

## Supplementary Information

### Reassessing endothelial-to-mesenchymal transition in mouse bone marrow: Insights from lineage tracing models

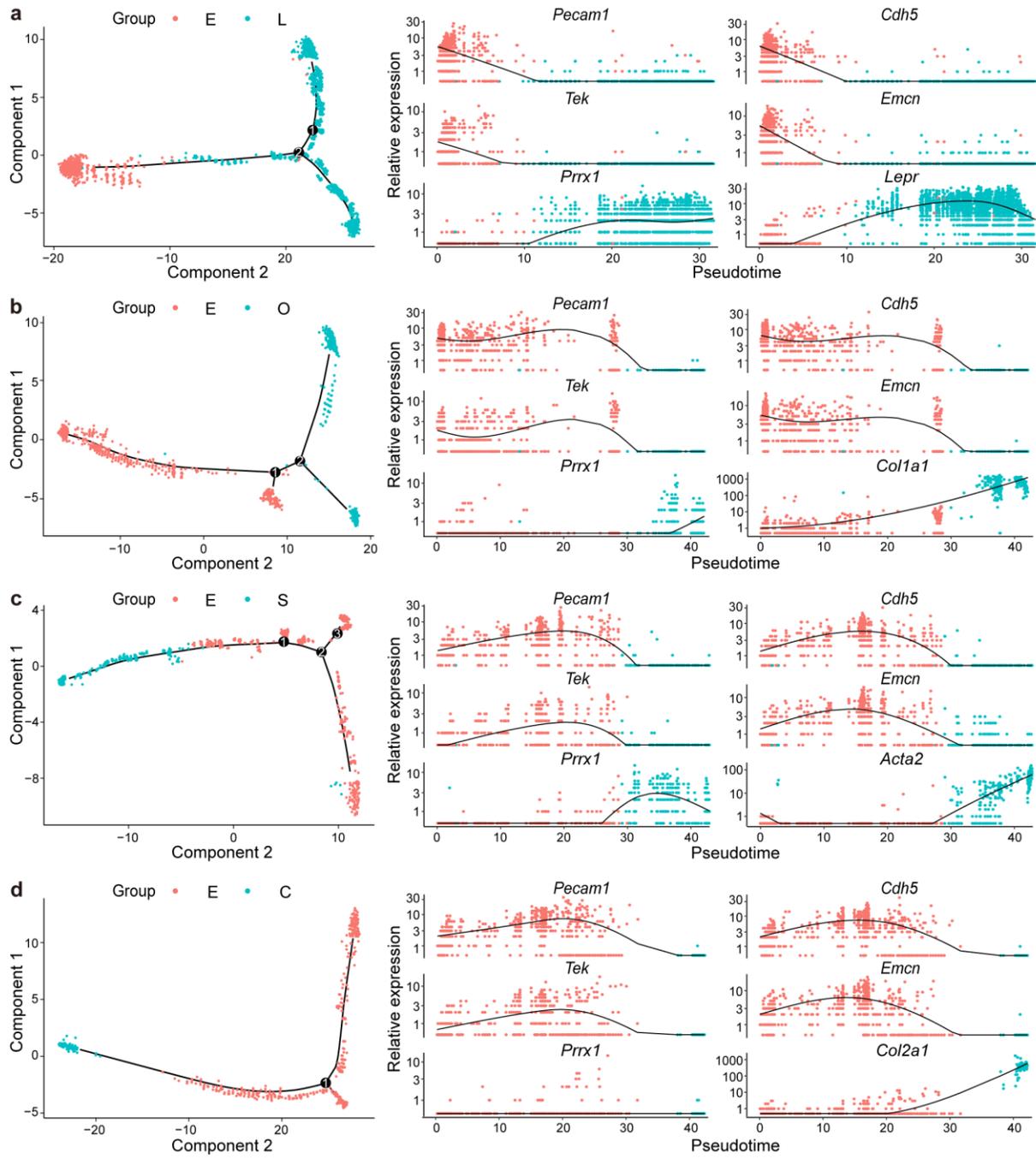
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