

Plant Gene Register

Nucleotide Sequence of a cDNA Encoding 3-Hydroxy-3-Methylglutaryl Coenzyme A Reductase from *Catharanthus roseus*¹

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HMGR³ (EC 1.1.1.34) performs the final enzymic step in mevalonate biosynthesis. In plants, mevalonate serves as a precursor for a number of important primary metabolites including ubiquinone, plastoquinone, carotenoids, ABA, and gibberellins. Additionally, mevalonate is used in the synthesis of a wide variety of secondary compounds such as phytoalexins, rubber, and alkaloids. We are interested in the role of HMGR in the synthesis of secologanin, the monoterpene component of *Catharanthus roseus* indole alkaloids.

C. roseus has been extensively studied because of the pharmaceutical importance of its antitumor bisindole alkaloids vincristine and vinblastine. Biosynthesis of these compounds begins with the conversion of tryptophan to tryptamine, catalyzed by tryptophan decarboxylase (5), followed by the condensation of tryptamine with secologanin to form strictosidine by strictosidine synthase (10). *C. roseus* cDNA clones for tryptophan decarboxylase (4) and strictosidine synthase (7) have been isolated and expressed in tobacco (6, 9). With the HMGR cDNA described here, it will now be possible to study the regulation of an enzyme in indole alkaloid biosynthesis that is not exclusively dedicated to this pathway but is also a key enzyme of primary metabolism.

A 444-bp HMGR fragment was generated from *C. roseus* cDNA by PCR using oligonucleotide primers (Table I) corresponding to two conserved regions within the active site of published plant and animal HMGR sequences. Because the predicted amino acid sequence of the PCR fragment showed 87% identity with the *Arabidopsis thaliana* HMGR (2), the fragment was used as a probe to screen a *C. roseus* seedling cDNA library. The complete nucleotide and derived amino acid sequence of a full-length *C. roseus* HMGR cDNA, pHMC3, is shown in Figure 1. The cDNA is 2636 bp, with an open reading frame encoding a protein of 601 amino acids having a calculated M_r of 64,106. The hydrophathy profile of

Table I. Characteristics of *C. roseus* HMGR cDNA pHMC3

Organism	<i>Catharanthus roseus</i> (L.) G. Don cv Little Delicata.
Location in Genome:	Nuclear genome.
Gene, Function, and Pathway:	HMGR (EC 1.1.1.34); reduction of 3-hydroxy-3-methylglutaryl CoA to mevalonate in terpene biosynthetic pathway.
Techniques:	PCR of HMGR active site region from <i>C. roseus</i> cDNA with oligonucleotide primers with the sequences: 5'-GATGCT-ATGGGGATGAATATG-3', and 5'-ACCTCGATAGATGGCA-TAGT-3'; cDNA library screening; double-stranded plasmid sequencing of both strands.
Method of Identification:	Amino acid sequence identity with <i>A. thaliana</i> (71.1%) and <i>H. brasiliensis</i> (76.6%) HMGRs.
Expression Characteristics:	Abundant 2.7-kb transcripts in all tissues.
Features of cDNA Structure:	Open reading frame of 1803 bp. Translation start at nucleotide 126 and stop at nucleotide 1929. (G + C) Content: 45.4%.
Structural Features of the Protein:	Deduced amino acid sequence of 601 residues; M_r 64,106; predicted isoelectric point 6.13; two potential transmembrane domains toward the N terminus; "PEST" cleavage sequences at amino acids 112 to 131 and 167 to 185; potential asparagine-linked glycosylation sites (N-X-S/T) at residues 86, 146, 149, 350, and 596.
Genbank/EMBL Accession No.:	M96068.

the predicted *C. roseus* HMGR indicates the presence of two hydrophobic domains located at residues 41 to 57 and 72 to 104, each of which is sufficient to span a membrane bilayer. Two N-terminal membrane spanning domains appear to be typical of other plant HMGRs (2, 3).

