 26.5	Mean normal values from 6 individuals with range figures
1	J. McQ. Male 79 yrs.	26/9/53 After concentration	2.2	9.5 16	1.1	23 38	0.41	1	0	65	57 49	8 16	7.47	7.22	28 28	30.5	28	Addisonian pernicious anemia. Parenteral vitamin B12 therapy started on 27/9/53
2	M. P. Female 30 yrs.	2/4/53 After concentration	0.90	5.5 19	1.5	10.5 38	0.52	1	0	57	50 41	7 16	7.4	7.14	32 32	32	28.5	Megaloblastic anemia with free HCl in gastric juice. Curve before treatment
3	M. M. Female 70 yrs.	9/5/53 After concentration	2.05	12 18	1.4	24 39	0.50	3	0.2	59	53 49	6 9.5	7.42	7.23	27 27	27.5	27	Addisonian pernicious anemia. Parenteral liver extract therapy started on 11/5/53
4	J. B. Male 56 yrs.	13/1/53 26/1/53 29/1/53 18/2/53	1.39 2.79 3.14 3.60	7.5 12 13 16.5	1.3 1.1 1.1 1.1	22 29 38	0.34 0.41 0.44	14 2.5 3.0 1	0.2	59 57	47	10	7.42 7.40	7.20	31 30 31 28.5	31.5 28.5	28	Addisonian pernicious anemia. Parenteral vitamin B12 therapy started on 6/1/53
5	J. A. Male 36 yrs.	23/2/53 28/2/53 17/3/53	1.51 1.85 3.38	8 10 14.5	1.3 1.4 1.1	18 25 40	0.44 0.40 0.36	2.5 31.5 2.5	0.5 1	51 62	47 50	4 12	7.35 7.44	7.24	28 26.5 26.5	26.5 28	28.5	Addisonian pernicious anemia. Parenteral vitamin B12 started on 23/2/53
6	A. D. Female 43 yrs.	8/2/53 17/2/53 6/3/53 16/3/53 17/3/53	2.3 2.62 3.06 3.42 3.51	10 12 15.5 14.5 14.5	1.1 1.1 1.1 1.1 1.0	19.5 30	0.51 0.48	20 3.6 1	0.2 0.25	52 56 62	48 48.5 52	4 7.5 10	7.36 7.39 7.44		29 29 29 31 30	27.5 31 31.5		Addisonian pernicious anemia. Parenteral vitamin B12 started on 2/2/53

[1] Column D—HbO2 vol. % = the oxyhemoglobin capacity in volume per cent.
Column J—(CO2)s vol. % = the total carbon dioxide of true plasma from oxygenated blood at a CO_2 partial pressure of 40 mm. Hg.
Column K—(CO2)o vol. % = the total carbon dioxide of oxygenated blood at a CO_2 partial pressure of 40 mm. Hg.
Column L—Δ(CO2)s-o = the difference between the total CO_2 of true plasma and oxygenated blood.
Column M—Blood pH_s = the plasma pH determined on true plasma from oxygenated blood at a CO_2 partial pressure of 40 mm. Hg.
Column N—Blood pH_o = the cell pH determined on oxygenated blood at a CO_2 partial pressure of 40 mm. Hg.
Columns O, P, Q—pO2 for Hb = HbO2, = the partial pressure of oxygen at which oxyhemoglobin saturation is 50% at the plasma pH and cell pH of the subject (column O), at standard plasma pH (column P), and at standard cell pH (column Q).