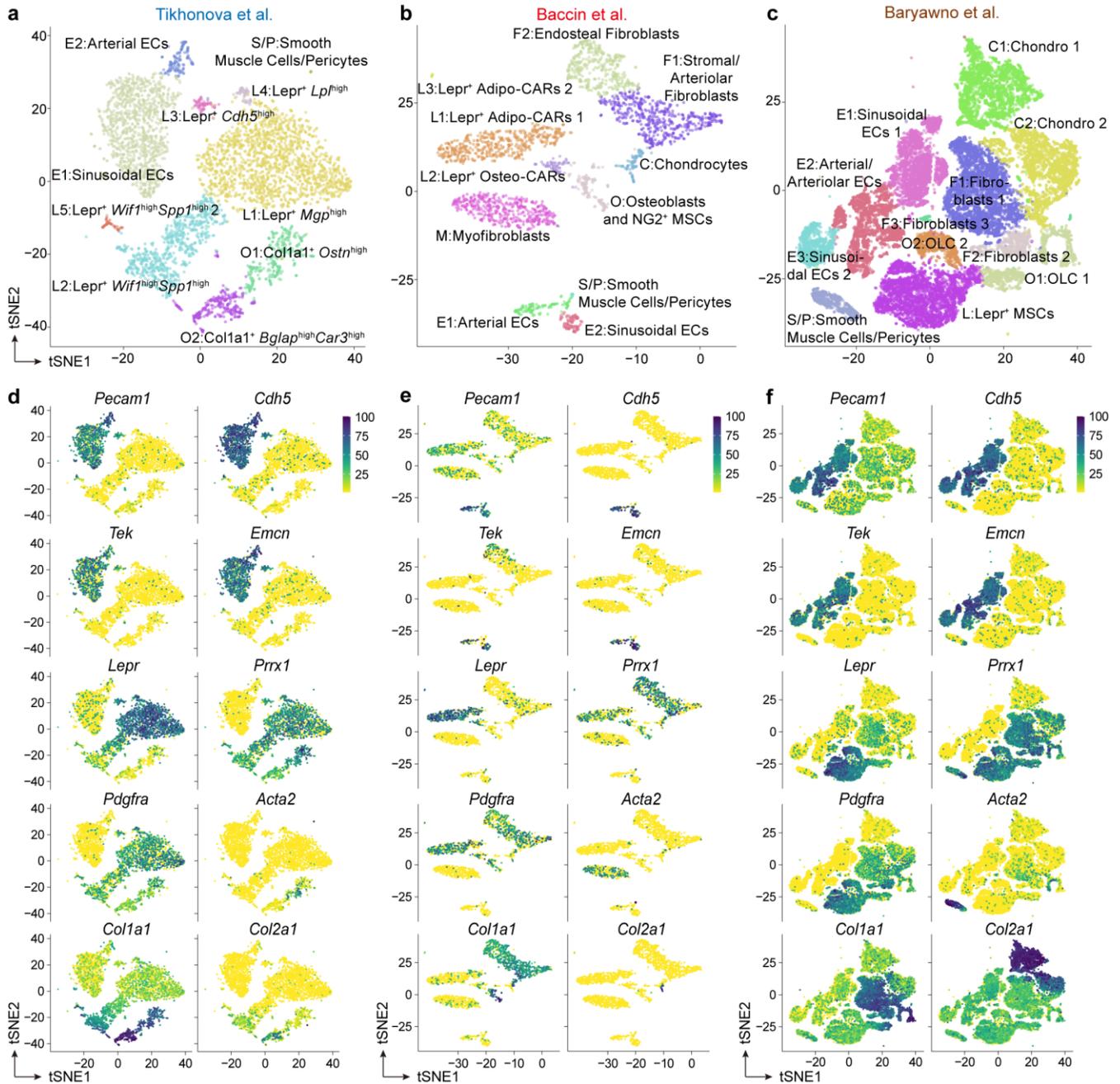
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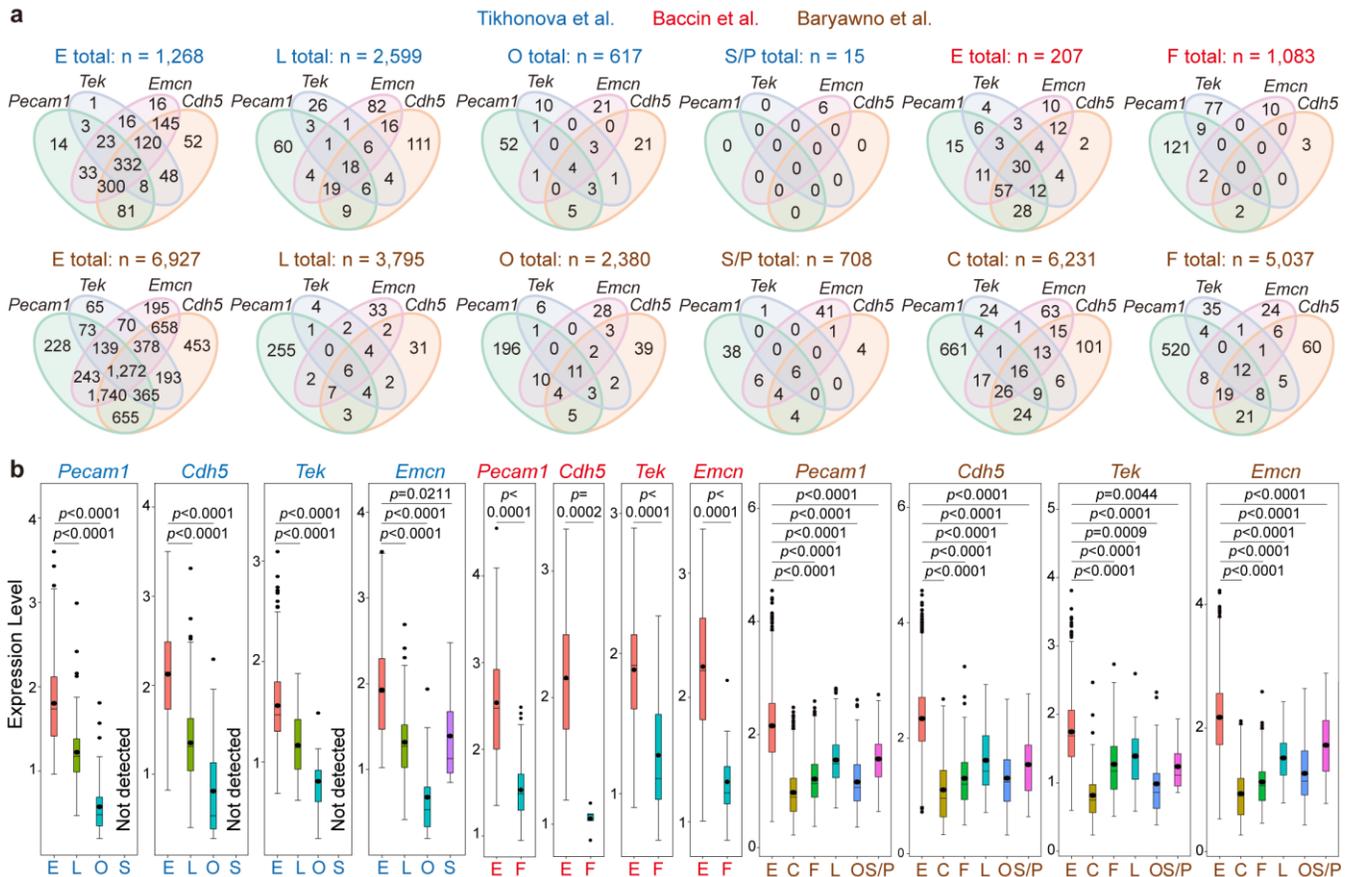


