

Supplementary Information

High-Throughput engineering of nonribosomal extension modules

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Table of Contents

Materials and methods	2
Molecular cloning	2
Production of tyrocidine synthetase proteins in <i>E. coli</i>	5
Protein purification	5
Supplementary figures	7
Protein sequences.....	14
DNA sequences.....	17
References.....	27

Materials and methods

All chemicals were purchased from Sigma Aldrich and enzymes from NEB unless otherwise stated. Primers (Table S1) were synthesized by Microsynth AG (Switzerland). Propargyl glycine was obtained from Bachem, whereas 4-propargyloxy phenylalanine was synthesized as described by Kries et al.¹ Media and buffer components, kits, and enzymes were used as received from specified commercial suppliers. Expression media and buffers were prepared using purified H₂O (Nanopure system, Barnstead).

Molecular cloning

The plasmids pSU18His-TycA and pTrc99a-TycB1, which respectively encode the C-terminally His₆-tagged TycA protein and the first module of TycB, were obtained from Grünewald et al.² Detailed descriptions of the construction of plasmids pSU18His-TycA, pSU18His-TycA_{pPhe}, pTrc99a-TycB1SrfTE_{P26G}, pTrc99a-TycB, pTrc99a-TycC, and pMG211-Sfp were reported previously.^{1,3,4} Plasmid pCT302⁵ was used for all yeast experiments but a NheI restriction site was replaced with an NdeI restriction site; the XhoI restriction site was also moved upstream of the c-myc tag, and all other XhoI sites were removed.⁶ The cloning protocol for generating pCTRB-TycA A-T and pCTRB- W227S TycA A-T, which contains the W227S mutation, was previously described.³

PCRs were conducted using Phusion HF Polymerase (NEB) and GC buffer according to the supplier's protocol (10 µL GC buffer 50-100 ng DNA template, 0.5 µM primer, 0.2 mM dNTPs and 0.5 µL Phusion Polymerase in 50 µL total volume). PCR products were purified by 1% agarose gel electrophoresis and isolated using the Zymoclean™ Gel DNA Recovery Kit (Zymo Research). The resulting PCR products were digested with the appropriate restriction enzymes in CutSmart Buffer at 37 °C for 1 h followed by a purification step using DNA Clean & Concentrator-5 Kit (Zymo Research). The restriction digest of the vector backbone (pSU18-His or pTr99a or pCTRB) vectors was conducted as described for the inserts but the incubation time was increased to 4 h. The vector backbone was also gel-purified and extracted with the Zymoclean™ Gel DNA Recovery Kit (Zymo Research). The ligation of digested PCR products and vector backbone (at a 6:1 ratio, 20-60 ng vector backbone) was performed at room temperature for 15 min using T4 DNA ligase. The ligation product was purified using the DNA Clean & Concentrator-5 Kit, and the plasmid was transformed into electrocompetent *E. coli* HM0079 cells. 0.8 mL SOC medium (0.5% yeast extract, 2% tryptone, 10 mM NaCl, 2.5 mM MgCl₂, 10 mM MgSO₄, 20 mM glucose) was immediately added to the cells. The transformed cells were grown at 37 °C, 230 rpm for 1 h. 100 µL of the cell solution was plated on LB agar plates supplemented with the appropriate antibiotic (sodium ampicillin for pTrc99a and chloramphenicol for pSU18-His). 5 mL LB pre-cultures containing sodium ampicillin (100 µg/mL) or chloramphenicol (30 µg/mL) were inoculated with a single *E. coli* colony and incubated at 37 °C, 230 rpm overnight. A 1.5 mL culture was centrifuged at 21,000 x g. Plasmids were extracted using the ZR Plasmid Miniprep Pure kit (Zymo Research) and sequenced at Microsynth AG (Balgach Switzerland).

Table S1: Primer list

Primer	Primer sequence 5' to 3'	Application
G79	TGCTCCTGTTGCAGGCTTTCC	Cloning TycB variant
G80	AAAGCCTGCAACAGGAGC	Cloning TycB variant
G103	GAGTGCCTGGTGGCCACCTACCTGCGAGATACAC	Cloning TycC variant
G112	GCAAAGATCGCTCGAGATG	Cloning TycB variant
G114	CTCTTTCATTGAGCTCCC	Cloning TycB variant
G150	AACATATCCGACACCGATGCGTC	Cloning TycB variant
G151	ATCGGTGTCGGATATGTTTGGC	Cloning TycB variant
P4n	TGGTGGTGGTTCTCATATGCACTTCAACGACACGGCC	Cloning TycB variant
P6n	ATAAGCTTTTGTTCCTCGAGAATGGCGGGCGCCGGCT	Cloning TycB variant
P16n	TGGTGGTGGTTCTCATATGTTCAACGGTGCGCACAAAG	Cloning TycC variant
P18n	ATAAGCTTTTGTTCCTCGAGGATCGGCACATACGTCTC	Cloning TycC variant
P30	CTCGAGGAACAAAAGCTTATT	Cloning TycC variant
P43	CATATGAGAACCACCACCA	General cloning
P44	GCCTTCGATGCCTTTGCCNNKACTTTCTTTACGTTGATTG	Library cloning
P45	GGCAAAGGCATCGAAGGC	Library cloning
P48	GCTCCATCCAGGCAGTCNNKCTCGGGGGCGAAAAGC	Library cloning
P49	GACTGCCTGGATGGAGC	Library cloning
P50	CGACGGAGAGCAGCGTCNNKGCCACCTACCTGCGAGA	Library cloning
P51	GACGCTGCTCTCCGTCG	Library cloning
P64	TGGTGGTGGTTCTCATATGTTCAACGATACGCACAGAG	Cloning TycB variant
P65	ATA AGC TTT TGT TCC TCG AGG ACA CCC TGC TCG CTC TC	Cloning TycB variant
P87	CCTTTGACGCTTTCGTTNNKTCCTTCTTTACGCCTGTGC	Library cloning
P88	AACGAAAGCGTCAAAGG	Library cloning
P89	CGATCTGGTCATCGTCAACNNKTACGGCCCCGACAGAAAG	Library cloning
P90	GTTGACGATGACCAGATCG	Library cloning
P91	CGACAGAAAGCAGTGTCNNKGCCACCTGGCAGCGC	Library cloning
P92	GACTGCTTTCTGTGTCG	Library cloning
P97	TGGTGGTGGTTCTCATATGGAACAGGCAGCCGGCG	Cloning TycB variant
P101	TGGTGGTGGTTCTCATATGGCCAAAGGGAATGTCTTCTCG	Cloning TycC variant
P104	GCTGCTGGTGGCGTTCCTGGATACGCACAGAGAATACC	Cloning TycB variant
P105	GAACGCCACCAGCAGC	Cloning TycB variant
P106	GGTATCCACCCGCCTGCATGCGTCAGGCTGGACG	Cloning TycB variant
P107	CAGGCGGGTGGATAACC	Cloning TycB variant

TycB variants

pCTRB-TycB2 A-T: To generate the plasmid for yeast surface display of the A and T domains of the second module of TycB, two PCR fragments were generated using the primer pairs H32/P43, P4n/P6n and the template pTrc99a-TycB, respectively (Table S1).

TycB2 gene library: To create space to accommodate pPhe, residues L1704, M1768, and M1803 in the active site of the TycB2 A domain were randomized using NNK codons. Initially, five fragments were generated using pTRc99a-TycB as a template and the following primer pairs: H32/P43, P4n/P45, P44/P49, P48/P51 and P50/P6n. In a second step, the first three fragments were assembled using primers H32 and P49 and the last two fragments using primers P48 and P6n. In a third step, the two assembled products were connected by overlap PCR using H32 and P6n (Table S1). The PCR product was then transformed into freshly prepared electrocompetent yeast cells together with the pCTRB vector backbone as described below.

pCTRB-M1768A/M1803I-TycB2 A-T: this plasmid was isolated from yeast after sorting the TycB2 library (see Sequencing plasmids from homologous recombination or library clones)

pTrc99a-M1768A/M1803I-TycB: To introduce the mutations that confer pPhe specificity into the full-length TycB gene, two fragments were produced by PCR using the primer pairs G112/G79 and G80/G114 and pCTRB-M1768A/M1803I-TycB2 AT and pTrc99a-TycB as templates, respectively (Table 5). The two fragments were subsequently assembled by PCR. The purified and assembled PCR product was digested with XhoI and SacI and incubated for 1 h at 37 °C. The digested insert was purified using the Clean and Concentrator-5 kit (Zymo Research). The purified insert and the pTrc99a vector backbone were ligated using T4 DNA ligase in a ratio of 1:6 at 16 °C for 16 h. The ligated plasmid was purified using the Clean and Concentrator-5 kit (Zymo Research) and transformed by electroporation into electrocompetent HM0079 cells. The cells were resuspended in 800 µL SOC medium which contains 0.5% yeast extract, 2% tryptone, 10 mM NaCl, 2.5 mM MgCl₂, 10 mM MgSO₄, 20 mM glucose. The suspension was grown for 1 h at 37 °C and plated onto LB-agar plates containing 100 µg/mL ampicillin. The plates were incubated at 37 °C for 16 h. 3 mL LB Miller medium were inoculated with a single colony and incubated at 37 °C, 230 rpm for 16 h. The plasmid was isolated using the ZymoPURE™ Plasmid Miniprep Kit and sequenced by Microsynth AG (Switzerland). A 20% (w/v) glycerol stock of the culture containing the plasmid was prepared and stored at -80 °C.

pCTRB-W2742S-TycB3 A-T: To generate the plasmid for yeast surface display of the A and T domains of the third module of TycB containing the W2742S mutation, three PCR fragments were generated using the primer pairs H32/P43 (template: pCTRB-TycA A-T), P64/G150 (template: pTrc99a-TycB), and G151/P65 (template: pTrc99a-TycB) (Table S1).

pCTRB-TycB3 C-A-T: To generate the plasmid for yeast surface display of the C, A and T domains of the third module of TycB, three PCR fragments were generated using the primer pairs H32/P43 (template: pCTRB-TycA A-T), P97/P65 (template: pTrc99a-TycB), and P30/H33 (template: pCTRB-TycA A-T) (Table S1).

pCTRB-W2742S-TycB3 C-A-T: To generate the plasmid for yeast surface display of the C, A, and T domains of the third module of TycB containing the W2742S mutation, three PCR fragments were generated using the primer pairs H32/P105 (template: pCTRB-TycB3 C-A-T), P104/P107 (template: pTrc99a-W2742S-TycB), and P106/H33 (template: pCTRB-TycB3 C-A-T) (Table S1).

TycC variants

pCTRB-TycC3 A-T: To generate the plasmid for yeast surface display of the A and T domains of the third module of TycC, two PCR fragments were generated using the primer pairs H32/P43 and P16n/P18n and the templates pCTRB-TycA A-T and pTrc99a-TycC, respectively (Table S1).

pCTRB-TycC6 C-A-T: To generate the plasmid for yeast surface display of the C, A and T domains of the last module of TycC, three PCR fragments were generated using the primer pairs H32/P43 (template: pCTRB-TycA AT), P101/G103 (template: pTrc99a-TycC), and P30/H33 (template: pCTRB-TycA AT) (Table S1).

