Supplementary material

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Supplementary methods

- 4 Preparation of formalin-fixed, paraffin-embedded (FFPE) tissues
- 5 Seven 10µm formalin-fixed, paraffin-embedded (FFPE) tissue slices were prepared for
- 6 protein analysis, essentially as previously described¹⁻³. In short, paraffin was removed with
- 7 xylene followed by rehydration with decreasing levels of ethanol/water and dried. The tissue
- 8 was dissolved in lysis buffer (5% SDS, 50 mM triethylammonium bicarbonate [TEAB], pH
- 9 8.5) and further processed using S-TrapTM micro spin columns (Protifi, Farmingdale,
- Huntington, NY, USA) essentially as previously described^{1,2,4}. A total of 74 samples (*i.e.*, 34
- 11 nt-FL, 20 st-FL, and 20 tFL) were prepared and finally dissolved in 0.1% formic acid at a
- 12 concentration of $0.5 \mu g/\mu L$. One μg of each sample was injected in triplicate, except for one
- sample injected in duplicate due to limitations of sample amount.

- 15 *LFQ nLC-MS/MS and database searches*
- Mass spectrometry was performed using an Orbitrap Fusion Tribrid mass spectrometer
- coupled to a Dionex Ultimate 3000 RSCL nano LC system (Thermo Fisher Scientific
- 18 Instruments, Waltham, MA, USA) using the universal method with settings as previously
- described^{1,4}. The 221 raw files were entered into MaxQuant version 1.6.6.0 (Max Planck
- 20 Institute of Biochemistry, Martinsried, Germany: https://maxquant.net/maxquant/) for label-
- 21 free quantification (LFQ) analysis⁵ using the UniProt *Homo sapiens* filtered and reviewed
- database (www.uniprot.org, downloaded 19 March 2021). MaxQuant settings were as
- previously described⁶. Further processing was performed in Perseus version 1.6.14.0 (Max
- 24 Planck Institute of Biochemistry, Martinsried, Germany: https://maxquant.net/perseus/) for

- 1 filtering and further statistical analysis⁷. The means of the Log₂ transformed LFQ values from
- 2 each technical replicate were calculated for each protein in each sample. At least two unique
- 3 peptides were required for identification. Proteins that were quantified in at least 70% of the
- 4 samples in each group were included in the downstream analyses. The median technical
- 5 coefficient of variation was calculated for each sample from proteins determined in triplicate.
- 6 The mean value was 13.6% (Range 9.72%-20.3%). P-values were calculated by two-tailed t-
- 7 test without further correction in order to not increase type 2 errors with the risk of
- 8 overlooking putative predictive markers.

- 10 Bioinformatic analysis
- Bioinformatic analysis was performed using both the STRING database (string-db.org) and
- with the use of QIAGEN IPA (QIAGEN Inc., https://digitalinsights.qiagen.com/IPA)8. For
- each protein, the corresponding UniProt ID were submitted to the software tools. When more
- than one UniProt protein ID were identified, only one representative, the first listed, was used
- in the analysis.
- The STRING analyses were performed based on the set of significantly differentially
- expressed proteins (p<0.05) with at least 20% difference in fold change, and the
- corresponding calculated fold-changes. Analyses were performed with input of UniProt IDs.
- 19 For the nt-FL vs st-FL analysis, one UniProt ID was not recognized in the Cytoscape
- 20 STRING software, resulting in a total of 241 proteins in the analysis. In the st-FL vs tFL
- 21 analysis, two UniProt IDs was not recognized in the STRING software, resulting in a total of
- 22 798 proteins included in the analyses. The database was assessed using the StringApp
- (version 1.7.0) in Cytoscape (version 3.9.0)⁹⁻¹¹. The minimum required interaction score was
- set to medium confidence, 0.4, with both functional and physical interactions included. For

