









Supplementary Figure 1. sp-tagged αSyn are secreted and internalized. (a) Representative WB analyses of whole cell lysates (WCL, 1:10 from the total lysate) and conditioned media (CM, 1:100 from the total CM) of human neuroblastoma SH-SY5Y cells expressing GFP or wt-, sp1- or sp2-aSyn. Stck, stacking gel. (b) Quantification of the PRM traces of cell lysates ("Cells") and conditioned media ("Media") of cells transfected with an empty vector (\emptyset) or expression vectors for wt-aSyn or sp2-aSyn. The PRM traces are shown in (c) where the tryptic peptides targeted for PRM experiments are also indicated. Bar plots show averaged counts per seconds (cps) of PRM traces corresponding to the indicated tryptic peptides. Data is shown as the media \pm SD. * p < 0.05compared to empty vector (one-way ANOVA followed by the post hoc Tukey's test n=4). (d) PRM traces of cell lysates ("Cells") and conditioned media ("Media") of cells transfected with an empty vector (Ø) or expression vectors for wt-aSyn or sp2-aSyn. The acetylated aSyn N-terminus tryptic peptide (sequence: MDVFMK) was targeted for PRM experiments. (e) Z-stack images of selected panels of figure 1c. (f) Representative WB of WCL of Cos7 cells expressing GFP or wt-, sp1- or sp2-aSyn and treated with vehicle or monensin for 16 h. (g) Quantification of the signal intensity of the aSyn immunoblots of (f). Two areas of the gel were quantified separately, the lower part containing aSyn monomers and the upper part containing the HMW species. Total signal intensity was calculated as the sum of monomers and HMW. Data is shown as the media ± SD. * and **, p < 0.05 respect GFP and monensin, respectively (one-way ANOVA followed by the post hoc Tukey's test, n=3). (h) PRM traces of cell lysates ("Cells") and conditioned media ("Media") of cells transfected with an empty vector (ϕ) or expression vectors for wt- α Syn or sp2- α Syn. The sp2 semi-tryptic peptide was targeted for PRM experiments. Cps, counts per second. RT, retention time. (i) Western blot analyses of whole cell lysates obtained from mammalian cells transexpressed with recombinant wt-aSyn and sp2-aSyn. Anti-aSyn antibodies were used to detect the transexpressed proteins whereas an anti-GAPDH antibody was used as loading control. (j) Z-stack images of selected panels of figure 1f.



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Supplementary Figure 2. sp2- α Syn is toxic, amyloidogenic and cell-to-cell transmitted.

(a) Thioflavin-T fluorescence of recombinant wt-αSyn (blue) and sp2-αSyn (orange) incubated in PBS buffer at 37 °C in agitation for the indicated times. On the right a magnification of the 0-24 h is shown. (b) Two dimensional [¹⁵N,¹H]-Heteronuclear Multiple Quantum Coherence (HMQC) NMR spectra of ¹⁵N-labeled wt- and sp2-αSyn incubated in PBS buffer for the indicated times. NMR measurements were carried out at 10 °C. (c) Two dimensional [¹⁵N,¹H]-HMQC NMR spectra of ¹⁵N-labeled wt- and sp2-aSyn transexpressed in mammalian cells and kept in the NMR tube for the indicated times ("in cell-NMR"). The reference spectra correspond to the same proteins in buffer (first black spectra) is shown ("PBS"). NMR measurements were carried out at 10°C. (d) Bar graph showing the luciferase activity recovered in CM of cells co-expressing the Gaussia luciferase and wt-, sp1-, sp2- α Syn or the control secreted protein sp2-VC. Data is shown as the media ± SD. * p < 0.005 (one-way ANOVA followed by the post hoc Tukey's test, n=5). (e) Cos7 cells expressing sp2-aSyn for 48 h were fixed and subjected to confocal microscopy analyses to visualize aSyn (green), aSyn phosphorylated at serine 129, P62 and ubiquitin (all them in red). DAPI (blue) was used for nuclei staining. Scale bar, 10 μ m. (f) Western blot analyses of cell lysates ("Cells") and conditioned media ("CM") of cells transfected with an empty vector (-) or expression vectors for wt- α Syn or sp2- α Syn. Antibodies against α Syn and α Syn phosphorylated at serine 129 were used to detect the modified forms of this protein, whereas an anti-HSC70 antibody were used as loading control. (g) Schematic representation of the bimolecular fluorescence complementation (BiFC) approach used to study sp2-aSyn cell-to-cell propagation, seeded oligomerization and secondary secretion. Note that for visualization purposes the sp2 peptide was not removed from the secreted sp2- α Syn. (h) Cos7 cells expressing wt-aSyn-VC, VN-wt-aSyn or the control proteins VN or VC were grown alone or co-incubated as indicated. After 48 h the cells were fixed and subjected to confocal microscopy analyses to quantify cells containing the reconstituted Venus protein. Very few fluorescent cells (counts) were observed in wt-aSyn-VC and VN-wt-aSynexpressing cells even in absence of the missing complementary GFP half (bars 3 and 4). Co-cultures of cells expressing wt-aSyn-VC and VN-wt-aSyn (bar 7) yielded few fluorescent cells which were more abundant compared to cells expressing these proteins alone. Data is shown as the media \pm SD. * p < 0.05 compared to VN and VC. ** p < 0.05 compared to all other conditions. (i) Cos7 cells expressing sp2-aSyn-VC, VN-wt-aSyn or the control protein VC were co-incubated with cells expressing VN-wt-aSyn, VN or VC. After 48 h the cells were fixed and subjected to confocal microscopy analyses to visualize the reconstituted Venus protein. Scale bar, 10 μ m. (j) Quantification of luciferase activity in acceptor cells expressing sp2-aSyn-VC (grey bars) or VN-wt-aSyn (black bars). Acceptor cells were treated with conditioned media containing VC-sp2-aSyn or VN-wt-aSyn. Data is shown as the media \pm SD. * p < 0.05 compared to Empty. ** p < 0.01 compared to cells lacking α Syn (squared bars). Unpaired, two tails distribution Student's t test, (n=5). (k) HeLa cell clones expressing VN-wt- α Syn or sp2- α Syn-VC were co-cultured as indicated. 24 h later the CM was collected and subjected to WB together with the cell lysates (Cells). Ø, empty lane