Production of tyrocidine synthetase proteins in *E. coli*

To produce tyrocidine synthetase, the procedure reported by Niquille and Folger et al.⁴ was adopted with minor adaptations. The constituent proteins of the synthetase were produced with an N-terminal (TycA) or C-terminal (TycB variants and TycC) His₆-tag in *E. coli* strain HM0079.² The cell cultures were supplemented with chloramphenicol (30 µg/mL, PanReac AppliChem) for TycA production or with ampicillin (100 µg/mL, PanReac AppliChem) for TycB and TycC production. Briefly, 5 mL LB Miller broth containing the appropriate antibiotic were inoculated with a single HM0079 colony transformed with the desired plasmid (pSU18His-TycA, pTrc99a-TycB, pTrc99-M1786A/M1803I-TycB or pTrc99a-TycC) and incubated overnight at 37 °C, 230 rpm. The culture was prepared in a 2 L baffled flask, containing 800 mL modified Studier medium.¹⁰ The modified Studier medium is based on LB Miller broth to which MgSO₄ (2 mM), glycerol (Acros, 1% (m/v)), Na₂HPO₄ (25 mM), KH₂PO₄ (FisherBio, 25 mM), NH₄Cl (50 mM), and Na₂SO₄ (5 mM) were added. The medium was additionally supplemented with the appropriate antibiotic, inoculated at a 1:500 ratio with the pre-culture and incubated at 37 °C, 190 rpm for 4-6 h until an OD₆₀₀ of around 2 was reached. At this point, protein expression was induced by adding isopropyl β-d-1-thiogalactopyranoside (PanReac AppliChem, 250 µM), and the culture was incubated at 20 °C, 190 rpm for 20 h. The cells were centrifuged at 5,000 x g, 4 h for 20 min and the pellet was stored at -20 °C.

Protein purification

The heterologously expressed tyrocidine synthetase proteins were purified as reported by Niquille and Folger et al.⁴ Briefly, to lyse the HM0079 cells containing the protein of interest,

the thawed pellet was resuspended in 30 mL Tris-HCl (50 mM) buffer pH 8.0, supplemented with NaCl (500 mM), and glycerol (10% (w/v)). Polymyxin (Apollo, 1 mg/mL), RNase A (AppliChem, 10 µg/mL), DNase (PanReac AppliChem 10 µg/mL), and lysozyme (PanReac, 1 mg/ml) were added to increase lysis efficiency. The solution was incubated at 4 °C for 20 min before sonication at 60% amplitude in a Q700 sonicator (Qsonica) equipped with a 1/4 microtip (sonication time: 4 cycles of 30 s each with a 30 s break in between). The solution was centrifuged at 15,000 x g for 20 min at 4 °C and the supernatant containing the soluble protein was applied to pre-equilibrated Ni-NTA columns containing 5 mL Ni-beads (Quiagen). The collected flow-through was added to the Ni-NTA column again. The proteins bound to the Ni-NTA were washed with 40 mL Tris-HCl (50 mM) buffer pH 8.0, containing NaCl (500 mM), imidazole (Apollo Scientific, 20 mM), and glycerol (10% (w/v)). The bound protein was eluted from the column by addition of 10 mL Tris-HCl (50 mM) buffer pH 8.0, supplemented with NaCl (500 mM), imidazole (300 mM), and glycerol (10% (w/v)). The buffer was exchanged to Bis-Tris propane (Apollo, 50 mM) pH 7.25, containing NaCl (100 mM), MgCl₂ (10 mM), glycerol (10% (w/v)) using Amicon-Ultra-15 centrifugal filters (Merck, MWCO=10,000). The proteins were aliquoted, flash frozen in liquid nitrogen and stored at -80 °C until further use. Protein concentration was determined from the absorption at 280 nm measured on a Nanodrop 2000 spectrophotometer (Thermo Fisher). Extinction coefficients were calculated by the <https://web.expasy.org/protparam/> webtool. Proteins were analyzed on 7.5% SDS-PAGE gels using a Phast system to assess purity.

MS/MS analysis of the propargylated tyrocidine analogue

Aliquots (10 µL) from the quenched biosynthetic reaction mixtures were analyzed on an Agilent 1290 Infinity II liquid chromatography system coupled to a Bruker maXisII-ESI-Q-TOF MS system (Bruker Daltonics) and equipped with an Agilent ZorbaxEclipse Plus C18 column (3.5 µm 3.0 x 50 mm). The mobile phase was a mixture of 0.1% (v/v) formic acid in water and 0.1% (v/v) formic acid in acetonitrile. A linear gradient from 2% to 98% acetonitrile over 10 min was used at a flow rate of 0.6 mL/min. The ESI-TOF mass spectrometer was calibrated routinely for LC-MS in the positive electrospray ionization mode using the Agilent-ESI-TOF tuning mix in the enhanced quadratic algorithmic mode. The offset of each spectrum in the chromatogram was corrected with methylstearate and an Agilent Tunemix-mass 622 (m/z 299.2945 and 622.0290 respectively). The optimized source conditions were 8.0 l/h drying gas (nitrogen 99,99% purity) at a temperature of 180 °C, 1.6 bar nebulizer pressure, capillary and endplate voltages of 4,500 and 500 V, respectively, 12,000 V TOF flight tube voltage, 3,183 V reflection voltage, 1,700 V pusher voltage, and 2,268 V MCP detector voltage. The resolving power of the instrument was around 100,000 at m/z 1375 with 4 Hz spectra rate, depending on the sample concentration and peak width. For LC-MS/MS experiments (collision-induced dissociation; CID), the “Auto-MSMS-Mode” with an automatic precursor selection (5 Highest MS Signals) was used; isolation width m/z 3 at m/z 1375) and variable collision energy (20-90 eV; ISCID 0.0 eV) with nitrogen (99.99%) as collision gas (flow rate 28%). Data Analysis 5.1 (Built 201.2.4019) (64bit) software (Bruker Daltonics, Germany) was used for data processing.

Supplementary figures

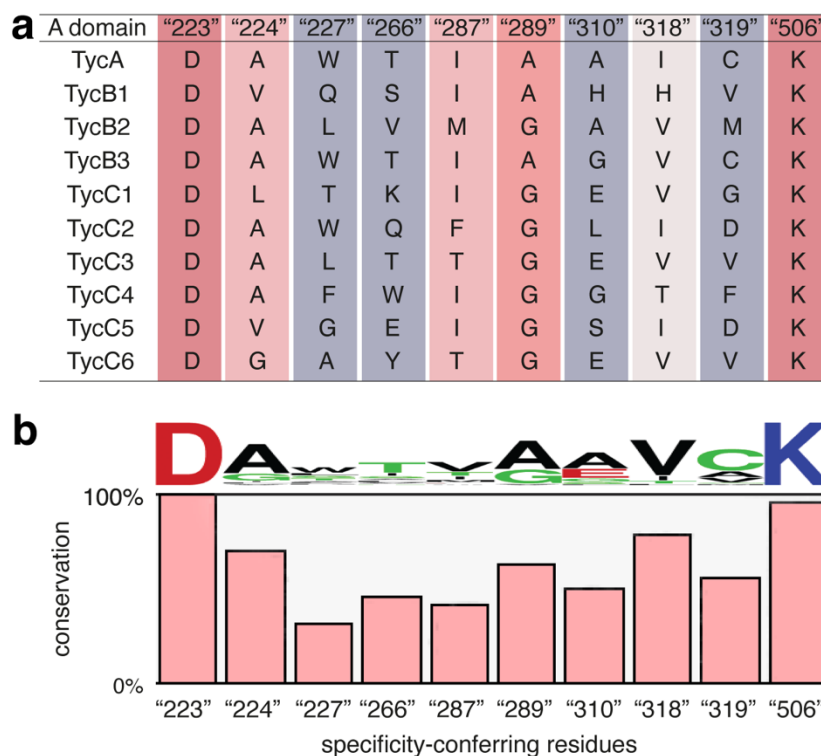


Figure S1: (a) Sequence alignment of the specificity determining residues of all ten A domains of tyrocidine synthetase. Dark red indicates high conservation and dark blue high variability. (b) Conservation of the specificity determining residues of 27 A domains, including TycA, TycB2, TycB3 and TycC3 from panel (a), that activate aromatic amino acids (twelve specific for Tyr, eleven for Phe, three for Trp, and one for phenyl glycine).^{13,14} TycA numbering is used. The sequence logo above the bar graph indicates the preferred residue(s).

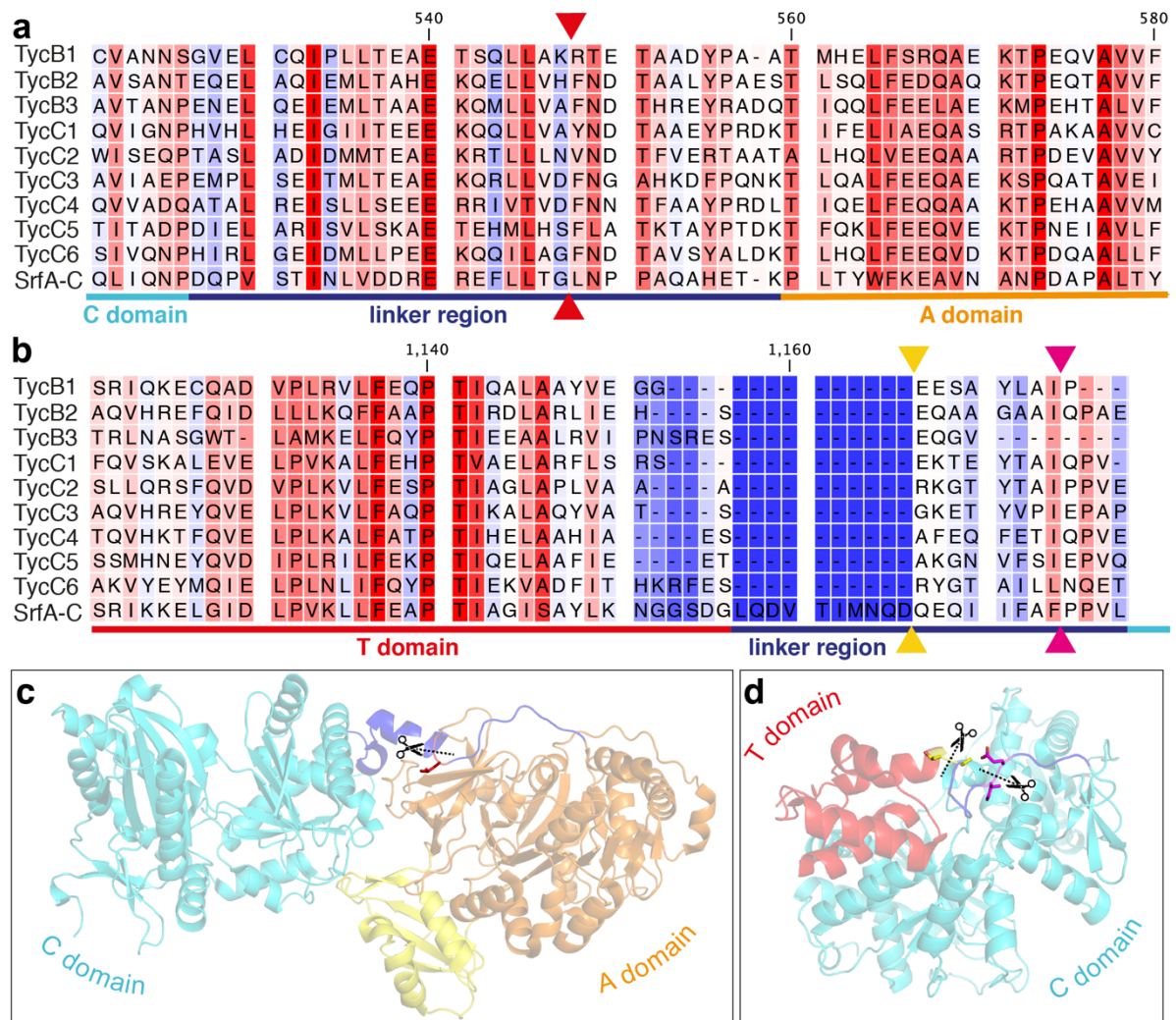


Figure S2. Cleavage sites for excising NRPS elongation modules for functional display on yeast. (a) Sequence alignment of the linker regions connecting the C and A domains of all TycB and TycC modules with structurally characterized SrfA-C, which served as a reference. The N-terminal excision site selected for the A-T constructs is indicated by red triangles. Dark red indicates high conservation, whereas dark blue indicates high variability. (b) The C-terminal excision site for the A-T and C-A-T constructs (magenta triangles) and the N-terminal excision site for C-A-T constructs (yellow triangles). (c) Crystal structure of SrfA-C (PDB: 2VSQ, residues 398-510),¹¹ showing the N-terminal excision site for the A-T constructs (red). The C domain is colored cyan, the A domain orange/yellow, and linker regions dark blue. (d) Crystal structure of the TycC5-TycC6 T-C bidomain (PDB: 2JGP)¹² showing the N-terminal excision site for C-A-T constructs (yellow) and C-terminal (magenta) excision site for A-T and C-A-T constructs. The T and C domains are colored red and cyan, respectively.

