

Supplemental information

**Circulating NK cells establish tissue residency upon
acute infection of skin and mediate accelerated
effector responses to secondary infection**

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Figure S1

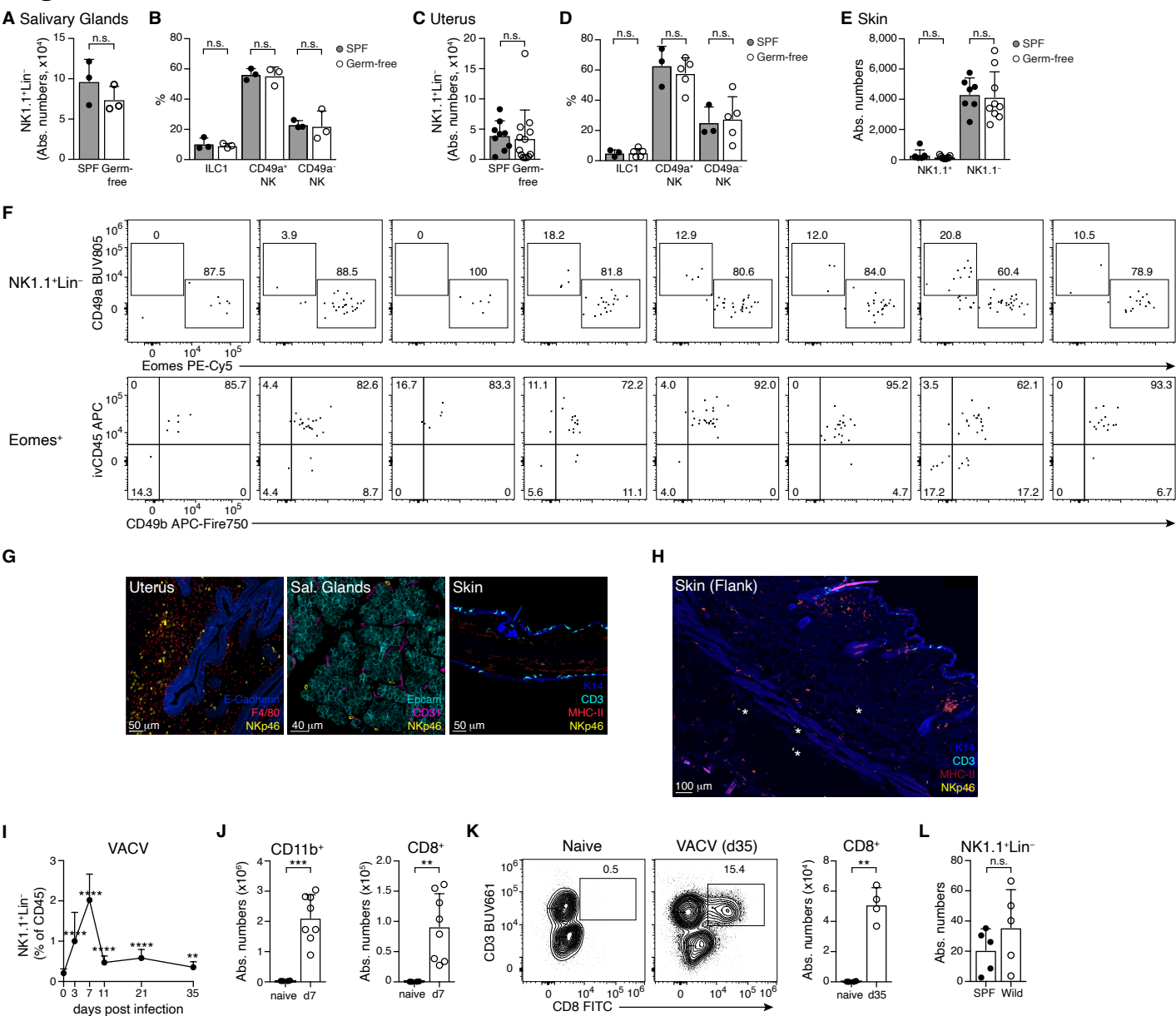


Figure S1: Immune populations in naïve and VACV-infected skin. Related to Figure 1.