**Supplementary Fig. 1** Intermediate cell states between ECs and BMSCs are identified in scRNA-seq analysis. **a–d.** Pseudotime trajectory analysis between ECs and LEPR<sup>+</sup> BMSC (**a**), ECs and osteolineage cells (**b**), ECs and SMCs/PCs (**c**), and ECs and chondrolineage cells (**d**) in the scRNA-seq of postnatal mouse bone marrow showing a subset of cells with a gradual decrease in endothelial markers and an increase in mesenchymal markers along the trajectories.

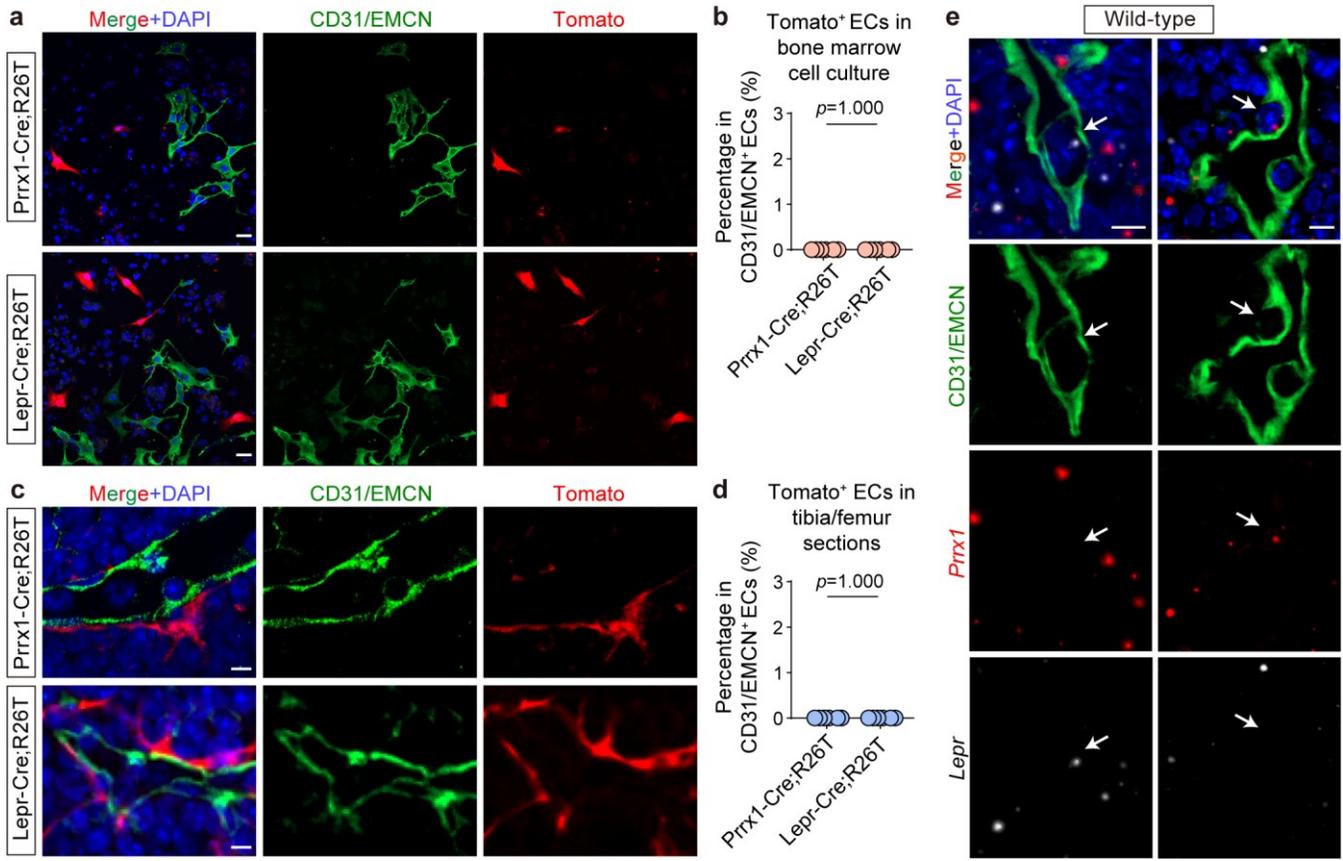


**Supplementary Fig. 2 BMSCs expressing endothelial markers are identified in reanalyzed scRNA-seq datasets. a–f.** t-SNE visualizations showing clusters of ECs and stromal cells (upper panels) and their expression of endothelial and mesenchymal markers (lower panels) in publicly available scRNA-seq datasets that characterized BMSCs: from lineage tracing models (Tikhonova et al.<sup>19</sup>) (a, d, n = 4,499 ECs and BMSCs, mice aged between 5–22 weeks); combining spatial transcriptomics (Baccin et al.<sup>22</sup>) (b, e, n

= 2,453 ECs and BMSCs, mice aged between 8–12 weeks); with several replicates (Baryawno et al.<sup>23</sup>) (**c**, **f**, n = 25,078 ECs and BMSCs, mice aged between 8–10 weeks). E, ECs. L, LEPR<sup>+</sup> cells. O, osteolineage cells. S/P, SMCs/PCs. C, chondrolineage cells. F, fibroblasts.