- 1 enrichment analysis, enriched terms were filtered to include only terms from UniProt
- 2 Keywords, Gene Ontology (GO) Biological Processes, GO cellular components, GO
- 3 molecular functions, Kyoto Encyclopedia of Genes and Genomes Pathways, Reactome
- 4 Pathways, and STRING clusters. The false discovery rate was set to 5%. The option to
- 5 remove redundant terms was enabled, and the redundancy cut-off was set to 0.50.
- The IPA analyses were performed with the combined set of identified proteins and the
- 7 corresponding calculated fold-changes. IPA recognized 2657 of the 2665 protein uploaded.
- 8 Analysis was performed with significantly differentially expressed proteins using p<0.05.
- 9 The IPA algorithms used are described by Krämer et al. Here, z-scores $\geq |2|$ are considered
- statistically significant, and a positive z-score indicates that the pathway is predicted to be
- activated, while a negative z-score indicates that the pathway is predicted to be inhibited.
- 12 Missing z-scores indicate a pathway for which the z-score cannot be calculated.
- 14 Immunohistochemical staining of selected proteins

- 15 Selected proteins from the MS-based proteomics results were evaluated using
- immunohistochemistry, namely caspase 3 (CASP3), induced myeloid leukemia cell
- differentiation protein (MCL1), BCL-2-associated X protein (BAX), B-cell lymphoma-extra-
- large (BCL-xL), and BCL2-like 13 (BCL-rambo). One nt-FL sample was excluded from the
- analyses due to insufficient lymphoma material available (n=33). Staining was performed on
- 20 4μm FFPE sections using the Ventana Benchmark Ultra automated staining system (Ventana
- 21 Medical Systems, Oro Valley, Arizona). Slides were deparaffinized with EZ Prep (Ventana,
- 950-102) followed by blocking of endogenous peroxidase activity using the OptiView DAB
- 23 IHC Detection Kit (Ventana, 760-700)¹²⁻¹⁴. Heat induced epitope retrieval (HIER) was
- performed by heating slides to 100°C in ULTRA Cell Conditioning Solution 1 (CC1,
- Ventana, 950-224). After HIER, incubation with primary antibody was performed. For all

- staining protocols see Supplemental Table S1. All antibodies were diluted in Tris-buffered
- 2 antibody diluent (pH 7.2, 15mmol/L NaN3 and stabilizing protein, Dako, Santa Clara,
- 3 California), followed by incubation at 37°C. Visualization was performed using the OptiView
- 4 DAB IHC Detection Kit with nuclear counterstaining by hematoxylin. Sections of appendix,
- 5 tonsil, liver, and pancreas were included on all slides as positive and negative controls.

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Digital image analysis

- 8 Stained slides were scanned at a magnification of x20 using the Hamamatsu Nanozoomer
- 9 2.0HT scanner (Hamamatsu, Shizouka, Japan). Expression levels of CASP3, MCL1, BAX,
- 10 BCL-xL, and BCL-rambo were quantified using Visiopharm Integrator system 2020.01
- 11 (Visiopharm A/S, Hoersholm, Denmark). As previously described¹²⁻¹⁴, areas for staining
- quantification were defined by manual outlining of regions of interest (ROI) on each digitized
- whole tissue section. Distinct areas of non-lymphoid tissue and technical artefacts were
- excluded. Analysis protocol packages (APPs) were designed to quantify the expression levels
- of each marker¹²⁻¹⁴. Staining quantification outputs were area fractions (AFs), defined as the
- stained area normalized to the total area within the ROI. Intrafollicular regions were manually
- outlined, guided by a consecutive parallel tissue section stained with PAX5 to identify B cell
- areas in the biopsy¹²⁻¹⁴. For the intrafollicular quantification, one sample was excluded from
- 19 the cohort due to the inability to define follicles based on the consecutive PAX5 staining
- 20 (n=52; nt-FL, n=32 and st-FL, n=20). Expression levels of CASP3, MCL1, BAX, and BCL-
- 21 xL were based on all positive staining, while expression levels of BCL-rambo were based on
- 22 strong-intensity staining.