(A-E) Numbers of Lin⁻NK1.1⁺ cells (A, C) or Lin⁻CD90⁺CD127⁺Il18ra⁺ ILC subsets (E) and frequency of Eomes⁻ ILC1 and Eomes⁺ NK subsets out of Lin⁻NK1.1⁺ cells (B and D) in indicated tissues of mice housed in SPF or germfree conditions. (F) Representative FACS plots show Lin⁻NK1.1⁺ (top row) and Eomes⁺ NK cells (bottom row) in naïve ears. Individual ears are shown. (G, H) Confocal immunofluorescence images of skin cryosections from indicated organs of naïve mice. (I) Frequency of Lin⁻NK1.1⁺ cells in ear skin of mice infected with VACV at the indicated times post infection. (J, K) Numbers and representative gating of CD11b⁺ cells and CD8⁺ T cells in naïve versus VACV-infected ear skin at indicated times post infection. (L) Absolute numbers of Lin⁻NK1.1⁺ cells in the ear skin of naïve mice housed in SPF or wildling conditions. Data are representative of three independent experiments with n=3 mice (A, B, D, G, H) or n=4-8 ears (F, J-L) per group. Data in (C, E, I) were pooled from two independent experiments with n=9-13 mice (C) or n=7-23 ears (E, I) per group. Error bars indicate mean ± SD. p values were calculated by unpaired Student's t test. **p<0.01, ***p<0.001, ****p<0.0001, n.s. not significant.

Figure S2

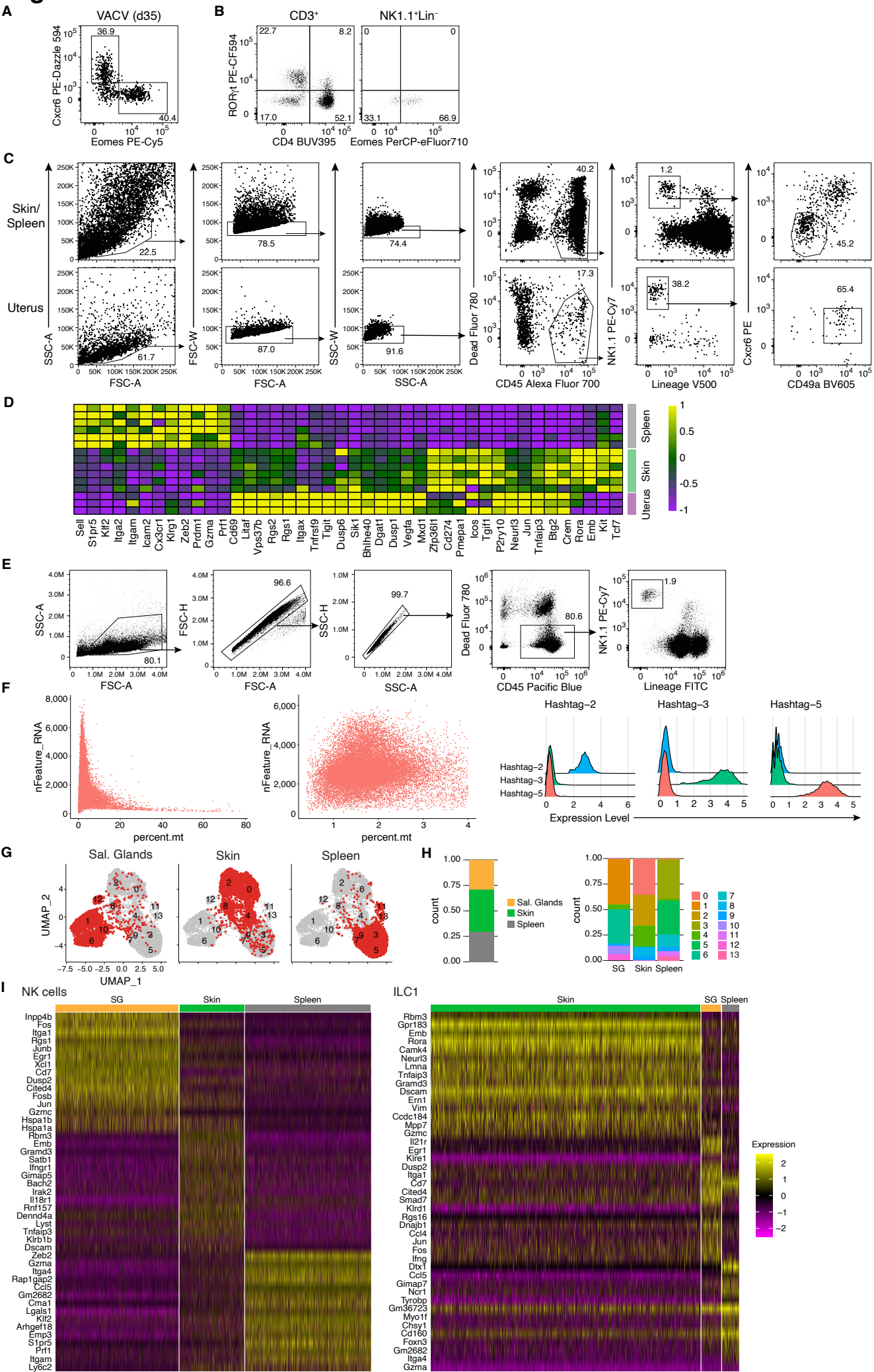


Figure S2: mRNA sequencing reveals unique and tissue-specific NK and ILC1 gene signatures. Related to Figure 2.