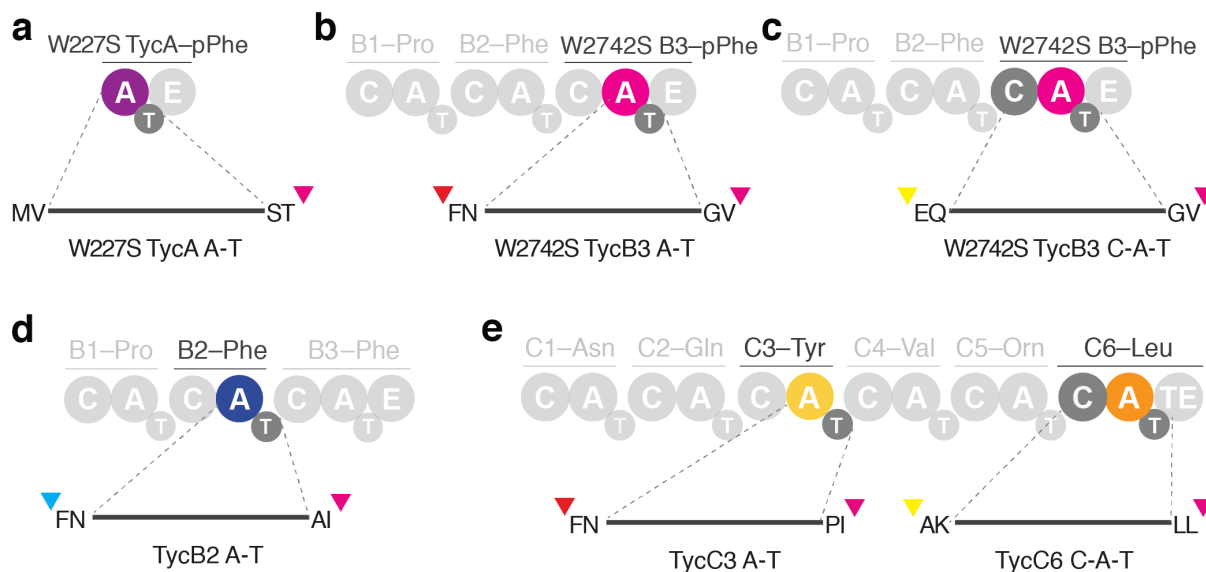


Figure S3. Tyrocidine synthetase domains/modules chosen for yeast display, showing their native context within the megaenzyme. The segment below the synthetase indicates the N- and C-terminal amino acids of the display constructs with triangles, colored as in Figure S2, indicating the cleavage sites chosen. (a) The previously described TycA A-T bidomain.³ Its A domain (purple) contained the W227S mutation that confers pPhe specificity. The corresponding (b) A-T and (c) C-A-T constructs from TycB3 are shown. The A domain of TycB3 (magenta) contained the W2742S mutation to accommodate pPhe. (d) The analogous A-T construct from TycB2 was engineered in high-throughput to switch its substrate specificity from L-Phe to pPhe. (e) The A-T construct from TycC3 (yellow) and the C-A-T construct from TycC6 (orange) are illustrated.

WT	NSVTWNRDEF	ALSVRDSGTL	SLSFAFDAFA	LTFFTLIVSG
v1
v2
v3
v4
v3

1704

WT	IQAVMLGGEK	LSPKLVQLCK	AMHPQMSVMN	AYGPTSSVM
v1G.....
v2A.....I.....
v3G.....V.....
v4C.....V.....
v3G.....V.....

1768

1803

Figure S4: TycB2 mutations that confer activity with pPhe.

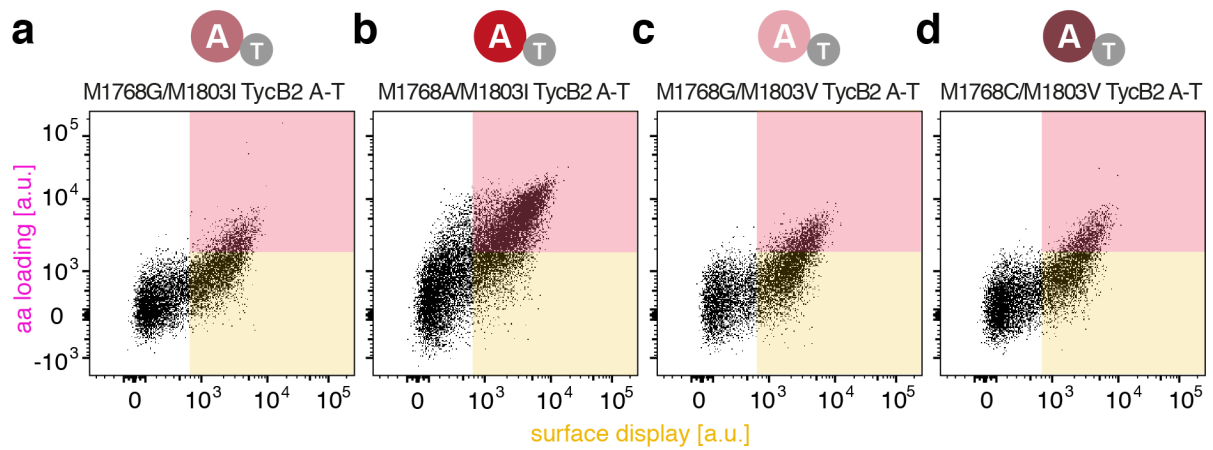


Figure S5: The activity of TycB2 variants 1 (a), 2 (b), 3 (c) and 4 (d) displayed on the surface of yeast. Assays were performed with 0.1 mM ATP and 0.5 mM pPhe in the presence of 0.5 mM Phe as a competitive inhibitor. The fraction of cells in the pink quadrant correlates with the ability of the displayed A domain to activate pPhe and transfer it to the ppan cofactor of the T domain tethered to the cell surface.

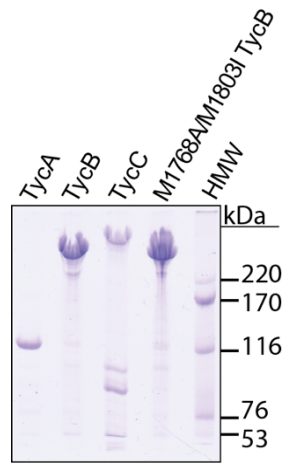


Figure S6: 7.5% SDS-PAGE gel showing purified His₆-tagged TycA, TycB, TycC and the variant M1768A/M1803I TycB produced in *E. coli* HM007. HMW= High molecular weight ladder (GE).

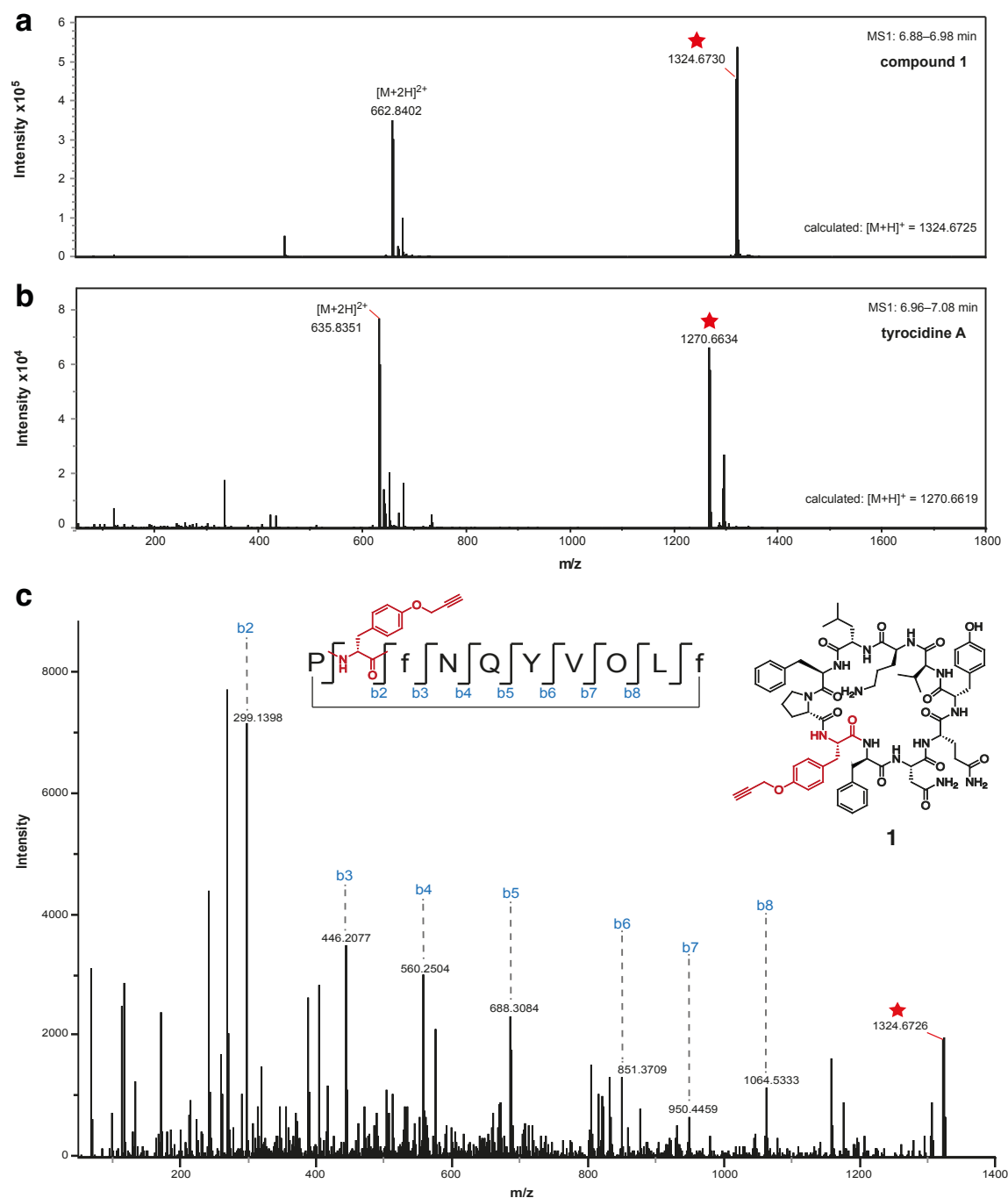


Figure S7: MS/MS characterization of the products of the *in vitro* biosynthetic reaction catalyzed by TycA, M1768A/M1803I TycB and TycC in the presence of pPhe. (a) The selected mass of the major product that elutes at a retention time of 6.88–6.98 (see Fig. 5 in the main text) is consistent with the propargyloxy-tyrocidine derivative **1** (calculated $[M+H]^+$ 1324.6725 Da; found 1324.6730 Da). (b) The side product that elutes at a retention time of 6.96–7.08 min is wildtype tyrocidine A (calculated $[M+H]^+$ 1270.6619; found 1270.6634 Da). (c) MS2 spectrum of the major product confirms its assignment to structure **1**, with the noncanonical pPhe incorporated site-specifically as the third residue in the natural product analogue. The observed fragmentation masses (b-ions) are shown in blue below the single letter code of the sequence, the sites of fragmentation are marked with dashed lines, and the parent ion is highlighted with a red star. O stands for Orn and f for D-Phe; all other capital single letters refer to L-amino acids.