- 5 Supplementary tables
- **Table S1:** Immunohistochemistry staining protocols
- **Table S2:** 2665 identified proteins from MS-based proteomics analysis
- **Table S3:** 795 identified proteins comparing st-FL and nt-FL samples
- **Table S4**: significantly differentially expressed proteins, nt-FL and st-FL
- **Table S5:** 800 significantly differentially expressed proteins comparing tFL and st-FL
- 11 samples
- **Table S6:** 63 significantly differentially expressed proteins comparing st-FL and nt-FL
- samples in high-risk group from *Analysis A*
- **Table S7:** 68 significantly differentially expressed proteins comparing st-FL and nt-FL
- samples in high-risk group from *Analysis B*
- **Table S8:** Complete STRING pathway analysis comparing nt-FL and st-FL samples
- **Table S9:** IPA pathway analysis comparing nt-FL and st-FL samples
- **Table S10:** IPA diseases and function pathway analysis comparing st-FL and tFL samples
- **Table S11:** STRING pathway analysis comparing st-FL and tFL samples
- 20 Table S12: IPA pathway analysis comparing st-FL and tFL samples
- **Table S13:** IPA diseases and function pathway analysis comparing st-FL and tFL samples

Supplemental Table S1: immunohistochemistry staining protocols

Target protein	HIER incubation (min)	Primary antibody	Primary antibody dilution	Antibody incubation (min)
CASP3	64	Polyclonal rabbit anti-human anti- CASP3 (HPA00264, Sigma-Aldrich, St. Louis, Missouri, USA)	1:750	32
MCL1	92	Polyclonal rabbit anti-human anti-MCL1 (HPA008455, Sigma-Aldrich)	1:300	32
BCL-xL	92	Monoclonal rabbit anti-human anti-BCL-xL (ab32370, Abcam, Cambridge, UK)	1:1000	32
BCL-rambo	64	Polyclonal rabbit anti-human anti-BCL-rambo (16612-1-AP, ProteinTech, Rosemont, Illinois, USA)	1:700	32
BAX	64	Monocloncal mouse anti-human anti-BAX (60267-1-Ig, ProteinTech)	1:2500	32

Supplementary Table S4: significantly differentially expressed proteins, nt-FL and st-FL

Fold change		C	Post de la companya d
(st-FL/nt-FL)	p-value	Gene name	Protein name
Upregulated			
2.5	0.038	MPO	Myeloperoxidase
2.1	0.025	RNASE3	Eosinophil cationic protein
2.0	0.014	FABP3	Fatty acid-binding protein, heart
1.9	0.003	MPG	DNA-3-methyladenine glycosylase
1.7 1.6	$0.041 \\ 0.010$	NTPCR DBI	Cancer-related nucleoside-triphosphatase Acyl-CoA-binding protein
1.6	0.010	SNRPC	U1 small nuclear ribonucleoprotein C
1.6	0.016	HN1	Hematological and neurological expressed 1 protein
1.5	0.010	PAX5	Paired box protein Pax-5
1.5	0.020	MCL1	Induced myeloid leukemia cell differentiation protein Mcl-1
1.5	0.031	EXOSC6	Exosome complex component MTR3
1.5	0.013	SRSF3	Serine/arginine-rich splicing factor 3