(A, B) Representative FACS plots of Lin⁻NK1.1⁺ (A, B right) and CD3⁺ T cells (B left) in ear skin of mice infected with VACV (A) or *S. aureus* (B). (C, E) Full gating strategy for NK cells sorted for bulk (C, D) and scRNA-seq analysis (E-I). (D) Heat map depicting z-score of significantly DEG in the indicated tissues. (F-I) scRNA-seq of hashtagged skin, spleen and SG Lin⁻NK1.1⁺ cells sorted from VACV-infected mice on d25 pi. (F) General QC, filtering of low-quality cells (feature counts > 1000 and percentage of genes mapped to mitochondrial genome < 8) and doublets, and demultiplexing of CD45 and Hashtag (HT) antibodies to identify tissue of origin. Single cell transcriptome visualization using a UMAP color-coded by hashtag (tissue origin) (G) and bar graph show the proportion of cells per tissue and cluster (H). (I) Heat map of the top 15 most DEG across NK cells (clusters 1, 3, 5, 6, 7, 9, 10, 13, left) and ILC1 (0, 2, 11, 12) in the indicated tissues. Data in (A, B) are representative of three independent experiments with n=4 ears per group.

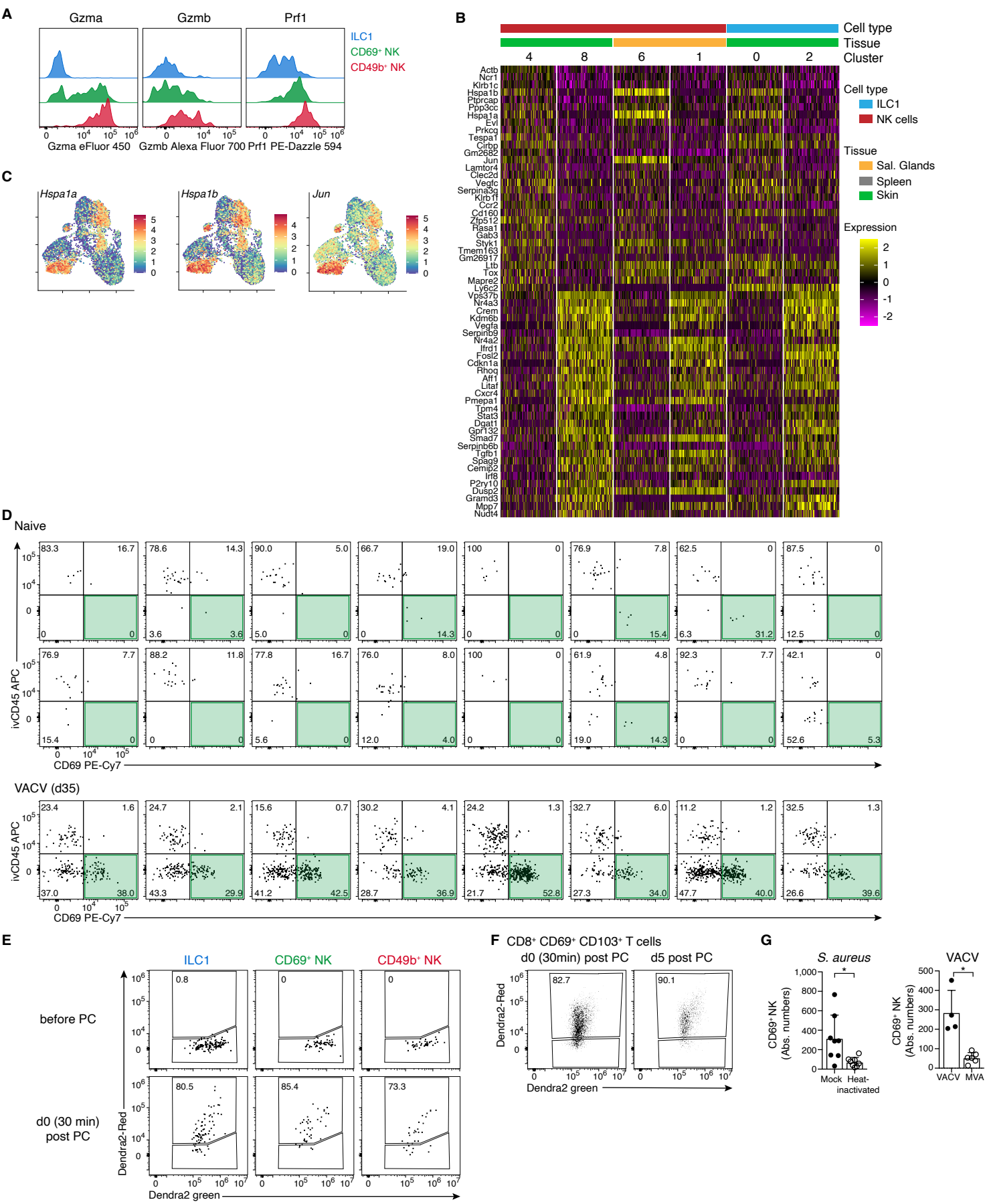
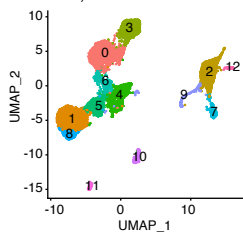
Figure S3