Protein sequences

Aga-HA-W2742S-TycB3-A-T-c-myc (protein sequence displayed on yeast, bold amino acid marks start of NRPS protein and highlighted in blue is W2742S mutation)

MQLLRCSFISVIA SVLAQELTTICEQIPSPLESTPYSLSSTTTILANGKAMQGVFEYYKSVTFVSNCGSHPSTTSKGSPI
INTQYVFKDNSSTIEGRYPYDVPDYALQASGGGGSGGGSGGGGSHMFNDTHREYRADQTIQQLFEEELAEKMPEH
TALVFEKRMSFRELNERANQLAAVLREKGVGPAQIVALLVERSAEMVIATLATLKAGGAFLPVDPDYPEERIRYM
LEDSQAKLVVTHAHLHLHKVSSQSEVVDVDDPGSYATQTDNLPCANTPSDLAYIIYTSGTTGKPKGVMLEHKGVAN
LQAVFAHHLGVTPQDRAGHFASISFDASVSDMFGPLLSGATLYVLSRDVINDFQRF AEYVRD NAITFLTLPTTYAIY
LEPEQVPSRLTLITAGSASSVALVDKWKEKVTYVNGYGPTTESTVCATLWKAKPDEPVEITITIGKPIQNTKLYIVDDQ
LQKAPGMGELCISGLSLARGYWNRPELTA EKFDVNDPFPVPGTKMYRTGDLARWLPDGTIEYLRIDHGVKIRGH
RVELGEVESVLLRYDVTKEAAAITHEDDRGQAYLCAYYVAEGEATPAQLRAYMENELPNYMVPAFFIQLEKMP
PNDKIDRKALPKPNQEENRTEQYAAPQTELEQLLAGIWADVLGIKQVGTQDNFFELGGDSIKAIQVSTRLNASGWT
LAMKELFQYPTIEEAALRVIPNSRESEQGVLEEQKLISEEDL

Aga-HA-W2742S-TycB3-C-A-T-c-myc (protein sequence displayed on yeast, bold amino acid marks start of NRPS protein and highlighted in blue is W2742S mutation)

MQLLRCSFISVIA SVLAQELTTICEQIPSPLESTPYSLSSTTTILANGKAMQGVFEYYKSVTFVSNCGSHPSTTSKGSPI
INTQYVFKDNSSTIEGRYPYDVPDYALQASGGGGSGGGSGGGGSHMEQAAGAAIQPAEPQAYYPVSSAQQRMY
LLHQLEGAGISYNTPGIIMLEGKLDREQLANALQALVDRHDILRTSFEMVGDELVQKIHDRVAVNMEYVTAEEQQI
DDLHFHAFVRPFDLSVPLLRLMSLVKLADERHLLLYDMHHAADAASITILFDELAELYQGRELP EMRIQYKDFAVW
QKALHESDAFKQQEAYWLSTFAGNITAVDVPTDFPRPAVKSFAGQVTLSDMQELLSALHELAAHTNTTLFMVLL
AAYNVLLAKYAGQDDIIVGTPISGRSRAELAPVVGFMFVHTLAIRNKPTAEKTFKQFLQEVKQNALDAFDHQDY PFE
SLVEKLGIPRDPGRNPLFDTMFILQNDLHAKTLDQLVYRYPYEDSALDVAKFDLSFHLTERETDLFLRLEYCTKLF
KQQTVERMAHHLFLQILRAVTANPENELQEIEMLTAAEKQMLLVAFNDTHREYRADQTIQQLFEEELAEKMPEHTAL
VFEKRMSFRELNERANQLAAVLREKGVGPAQIVALLVERSAEMVIATLATLKAGGAFLPVDPDYPEERIRYMLED
SQAKLVVTHAHLHLHKVSSQSEVVDVDDPGSYATQTDNLPCANTPSDLAYIIYTSGTTGKPKGVMLEHKGVANLQA
VFAHHLGVTPQDRAGHFASISFDASVSDMFGPLLSGATLYVLSRDVINDFQRF AEYVRD NAITFLTLPTTYAIYLEPE
QVPSRLTLITAGSASSVALVDKWKEKVTYVNGYGPTTESTVCATLWKAKPDEPVEITITIGKPIQNTKLYIVDDQLQK
APGMGELCISGLSLARGYWNRPELTA EKFDVNDPFPVPGTKMYRTGDLARWLPDGTIEYLRIDHGVKIRGHRVEL
GEVESVLLRYDVTKEAAAITHEDDRGQAYLCAYYVAEGEATPAQLRAYMENELPNYMVPAFFIQLEKMP
PNDKIDRKALPKPNQEENRTEQYAAPQTELEQLLAGIWADVLGIKQVGTQDNFFELGGDSIKAIQVSTRLNASGWT
LAMKELFQYPTIEEAALRVIPNSRESEQGVLEEQKLISEEDL

Aga-HA-TycC3-A-T-c-myc (protein sequence displayed on yeast, bold amino acid marks start of NRPS protein)

MQLLRCSFISVIA SVLAQELTTICEQIPSPLESTPYSLSSTTTILANGKAMQGVFEYYKSVTFVSNCGSHPSTTSKGSPI
INTQYVFKDNSSTIEGRYPYDVPDYALQASGGGGSGGGSGGGGSHMFNGAHKDFPQNKTLLQALFEEQAEKSPQA
TAVEISGQPLSYQELNERANQLAATLRERGVQPDQPVGIMANRSVEMVVGILAILKAGGAYVPIDPEYPEERVAYM
LTDCQARLVLTQKHLGAKLGSSVTAECYLDDESNGVHRSNLQPINTASDLAYIIYTSGTTGKPKGVMVEHRGIV
NNVLWKKAEYQMKVGDRSLLSLSFAFDVLSFFTPVLSGATVVLAEDEEAKDPVSLKLLIAASRCTLMTGVPSLF
QAILECSTPADIRPLQTVTLGGEKITAQLVEKCKQLNPDLVIVNEYGPTSESSVATWQRLAGPDAAITIGRPIANTSL
YIVNQYHQLPQIGVVEICIGGRGLARGYWNKPALTEEFVSHPF AAGERMYKTGDLGKWLDPGTIEYIGRIDEQV
KVRGYRIEIGIEIESALLAEKLTAAVVVVYEDQLQGSALAA YFTADEQLDVTKLWSHLSKRLPSYMPAHFVQLDQ
LPLTPNGKVDKALPKPEGKPVTEAQYVAPTNAVESKLAIEWVERVLGVSGIGILDNFFQIGGHSLKAMAVAAQVHR
EYQVELPLKVLFAQPTIKALAQYVATSGKETYVPILEEQKLISEEDL

Aga-HA-TycC6-C-A-T-c-myc (protein sequence displayed on yeast, bold amino acid marks start of NRPS protein)

MQLLRCSFISVIAVLAQELTTICEQIPSPLESTPYSLSSTTILANGKAMQGVFEYYKSVTFVSNCGSHPSTTSKGGSP
INTQYVFKDSSSTIEGRYPYDVPDYALQASGGGGGGGGGGGGSHMAKGNVFSIEPVQKQAYYPVSSAQKRMYYI
LDQFEGVGISYNMPSTMLIEGKLERTRVEAAFQRLIARHESLRTSFAVNGEPVQNIHEDVPFALAYSEVTEQEARE
LVSSLVQPFDELVAPLIRVSLKIGEDRYVLFDTMHHSISDGVSSGILLAEVWVQLYQGDVLPRLIQYKDFAVWQQE
FSQSAAFHKQEAYWLQTFADDIPVLNLPDFTFRPSTQSFAGDQCTIGAGKALTEGLHQLAQATGTTLYMVLLAAY
NVLLAKYAGQEDIIVGTPITGRSHADLEPIVGMFVNTLAMRNKQREKTFSEFLQEVKQNALDAYGHQDYPFEEVLV
EKLAIARDLSRNPLFDTVFTFQNSTEEVMTLPECTLAPFMTDETGGHAKFDLTFSAATEEREEMTIGVEYSTSLFTRET
MERFSRHFLTIAASIVQNPHIRLGEIDMLLPEEKQQLAGFNDAVSYALDKTLHQLFEEQVDKTPDQAALLFSEQSL
TYSELNERANRLARVLRKAGVGPDRLLVAIMAERSPEMVGILGILKAGGAYVPVDPGYPQERIQYLLSNAALLS
QAHLPLLAQVSSPECLDLNAELDAGLSGNLPAVNQPTDLAYVIYTSGTTGKPKGVMIPHQGIVNCLQWRDE
YGFGPSDKALQVFSFAFDGFVASFAPLLGGATCPLQEAADKDPVALKKLMAATEVTHYYGVPSLQAILDCSTT
TDFNQLRCVTLGGKELPVQLVQKTKKHPAIEINNEYGPTENSVVTTISRSEAGQAITGRPLANVQVYIVDEQHHL
QPIGVVVELCIGGAGLARGYLKPELTAEKFVANPFRPGERMYKTGDLVKWRDGTIEYIGRADEQVKVRGYRIE
GEIESAVLAYQGIDQAVVVARDDDATAGSYLCAVFVAATAVSVSGLRSHLAKELPAYMIPSYFVELDQLPLSANG
KVDRKALPKPQQSDATTREYVAPRNATEQQLAAIWQEVLGVEPIGTDQFFELGGHSLKATLLIAKVVEYMQIELPL
NLIFQYPTIEKVADFIHKRFESRYGTAILLLEQKLISEEDL

Aga-HA-TycB2-A-T-c-myc (protein sequence displayed on yeast, bold amino acid marks start of NRPS protein and highlighted in red are target sites for mutagenesis)

MQLLRCSFISVIAVLAQELTTICEQIPSPLESTPYSLSSTTILANGKAMQGVFEYYKSVTFVSNCGSHPSTTSKGGSP
INTQYVFKDSSSTIEGRYPYDVPDYALQASGGGGGGGGGGGGSHMFNDTAALYPAESTLSQLFEDQAQKTPEQ
TAVVFGDKRLTYRELNERANQLAHTLRKAGVQAEQSVGIMAQRSLEMAIGIHAAILKAGGAYVPIDPDYPNERIAYM
LEDCEARLVLTQQQLAEKMTANVECLYLDEEGSYSPQ TENIEPIHTAADLAYIIYTSGTTGRPKGVMVEHRGIVNSV
TWNRDEFALSVRDSGTLSSFAFDALFTFTLIVSGSTVVLMPDHEAKDPIALRNLIAAWECYSVVFPSPMFQAIL
ECSTPADIRSIQAVMLGGKLSPKLVQLCKAMHPQMSVMNA YGPTESSVMATYLRDTPDQIPITGRPIANTAIYIV
DQHHQLLPVGVVGEICIGGHGLARGYWKPELTAEKFVANPAVGERMYKTGDLGRWLHDGTIDFIGRVDQIKV
RGYRIEVEIEAVLLAYDQTNEAIVVAYQDDRGSYLAAYVTGKTAIESELRAHLLRELPAVMPTYLIQLDAFPL
TPNGKVDRKALPKPEGKPATGAAYVAPATEVEAKLVAIWENALGISGVGLDHFELGGHSLKAMTVVAQVHRE
FQIDLLKQFFAAPTIRDLARLIEHSEQAAGAAILEEQKLISEEDL