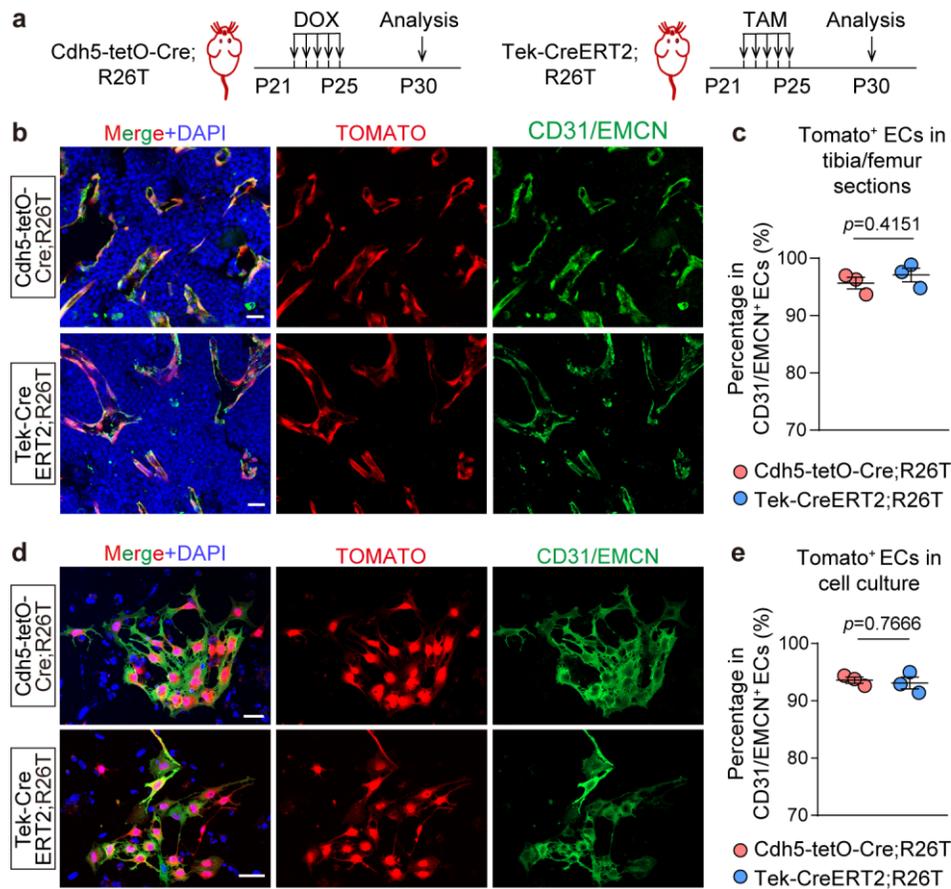


**Supplementary Fig. 3 BMSCs primarily present single endothelial markers at low levels in reanalyzed scRNA-seq datasets. a–b.** The co-expression (a) and the transcript levels of endothelial markers (b) in EC and stromal cells expressing these endothelial markers in the reanalyzed scRNA-seq datasets. Statistical significance was determined by two-tailed Wilcox rank-sum test. Source data are provided as a Source Data file.