Figure S3: Skin NK cells from naïve and infected mice. Related to Figure 3.

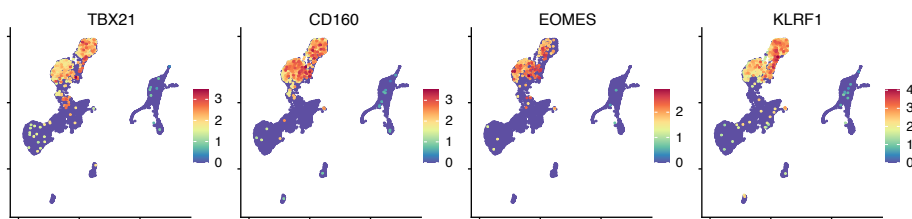
(A) Representative histograms show marker protein expression by indicated skin NK subsets 4 weeks post VACV infection. (B) Heat map showing gene expression of top 30 DEG between cluster 4 and 8 for all clusters. (C) UMAP visualization of selected gene expression, correlating with subclustering of trNK (cl4+8, cl1+6) and ILC1 (cl0+2). (D) FACS plots show frequency of Eomes⁺ NK subsets from individual naïve (top) and VACV-infected (bottom) ear skins after intravascular labelling with anti-CD45 (ivCD45). (E, F) Representative FACS plots show Dendra2^{Red+} fraction before and 30 mins (d0) or 5 days post photoconversion (PC) among indicated subtypes of Lin⁻NK1.1⁺ cells (E), or CD8⁺CD69⁺CD103⁺ tissue-resident T cells (F). (G) Absolute numbers of ear skin CD69⁺ NK cells 4 weeks post *S. aureus*, heat-inactivated *S. aureus*, VACV-wt or non-replicating VACV (MVA) infection. Data are representative of three (A, D, G) or two (B, C, E, F) independent experiments with n=4-16 ears per group. Error bars indicate mean ± SD. p values were calculated by unpaired Student's t test. *p<0.05.

Figure S4

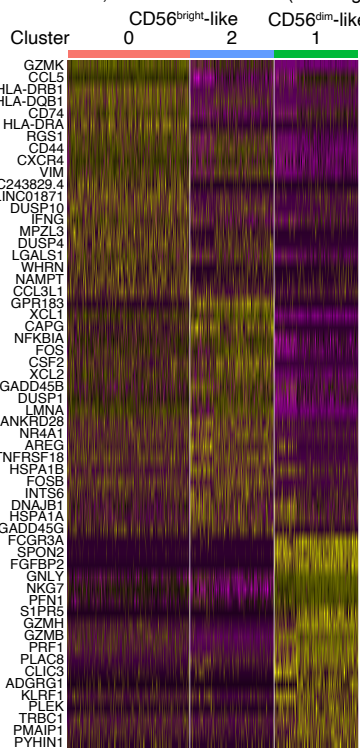
A Alkon et al., human skin



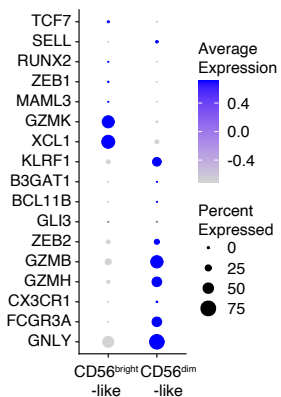
B



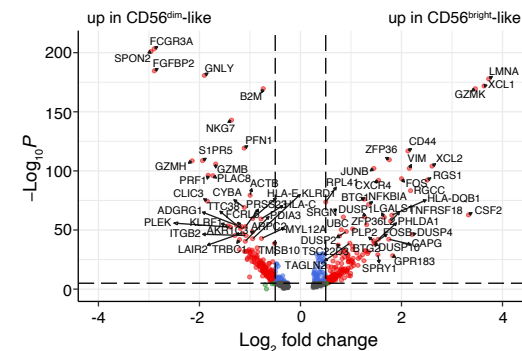
C Alkon et al., human skin NK cells (from Figure 4A)



