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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For	all statistical an	alyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.				
n/a	Confirmed					
	The exact	sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement				
	A stateme	ent on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly				
	The statist	tical test(s) used AND whether they are one- or two-sided on tests should be described solely by name; describe more complex techniques in the Methods section.				
\boxtimes	A description of all covariates tested					
\boxtimes	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons					
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)					
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.					
\boxtimes	For Bayes	ian analysis, information on the choice of priors and Markov chain Monte Carlo settings				
\boxtimes	For hierar	chical and complex designs, identification of the appropriate level for tests and full reporting of outcomes				
\times	Estimates	of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated				
	'	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.				
So	ftware an	d code				
Poli	cy information	about <u>availability of computer code</u>				
Da	ata collection	Olympus CellsSens Standard 2.1 Thermo Fisher Scientific EPU				
Da	ata analysis	CHIMERA (http://www.cgl.ucsf.edu/chimera) COOT (https://www2.mrc-lmb.cam.ac.uk/personal/pemsley/coot/) Gctf (https://en.wikibooks.org/w/index.php?title=Software_Tools_For_Molecular_Microscopy&stable=0#Gctf) RELION3.0 (https://www2.mrc-lmb.cam.ac.uk/relion) MotionCor2 (https://msg.ucsf.edu/em/software/motioncor2.html) Gautomatch (https://www2.mrc-lmb.cam.ac.uk/research/locally-developed-software/zhang-software/#gauto)				

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

OriginPro v8.5 (https://www.originlab.com/)

GraphPad Prism v9.3 (https://www.graphpad.com:443/)

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

The atomic coordinates for the TcsL at pH 7.4 and at the pH 5.0 in its open and closed conformations have been deposited in the Protein Data Bank (PDB) with the accession codes 8JB5, 8X2H, and 8X2I, respectively. The EM maps have been deposited in the EMDB with the accession codes EMD-36141, EMD-38010, and EMD-38011 for the TcsL at pH 7.4 and 5.0 in its open and closed conformations, respectively. The other data that support the findings of this study are available in a publicly accessible repository. Source data are provided with the paper.

Research involving human participants, their data, or biological material

Policy information aband sexual orientation		with <u>human participants or human data</u> . See also policy information about <u>sex, gender (identity/presentation), thnicity and <u>racism</u>.</u>		
Reporting on sex and gender		N/A		
Reporting on race, ethnicity, or other socially relevant groupings		N/A		
Population characteristics		N/A		
Recruitment		N/A		
Ethics oversight		N/A		
Note that full information	on on the appro	oval of the study protocol must also be provided in the manuscript.		
Field-spec		•		
	e below that is	the best fit for your research. If you are not sure, read the appropriate sections before making your selection.		
Life sciences	B	ehavioural & social sciences		
For a reference copy of the	e document with a	all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>		
Life scienc	ces stu	ıdy design		
All studies must discl	ose on these	points even when the disclosure is negative.		
'	For all cell rounding experiments: n=6 for each group. For pathological assay: n=10 mice per group. For toxin challenge assay: n=5 mice per group. Sample size was determined based on previous study and knowledge. Each sample size was selected so that a reasonable researcher would conclude that the size is sufficient to draw a statistical conclusion. For in vitro studies, at least three biological replicates were performed. For in vivo toxicological studies for large clostridial toxins, 5-10 mice per group is considered sufficient. (Tao et al. Nature 2016; Chen et al. Nat Commun 2021; Luo et al. Cell 2022; etc.)			
Data exclusions	No data exclusio	ons.		
Replication	All experiments were replicated at least three times. All attempts at replication are successful.			
Randomization	Samples were allocated into experimental groups randomly.			
	The stained tissue sections were scored blinded by two pathologists. For other experiments, blinding was not performed as virtually those data are quantitative and would not easily subject to operator by			

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experime	ntal sy	ystems Methods	
n/a Involved in the study		n/a Involved in the study	
Antibodies		ChIP-seq	
Eukaryotic cell lines		Flow cytometry	
Palaeontology and a	ırchaeol	ogy MRI-based neuroimaging	
Animals and other o	rganism	S	
Clinical data			
Dual use research of	f concer	1	
Plants			
Eukaryotic cell lin	es		
Policy information about <u>ce</u>	ell lines	and Sex and Gender in Research	
Cell line source(s)		HeLa (H1, CRL-1958) and A549 (CCL-185) cells were originally obtained from ATCC.	
Authentication		HeLa and A549 cells were authenticated via STR profiling (Shanghai Biowing Biotechnology Co. LTD, Shanghai, China).	
Mycoplasma contaminati	on	These cells were tested negative for mycoplasma contamination.	
Commonly misidentified lines (See ICLAC register)		No commonly misidentified lines were used in this study.	
Animals and othe	r res	earch organisms	
Policy information about <u>st</u> Research	<u>udies ir</u>	volving animals; ARRIVE guidelines recommended for reporting animal research, and Sex and Gender in	
<u>Nescaren</u>			
Laboratory animals	BALB/c mice (6-8 weeks, male, specific-pathogen-free) were purchased from Laboratory Animal Resources Center at Westlake University (Hangzhou, China). Mice were housed in specific-pathogen-free micro-isolator cages with free access to drinking water and food during the experiments. All mice had a 12-hour cycle of light/darkness (7am-7pm), housed at 20-24°C with 40-60% of		
	humidi	ty, and monitored under the care of full-time staff.	
Wild animals	No wild	No wild animals were used in this study.	
Reporting on sex	The mice used in this study are all male. Physiological cycles of female animals may affect the stability of toxicological experiments; thus male mice were chosen.		
Field-collected samples	No field-collected samples were used in this study.		
Ethics oversight	All animal procedures reported herein were performed following the institutional guidelines and approved by the Institutional Animal Care and Use Committee at Westlake University (IACUC Protocol #19-010-TL and #22-018-TL-5).		
Note that full information on th	he appro	oval of the study protocol must also be provided in the manuscript.	
Plants			
Seed stocks	N/A		
Novel plant genotypes	N/A		
	N1/2		
Authentication	N/A		