M1768A/M1803I-TycB-His (best protein variant isolated from the library for activation of *4-propargyloxy phenylalanine*. The mutated residues are highlighted in red)

MSVFSKEQVQDMYALTPMQEGMLFHALLDQEHNSHLVQMSISLQGDLDVGLFTDSLHVLVERYDVFRTLFLYEK
LKQPLQVVLKQRPPIEFYDLSACDESEKQLRYTQYKRADQERTFHLAKDPLMRVALFQMSQHDYQVIWSFHILM
DGWCFSIIFDILLAIYLSLQNKTALESLEPVQPYSRFINWLEKQNKQAALNYWSDYLEAIEYEQKTTLPKKEAAFAKAF
QPTQYRFLSNRTLKQLGTIASQNVTLSTVIQTIWGVLLQKYNAAHDLVFGSVVSGRPTDIVGIDKMVGLFINTIPF
RVQAKAGQTFSELLQAVHKRTLQSQPYEHVPLYDIQTQSVLKQELIDHLLVIENYPLVEALQKKALNQQIGFTITAV
EMFEPTNYDLTVMVMPKEELAFRFDYNAALFDEQVQKLAGHLQQIADCVANNSGVELCQIPLLTEAETSQLLAK
RTETAADYPAATMHELFSRQAEKTPEQVAVVFADQHLTYRELDEKSNQLARFLRKKGIGTGSLVGTLLDRSLDMI
VGILGVLKAGGAFVPIDPELPAERIAYMLTHSRVPLVVTQNHLRAKVTTPTETIDINTAVIGESRAPIESLNQPHDLF
YIIYTSGTTGQPKGVMLEHRNMANLMHFTFDQTNIAFHEKVLQYTTCSFDVCYQEIFSTLLSGGQLYLITNELRRHV
EKLFAFIQEKQISILSLPVSFLKIFNEQDYAQSFPKCVKHIITAGEQLVVTHELQKYLQRHVRFLHNHYGPSETHVVT
TCTMDPGQAIPELPPIGKPISTNGIYLDEGLQLKPEGIVGELYISGANVGRGYLHQPELTAEKFLDNPYQPGERMYR
TGDLARWLPDGGLEFLGRIDHQVKIRGHRIELGEIESRLNHPAIEAVVIDRADETGGKFLCAYVVLQKALSDEEM
RAYLAQALPEYMIPSFVTLERIPVTPNGKTDRRALPKPEGSAKTKADYVAPTTELEQKLVAIWEQILGVSPIGIQDH
FFTLLGGHSLKAIQLISRIQKQADVPLRVLFQOPTIQAALAYVEGGEESAYLAIPQAEPQAYYPVSSAQKRMILNQL
LDPHSTVYNLPVAMILEGTLDKARLEHAISNLVARHESLRTSFHTINGEPVSRIHEQGHLPIVYLETAEQVNEVILG
FMQPFDLVTAPLCRVGLVLAENRHVLIIDMHHSISDGVSSQLILNDFSRLYQNKALPEQRIHYKDFAVWEKAWTQ
TTDYQKQEKYWLDRFAGEIPVLNLPMDYPRPAVQSFEGERYLFRTKQLLESQDVAQKTGTTLYMVLLAAYHVL
LSKYSQDDVMIGTVTAGRVHPDTEMTGMFVNTLAMRNQSAPTQFRQFLLEVKDNTLAAFEHGQYPFEELVEK
LAIQRNRSRNPLFDTLFIQNMDADLIELDGLTVTPYVPEGEVAKFDLSLEASENQAGLSFCFEFCTKLFARETIERM

SLHYLQILQAVSANTEQELAQIEMLTAEHEKQELLVHFNDTAALYPAESTLSQLFEDQAQKTPEQTAVVFGDKRLTY
RELNERANQLAHTLRAGVQAEQSVGIMAQRSLEMAIGHAILKAGGAYVPIDPDYPNERIAYMLEDCEARLVLTQ
QQLAEKMTANVECLYLDEEGSYSPQTENIEPIHTAADLAYIIYTSGTTGRPKGVMVEHRGIVNSVTWNRDEFALSV
RDSGTLSLSFAFDALFTFFTLIVSGSTVVLMPDHEAKDPIALRNLIAAWECYSYVVFVPSMFQAILECSTPADIRSIQA
VALGGEKLSPKLVQLCKAMHPQMSVMNAYGPTESSVIATYLRDTQPDQPITIGRPIANTAIYIVDQHHQLLPVGVV
GEICIGHGLARGYWKPELTAEFVANPAVPERMYKTGDLGRWLHDGTIDFIGRVDDQIKVRGYRIEVGEIEAV
LLAYDQTNEAIVVAYQDDRGDSYLAAYVTGKTAIEESELRAHLLRELPAVMVPTYLIQLDAFPLTPNGKVDRKALP
KPEGKPATGAAYVAPATEVEAKLVAIWENALGISGVGLDHFELGGHSLKAMTVVAQVHREFQIDLLKQFFAA
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MHHAADAASITILFDELAELYQGRELPEMRIQYKDFAVWQKALHESDAFKQQEAYWLSTFAGNITAVDVPTDFPR
PAVKSFAGGQVTLSDQELLSALHELAHAHTNTLFLMVLAAYNVLLAKYAGQDDIIVGTPISGRSRAELAPVVG
FVHTLAIRNKPTAEKTFKQLQEVKQNALDAFDHQDYPFESLVEKLGIPRDPGRNPLFDTMFILQNDLHAKTLDQL
VYRPPYESDSALDVAKFDLSFHLTERETDLFLRLEYCTKLFKQQTVERMAHHFLQILRAVTANPENELQEIEMLTAA
EKQMLLVAFNDTHREYRADQTIQQLFEELAEKMPHEHTALVFEEKRMSFRELNERANQLAAVLREKGVGPAQIVAL
LVERSAEMVIATLTLKAGGAFLPVDPDYPEERIRYMLEDSQAKLVVTHAHLHKVSSQSEVVVDVDDPGSYATQT
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PTESTVCATLWKAKPDEPVETITIGKPIQNTKLYIVDDQLQLKAPGQMGELCISGLSLARGYWNRPELTAEFVDNP
FVPGTKMYRTGDLARWLPDGTIEYLGRIDHQVKIRGHRVELGEVESVLLRYDVTVEAAAITHEDDRGQAYLCAYY
VAEGEATPAQLRAYMENELPNYMVPAFFIQLEKMPPLTPNDKIDRKALPKPNQEENRTEQYAAPQTELEQLLAGIW
ADVLGKQVGTQDNFFELGGDSIKAIQVSTRNLASGWTLAMKELFQYPTIEEAALRVIPNSRESEQGVVEGEIALPI
QKWWFANNFTDRHHWNQAVMLFREDGFDEGLVRQAFQQIVEHHDALRMVYKQEDGAIKQINRGLTDERFRFYSY
DLKNHANSEARILELSDQIQSSIDLEHGPLVHVALFATKDGDLHLLVAIHHLVVDGVSWRILFEDFSSAYSQALHQE
IVLPKKTDSFKDWAAQLQKYADSDELLREVAYWHNLETTTTTAAALPTDFVTADRKQKHTRTSLFALTPQTENLL
RHVHHAYHTEMNDLLLALGLAVKDWHTNGVVINLEGHGREDIQNEMNVTRTIGWFTSQYPVLDMEKAEDL
PYQIKQTKENLRIPKKGIGYEILRTLTTSQLQPPLAFTLRPEISFNLYLGGFESDGKTGGFTSPLGTGQLFSPESERV
LLDISAMIEDGELRISVGYRSLQYEEKTIASLADSYRKHLLGIEHCMAKEEGEYTPSDLGDEELSMEELENILEWIGS
RSHHHHHH

DNA sequences

pCTRB vector backbone

TAGGTCGAGATCTGATAACAACAGTGTAGATGTAACAAAATCGACTTTGTTCCCACTGTACTTTTAGCTCGTAC
AAAATACAATATACTTTTCATTCTCCGTAACAACATGTTTTCCCATGTAATATCCTTTTCTATTTTTTCGTTCC
GTTACCAACTTTACACATACTTTATATAGCTATTCACCTTCTATACACTAAAAAACTAAGACAATTTTAATTTTG
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GGCTGAACGCGGGGTTCTGTCACACAGCCAGCTTGGAGCGAAGCAGCACTACACCGAATGAGATACCTACAG
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ACCTCTGACTTGAGCGTCGATTTTTGTGATGCTCGTCAGGGGGGCCGAGCCTATGGA AAAACGCCAGCAACGC
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ATTACTTCTTATTCAAATGTAATAAAGATCGAATTCTACTTCATACATTTTCAATTAAG (Open reading frame)

Aga-HA-W2742S-TycB3-A-T-c-myc (ORF of pCTRB-W2742S-TycB3 A-T with the start codon in bold)

ATGCAGTACTTCGCTGTTTTTCAATATTTTCTGTTATTGCTTCAGTTTTAGCACAGGAACTGACAACATATATGC
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AGGACTTG

Aga-HA-W2742S-TycB3-C-A-T-c-myc (ORF of pCTRB-W2742S-TycB3 C-A-T with the start codon in bold)