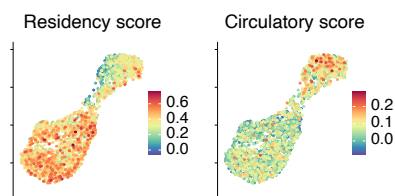
D Alkon et al., human skin, NK cells



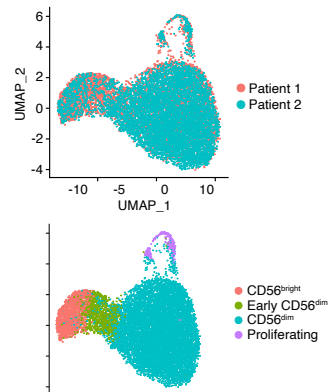
E Alkon et al., human skin, NK cells



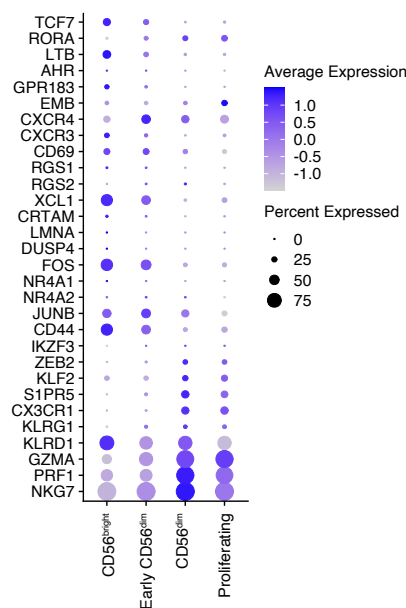
F



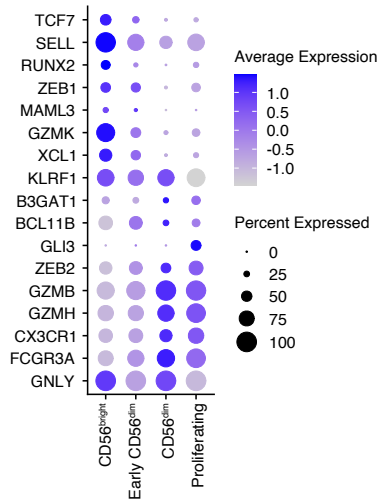
G Ruckert et al., human PBMC



H Ruckert et al., human PBMC



I Ruckert et al., human PBMC



J Ruckert et al., human PBMC

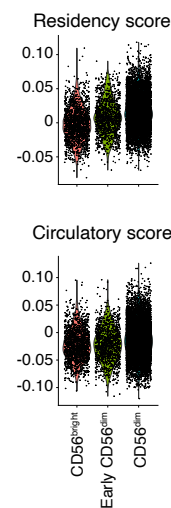


Figure S4: Gene expression analysis of human NK cells from skin and peripheral blood. Related to Figure 4.

Analysis of single-cell transcriptome of healthy human skin data generated by Alkon et al.¹ (A-F), and human PBMC data generated by Ruckert et al.² (G-I). (A and B) UMAP visualization delineating NK clusters, identified based on expression of TBX21, EOMES, CD160 and KLRF1. (C-E) Analysis of human NK cells re-clustered from cluster 0 and 3 as in Figure 4. Heatmap (C) and volcano plot (E) show DEG across indicated NK clusters. (D, H) Bubble plot representation of z-score of selected marker genes of indicated NK clusters. (F) UMAP of a score based on genes associated to tissue-resident memory of T cells. (G) UMAP visualization based on origin or on identified clusters. (I) Violin plot visualization of residency and circulating scores, based on genes associated to tissue-resident memory of T cells as in Figure 4E, in the indicated subsets.