TABLE I—*Continued*

Case no.	Patient	Date	R.B.C. mill./cu.mm.	HbO$_2$ Vol.%	Color index	Packed cell volume	HbO$_2$/P.C.V.	Retics. %	CO Vol.%	(CO$_2$)$_?$ Vol.%	(CO$_2$)$_?$ Vol.%	Δ(CO$_2$)$_{?-?}$	Blood pH$_?$	Blood pH$_?$	pO$_2$ for Hb = HbO$_2$ Blood pH$_?$ and pH$_?$	pH$_?$=7.4	pH$_?$=7.22	Comments
	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	
7	M. C. Female 58 yrs.	3/3/53 16/3/53	1.77 1.47	8.5 8.5	1.2 1.5	19 18	0.44 0.47	2	0	62 58.5	56 53	6 4.5	7.44 7.41		30 29	31.5 29.5		Addisonian pernicious anemia. Oral therapy with Hog's stomach and vitamin B$_{12}$ started on 9/3/53
8	J. L. Male 67 yrs.	1/9/52 8/9/52 7/11/52 5/2/53	1.75 2.6 4.49 5.02	11.5 14.5 20 20	1.6 1.4 1.1 1.0	23 31 43 46	0.50 0.47 0.47 0.44	5.2 12	0.25 0.5	66 57	58.5 45	7.5 12	7.47 7.40		30 30 27.5 27	32 27		Addisonian pernicious anemia. Oral therapy with Hog's stomach and vitamin B$_{12}$ started on 2/9/53. Subsequent parenteral vitamin B$_{12}$
9	M. F. Female 51 yrs.	23/7/52 24/7/52 1/8/52 28/8/52 24/2/53	1.43 1.5 3.83 3.82	7.8 7.5 9.5 15.7 18	1.4 1.6 1.0 1.2	16 16 21 33 41.5	0.49 0.47 0.45 0.44 0.45	2.2 1 20.4 1	0.2	59 58	54 53	5 5	7.41 7.40		28.5 28.5 28.5 28.5 27	29 28.5		Addisonian pernicious anemia. Oral therapy with Hog's stomach and vitamin B$_{12}$ started on 25/7/52 and continued during period of study
10	M. R. Female 59 yrs.	8/8/52 14/8/52 5/9/52	2.22 2.32 3.8	10 10.2 17	1.1 1.1 1.1	21.5 24 36.5	0.47 0.43 0.46	1 18.5 1	0	62 64	56 58	6 6	7.44 7.45		28.5 28.5 29	30 30.5		Addisonian pernicious anemia. Parenteral vitamin B$_{12}$ started on 9/8/53
11	A. N. Female 43 yrs.	19/1/53 30/1/53 17/2/53	2.91 3.69 4.21	13.3 17.3 17.3	1.2 1.2 1.0	38 39	0.45 0.44	14.8 2.8 1	0 0.5	59	54	5	7.42		31 31 28.5	31.5		Addisonian pernicious anemia with subacute combined degeneration of cord. Parenteral vitamin B$_{12}$ started on 9/1/53
12	M. D. Female 40 yrs.	6/3/53	4.11	18	1.1	39	0.46	1	0.5	57	48	9	7.40	7.22	27	27	27	Addisonian pernicious anemia. Maintenance therapy with parenteral vitamin B$_{12}$ for 3 months
13	A. B. Male 78 yrs.	27/1/53	4.83	20.5	1.0	47	0.44		0.5	55.5	45	10.5	7.38	7.20	27	26.5	26.5	Addisonian pernicious anemia. Maintenance therapy with parenteral liver extract and vitamin B$_{12}$ since August, 1948
14	J. E. B. Male 85 yrs.	28/1/53	4.55	20	1.0	43	0.46		0.5						26			Addisonian pernicious anemia. Maintenance therapy with parenteral vitamin B$_{12}$ since May, 1951

TABLE II

Hypochromic anemia, secondary anemia, aplastic anemia and hemolytic anemia

Case no.	Patient (A)	Date (B)	R.B.C. mill./cu. mm. (C)	HbO$_2$ Vol. % (D)	Color index (E)	Packed cell volume (F)	HbO$_2$/P.C.V. (G)	Retics. % (H)	CO Vol. % (I)	(CO$_2$)$_a$ Vol. % (J)	(CO$_2$)$_b$ Vol. % (K)	(CO$_2$)$_{a-b}$ (L)	Blood pH$_a$ (M)	Blood pH$_e$ (N)	pO$_2$ for Hb = HbO$_2$			Comments
															Blood pH$_a$ and pH$_e$ (O)	pH$_a$ = 7.4 (P)	pH$_e$ = 7.22 (Q)	
1	J. McL. Male 68 yrs.	27/2/53 After concentration	3.77	8 15	0.53	21 39	0.38	2	0.5	57.5	51 42	6.5 15.5	7.4	7.16	29 29	29	27	Nutritional hypochromic anemia
2	M. B. Female 41 yrs.	13/2/53	2.71	7	0.65	18	0.40	2	0	54	47	7	7.37		30	29		Nutritional hypochromic anemia
3	J. D. Male 70 yrs.	16/3/53	2.92	10	0.85	23	0.43	2	0.2	58	50	8	7.41		32	32		Nutritional hypochromic anemia
4	A. W. Female 51 yrs.	23/2/53 After concentration 18/3/53 24/3/53	3.1 3.58 3.79	9 15 13.5 15.2	0.73	25 37.5 33	0.36	2	0.2	52 58 55	47 42 50 47	5 10 8 8	7.36 7.4 7.38	7.12	32 31.5 34 32	30 34 31	28	Hypochromic anemia secondary to hereditary telangiectasis
5	W. McC. Male 33 yrs.	24/2/53	3.10	8.5	0.70	22	0.40	2	0.2	52	47	5	7.35		27	25		Hypochromic anemia secondary to hemorrhoids
6	W. M. Male 42 yrs.	15/9/53	3.42	7	0.51	19	0.36	1.3	0	59	54	5	7.41		32	32.5		Hypochromic anemia secondary to hemorrhoids
7	A. F. Male 73 yrs.	5/3/53	2.15	9.5	1.11	24	0.40	12	0	56	52	4	7.39		28.5	28.5		Severe scurvy due to dietary inadequacy
8	J. S. Male 28 yrs.	19/2/53	1.85	7	0.95	13	0.54	5	0	34	31	3	7.14		32	25		Chronic glomerulonephritis. Blood urea 240 mg. %