ATGCAGTACTTCGCTGTTTTTCAATATTTTCTGTTATTGCTTCAGTTTTAGCACAGGAACTGACAACATATATGC
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CGCTCGTATTGAAAGAAAAGCGCATGTCGTTCCGGGAGCTCAATGAAAGAGCGAACAGCTCGCAGCCGTTTT
GCGGGAAAAAGGAGTCGGGCCAGCGCAGATCGTCGCTTTGCTGGTAGAGCGTTCCGCGGAGATGGTCATTGC
CACGCTTGCCACGTTAAAAGCGGGCGGCGCCTTTTTGCCCGTCGATCCTGATTATCCGGAAAGAGCGAATCCGC
TACATGCTGGAGGACAGCCAGGCAAAAATGGTGGTGACCCATGCGCACTTGTGCACAAAAGTGAGCAGTCAG
TCCGAAGTCGTTGATGTGGATGACCCCTGGAAGTACGCAACACAGACAGACAACCTGCCGTGCGCAAAACACA
CCGTCGTATTGGCTTATATCATTTACACGTCGCGTACGACGGGCAAGCCAAAAGCGCTCATGTGGAGCACA
AAGGGGTAGCAATCTGCAAGCGGTATTTGCCCATCATCTAGGCGTCACGCCGCAAGATCGGGCAGGGCATT
TGCCAGCATCTGTTTGCAGCATCGGTGTCGGATATGTTTGGCCCGTTGCTGTCGGGAGCGACCTTGTACGTCT
TGTCCCAGACGTCATCAACGATTTTCAACGATTTCGCCGAATACGTTTCGCGATAACGCGATCACCTTCCTCACT
TTGCCGCCGACGTACGCGATTTATCTGGAGCCGGAGCAGGTGCCGTCGTTACGCACCCTGATTACAGCCGGAT
CGGCTTCTCCGTTGCATTGGTGGATAAATGGAAAGAAAAAGTACCTATGTCAATGGATACGGCCCAACAGA
GAGCACCGTTTGCAGCACACTGTGGAAGCCAAACCGGATGAGCCAGTCGAAACGATCACGATTGGCAAAC
GATTCAGAACACCAAGCTGTACATCGTGGATGACCAGTTGCAAGTTGAAAGCGCCGGGCGAGATGGGAGAACT
GTGCATCAGCGGCTTGTGCTGGCGAGAGGCTATTGGAATCGTCCAGAGCTGACCGCCGAGAAGTTCGTCGAC
AACCCGTTTGTGCCAGGAACAAAGATGTACCGGACAGGGCAGCTGGCAAGATGGCTGCCAGATGGAACATC
GAGTATCTGGGCAGAATCGATCACCAGTGAATAATCGCGGACATCGTGTGGAACCTCGGCGAAGTGGAAAGC
GTGCTGCTGCGGTATGACACGGTCAAAGAGGCGGCCGCATCACACATGAGGACGACCGCGCCAAGCTTAC
TTGTGCGCCTACTACGTAGCGGAGGGAGAAGCCACGCCTGCGCAACTTCGAGCCTATATGGAAAACGAGTTG
CCGAACACATGTTTCCCGCCTTCTTATCCAGTTGAAAAGATGCCGCTGACCCGAATGACAAGATTGACC
GAAAAGCGCTGCCGAAAGCCGAAAGGAGGAGAACCGGACTGAGCAATATGCACCGCGCAACCCGAGCTG
GAACAGTTGCTGGCTGGCATCTGGGCAGATGTAAGGATCAAGCAAGTCGGGACGCAAGACAACCTTCTTTG
AATTGGGCGGCGATTGATTAAGCGATCCAGGTATCCACCCGCTGAATGCGTCAGGCTGGACGCTTGCAT
GAAAGAAGCTTCCAGTACCCGACGATTGAAGAAGCTGCTCTGCGCGTCATCCCGAACAGCCGAGAGAGCGA
GCAGGGTGTCTCGAGGAACAAAAGCTTATTTCTGAAGAGGACTTG

Aga-HA-TycC3-A-T-c-myc (ORF of pCTRB-TycC3 A-T with the start codon in bold)

ATGCAGTTACTTCGCTGTTTTTCAATATTTTCTGTTATTGCTTCAGTTTTTAGCACAGGAACTGACAACTATATGC
GAGCAAATCCCCTACCAACTTTAGAATCGACGCCGACTCTTTGTCAACGACTACTATTTTGGCCAACGGGA
AGGCAATGCAAGGAGTTTTTGAATATTACAAATCAGTAACGTTTGTGAGTAATTGCGGTTCTCACCCCTCAAC
AACTAGCAAAGGCAGCCCATAAACACACAGTATGTTTTAAGGACAATAGCTCGACGATTGAAGGTAGATA
CCCATACGACGTTCCAGACTACGCTCTGCAGGCTAGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGT
GGT
AACAAGCGAAAAGTCGCCGACGCAACAGCCGTGGAATCAACGCGGACGCCCTGTCTTACAGGAGCTGA
ATGAGCGAGCCAACCAGCTTCCGCTACGCTGCGGGAGCGGGAGTACAGCCTGACCAACCTGTAGGGATTA
TGGCGAACCGCTCTGTGGAGATGGTCTGCGCATCCTCGCCATCTTGAAGCAGGCGGAGCTTACGTGCCGAT
CGACCCGGAATATCCGGAGGAGCGTGTGCGCTACATGCTGACGGATTGCCAAGCCCGCCTGGTGTGACGCA
AAAGCATCTGGGAGCGAAGCTTGGTTCCAGCGTGACCGCGAATGCCTGTATCTCGACGACGAGAGCAACTA
TGGTGTGACCCGCTCGAATTTGCAGCCGATCAATACCGCTTCCGATCTGGCTTACATCATCTACACATCGGGTA
CGACTGGCAAGCCAAAAGGGTCAATGGTTCGAGCACCCGGGATCGTCAACAACGTCGTGTGGAAGAAAAGCGG
AGTACCAAATGAAGTTGGCGACAGAAGCTTGTGCTCTGTCCTTTGCTTTGCTTTGCTTTGCTTTGCTTTGCTTT
TTACGCTGTGCTTTCCGGGGCAACTGTCGTAAGGCGGAGGATGAAGAAGCCAAGGACCCAGTCTCTTTGAA
AAAGCTCATCGCCGCTTCGCGCTGCACCTTGTGACAGGCGTGCCGAGCTTGTCCAGGCCATTCTGGAATGC
AGCACGCCAGCGGATATCCGTCGCTGCAAAACCGTCACTCGGCGGAGAAAAAATTACGGCGCAGCTTGT
GAAAAATGCAAGCAGCTGAATCCCGATCTGGTCACTGCAACGAGTACGGCCGACAGAAAAGCAGTGTGCTG
GCCACCTGGCAGCGCCTTGCGGGTCCGGATGCTGCCATCACCATCGGGCGGCGGATTGCCAACACCAGCCGT
ACATCGTGAACCAATATACACAGCTACAGCAATCGGCGTGGTGGTGGGAGATTTGCATCGGCGGCCCGCTT
GGCAGAGGCTATTGGAACAAGCCAGCGCTACGGAAGAGAAGTTGCTTTCCCATCCGTTTGCAGGAGGCGA
GCGCATGTACAAGACGGGCGATCTTGGCAAGTGGCTCCCGGACGGAACGATTGAATACATTGGGCGCATCGA
CGAACAGTCAAAGTCCGAGGCTACCGAATTGAAATCGGCGAGATCGAGTCGGCTCTGCTGGCTGCGGAAAA
GCTGACAGCGGCTGTTGTGGTCTGCTATGAGGATCAGCTTGGCCAGTCGGCTCTGGCAGCGTATTTACCGCC
GACGAACAGCTTGTGTCACGAAGCTGTGGTTCGATCTGTCGAAGCGACTCCCGTCGTACATGATTCTGCGC

ATTTTGTGCAGCTCGATCAGCTTCCGCTTACGCCAAACGGCAAAGTCGACAAGAAAGCCTTGCCGAAGCCAGA
AGGCAATGCCCGTAACCGAAGCGCAATATGTCGCGCCGACAAATGCGGTGAAAGCAAGCTGGCAGAGATTTG
GGAACGCGTGCTCGGGGTTAGCGGCATCGGCATTCTCGACAACTTTTCCAGATCGGCGGACATTCCTTGAAA
GCGATGGCTGTCGTGCACAGGTGCATCGCGAGTATCAGGTTGAGCTTCCGCTGAAAGTGTCTGTCGCGCAGC
CTACGATCAAGGCGTGGCCCAGTATGTCCACGAGCGGAAAAGAGACGTATGTGCCGATCCTCGAGGAAC
AAAAGCTTATTTCTGAAGAGGACTTG

Aga-HA-TycC6-C-A-T-c-myc (ORF of pCTRB-TycC6 C-A-T with the start codon in bold)

ATGCAGTTACTTCGCTGTTTTTCAATATTTTCTGTTATTGCTTCAGTTTTTAGCACAGGAACTGACAACCTATATGC
GAGCAAATCCCCTACCAACTTTAGAATCGACGCCGACTCTTTGTCAACGACTACTATTTTGGCCAAACGGGA
AGGCAATGCAAGGAGTTTTTGAATATTACAAATCAGTAACGTTTGTAGTAATTGCGGTTCTCACCCCTCAAC
AACTAGCAAAGGCAGCCCATAAAACACACAGTATGTTTTAAGGACAATAGCTCGACGATTGAAGGTAGATA
CCCATACGACGTTCCAGACTACGCTCTGCAGGCTAGTGGTGGTGGTGGTTCTGGTGGTGGTGGTTCTGGTGGT
GGTGGTTCTCATATGGCCAAAGGGAATGTCTTCTCGATCGAGCCTGTGCAAAAGCAAGCGTACTATCCGGTCT
CCTCGGCACAAAAGCGCATGTACATCTCGATCAATTTGAGGGAGTCGGCATCAGCTACAACATGCCGTGAC
TATGCTGATCGAAGGCAAGCTGGAGCGAACACGGGTAGAAGCGGCGTTCCAGCGCTTGATTGCGCGACATGA
AAGCCTGCGCACTTCGTTTCCGTCGTCACGGAGAGCCTGTGCAAAACATTACAGGAGGACGTTCCGTTTGGC
CTTGCTATTCCGGAAGTACAGAACAGGAGGCGCGCAACTCTGTTTCTCTGTCGAGCCGTTGATCCTGATCG
AGGTCGCAACACTCATCCGCGTGTCTGCTGAAAATCGGCGAGGATCGTTACGTGCTCTTACCGACATGCA
TCACAGCATTTCCGATGGCGTATCCTCCGGCATTCTTTTTGGCAGAGTGGGTGACGCTGTACCAGGTTGACGTT
TGCCGGAGCTGCGTATCCAGTACAAGGACTTTGCTGTGTGGCAACAAGAGTTTTCCAGTCGGTGCCTTCCA
CAAGCAGGAAGCGTACTGGTTGCAAACGTTTGGCGATGACATTCTGTGCTGAACTTGCCGACCGATTTACC
CGCCCCAGCACCCAAAGCTTTGCCGGGGATCAGTGCACGATCGGCGCGGGCAAAAGCGTACCGGAAGGCTTG
CACCAGTTGGCGCAGGCGACGGGAACGACTTTGTACATGGTTTTTCTGCCGCGTACAACGTGTGCTCGCCA
AGTATGCCGGGAGGAGACATCATCGTCGGCACGCCGATTACAGGCAGATCCCATGCCGATCTCGAACC
TCGTCGGCATGTTCTGTGAACACCTTGGCGATGCGAAAACAACCGCAGCGCGAAAAGACTTTTAGCGAGTTTT
GCAAGAAGTCAAGCAAAATGCGCTGGATGCGTACGGCCATCAGGATTACCCGTTTGAAGAAGTGGTGGAAAA
GCTCGCATCGCGCGGATTTGAGCCGAAATCCGCTGTTGACACCGTGTTCAGTTCCAAAACAGCAGCGGAA
GAGGTCATGACGCTGCCTGAATGCACGTTGCGCCGTTTATGACGGACGAAACAGGCCAGCAGCCAAGTTC
GACTTGACTTTCAGCGCTACGGAAGAGCGGGAAGAAATGACGATTGGCGTGGAGTACAGCACAAGCTTGTT
ACGCGGAAACGATGGAACGGTTTCAGCCGCACTCTCTGACGATTGTCAGCGAGCCTGTGCAAAATCCGCA
ATCCGCTGCGGAGATCGACATGCTTTTTGCCAGAGAATAACAGCAGATTTTGGCCGGTTCAACGATACGG
CAGTCAGCTATGCGCTGGACAAAACGCTGACCAGATTTGAAAGAGCAGGTGACAAAACCCGATCAGG
CAGCGCTTCTCTTGTAGCGAGCAATCGCTGACGTACAGCGAACTGAACGAGCGAGCAAACAGACTGGCAAGGG
TCCTGCGCGCAAAGGAGTCGGACCGGACCCTGTTGTTAGCGATCATGGCGGAGCGCTCGCCGAAATGGTGA
TCGGTATTCTCGGTATTTTGAAGGCAGCGCGCTTATGTTCCCGTCGATCCCAGGTATCCGCAAGAGCGCATT
CAGTACCTGCTCGAAGATAGCAACCGCAGCCCTGCTGCTCAGCCAGGCGCATCTGTTGCCGCTGTTGGCCAGG
TGTCAAAGCGAGCTGCCGGAGTGCCTTGATCTGAACGCTGAACTGGATGCCGGACTGAGCGGTCCAACCTGCC
AGTGTCAACCAACCGACTGACCTTGCTTGCCTACGTATCTATACATCCGGTACGACCGGAAGCGGAGGTTGTC
ATGATCCCGCATCAAGGAATCGTGAATGCTTGCAGTGGAGAAGAGAGCAATAACGGTTCGGGCGGATGAC
AAGGCGTGTGCAAGTGTCTCTTTGCCCTTCGACGGTTTTGTAGCCAGCTTGTTCGCTCCGCTGCTCGGAGGGG
AACGTGCGTGTGCGCAAGAAGCAGCTGCCAAAGACCCGGTCGCGCTGAAAAAACTGATGGCCGCAACCGGA
AGTCACCCATTAACGCGCTACCGAGTCTGTTCCAGGCCATTCTCGATTGCTCGACGACAACCGACTTCAATC
AGTTGCGTTGCGTCACTTTGGGCGCGGAGAAGCTGCTGTCAGCTTGTGCAAAAACAAGGAAAAGCAGT
CGGCAATCGAGATCAACAACGAGTACGGCCCCAGGAAACAGCGTCTCACCACCTCTCGCGCTCGATTG
AAGCGGGCAAGCAGTACGATTTGGCCAGCCGTTGCCAACGTCGCAAGTCTACATTGTAGATGAGCAGCATC
ACTTGCAGCCGATTGCGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGG
ACCGGAGCTGACCGCAGAGAAGTTTTGTGCAAAATCCGTTCCGACCAGGCGAGCGCATGTACAAAACAGGCGA
CTTGGTAAAAATGGCGGACGGATGGCACGATCGAGTACATCGGCCGCGCAGACGAACAGGTCAAGGTGAGAGG
GTATCGCATCGAGATCGGCGAGATCGAGAGCGCCGACTCGCTTACCAGGGCATCGATCAAGCGGTGGTCTGT
GCGCGAGACGATGACGCTACGGCTGGTTTCTATCTTTGCGCTACTTTGTGCGAGCAACAGCCGTGTCCGTATC
CGGCTTGAAGCCATCTGGCCAAAGAGCTGCTTGCATGATTCCGAGCTATTTCTGTCGAGCTGGATCAG
CTGCCGTTTTCCGCAATGGAAAAGTGGATCGCAAAGCTTTGCCGAAAGCCGCAACAGTCCGATGCGACCAGC
CGCAATACGTGGCCCGAGGAATGCGACCGAACAGCAACTGGCAGCCATCTGGCAAGAAGTTTTGGGAGTAG
AGCAATCGGCATCACCGACCAGTTCTTTGAACTCGGAGGACATTCCTAAAAGCTACGCTGTTGATTGCCAA
AGTGATGAGTACATGCAAATCGAGCTGCCGCTGAATCTCATCTTCCAGTATCCGACGATCGAAAAGGTGGCC
GATTTTCATCACGCATAAGCGCTTTGAGAGCAGATACGGCACAGCCATTTTGTTACTCGAGGAACAAAAGCTT
ATTTCTGAAGAGGACTTG