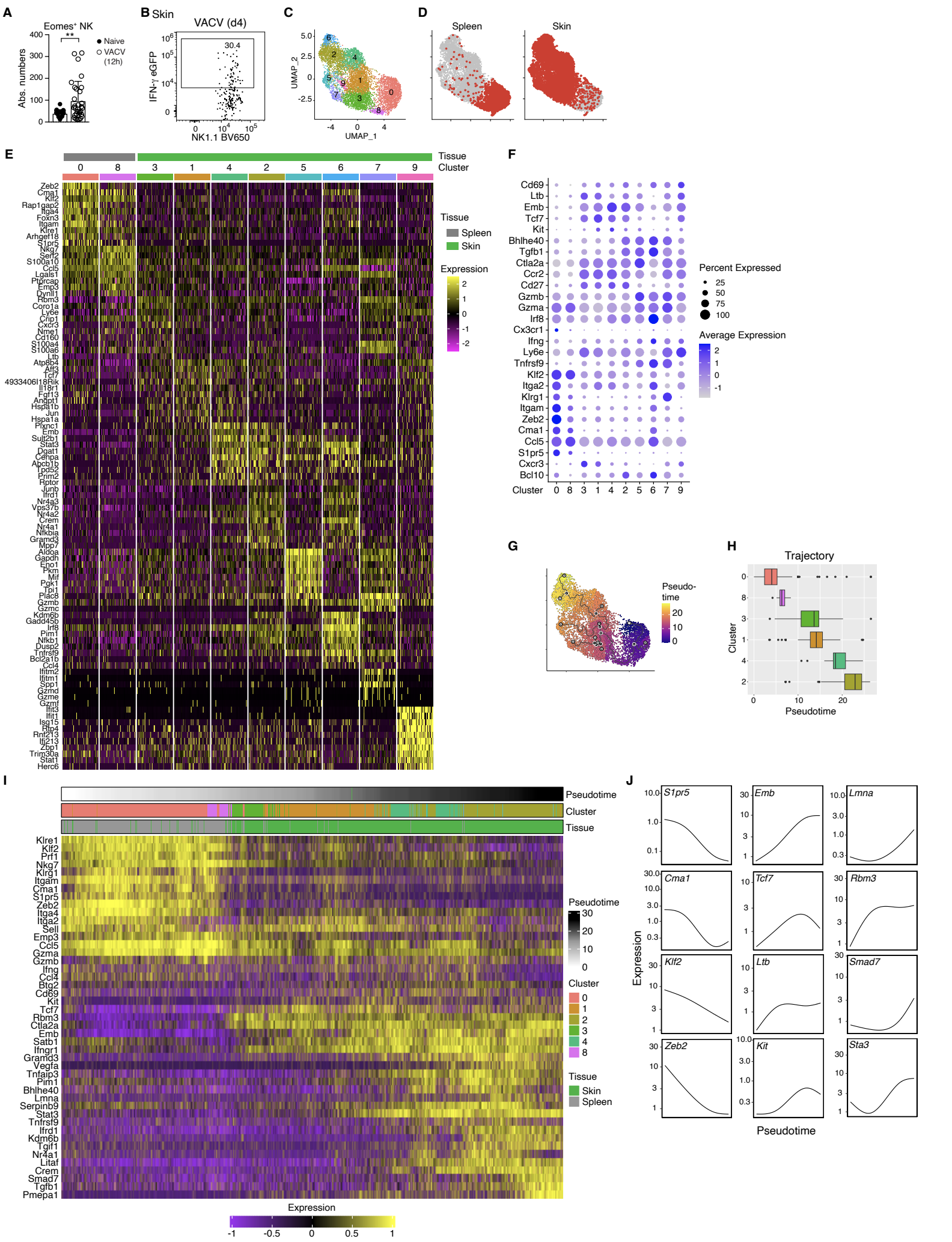
Figure S5

Figure S5: Rapid recruitment and activation of NK cells VACV infected skin. Related to Figure 5.

(A) Absolute numbers of Eomes⁺ NK cells from naïve ear skins or 12h post VACV-wt infection. (B) Representative FACS plot showing IFN- γ (eGFP)-expressing NK cells in ear skin on d4 post VACV infection. (C-F) Single cell mRNA-Seq analysis of spleen and skin NK cells sorted on d8 post VACV infection. (C, D) UMAP visualisation of clusters (C) and tissue origin (D). (E) Heatmap showing top 10 most DEG across clusters. (F) Bubble plot representation of z-score of selected marker genes between NK cell clusters. (G, H) Pseudotime and trajectory analysis using Monocle3. (I, J) Expression of selected genes in NK cells along pseudotime. Data in (A) are pooled from three independent experiments with n=18 or n=30 ears per group. Data in (B) are representative of two independent experiments with n=4 ears.