TABLE II—*Continued*

Case no.	Patient (A)	Date (B)	R.B.C. mill./cu. mm. (C)	HbO₂ Vol. % (D)	Color index (E)	Packed cell volume (F)	HbO₂ P.C.V. (G)	Retics. % (H)	CO Vol. % (I)	(CO₂)ₛ Vol. % (J)	(CO₂)ₒ Vol. % (K)	(CO₂)ₛ₋ₒ (L)	Blood pHₛ (M)	Blood pHₒ (N)	pO₂ for Hb = HbO₂			Comments
															Blood pHₛ and pHₒ (O)	pHₛ = 7.4 (P)	pHₒ = 7.22 (Q)	
9	C. R. Male 73 yrs.	24/4/53 After concentration	2.17	9 16	1.03	20 36	0.45	5	0.2	56	48.5 44	7.5 12	7.39	7.15	32 32	31.5	28.5	Reticulosis;
10	D. A. Male 16 yrs.	7/4/53	1.51	7.5	1.25	14	0.53	1	0	63	57	6	7.44		29.5	31		Acute leukemia
11	J. McG. Male 16 yrs.	4/3/53 After concentration	1.76	9 18.5	1.27	20 40	0.45	1	0.2	53	48.5 43	4.5 10	7.37	7.17	28 28	27	26	Aplastic anemia of recent onset
12	M. T. Female 53 yrs.	25/5/53 After concentration	1.82	7.5 17	1.04	16 36	0.47	1	0.2	59.5	55 47	4.5 12.5	7.42	7.17	30 30	31	28	Aplastic anemia of several years' duration. Numerous transfusions
13	C. M. Female 56 yrs.	6/7/53 After concentration	2.24	10 17.5	1.0	22 39	0.45	12.5	0.2	59	52 45	7 14	7.41	7.16	28	28.5 28.5	26	Congenital acholuric jaundice. Microspherocytosis prominent
14	I. N. Male 8 yrs.	10/4/53	3.86	17.5	1.14	39	0.45	10	0	75	47.5	27.5	7.4	7.21	27	27	26.5	Congenital acholuric jaundice. Microspherocytosis prominent
15	A. W. Female 29 yrs.	8/6/53	1.06	5.5	1.34	11	0.50	20	0.25	52	50	2	7.36		27	25.5		Acquired hemolytic anemia. Microspherocytosis prominent

TABLE III

Oxyhemoglobin saturation of laked blood solutions derived from patients with anemia

	Normal blood	Pernicious anemia in relapse (Table I, Case 8)	Hypo-chromic anemia (Table II, Case 4)	Pernicious anemia after treatment (Table I, Case 10)
pO₂ mm. Hg		HbO₂ Volume %		
15	28	29	29	28
20	50	50	48	50.5
30	78	80	78	79
40	86	87	85	85

by Dill in the paper of Keys, Hall, and Barron (6) and Dill, Graybiel, Hurtado, and Taquini (7)).

pK'c was obtained from the alignment chart and procedure given in the paper of Keys, Hall, and Barron (6) modified in such a manner for the pK'c to be 5.98 for normal blood (8, 9) giving a cell pH of 7.20 to 7.25. The other values for pK'c derived from this alignment chart were changed proportionately.

Correction of the position of the curve according to the plasma pH and cell pH was carried out in curves drawn on logarithmic co-ordinates by a slight modification (10) of the procedure outlined by Dill in the paper of Keys, Hall, and Barron (6).