1.5	0.040	RABIF	Guanine nucleotide exchange factor MSS4
1.5	0.005	CDC26	Anaphase-promoting complex subunit CDC26
1.5	0.028	LYZ	Lysozyme C
1.5	0.003	SARNP	SAP domain-containing ribonucleoprotein
1.5	0.049	TRMT61A	tRNA (adenine(58)-N(1))-methyltransferase catalytic subunit TRMT61A
1.5	0.017	CCAR1	Cell division cycle and apoptosis regulator protein 1
1.5 1.5	0.009	ALYREF NUP35	THO complex subunit 4
1.5	$0.014 \\ 0.010$	PSME3	Nucleoporin NUP53 Proteasome activator complex subunit 3
1.3	0.010	ARL2	ADP-ribosylation factor-like protein 2
1.4	0.019	BAK1	Bcl-2 homologous antagonist/killer
1.4	0.019	LYPLAL1	Lysophospholipase-like protein 1
1.4	0.010	SRSF1	Serine/arginine-rich splicing factor 1
1.4	0.002	TRPT1	tRNA 2-phosphotransferase 1
1.4	0.011	SRA1	Steroid receptor RNA activator 1
1.4	0.027	PABPC4	Polyadenylate-binding protein 4
1.4	0.014	ISG20	Interferon-stimulated gene 20 kDa protein
1.4	0.012	CHERP	Calcium homeostasis endoplasmic reticulum protein
1.4	0.007	KPNA2	Importin subunit alpha-1
1.4	0.001	SRSF10	Serine/arginine-rich splicing factor 10
1.4 1.4	$0.007 \\ 0.025$	SAE1 MMP0	SUMO-activating enzyme subunit 1
1.4	0.023	<i>MMP9</i> SNRPB	Matrix metalloproteinase-9 Small nuclear ribonucleoprotein-associated proteins B and B
1.4	0.006	PLD3	Phospholipase D3
1.4	0.012	RMDN3	Regulator of microtubule dynamics protein 3
1.4	0.007	BAX	Apoptosis regulator BAX
1.4	0.010	NPEPL1	Probable aminopeptidase NPEPL1
1.4	0.029	NDUFS8	NADH dehydrogenase [ubiquinone] iron-sulfur protein 8, mitochondrial
1.4	0.026	CDK6	Cyclin-dependent kinase 6
1.4	0.023	BCS1L	Mitochondrial chaperone BCS1
1.4	0.028	SNRPG	Small nuclear ribonucleoprotein G
1.4	0.024	SRSF2	Serine/arginine-rich splicing factor 2
1.4	0.045	DR1	Protein Dr1
1.4 1.4	0.044 0.025	FUCA1 SEC23B	Tissue alpha-L-fucosidase Protein transport protein Sec23B
1.4	0.023	ACOT7	Cytosolic acyl coenzyme A thioester hydrolase
1.4	0.040	CIRBP	Cold-inducible RNA-binding protein
1.4	0.008	MTMR14	Myotubularin-related protein 14
1.4	0.019	HTRA2	Serine protease HTRA2, mitochondrial
1.4	0.036	NDUFS3	NADH dehydrogenase [ubiquinone] iron-sulfur protein 3, mitochondrial
1.4	0.002	TBC1D13	TBC1 domain family member 13
1.4	0.044	CSTF2	Cleavage stimulation factor subunit 2
1.4	0.002	SNRPD1	Small nuclear ribonucleoprotein Sm D1
1.4	0.010	NAA40	N-alpha-acetyltransferase 40
1.4	0.043	COX5A	Cytochrome c oxidase subunit 5A, mitochondrial
1.4	0.005	ATXN10	Ataxin-10
1.3	0.015	LEMD2	LEM domain-containing protein 2
1.3	$0.004 \\ 0.006$	FBXO22	F-box only protein 22 ADP-ribosylation factor-like protein 1
1.3	0.000	ARL1	ADI -Hoosyiation factor-like protein i