Figure S6

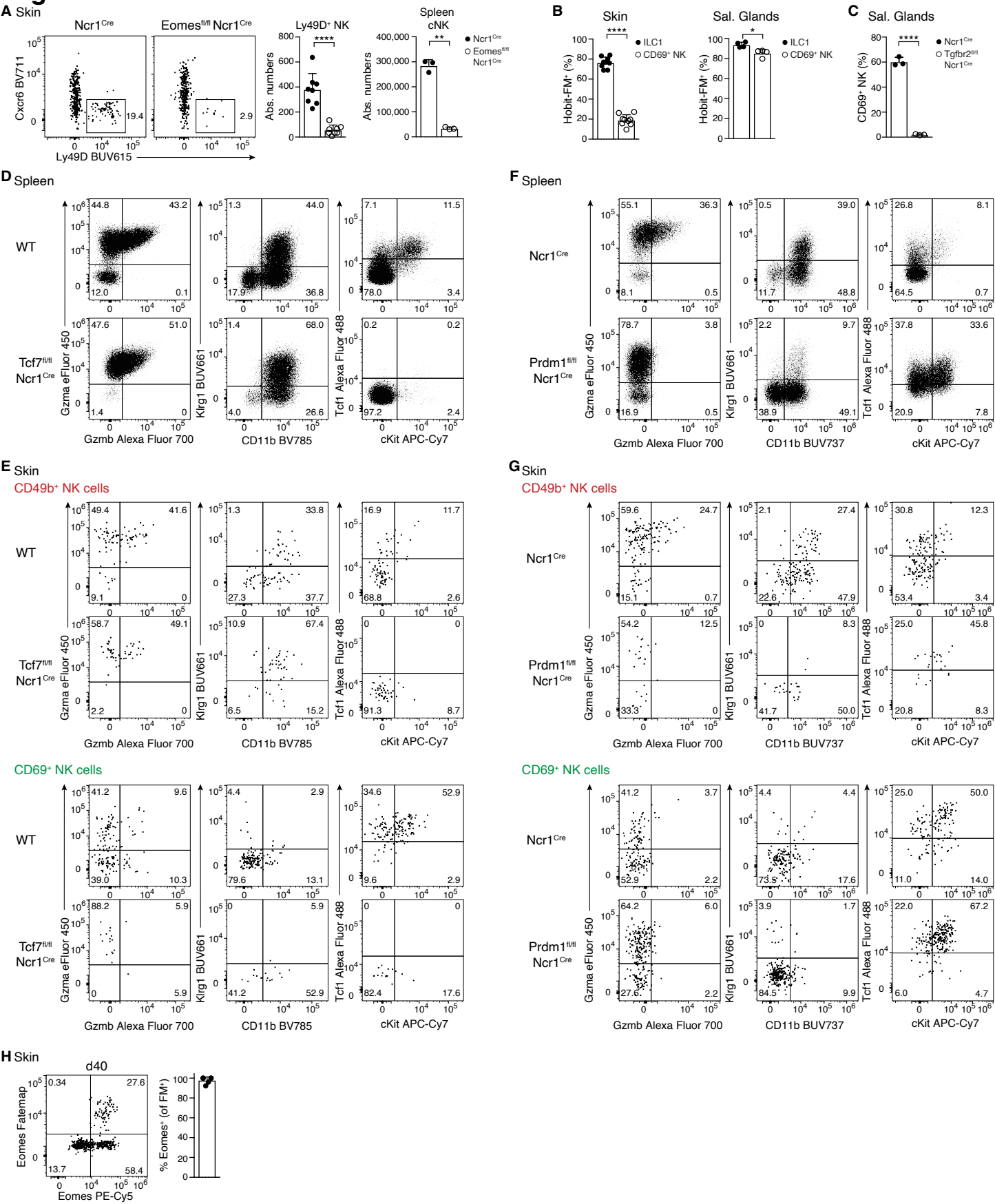


Figure S6: Eomes, Tcf1 and Blimp1 regulate the differentiation of circulating and skin-resident NK cells. Related to Figure 6.

Analysis of NK cell subsets (A-G) and ILC1 (B) in the indicated organs of Eomes^{fl/fl} Ncr1^{cre} (A), Hobit-FM (B), Tgfb2^{fl/fl} Ncr1^{cre} (C), Tcf7^{fl/fl} Ncr1^{cre} (D, E) and Prdm1^{fl/fl} Ncr1^{cre} (F, G) and indicated control mice on d30 post VACV infection. Spleen (A) and SG (B, C) were analyzed from naïve mice. (A) Alternative gating strategy for ear skin NK cells in absence of Eomes expression as Lin⁻NK1.1⁺Ly49D⁺ cells. Spleen NK cells were gated as Lin⁻NK1.1⁺CD49a⁻CD127⁻ cells. (H) Representative FACS plot showing Lin⁻NK1.1⁺ of Eomes-FM mice on d40 post infection. Bar graphs shows frequency of Eomes⁺ cells within Lin⁻NK1.1⁺ FM⁺ cells. Data are representative of two (A, B, D, E, H) or three (C, F, G) independent experiments with n=3-4 mice (A-D, F, H) or n=8-9 ears (A, B, E, G) per group. Error bars indicate mean ± SD. p values were calculated by unpaired Student's t test (A-C). *p < 0.05, ****p < 0.0001.

