

227 **Supplementary Methods**

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229 **Datasets**

230 A multifasta file of phage genomes was downloaded from INPHARED
231 (<https://github.com/RyanCook94/inphared>; September 2023)⁶. Stop codon reassignment of
232 INPHARED genomes was predicted using Prodigal-gv v2.11.0
233 (<https://github.com/apcamargo/prodigal-gv>), a fork of Prodigal written to improve viral gene
234 calling⁸. Those predicted to use translation table 4 or 15 were retained for downstream
235 analysis.

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237 The Unified Human Gut Virome Catalog (UHGV) was filtered for high quality and complete
238 vOTUs deemed to be a “high confidence” virus and predicted to use either translation table
239 4 or 15 (<https://github.com/snayfach/UHGV>). Stop codon reassignment had already been
240 predicted for UHGV vOTUs using Prodigal-gv and is available in the UHGV metadata.

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242 **Prokka**

243 A fork of Prokka v1.14.5¹¹ was written that incorporates an initial stage of ORF prediction
244 using Prodigal-gv v2.11.0 (<https://github.com/apcamargo/prodigal-gv>)⁸. A first gene calling
245 step is used to infer the genetic code most likely adopted by the genome, then the predicted
246 genetic code is used to perform the translation FASTX::Seq, which we updated to accept
247 code 15 (metacpan.org/pod/FASTX::Seq)¹⁶. The code for this is available at
248 (github.com/telatin/metaprokka). We included publicly available HMMs of the PHROGs
249 database in our Prokka-gv annotations
250 (http://s3.climb.ac.uk/ADM_share/all_phrogs.hmm.gz)¹⁷. The fork is installable from
251 Bioconda as ‘metaprokka’.

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253 **Pharokka**

254 Pharokka v1.5.0¹² was updated to include support for pyrodigal-gv implementing pyrodigal-
255 gv as a gene predictor. This is specified by using ‘-g prodigal-gv’ when running Pharokka. The
256 updated code is available on GitHub (<https://github.com/gbouras13/pharokka>). Pharokka
257 uses tRNAscan-SE for predicting tRNAs¹⁴.

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259 **Statistical Analyses and Data Visualisation**

260 To test for significance in differences of results, a simple paired T test was performed in R
261 v4.2.2¹⁸ and P-values were adjusted using the Benjamini-Hochberg procedure¹⁹. Figure 1 was
262 produced using ggplot2 v3.4.2²⁰.

263 **Supplementary Results**

264 **Prokka-gv Annotations**

265 For Prokka-gv, the largest differences were observed for sequences predicted to use
266 translation table 15, for which Prokka-gv increased the median gene length (median of per
267 genome medians) from 276 to 396 bp for UHGV sequences (43.5% increase), and from 309
268 to 483 bp for INPHARED sequences (56.3% increase). This was also reflected in an increase
269 of median coding capacity from 66.6% to 86.7% for UHGV, and from 69.2% to 87.3% for
270 INPHARED. As it is commonly used as a phylogenetic marker for bacteriophages, we
271 investigated how commonly the major capsid protein (MCP) could be identified with and
272 without predicted stop codon reassignment¹⁵. For sequences predicted to use translation
273 table 15, the MCP could be identified on 382/715 (53.4%) sequences with Prokka and this
274 was marginally increased to 386/715 (53.9%) with Prokka-gv.

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276 When investigating the sequences for which translation table 4 was predicted, a substantial
277 increase was also observed for UHGV sequences, with Prokka-gv increasing median median
278 gene length from 319 to 460 bp (44.2%), resulting in an increase of coding capacity from
279 78.4% to 91.4%. However, the same was not observed for INPHARED sequences predicted to
280 use translation table 4. These sequences observed a modest increase in median median
281 gene length from 573 to 584 bp (1.8%) for Prokka-gv. Median coding capacity was not
282 increased with Prokka and Prokka-gv both obtaining 86.2%.