Supplementary Figure 3. Neuron-derived extracellular aSyn triggers a robust LB-like pathology in the substantia nigra. (a) Representative IHC images of coronal sections of cortex, striatum and the CA2 region of the hippocampus (CA2) of wild type mice transduced with AAV-sp2-aSyn or AAV-wt-aSyn for 4 months. Both contralateral (Contra) and ipsilateral (Ipsi) hemispheres are shown. Arrows, aSyn-positive inclusions found in neuronal bodies and fibers. Scale bar, 100 µm. (b) Representative IHC images of consecutive coronal sections of brains of wild type mice (n=6) transduced with an empty AAV9 (grey, Empty) or an AAV9 encoding GFP (green, GFP). Animals were sacrificed at 4 months post injection. Approximate rostrocaudal coordinates taking the bregma as reference are indicated on the right. Only ipsilateral hemispheres are shown. Arrows, substantia nigra. (c) Control IHC images of substantia nigra of wild type mice transduced with AAV-GFP, AAV-sp2- or AAV-wt-αSyn in for 4 months. Primary antibody was omitted in these control experiments. Scale bar, 100 µm. (d) Representative fluorescence images of mice expressing eGFP for 4 months in the DMN. The area where the virus was administrated and the substantia nigra are shown. Nuclei were stained with DAPI. Scale bar, 100 µm.



Supplementary Figure 4. Lewy body-like pathology elicited by sp2- and wt-αSyn. (a) Representative double immunofluorescence images of mice expressing GFP for 4 months. Immunostaining was carried out with antibodies specific for ubiquitin (Ub) and aSyn phosphorylated at serine 129. Nuclei were stained with DAPI. Scale bar, 10 µm. (b) Lower magnification images showing immunofluorescence analyses of brains expressing sp2- and wt- α Syn for 4 months. Immunostaining was carried out with an antibody that recognizes aSyn phosphorylated at serine 129. a higher magnification showing the substantia nigra is shown in the bottom. Nuclei were stained with DAPI. (c) Representative WB of brain tissue enriched of midbrain from mice transduced with AAV-GFP, AAV-wt- or AAV-sp2aSyn. An anti-aSyn antibody (clone LB509) that recognizes human but not mouse aSyn was used for the immunoblot. Note that in this experiment we used four times more material from the sp2-aSyn mice due to lesser overall amount of αSyn compared to wt-αSyn-expressing mice (see GAPDH). (d) Representative WB of brain tissue enriched of midbrain from mice transduced with AAV-wt- or AAV-sp2-aSyn and subjected to sequential extraction with the indicated buffers. The LB509 anti-aSyn antibody was used. Stck, stacking gel. Four times more material from the sp2- α Syn mice was used due to lesser overall amount of α Syn compared to wt- α Syn-expressing mice.



O Injection site

Supplementary Figure 5. sp2-αSyn spreading in the mammalian brain. (a) Representative IHC images of coronal brain sections of hippocampus and striatum of wild type mice that received a single injection of an AAV9 encoding sp2- or wt-aSyn in the cortex. Animals were sacrificed at 4 months post-surgery. In the upper part a schematic representation of the anatomical structures analyzed that includes approximate rostro-caudal coordinates with the bregma as reference. Contralateral (Contra) and ipsilateral (Ipsi) hemispheres are shown. Scale bar: 200 µm.



Supplementary Figure 6. sp2- α Syn triggers neuronal death and neuroimmflamation. (a and b) Representative immunofluorescence images of the substantia nigra of mice transduced with GFP, sp2- or wt- α Syn AAVs for 4 months. Human α Syn (green) was co-immunostained with the marker for astrocytes GFAP (a) or microglia Iba1 (b). Arrows indicate co-localization between the glial marker and α Syn in astrocytes or microglia. DAPI was used for nuclear staining. Scale bar: 100 µm. (C) Representative WB of brain tissue from mice transduced with AAV-GFP (n=3) or AAV-sp2- α Syn (n=3) for 4 months.



Supplementary Figure 7. Locomotor deficits elicited by sp2- α Syn. (a-c) Plots of the total distance travelled (a), time (in percentage) that the animals occupied the center (b) or distance travelled in the center (c) derived from the open field test (20 minutes sessions). Mice expressing GFP (n=6), sp2- (n=6) or wt- α Syn (n=5) for 4 months were used in this study. Data is shown as the media \pm SD. # *p* < 0.05 (one-way ANOVA followed by the *post hoc* Dunnett's test).

Western blotting, uncropped blots





Figure 1d





Figure 1e

Supplem. Figure 1i

Supplem. Figure 1a





Supplem. Figure 2f















Figure 2d





Supplem. Figure 2k





Supplem. Figure 4d



Supplem. Figure 4c







Supplem. Figure 6c