Figure S7

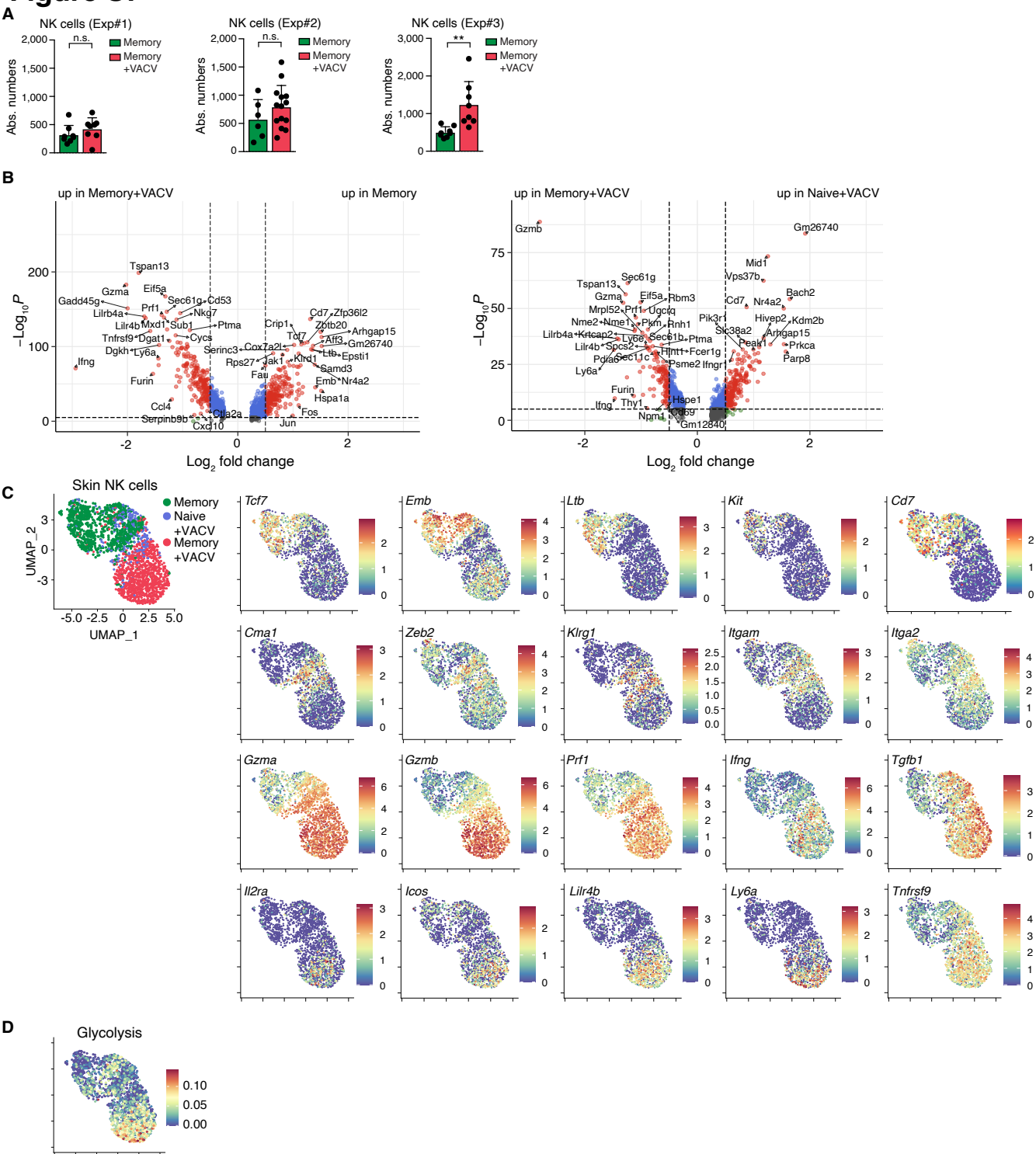


Figure S7: Skin-resident NK cells rapidly reprogram into effector cells during secondary infection. Related to Figure 7.

Analysis of ear skin NK cells as in Figure 7A. (A) Absolute numbers of Eomes⁺ NK cells. Three independent experiments with n=6-14 ears per group are shown. (B) Volcano plot showing DEG. (C, D) Single cell mRNA-Seq analysis as indicated in Figure 7G. UMAP visualization of selected marker gene expression (C) or glycolytic score (D). Error bars indicate mean \pm SD. p values were calculated by unpaired Student's t test. **p<0.01, n.s. not significant.