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Supplementary Materials for

Engineered probiotics biofilm enhances osseointegration via immunoregulation and anti-infection

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Fig. S1. Preparation procedure and surface characterization of *L. casei* biofilm. (A) Preparation procedure of *L. casei* biofilm modified Ti implants. (B) Surface morphology of Ti and AHT-Ti. (C) Surface morphology and Live/dead (green/red) staining of *L. casei* biofilm modified Ti. (D) Number of bacterial colonies on the surface of Ti and AHT-Ti. (E) XPS of Ti, AHT-Ti and *L. casei*-AHT-Ti. n = 3 independent experiments per group, ***P* < 0.01.



Fig. S2. SEM images of *L. casei*-AHT-Ti after immersing in PBS solution for 7, 14 and 28 days.



Fig. S3. SEM images of *L. casei*-AHT-Ti scratched by a 200 μ L pipette tip and a 5 mL syringe needle, respectively.



Fig. S4. The secretion of bacteriocin from the biofilm of *L. casei*-AHT-Ti-1 (1 day), *L. casei*-AHT-Ti-2 (2 days) and *L. casei*-AHT-Ti (3 days) by a Tricine-SDS-PAGE technique.



Fig. S5. **Original phenotypes of Raw 246.7 macrophages.** (A) Characterization of Raw 246.7 macrophages phenotypes *via* CD11b, F4/80, (B) CD11c and CD206 (gated on CD11b⁺F4/80⁺).



Fig. S6. Gene expression of TNF- α , OSM and IL-10 after 72 h treatments. n = 3 independent experiments per group, **P* < 0.05, ***P* < 0.01 and ****P* < 0.001.







Fig. S7. Up-regulated and down-regulated genes GO enrichment analysis in *L. casei*-AHT-Ti.



Fig. S8. Micro-CT results without MRSA infection and immunohistochemical staining for macrophages *in vivo*. (A) Micro-CT results without MRSA infection. (B) Bone volume (BV) / tissue volume (TV) values of Ti, AHT-Ti and *L. casei*-AHT-Ti. (C) Immunohistochemical staining of iNOS (M1) and TGF β (M2) for macrophages, respectively (Scale bar, 100 µm). (D) Mean optical density of iNOS and (E) TGF β expression, respectively. The calculated area is 50 µm around the implant. n = 3 independent experiments per group, **P* < 0.05, ***P* < 0.01 and ****P* < 0.001.

Gene	Primers (F, Forward; R, Reverse)
TNF-α	F: 5'-GACGTGGAACTGGCAGAAGAG-3' R: 5'-TTGGTGGTTTGTGAGTGTGAG-3'
OSM	F: 5'-CCCGGCACAATATCCTCGG-3' R: 5'-TCTGGTGTTGTAGTGGACCGT-3'
IL-10	F: 5'-GCTCTTACTGACTGGCATGAG-3' R: 5'-CGCAGCTCTAGGAGCATGTG-3'
ALP	F: 5'- AGCGACACGGACAAGAAGC-3' R: 5'- GGCAAAGACCGCCACATC-3'
OCN	F: 5'- AAGCCCAGCGACTCTGAGTCT-3' R: 5'- CCGGAGTCTATTCACCACCTTACT-3'
RUNX2	F: 5'- AATGCCTCCGCTGTTATG-3' R: 5'- TTCTGTCTGTGCCTTCTTG-3'
COL-I	F: 5'-CCTGAGCCAGCAGATTGA-3'
	R: 5'-TCCGCTCTTCCAGTCAG-3'
GAPDH	F: 5'- GCCTCGTCTCATAGACAAGATGGT-3' R: 5'- GAAGGCAGCCCTGGTAACC-3'

Table S1. Primers of tested genes by quantitative polymerase chain reaction (qPCR).