Buffered solutions of laked blood of pH 7.4 were prepared according to the methods described by Brooks (11) and Darling and Roughton (12).
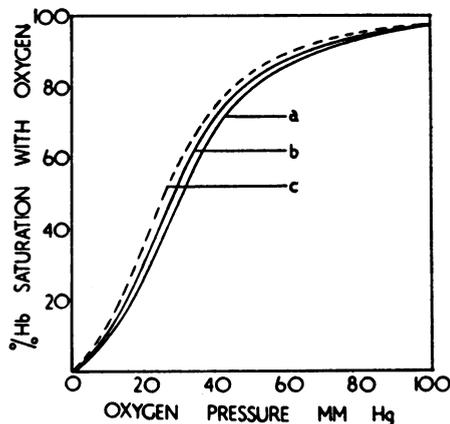
FIG. 1. OXYGEN DISSOCIATION CURVES FROM ANEMIC AND NORMAL SUBJECTS

Curve a—Addisonian pernicious anemia in relapse, hypochromic anemia, secondary anemia. At serum pH 7.4

Curve b—Addisonian pernicious anemia in relapse, hypochromic anemia, secondary anemia. At cell pH 7.22

Curve c—Normal blood at serum pH 7.4 and cell pH 7.22

RESULTS

The results are presented in Tables I, II, and III and in Figure 1. The position of the oxygen dissociation curve is indicated by the partial pressure of oxygen at which 50 per cent of the hemoglobin is oxygenated.

In Table I are shown the mean values obtained from six normal individuals, which have been presented in detail elsewhere (10), and the individual data of eleven patients with megaloblastic anemia in relapse and three in therapeutic remission. Of the patients in relapse the position of the oxygen dissociation curve at the plasma and cell pH of the subject (column O) was displaced to the right of normal in all but one (case 3). After correction to the standard plasma pH of 7.4 (Column P and curve a of Figure 1) this shift to the right was more pronounced except in cases 5 and 6. In these two cases the curve was nearly normal at standard plasma pH. Serial observations (cases 4, 8-11) show that the displacement to the right of the oxygen dissociation curve did not alter during the earlier response to treatment, although by the time the red cell count had reached about the four million per cubic mm. level, between four and six weeks, some lessening of the displacement was apparent. In the two patients in whom the initial oxygen dissociation curves were normal at standard plasma pH (cases 5 and 6) a shift to the right developed during the early stages of treatment. The presence of reticulocytes (column H) did not appear to influence the position of the oxygen dissociation curve. The curve was normal in position in the three patients in therapeutic remission (cases 12–14). The cell pH (column N) was determined in seven of the cases, of whom three were in relapse, two were during treatment, and two were in therapeutic remission. Of the cases in relapse, the difference between the cell and plasma pH was increased in those showing a shift of the oxygen dissociation curve to the right at standard plasma pH (cases 1 and 2), and normal in the case showing a normal oxygen dissociation curve at standard plasma pH (case 3). In the patients (cases 4 and 5) in whom the cell pH was determined when the red cell count had reached between three and four millions per cubic mm., at which time the oxygen dissociation curve was still slightly displaced to

the right at standard plasma pH, the difference between the cell and plasma pH was normal. After correction to standard cell pH the position of the oxygen dissociation curve was unaltered (column Q). The cell pH was normal in the two patients in therapeutic remission (cases 12 and 13) both of whom showed a normal position of the oxygen dissociation curve.

In Table II are shown the results obtained in patients with nutritional hypochromic anemia, chronic post-hemorrhagic anemia, secondary anemia, aplastic anemia, and hemolytic anemia. With the exception of the case of anemia associated with chronic nephritis (case 8), in which there was an acidosis, the plasma pH of the subjects was normal; in the majority of cases the values were in the lower range of normal. The position of the oxygen dissociation curve at the plasma and cell pH of the subject was displaced to the right of normal in all the patients with the exception of one patient with post-hemorrhagic anemia (case 5), one patient with aplastic anemia (case 11), and all three patients with hemolytic anemia (cases 13–15). The displacement to the right was still present after correction to the standard plasma pH of 7.4 (Figure 1, curve a) excepting the patient with chronic nephritis whose curve was to the right of normal at blood plasma pH but normal in position at standard plasma pH. The cell pH was low in six of the seven cases in which it was determined and normal in the remaining case (case 14), a patient with hemolytic anemia. After correction to the standard cell pH the position of the oxygen dissociation curve was normal in the two patients with hemolytic anemia (cases 13, 14) and in one patient with aplastic anemia (case 11) but was still slightly to the right (Figure 1, curve b) in the patients with hypochromic anemia (cases 1, 4), secondary anemia (case 9), and the remaining patient with aplastic anemia (case 12).