As in yeast (1), *A. thaliana* (2), and *Hevea brasiliensis* (3), "PEST" cleavage sequences are present in *C. roseus* HMGR (residues 112–131 and 167–185 in Fig. 1). PEST sequences are present in many short-lived proteins (8), although the half-life of HMGR has not yet been determined in plants. Amino acid sequence comparisons of *C. roseus* HMGR with sequences from plant, animal, and fungus HMGRs indicate

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³ Abbreviations: HMGR, 3-hydroxy-3-methylglutaryl-CoA reductase; PCR, polymerase chain reaction.

1 CCACAACCACCGTACCAGCTCCGCTCCTCCCTCCTAATCTCCGCCACTGCCACCCC
 61 ATTCTTTCTCTTCTCCCTACCACCCCCACCCCTTCTCACTAGACCTTTGTGCAAAAC
 121 AGAAAATGGACTCGCCGGGATCACCAACTGTGACGGCAAGGCTGTCGGGGTGAAC
 M D S R R R S P T V T A K A A A G E L
 181 TGCCATTGGCACCTCAGAAAGCCAAATCAACAGCCTTCCATTCTAGGCTTCCGATG
 P L A P H E G Q N Q Q P S I P R S S D V
 241 TTTTACCTTTGGCTCTGTATCTCGCAATGGGGTTTCTTACCTGTTTCTCTGTGTA
 L P L P L Y L A N G V F F T L F F S V M
 301 TGTACTTTCTTGACAGGTGGCGTGAAGATCCGAAATGCTACTCTCTCCATGTCG
 Y F L L T R W R E K I R N A T P L H V V
 361 TCACCTTTCTGAGCTTCCGCTTGGCTCATTAATCGCTTCTGTGATTTATCTGTGA
 T L S E L A A L A S L I A S V I Y L V S
 421 GTTTCTTTGGGCTGACTTTGTTCAGTCTTAATTTACAAGCCTAATAATGAAGGTTGGG
 F F G L D F V Q S L I Y K P N N E G W E
 481 AAATTGAAGAAGAAATCTTATGGTGAAGATAGCCGCAACGCTACTAATGTACTACC
 I E E E I L M V E D S R N G T N C T T L
 541 TAGGTTGCGCTGTCCTCCACATCTGTTCCTCAAAAATGCCAGTGTCTTCTCAACAAC
 G C A V P P P S V P K I A P V V P Q Q P
 601 CTTCTAAAATGGTCATAGAGAGCCCTCCCTCTCATTACCCCAAAAATCAGAGG
 S K M V I I E K P A P I I T P O N S E E
 661 AAGATGAGGATATTCAAGCGGTGGTTCGAAAAATTCCTTCTGATTTCTCGAGT
 D E D I I K A V V A G K I P S Y S L E S
 721 CAAAATGGGGATGCAAGAGAGCTGGAATTCGTAGGAAGCGCTCAGAGGATCA
 K L G D C K R A A G I R R E A L Q R I T
 781 CAGGGAAGTCATGGAGGGTACCTCTTGGGGTTTGATTATGCATCTATTTGGGTC
 G K S L E G L P L E G F D Y A S I L G Q
 841 AGTGTGTGAGATCGCGGTGGTGTATGTACAGTTCGCGGTGGAATAGCTGGGCTTTGT
 C C E M P V G Y V Q L P V G I A G P L L
 901 TGCTGATGGGAGAGATATATGTGCCAATGGCTACTACAGAAGGGTGTAGTGGSTA
 L D G R E Y M L P M A T T E G C L V A
 961 GTACTAACAGGGTGTAAAGCTATCTTGCATCTGGTGGGCAAAATAGCCTTTTGTCTCA
 T N R G C K A I L A S G G A N S V L L R
 1021 GAGATGGAATGACTAGGGCTCTGTGTGAGTGTGGAAGTCAAAAAGAGCCCTGAGC
 D G M T R A P V R F G T A K R A E L
 1081 TCAAGTTTACATGGAGGATACCCAGAATTTGAGACTATATCTGTAGTTTCAACAAT
 K F Y H E D T Q N F E T I S V V F N K S