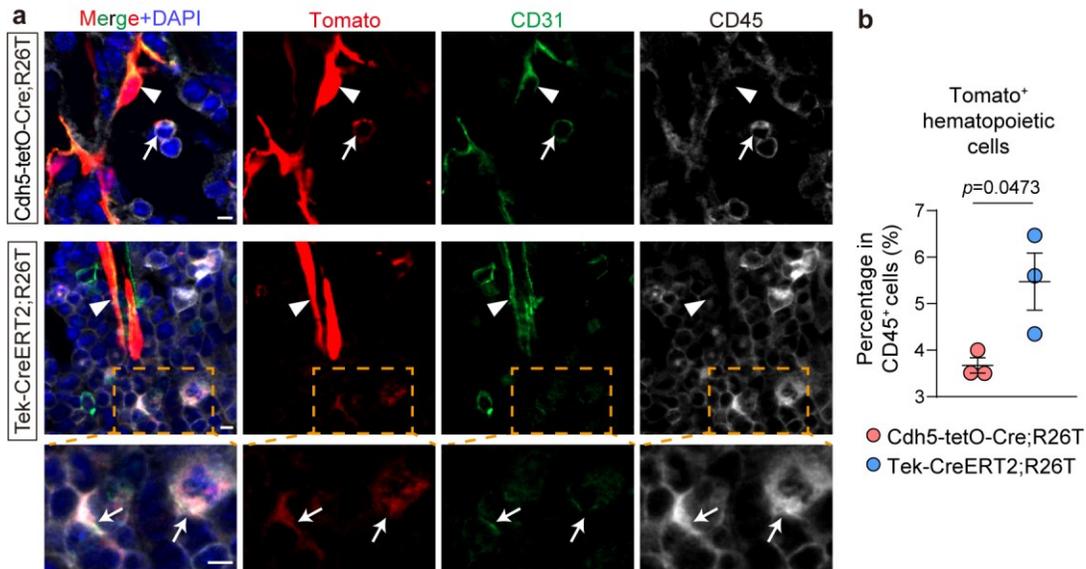
**Supplementary Fig. 4 Bone marrow ECs are not labeled by *Prrx1-Cre* and *Lepr-Cre* transgenic mouse models.** **a.** Immunostaining analysis of EGM-cultured bone marrow cells (**a, b**) and tibia/femur sections (**c, d**) revealing the absence of Tomato<sup>+</sup> cells in CD31/EMCN<sup>+</sup> ECs from Prrx1-Cre;R26T ( $n = 6$  biologically independent animals) and Lepr-Cre;R26T ( $n = 6$  biologically independent animals) mice. **e.** RNAscope ISH analysis of tibia/femur sections from wild-type mice ( $n = 3$  biologically independent animals) demonstrating the presence of *Prrx1* and the *ObRb* splice of LEPR in a small number of CD31/EMCN<sup>+</sup> ECs.

Scale bars: 5  $\mu$ m. Data represent the mean  $\pm$  S.E.M.. Statistical significance was determined by two-tailed unpaired Student's *t*-test. Source data are provided as a Source Data file.



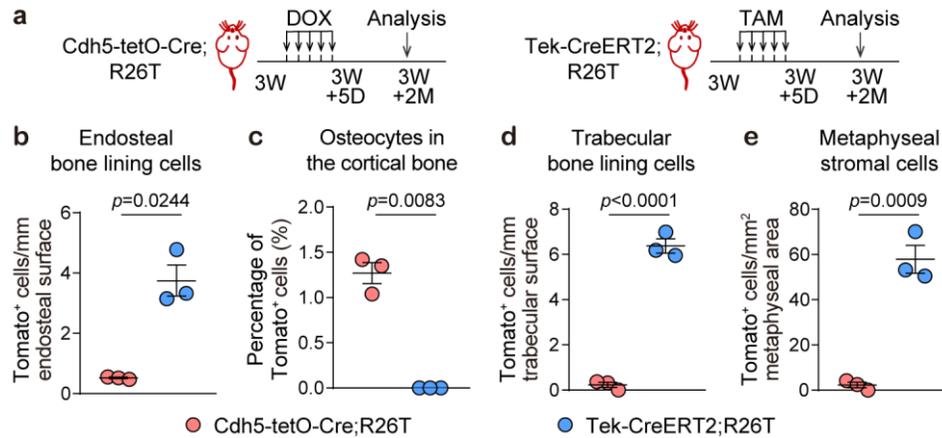
**Supplementary Fig. 5 EC lineage tracing models effectively label bone marrow ECs. a.** Diagrams of protocols. **b–e.** Immunostaining analysis of the tibia/femur sections (**b, c**) and EGM-cultured bone marrow cells (**d, e**) from Cdh5-tetO-Cre;R26T ( $n = 3$  biologically independent animals) and Tek-CreERT2-R26T ( $n = 3$  biologically independent animals) mice showing the effectiveness of these models in labeling bone marrow CD31/EMCN<sup>+</sup> ECs.

Scale bars: 10  $\mu$ m. Data represent the mean  $\pm$  S.E.M.. Statistical significance was determined by two-tailed unpaired Student's *t*-test. Source data are provided as a Source Data file.