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1.2 U.U54 KPNA5 Importin subunit alpha-4				
	1.2	0.034	KPNA3	importin subunit alpha-4

1.2	0.011	PFDN2	Prefoldin subunit 2
1.2	0.017	PAAF1	Proteasomal ATPase-associated factor 1
1.2	0.029	COASY	Bifunctional coenzyme A synthase
1.2	0.029	PPAT	Amidophosibosyltransferase
1.2	0.037	SNAP29	Synaptosomal-associated protein 29
1.2	0.017	DTYMK	Thymidylate kinase
1.2	0.011	CHMP2A	Charged multivesicular body protein 2a
1.2	0.039	NECAP1	Adaptin ear-binding coat-associated protein 1
1.2	0.002	G3BP1	Ras GTPase-activating protein-binding protein 1
1.2	0.041	BTF3	Transcription factor BTF3
Downregulated			
0.4	0.005	<i>IQGAP2</i>	Ras GTPase-activating-like protein IQGAP2
0.5	0.004	UGGT1	UDP-glucose:glycoprotein glucosyltransferase 1
0.5	0.001	DOCK11	Dedicator of cytokinesis protein 11
0.5	< 0.001	ACTB	Actin, cytoplasmic 1
0.5	0.029	AP2A2	AP-2 complex subunit alpha-2
0.6	0.011	AKAP13	A-kinase anchor protein 13
0.6	< 0.001	MAP2K1	Dual specificity mitogen-activated protein kinase kinase 1
0.6	0.040	RBM25	RNA-binding protein 25
0.6	0.018	EHD3	EH domain-containing protein 3
0.6	0.014	STT3B	Dolichyl-diphosphooligosaccharid-protein glycosyltransferase STT3B
0.6	0.027	PLXNB2	Plexin-B2
0.7	0.026	CKAP5	Cytoskeleton-associated protein 5
0.7	0.013	FLNB	Filamin-B
0.7	0.028	KTN1	Kinectin
0.7	0.039	IGHG1	Ig gamma-1 chain C region
0.7 0.7	0.005	ADD1	Alpha-adducin
0.7	0.025	ACAP2 NIPSNAP1	Arf-GAP coiled-coil, ANK repeat and PH domain-containing protein 2
0.7	0.020 0.043	GLG1	Protein NipSnap homolog 1 Golgi apparatus protein 1
0.7	0.043	SBF1	Myotubularin-related protein 5
0.7	0.048	LMAN1	Protein ERGIC-53
0.7	0.002	SRP54	Signal recognition particle 54 kDa protein
0.7	0.027	ADD3	Gamma-adducin
0.7	0.004	HMHA1	Minor histocompatibility protein HA-1
0.7	0.033	SERPINC1	Antithrombin-III
0.7	0.014	SLC1A5	Neutral amino acid transporter B(0)
0.7	0.016	PALD1	Paladin
0.7	0.010	RPN2	Dolichyl-diphosphooligosaccharide-protein glycosyltransferase 2
0.7	0.026	PDIA4	Protein disulfide-isomerase A4
0.7	0.047	ATP2B1	Plasma membrane calcium-transporting ATPase 1
0.7	0.009	ABCE1	ATP-binding cassette sub-family E member 1
0.7	0.046	RPS6KA3	Ribosomal protein S6 kinase alpha-3
0.7	0.005	CD81	CD81 antigen
0.7	0.011	CTTN	Src substrate cortactin
0.7	0.027	PIK3AP1	Phosphoinositide 3-kinase adapter protein 1
0.7	0.011	PACS1	Phosphofurin acidic cluster sorting protein 1
0.7	0.036	OSBPL8	Oxysterol-binding protein-related protein 8
0.7	0.016	SEPT7	Septin-7
0.7	0.005	ELMO1	Engulfment and cell motility protein 1
0.7 0.7	0.001	GNPDA2	Glucosamine-6-phosphate isomerase 2 Ethylmalonyl-CoA decarboxylase
0.7	$0.002 \\ 0.030$	ECHDC1 ITGB1	Integrin beta-1
0.7	0.030	NT5C2	Cytosolic purine 5-nucleotidase
0.7	0.040	PDCD4	Programmed cell death protein 4
0.7	0.003	CD2AP	CD2-associated protein
0.7	0.012	HERC4	Probable E3 ubiquitin-protein ligase HERC4
0.7	0.038	ITGAL	Integrin alpha-L
0.7	0.008	GCLC	Glutamatecysteine ligase catalytic subunit
0.7	0.018	TES	Testin
0.7	0.014	BRK1	Protein BRICK1
0.7	0.049	MAGED2	Melanoma-associated antigen D2
0.7	0.023	SLC48A1	Heme transporter HRG1
0.7	0.025	NCKAP1L	Nck-associated protein 1-like
0.8	0.042	AP1M1	AP-1 complex subunit mu-1
0.8	0.012	CDK5RAP3	CDK5 regulatory subunit-associated protein 3
		·	