Aga-HA-TycB2-A-T-c-myc (ORF of pCTRB-TycB2 A-T with the start codon in bold)

ATGCAGTTACTTCGCTGTTTTTCAATATTTTCTGTTATTGCTTCAGTTTTTAGCACAGGAACTGACAACCTATATGC
GAGCAAATCCCCTACCAACTTTAGAATCGACGCCGACTCTTTGTCAACGACTACTATTTTGGCCAAACGGGA
AGGCAATGCAAGGAGTTTTTGAATATTACAAATCAGTAACGTTTGTAGTAATTGCGGTTCTCACCCCTCAAC

AACTAGCAAAGGCAGCCCCATAAACACACAGTATGTTTTTAAGGACAATAGCTCGACGATTGAAGGTAGATA
CCCATACGACGTTCAGACTACGCTCTGCAGGCTAGTGGTGGTGGTGGTTCGGTGGTGGTGGTTCGGTGGT
GGTGGTTCATATGCACTTCAACGACACGGCCGCCCTGTATCCAGCGGAGAGCACGCTGTGCGAGCTGTTTG
AAGATCAGGCACAGAAAACCTCTGAGCAAACCGCCGTCGTCTTCGGTGACAAACGACTGACGTACCCGCGAAC
TGAACGAGCGGGCCAAACAGCTCGCGCACACTTTGCGGGCAAAGGCGTGCAGGCTGAGCAAAGCGTAGGGA
TCATGGCGCAAAGATCGTTAGAGATGGCGATCGGAATCATCGCTATTCTCAAAGCGGGCGGGCGTATGTGCC
GATCGATCCGGATTATCCGAATGAGCGGATTGCTTACATGCTGGAAGATTGCGAAGCCCGTCTGGTGTGACC
CAGCAGCAGCTCGCCGAAAAGATGACCGCAAACGTGGAATGCCTGTATCTGGATGAGGAGGGCAGCTACTCG
CCTCAGACGGAACATCGAGCCGATCCATACCGCTGCTGATCTCGCTTACATCATCTACACATCCGGTACGA
CAGGCAGGCCAAAAGGCGTCATGGTAGAGCATCGGGGAATCGTCAACAGTGTGACGTGGAACAGGGACGAGT
TTGCCCTTTCTGTCCGGGACAGTGGAACGCTGTCGCTATCTTTTGCCTTCGATGCCTTTGCCCTTACTTTCTTTA
CGTTGATTGTATCAGGCTCCACGGTCTGCTGATCGCGGATCACGAAGCCAAAGATCCGATCGCGCTACGCAA
CCTGATTGCCGCTTGGGAATGCAGCTACGTCGTTTTCTGTCGCCAGTATGTTCCAGGCATATTGGAGTGCAGCA
CTCCGGCAGACATCCGCTCCATCCAGGCAGTCATGCTCGGGGCGAAAAGCTGTGCGCCGAAGCTTGTTACGCT
GTGCAAAGCGATGCATCCGAGATGAGCGTGATGAATGCATACGGCCCGACGGAGAGCAGCGTCATGGCCAC
CTACCTGCGAGATACACAGCCAGATCAGCCGATCACCATCGGGCGGCCGATTGCCAACACCGCCATTACATC
GTAGACCAGCACCATCAACTGCTGCCTGTGCGGGTGGTAGGGGAAATCTGCATCGGCGGTACAGGCTTGCGCG
GGGGCTATTGGA AAAAGCCGGAGCTTACTGCGGAGAAATTCGTGGCCAATCCAGCTGTTCCGGGAGAGCGCA
AATCAAAAACAGGCGATCTGGGCAGATGGCTCCACGACGGCAGGATGATTTTATAGGCCGCGTCGATGACCA
AATCAAGGTGAGAGGATACCGGATTGAGGTGCGGGAGATTGAAGCGTTTTGCTCGCTTACGATCAGACGAA
TGAAGCTATCGTCGTCGCTTATCAGGACGATCGCGGCGATTCTATCTGGCTGCGTATGTCACGGGAAAAACG
GCGATAGAGGAATCCGAGCTTCGCGCGCATCTGTTGCGAGAGCTTCCGGCCTACATGGTGCCGACCTACCTGA
TTCAACTGGACGCTTTCCCGCTCAGCCAAACGGCAAGGTCGACCGCAAGGCAAGGCAAGCCGGAAGGAA
AGCCTGCAACAGGAGCAGCTTATGTCGCACCCGCTACAGAAGTGGAGGCGAAGCTGGTGCCTATTGGGAGA
ATGCGCTGGGGATTTCGGCGTTCGGGGTGTGGATCACTTTTTTGGAGCTGGGCGGTCATTCTTGAAGCGGAT
ACGGTTGTGGCGCAAGTGCATCGCGAGTTCAAATCGACCTTTTGTGAAGCAGTTTTTTCGACGCGCAACCAT
CCGGGACTTGGCCCGCTTGATCGAACATAGCGAACAGGCAGCCGGCGCCGCCATTCTCGAG**GAACAAAAGCT**
TATTCTGAAGAGGACTTG

Aga-HA-M1768A/M1803I-TycB2-A-T-c-myc (ORF of pCTR B-M1768A/M1803I-TycB2 A-T with the start codon in bold)

ATGCAGTACTTCGCTGTTTTTCAATATTTTCTGTTATTGCTTACGTTTTAGCACAGGAACTGACA ACTATATGC
GAGCAAATCCCCTACCAACTTTAGAATCGACGCCGACTCTTTGTCAACGACTACTATTTTGCCAACGGGA
AGGCAATGCAAGGAGTTTTTGAATATTACAAATCAGTAACGTTTGTGAGTAATTGCGGTTCTACCCCTCAAC
AACTAGCAAAGGCAGCCCCATAAACACACAGTATGTTTTTAAGGACAATAGCTCGACGATTGAAGGTAGATA
CCCATACGACGTTCAGACTACGCTCTGCAGGCTAGTGGTGGTGGTGGTTCGGTGGTGGTGGTTCGGTGGT
GGTGGTTCATATGCACTTCAACGACACGGCCGCCCTGTATCCAGCGGAGAGCACGCTGTGCGAGCTGTTTG
AAGATCAGGCACAGAAAACCTCTGAGCAAACCGCCGTCGTCTTCGGTGACAAACGACTGACGTACCCGCGAAC
TGAACGAGCGGGCCAAACAGCTCGCGCACACTTTGCGGGCAAAGGCGTGCAGGCTGAGCAAAGCGTAGGGA
TCATGGCGCAAAGATCGTTAGAGATGGCGATCGGAATCATCGCTATTCTCAAAGCGGGCGGGCGTATGTGCC
GATCGATCCGGATTATCCGAATGAGCGGATTGCTTACATGCTGGAAGATTGCGAAGCCCGTCTGGTGTGACC
CAGCAGCAGCTCGCCGAAAAGATGACCGCAAACGTGGAATGCCTGTATCTGGATGAGGAGGGCAGCTACTCG
CCTCAGACGGAACATCGAGCCGATCCATACCGCTGCTGATCTCGCTTACATCATCTACACATCCGGTACGA
CAGGCAGGCCAAAAGGCGTCATGGTAGAGCATCGGGGAATCGTCAACAGTGTGACGTGGAACAGGGACGAGT
TTGCCCTTTCTGTCCGGGACAGTGGAACGCTGTCGCTATCTTTTGCCTTCGATGCCTTTGCCCTTACTTTCTTTA
CGTTGATTGTATCAGGCTCCACGGTCTGCTGATCGCGGATCACGAAGCCAAAGATCCGATCGCGCTACGCAA
CCTGATTGCCGCTTGGGAATGCAGCTACGTCGTTTTCTGTCGCCAGTATGTTCCAGGCATATTGGAGTGCAGCA
CTCCGGCAGACATCCGCTCCATCCAGGCAGTCGCGCTCGGGGCGAAAAGCTGTGCGCCGAAGCTTGTTACGCT
GTGCAAAGCGATGCATCCGAGATGAGCGTGATGAATGCATACGGCCCGACGGAGAGCAGCGTCATTGCCAC
CTACCTGCGAGATACACAGCCAGATCAGCCGATCACCATCGGGCGGCCGATTGCCAACACCGCCATTACATC
GTAGACCAGCACCATCAACTGCTGCCTGTGCGGGTGGTAGGGGAAATCTGCATCGGCGGTACAGGCTTGCGCG
GGGGCTATTGGA AAAAGCCGGAGCTTACTGCGGAGAAATTCGTGGCCAATCCAGCTGTTCCGGGAGAGCGCA
TGTACAAAACAGGCGATCTGGGCAGATGGCTCCACGACGGCAGGATGATTTTATAGGCCGCGTCGATGACCA
AATCAAGGTGAGAGGATACCGGATTGAGGTGCGGGAGATTGAAGCGTTTTGCTCGCTTACGATCAGACGAA
TGAAGCTATCGTCGTCGCTTATCAGGACGATCGCGGCGATTCTATCTGGCTGCGTATGTCACGGGAAAAACG
GCGATAGAGGAATCCGAGCTTCGCGCGCATCTGTTGCGAGAGCTTCCGGCCTACATGGTGCCGACCTACCTGA
TTCAACTGGACGCTTTCCCGCTCAGCCAAACGGCAAGGTCGACCGCAAGGCAAGGCAAGCCGGAAGGAA
AGCCTGCAACAGGAGCAGCTTATGTCGCACCCGCTACAGAAGTGGAGGCGAAGCTGGTGCCTATTGGGAGA
ATGCGCTGGGGATTTCGGCGTTCGGGGTGTGGATCACTTTTTTGGAGCTGGGCGGTCATTCTTGAAGCGGAT
ACGGTTGTGGCGCAAGTGCATCGCGAGTTCAAATCGACCTTTTGTGAAGCAGTTTTTTCGACGCGCAACCAT
CCGGGACTTGGCCCGCTTGATCGAACATAGCGAACAGGCAGCCGGCGCCGCCATTCTCGAG**GAACAAAAGCT**
TATTCTGAAGAGGACTTG