 1141 CGAGCAGATTGCAAAATGCAAGGCTCAATGTGCAATTCAGGGGAAGAATTTGTACA
 S R F A K L Q S V Q C A I A G K N L Y I
 1201 TAAGTTTAGCTCAGACTGGTGTGCAATGGGATGCAATGGTCTCAAGGGCGTAC
 R F S C S T G D A M G M N M V S K G V Q
 1261 AAAATGTTCTTGAATTTCTCAGACTGATTATCTGATATGGATGCTCTGGGATTTCTG
 N V L E F L Q T D Y P D M D V L G I S G
 1321 GAAATTTCTGTGTCGACAAGAAACAGCAGCAGTCACTGGATTGAAGGAAGGTAAT
 N F C A D K K P A A V N W I E G R G K S
 1381 CTGTAGTTGTGAGGCTATCATAAAGGAGAGATTGTAAGACGGTATTGAAGCAGAAG
 V V C E A I I K E E I V K T V L K T E V
 1441 TTGCGCCCTTATTGAGCTAACATGGTAAAAATCTCGCTGGAATGCAATTTGCTGGAG
 A A L I E L N M V K N L A G S A I A G A
 1501 CCCTGGTGGATCAATGCTCAGCCAGCAATATAGTCTCTGCATATTTATTTGCCACCG
 L G G F N A H A S N I V S A I F I A T G
 1561 GCCAAGATCTGTCAGAAATGGAGAGCTCTCAGTGTACTATGATGGAGGCTGCA
 Q D P A Q N V E S S Q C I T M M E A V N
 1621 ATGATGGCAAGATCTCATATTTCTGTCCATGCTTCTATTGAGGTGGGACAGTTG
 D G K D L H I S V T M P S I E V G T V G
 1681 GTGAGGTAACAATGCAATCTCAGTCACTGTTGAAGTGTGAGGAAAGAGGTTG
 G G T Q L A S Q S A C L N L L G V K G A
 1741 CCAGCAAGACTCACCTGGTCAAAATCAAGGCTCTGGCTACCATTTGCTGTTGCTCAG
 S K D S P G A N S R L L A T I V A G S V
 1801 TTTCTCAGGTTAGCTTTCCCTATGCTCCATTCTGCAGGCCAAGCTCGTTAGGAGCC
 L A G E L S L M S A I S A G Q L V R S H
 1861 ACATGAAATCAACCGTTCTAGCAAGGATATAACCAACATAGCCTCTCTCAGTTAGAGA
 M K Y N R S S K D I T N I A S S Q L E S
 1921 GTGATAGTTAAATCAATCCCTCCCTCCCTTAAACCCACCTTGTAGTTAGT
 D S *
 1981 ATAGGCCATAGAAGAAATCCCAAAAAAAGGTTATGAAGTTAATAAA
 2041 CAAATCAAGAAAGGGTGGTGAATAAAAAAATATATAAACCCCAAAAAAAGG
 2101 AAAGAGAATAGGGTGTATTGGTGGTGAATGATATGTGAGAAGGAGAAATGAAGGAAGA
 2161 AACAGACAAAGGCAACATCATCTGTTGCAAGAGGGAGTCATGTGATGCTGCTACCT
 2221 CTCTTCT
 2281 GAACAGTAGCTGGCTGGCTGGCTGGCTGGCTGGCAAGAAAGAAATCTGTTTGTAA
 2341 TTATTTGTATCTTTGTAATGTCATTTTGGCATCAAGAAATGGAAGAGAAAGAGGAGTT
 2401 AAGTTAGAGTTTGGTGGCATTATGTCAGAAAGGAAAGATTTACTCTTAATTTTATGAT
 2461 GTTTTAAATAATCTCTCTCTCAGTGTCTCTCTCTCTCTCTCTCTCTCTCTCTCTCT
 2521 GTTTGTTTAAACAGCAGTATTAGTAGTAATTAAGACAGACATTCAAAAACATGAG
 2581 ACTCTGATGTCTTCAAGTTTGTGTAATGAAAAATGTTTTATCTTCAAAA

Figure 1. Nucleotide and deduced amino acid sequence of *C. roseus* HMGR cDNA (pHMC3). PCR primer sites are underlined in the nucleotide sequence. Potential PEST sequences are underlined in the amino acid sequence.

a high degree of similarity in the C-terminal region, which contains the catalytic site.

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