0.8	0.015	PIK3R4	Phosphoinositide 3-kinase regulatory subunit 4
0.8	0.020	DCTN4	Dynactin subunit 4
0.8	0.026	SRPR	Signal recognition particle receptor subunit alpha
0.8	0.043	GGT5	Gamma-glutamyltransferase 5
0.8	0.018	DGKA	Diacylglycerol kinase alpha
0.8	0.029	ATP2A3	Sarcoplasmic/endoplasmic reticulum calcium ATPase 3
0.8	0.038	TRIM25	E3 ubiquitin/ISG15 ligase TRIM25
0.8	0.032	OAS2	2-5-oligoadenylate synthase 2
0.8	0.026	BCAP31	B-cell receptor-associated protein 31
0.8	0.034	PIP4K2A	Phosphatidylinositol 5-phosphate 4-kinase type-2 alpha
0.8	0.013	METAP2	Methionine aminopeptidase 2
0.8	0.003	HSPA1A	Heat shock 70 kDa protein 1A
0.8	0.007	MSN	Moesin
0.8	0.035	SRP72	Signal recognition particle subunit SRP72
0.8	0.034	VASP	Vasodilator-stimulated phosphoprotein
0.8	0.040	LANCL1	LanC-like protein 1
0.8	0.038	FKBP2	Peptidyl-prolyl cis-trans isomerase FKBP2
0.8	0.001	CCNY	Cyclin-Y
0.8	0.011	ABCF3	ATP-binding cassette sub-family F member 3
0.8	0.002	PDIA3	Protein disulfide-isomerase A3
0.8	0.016	LMAN2	Vesicular integral-membrane protein VIP36
0.8	0.034	SPG11	Spatacsin
0.8	0.030	CYB5B	Cytochrome b5 type B
0.8	0.011	DENND2D	DENN domain-containing protein 2D
0.8	0.044	DNAJB11	DnaJ homolog subfamily B member 11
0.8	0.006	NRAS	GTPase NRas
0.8	0.005	DRG2	Developmentally-regulated GTP-binding protein 2
0.8	0.006	TSG101	Tumor susceptibility gene 101 protein
0.8	0.021	KDSR	3-ketodihydrosphingosine reductase
0.8	0.006	TMED10	Transmembrane emp24 domain-containing protein 10
0.8	0.020	RPN1	Dolichyl-diphosphooligosaccharide-protein glycosyltransferase 1
0.8	0.007	GEMIN5	Gem-associated protein 5
0.8	0.031	DSCR3	Down syndrome critical region protein 3
0.8	0.003	ADH5	Alcohol dehydrogenase class-3
0.8	0.029	NGLY1	Peptide-N(4)-(N-acetyl-beta-glucosaminyl)asparagine amidase
0.8	0.025	SMYD5	SET and MYND domain-containing protein 5
0.8	0.008	EPN1	Epsin-1
0.8	0.029	PRKCSH	Glucosidase 2 subunit beta
0.8	0.038	TOR1A	Torsin-1A
0.8	0.019	CUL5	Cullin-5
0.8	0.015	SEPT6	Septin-6
0.8	0.041	GLTP	Glycolipid transfer protein
0.8	0.011	NCK1	Cytoplasmic protein NCK1
0.8	0.023	PPIB	Peptidyl-prolyl cis-trans isomerase B
0.8	0.031	TFAM	Transcription factor A, mitochondrial
0.8	0.023	PTPN11	Tyrosine-protein phosphatase non-receptor type 11
0.8	0.028	<i>IQGAP1</i>	Ras GTPase-activating-like protein IQGAP1
0.8	0.020	SEPT9	Septin-9
0.8	0.046	PAK2	Serine/threonine-protein kinase PAK 2;PAK-2p27;PAK-2p34

1 Supplementary figures

2 Figure S1: Unsupervised clustering based on all identified proteins

- 3 (A) Principal component analysis based on all 2665 identified proteins comparing st-FL and
- 4 nt-FL samples. Abbreviations: nt-FL, non-transforming FL; PC, principal component; st-FL,
- 5 subsequently-transforming FL.

6

7 Figure S2: Re-analysis of high-risk patients from *Analysis A* and *Analysis B*

- 8 (A) 3D PCA based on differentially expressed proteins at p<0.05 comparing nt-FL and st-FL
- 9 from the high-risk group identified in *Analysis A*. (B) Hierarchal clustering with input of
- differentially expressed proteins at p<0.05 identified in the high-risk group in *Analysis A*. (C)
- 3D PCA based on differentially expressed proteins at p<0.01 comparing nt-FL and st-FL
- from the high-risk group identified in *Analysis B*. (D) Hierarchal clustering based on
- differentially expressed proteins at p<0.01 identified in the high-risk group in *Analysis B*.
- Abbreviations: nt-FL, non-transforming samples; PC, principal component; st-FL,
- subsequently-transforming samples.

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Figure S1

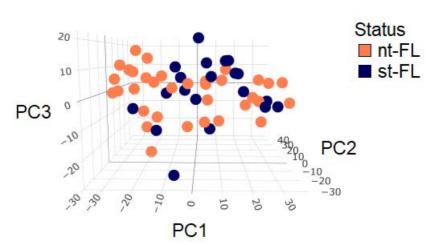


Figure S2