pTrc99a-M1768A/M1803I-TycB-His (The TycB start codon is in bold)

CTAGAGTCGACCTGCAGGCATGCAAGCTTGGCTGTTTTGGCGGATGAGAGAAGATTTTCAGCCTGATACAGAT
TAAATCAGAACGCAGAAGCGGTCTGATAAAACAGAATTTGCCTGGCGGCAGTAGCGCGGTGGTCCCACCTGA
CCCCATGCCGAACTCAGAAGTGAAACGCCGTAGCGCCGATGGTAGTGTGGGGTCTCCCCATGCGAGAGTAGG
GAACTGCCAGGCATCAAATAAAACGAAAGGCTCAGTCGAAAGACTGGGCCTTTCGTTTTATCTGTTGTTTGTCTC
GGTGAACGCTCTCCTGAGTAGGACAAATCCGCCGGAGCGGATTTGAACGTTGCGAAGCAACGGCCCCGGAGG
GTGGCGGGCAGGACGCCCCGATAACTGCCAGGCATCAAATTAAGCAGAAGGCCATCCTGACGGATGGCCT
TTTTGCGTTTTCTACAACTCTTTTGTATTATTTTCTAAATACATTCAAATATGTATCCGCTCATGAGACAATAAC
CCTGATAAATGCTTCAATAATATTGAAAAAGGAAGAGTATGAGTATTCAACATTTCCGTGTCGCCCTTATTCCC
TTTTTTCGGGCATTTTGCCTTCCTGTTTTTGTCTACCCAGAAAACGCTGGTGAAAAGTAAAAGATGCTGAAGATCA
GTTGGGTGCACGAGTGGGTACATCGAACTGGATCTCAACAGCGGTAAGATCCTTGAGAGTTTTCGCCCCGAA
GAACGTTTTCCAATGATGAGCACTTTTAAAGTCTGCTATGTGGCGCGGTATTATCCCGTGTGACGCCGGCA
AGAGCAACTCGGTCGCCGCATACACTATTCTCAGAATGACTTGGTTGAGTACTACCAGTCACAGAAAAGCAT
CTTACGGATGGCATGACAGTAAGAGAATTATGCAGTGTGCCATAACCATGAGTGATAAACTGCGGCCAACT
TACTTCTGACAACGATCGGAGGACCGAAGGAGCTAACCGCTTTTTTGCACAACATGGGGGATCATGTAACCTCG
CCTTGATCGTTGGGAACCGGAGCTGAATGAAGCCATACCAAACGACGAGCGTGACACCACGATGCCTACAGC
AATGGCAACAACGTTGCGCAAACCTATTAACCTGGCGAACTACTTACTTAGCTTCCCGGCAACAATTAATAGAC
TGGATGGAGGCGGATAAAGTTGCAGGACCACTTCTGCGCTCGGCCCTTCCGGCTGGCTGGTTATTGCTGATA
AATCTGGAGCCGGTGAGCGTGGGTCTCGCGGTATCATTGCAGCACTGGGGCCAGATGGTAAGCCCTCCCGTAT
CGTAGTTATCTACACGACGGGGAGTCAGGCAACTATGGATGAACGAAATAGACAGATCGTGTAGATAGGTGC
CTCACTGATTAAGCATTGGTAACTGTCAGACCAAGTTTACTCATATATACTTTAGATTGATTTAAAACCTTCATT
TTTAATTTAAAAGGATCTAGGTGAAGATCCTTTTTGATAATCTCATGACCAAAAATCCCTAACGTGAGTTTTTCG
TTCCACTGAGCGTCAGACCCCGTAGAAAAGATCAAAGGATCTTCTTGAGATCCTTTTTTCTGCGCGTAATCTG
CTGCTTGCAAACAAAAAACCCAGCTACCAGCGGTGGTTTTGTTTGGCGGATCAAGAGCTACCAACTTTTTT
CCGAAGGTAACGGCTTCAGCAGAGCGCAGATACCAATACTGCTCTTAGTGTAGCCGTAGTTAGGCCACC
ACTTCAAGAACTCTGTAGCACCGCTACATACCTCGCTCTGCTAATCCTGTTACCAGTGGCTGCTGCCAGTGGC
GATAAGTCGTGTCTTACCGGTTGGACTCAAGACGATAGTTACCGGATAAGGCGCAGCGGTGCGGGCTGAACG
GGGGTTCGTGCACACAGCCAGCTTGGAGCGAACGACCTACACCGAACTGAGATACCTACAGCGTGAGCTA
TGAGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCGGACAGGTATCCGGTAAGCGGCAGGGTTCGGAACAGG
AGAGCGCACGAGGGAGCTTCCAGGGGAAACGCTGGTATCTTTATAGTCTGTCGGGTTTCGCCACCTCTGA
CTTGAGCGTCGATTTTTGTGATGCTCGTCAGGGGGCGGAGCCTATGGAAAAACGCCAGCAACGCGGCCTTTT
TACGGTTCTGGCCTTTTGTGGCCTTTTGTCTACATGTTCTTTCTGCGTTATCCCCTGATTCTGTGGATAACC
GTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCCGACGCCGAACGACCGAGCGCAGCGAGTCAGTGAGCG
AGGAAGCGGAAGAGCGCCTGATGCGGTATTTTCTCCTTACGCATCTGTGCGGTATTTACACCCGCATATGGTG
CACTCTCAGTACAATCTGCTCTGATGCCGCATAGTTAAGCCAGTATAACTCCGCTATCGCTACGTGACTGGGT
CATGGCTGCGCCCCGACACCCGCCAACACCCGCTGACGCGCCCTGACGGGCTTGTCTGCTCCCGGCATCCGCT
TACAGACAAGCTGTGACCGTCTCCGGGAGCTGCATGTGTCAGAGTTTTTACCCTCATCACCGAAACGCGCGA
GGCAGCAGATCAATTCGCGCGCGAAGGCGAAGCGGCATGCATTTACGTTGACACCATCGAATGGTGCAAAAC
CTTTCGCGGTATGGCATGATAGCGCCCGGAAGAGAGTCAATTCAGGGTGGTGAATGTGAAACCAGTAACGTTA
TACGATGTCGACAGATATGCCGGTGTCTCTTATCAGACCGTTTTCCCGCTGGTGAACCAGGCCAGCCACGTTT
CTGCGAAAACGCGGGAAAAAGTGGAAGCGGCGATGGCGGAGCTGAATTACATTCCAACCGCTGGCACAAC
AACTGGCGGGCAAACAGTCTGTTGCTGATTGGCGTTGCCACCTCCAGTCTGGCCCTGCACGCGCCGTCGCAAAAT
TGTCGCGGCGATTAAATCTCGCGCCGATCAACTGGGTGCCAGCGTGGTGGTGTGATGGTAGAACGAAGCGGC
GTCGAAGCCTGTAAAGCGGCGGTGCACAATCTTCTCGCGCAACGCGTCAGTGGGCTGATCAATTAATATCCGC
TGGATGACCAGGATGCCATTGCTGTGGAAGCTGCCTGCACTAATGTTCCGGCGTTATTTCTTGATGCTCTGAC
CAGACACCCATCAACAGTATTATTTTCTCCCATGAAGACGGTACGCGACTGGGCGTGGAGCATCTGGTTCGCAT
TGGGTACCAGCAAATCGCGCTGTTAGCGGGCCATTAAGTTCTGTCTCGGCGCGTCTGCGTCTGGCTGGCTG
GCATAAATATCTCACTCGCAATCAAATTCAGCCGATAGCGGAACGGGAAGGCGACTGGAGTGCCATGTCCGG
TTTTCAACAAACCATGCAAATGCTGAATGAGGGCATCGTTCCCACTGCGATGCTGGTTGCCAACGATCAGATG
GCGCTGGGCGCAATGCGCGCCATTACCGAGTCCGGGCTGCGCGTTGGTGGCGGATATCTCGGTAGTGGGATACG
ACGATACCGAAGACAGCTCATGTTATATCCCGCCGTTAACACCATCAAACAGGATTTTCGCCTGCTGGGGCA
AACCAGCGTGGACCGCTTGTGCAACTCTCTCAGGGCCAGGCGGTGAAGGGCAATCAGCTGTTGCCGCTCTCA
CTGGTGA AAAAGAAAAACCCTGGCGCCAATACGCAAACCGCCTCTCCCCGCGGTTGGCCGATTCATTAA
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GAATTGATCTGGTTTACAGCTTATCATCGACTGCACGGTGCACCAATGCTTCTGGCGTCAGGCAGCCATCGG
AAGCTGTGGTATGGCTGTGAGGTGCTAAATCACTGCATAATTCGTGTGCTCAAGGCGCACTCCCCTTCTGG
ATAATGTTTTTTCGCGGACATCATAACGGTTCTGGCAAATATTCTGAAATGAGCTGTTGACAATTAATCATCC
GGCTCGTATAATGTGTGGAATTGTGAGCGGATAACAATTTACACAGGAAAGGTACCATGAGTGTATTTAGCA

AAGAACAAGTTCAGGATATGTATGCGTTGACCCCGATGCAAGAGGGGATGCTGTTTTACGCCTTGCTCGATCA
AGAGCACAACCTCGCATCTGGTACAGATGTCGATTTTCGTTGCAGGGCGATCTTGACGTTGGGCTATTTACGGAT
AGCCTGCATGTGCTGGTAGAGAGATACGATGTATTCCGCACGTTGTTTCTCTATGAAAAGCTGAAGCAGCCTT
TGCAAGTTGCTTGAAGCAACGGCCTATTCCGATCGAATTTTACGACTTGTCTGCCTGCGACGAGTCCGAGAA
ACAACCTTCGTATACGCAATACAAAAGGGCGGATCAGGAGCGGACGTTTCATCTGGCAAAAGACCCGTTGAT
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