Table III shows that the oxygen dissociation curves of hemoglobin solutions (buffered solutions of laked blood) from patients with pernicious anemia and hypochromic anemia are the same as that of hemoglobin solutions of normal blood.

DISCUSSION

Odaira (1) states that in anemia there is a marked acidosis and that as a result there is a shift to the right of the oxygen dissociation curve.

The techniques employed by Odaira are open to criticism, however, and many authors (3, 4, 13–16) have failed to demonstrate an acidosis in pernicious anemia but have, in fact, shown a tendency to an alkalosis. Stadie and Martin (17), obtained a normal oxygen dissociation curve in a single patient with pernicious anemia but they do not give the plasma pH at which the curve was performed. Richards and Strauss (2) reported that the oxygen dissociation curves in six patients with pernicious anemia and five patients with secondary anemia were in the normal position at the normal plasma pH of the blood (7.4), and that only at the abnormal plasma pH of 7.64 was there a shift to the right. Several of the cases studied by Richards and Strauss had received transfusions of whole blood within a few days prior to the determination of the oxygen dissociation curves. In discussing the cause of the shift observed at plasma pH 7.64, they suggest that it could be due, in the patients with pernicious anemia, to an increased concentration of hemoglobin within the red cell, which would have the result, at this high plasma pH, of increasing the value $(-\log r)$ as developed by Van Slyke, Wu, and McLean (18, 19) and so increasing the difference between the plasma and cell pH. The hypothesis is admitted to be invalid in hypochromic anemia in which there is a shift of the oxygen dissociation curve to the right. Dill and his co-workers (3) report that in one patient with pernicious anemia the oxygen dissociation curve performed before treatment was normal but the subsequent curves during treatment with liver orally, and single curves in four other cases of pernicious anemia during treatment, were displaced to the right of normal. No curves were determined after recovery from anemia. Dill and his co-workers by indirect methods found the value rH to be smaller than normal and so conclude that the difference between plasma and cell pH is increased, the cell pH being relatively more acid than normal. Both Dill and his co-workers and Henderson (20) were unable to relate the observed shift in the oxygen dissociation curve to altered osmotic or electrolytic relationships between cell and plasma. Isac, Matthes, and Yamanaka (4) found that the oxygen dissociation curves in four patients with pernicious anemia, four patients with secondary anemia, one patient with aplastic anemia, and one patient with hemolytic

anemia were displaced to the right of normal. These authors, using indirect methods like Dill and his co-workers, were unable to demonstrate any decrease in the cell pH.

The present observations confirm that there is a shift to the right of the oxygen dissociation curve at standard plasma pH in the majority of patients with hypochromic anemia, secondary anemia, and megaloblastic anemia in relapse, thus providing for the anemic patient an increased yield of oxygen to the tissues. The time taken for the shift in the curve to lessen following treatment was in the patients with megaloblastic anemia some four to six weeks but it should be noted that many of these patients were the subjects of an investigation of the value of oral vitamin B_{12} in the therapy of pernicious anemia and were not always showing optimal hematological responses. It is not apparent why two of the patients with megaloblastic anemia had a normal oxygen dissociation curve initially but developed a shift to the right during the course of treatment. A similar development was observed by Dill and his co-workers. The present investigation does not confirm that there is a shift to the right of the oxygen dissociation curve in hemolytic anemia (4).

The beneficial effect of the displacement of the oxygen dissociation curve is obvious but the mechanism responsible for it is not apparent. A reduction of the plasma pH of the subject would displace the curve to the right but there is no evidence that acidosis is present and moreover the displacement of the curve is still present after correction to standard plasma pH. The presence of carboxyhemoglobin causes a shift to the left of the oxygen dissociation curve (17) and the position of the oxygen dissociation curve in the normal subject may be influenced to a small extent by the carboxyhemoglobin present (21). We have found smaller amounts of carboxyhemoglobin in the anemic subjects than in normal subjects but the quantities involved are so small that they could not produce the observed displacement of the oxygen dissociation curve. Alteration in the concentration of hemoglobin within the red cell also cannot be the factor responsible for the displacement of the oxygen dissociation curve since the displacement is demonstrable in both pernicious anemia and hemoglobin deficiency anemia. The effect of the increased plasma volume in anemia can

likewise be excluded as the cause of the displacement since artificial dilution with fresh plasma from the same individual does not influence the position of the oxygen dissociation curve (2, 5).