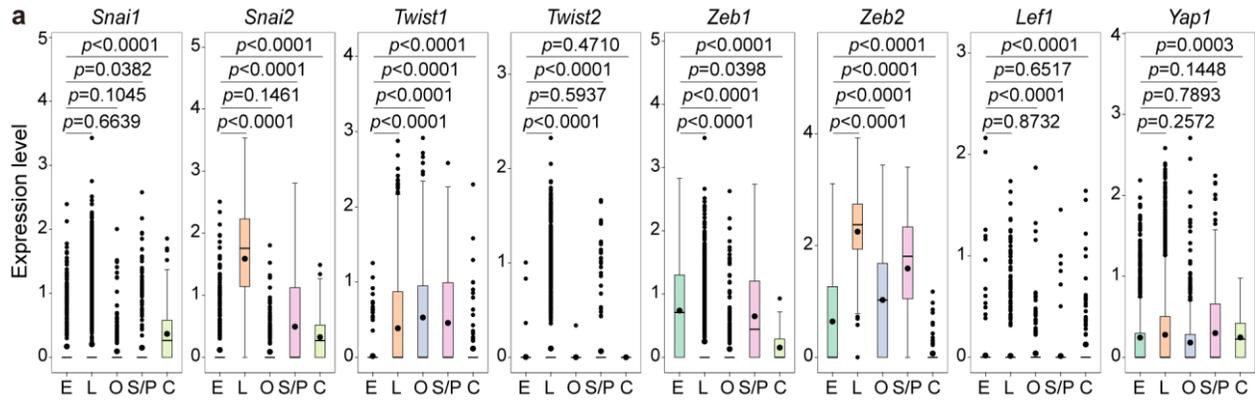
**Supplementary Fig. 6 EC lineage tracing models label small subsets of bone marrow hematopoietic cells. a–b.** Immunostaining analysis showing the presence of Tomato<sup>+</sup> hematopoietic cells (arrows) and ECs (arrowheads) in tibia/femur sections from Cdh5-tetO-Cre;R26T ( $n = 3$  biologically independent animals) and Tek-CreERT2-R26T ( $n = 3$  biologically independent animals) mice.

Scale bars: 5  $\mu$ m. Data represent the mean  $\pm$  S.E.M.. Statistical significance was determined by two-tailed unpaired Student's  $t$ -test. Source data are provided as a Source Data file.

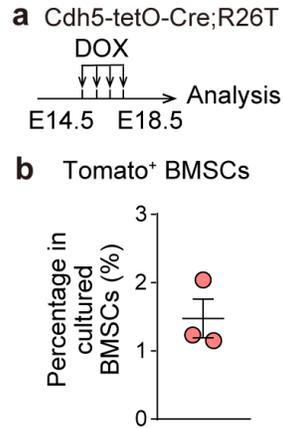


**Supplementary Fig. 7 Spatial distribution of endothelial marker-expressing BMSCs does not change in a two-month chase after doxycycline/tamoxifen administration.** **a.** Diagrams of protocols. Notes on the timeline indicate the age of mice. **b–e.** Quantification of Tomato<sup>+</sup> endosteal bone lining cells (**b**), osteocytes in the cortical bone (**c**), trabecular bone lining cells (**d**), and metaphyseal stromal cells (**e**) traced in the Cdh5-tetO-Cre;R26T ( $n = 3$  biologically independent animals) and Tek-CreERT2-R26T ( $n = 3$  biologically independent animals) models.

Data represent the mean  $\pm$  S.E.M.. Statistical significance was determined by two-tailed unpaired Student's  $t$ -test (**b–c** adjusted for unequal variances with Welch's test). Source data are provided as a Source Data file.



**Supplementary Fig. 8 EndoMT-related genes are upregulated in BMSCs than ECs. a.** Boxplots showing the expression of EndoMT-related genes among ECs and stromal cell subtypes in the scRNA-seq analysis of collagenase-digested bone and bone marrow cells from 5-week-old wild-type mice. Statistical significance was determined by two-tailed Wilcox rank-sum test. Source data are provided as a Source Data file.



**Supplementary Fig. 9 EC lineage tracing model labels small subsets of BMSCs during embryonic development.** **a.** Diagrams of protocols. **b.** Flow cytometry analysis showing the percentage of Tomato<sup>+</sup> cells within cultured BMSCs from E18.5 Cdh5-tetO-Cre;R26T ( $n = 3$  biologically independent samples) mice administered with doxycycline from E14.5 to E18.5.

E, embryonic. Data represent the mean  $\pm$  S.E.M.. Source data are provided as a Source Data file.