It is now accepted (22, 23) that many different kinds of hemoglobin, at least some of which may influence the position of the oxygen dissociation curve, may be present in pathological states. It is unlikely, however, that an abnormality of hemoglobin can be responsible for the observed displacement of the oxygen dissociation curve in anemia since the oxygen dissociation curves of hemoglobin solutions of anemic blood are the same as those of normal blood.

The position of the oxygen dissociation curve is dependent basically upon the cell pH and it is unfortunate that there is no entirely satisfactory method by which the cell pH can be determined directly. Hampson and Maizels (24) using a direct method at room temperature (glass electrode determinations on red cells hemolysed by repeated exposure to low temperature) found that the difference between the cell pH and plasma pH in pernicious anemia was greater than normal while in hypochromic anemia it was less than normal. Maizels (25), in referring to cell pH determinations, expresses doubts about his results partly because of the many manipulations involved and partly because the determinations were made at room temperature in conditions far removed from those in which red cells exist in the body. The indirect method for determining cell pH, in which the value "r" as developed by Van Slyke, Wu, and McLean (18) ($- \log r = pH_s - pH_c$) is determined, or the nomogram developed by Dill (see 6) is used, is superior to the direct method particularly as regards comparative studies, although the Henderson-Hasselbalch equation is not a strict physical chemical equation. If the indirect method, however, is applied to anemic blood where the cell phase is small, there is the disadvantage that experimental errors are multiplied. To avoid this disadvantage the cell pH was determined after concentrating the anemic blood samples by removal of plasma under oil to bring the packed cell volume into the normal range. This manipulation permits more accurate study.

Using this indirect method the cell pH was low relative to the plasma pH in cases of pernicious anemia in relapse and cases of hypochromic anemia

which showed a displacement of the oxygen dissociation curve to the right. On the other hand, in cases of pernicious anemia in therapeutic remission having the oxygen dissociation curve normal in position, the cell pH was normal. The cell pH was also normal in the case of pernicious anemia in relapse which showed a normal oxygen dissociation curve. In the cases in which the cell pH was low correction of the oxygen dissociation curve to the standard cell pH largely, but not completely, corrected the displacement to the right, suggesting that while a low cell pH is the main factor in producing the displacement there is also a further factor. In both cases of aplastic anemia the cell pH was also found to be low. Correction of the position of the oxygen dissociation curve in these cases to the standard cell pH wholly eliminated the displacement of the curve in the case of recent onset and partly corrected it in the case of longstanding. This patient had received numerous blood transfusions over several years and it would be unwise to draw any conclusions from examining this individual's oxygen dissociation curve since it would appear that one was in fact examining the oxygen dissociation curve of many donors. The cell pH was low in one of the two cases of hemolytic anemia in which the cell pH was determined and normal in the other. In the former case, correction of the position of the curve to the standard cell pH wholly corrected the slight displacement of the oxygen dissociation curve that was present at the cell pH of the subject. The cases of pernicious anemia studied during recovery showed that the oxygen dissociation curve is still slightly abnormal when the red cell count is about the 4 million per cubic mm. level, at which time there is still a degree of macrocytosis. In the cases in complete therapeutic remission, where there is no macrocytosis, there is no displacement of the oxygen dissociation curve.

While a lowered cell pH thus accounts for the major part of the displacement of the oxygen dissociation curve the results suggest that there is in addition a further factor. Measurements of the red cell diameter thickness ratio were not made in this study but the observed relationship between displacement of the oxygen dissociation curve and shape of the red cell in stored red blood cells (5, 10) indicates that such measurements might use-

fully be done in further studies in gas transport in anemia.

SUMMARY

1. The oxygen dissociation curves in twenty-nine cases of anemia have been studied.

2. In ten of the eleven cases of megaloblastic anemia in relapse and in nine of the ten cases of hypochromic anemia and secondary anemia there was a slight displacement of the oxygen dissociation curve to the right. No shift to the right was present in three cases of hemolytic anemia or in one of two cases of aplastic anemia.

3. The displacement of the oxygen dissociation curve persisted during the period of recovery in the cases of megaloblastic anemia but was not present in cases in therapeutic remission. The position of the curve was not influenced by the presence of reticulocytes.

4. The major part of the displacement appears to be due to an alteration in the cell pH.

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