

Small Molecule Screen Identifies Non-Catalytic

USP3 Chemical Handle

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Table S1: Compounds smiles strings and catalog numbers

Compound #	Catalog_Number	SMILES
1	STL102634	<chem>C(C([O-])=O)c1cccnc1</chem>
2	STK256632	<chem>C(Cn1cccn1)C([O-])=O</chem>
3	STK661679	<chem>C(Cc1ccno1)C([O-])=O</chem>
4	Z57127332	<chem>C(Cc1ccccc1)C(O)=O</chem>
5	STK802016	<chem>C(C([O-])=O)Nc1ncccn1</chem>
6	IF_02_176	<chem>C(Cc1cnccn1)C(O)=O</chem>
7	SY019802	<chem>C(Cc1nccs1)C(O)=O</chem>
8	A0954	<chem>C(C(O)=O)c1csc(N)n1</chem>
9	STK891980	<chem>C(COc1ccccc1)C([O-])=O</chem>
10	STL193409	<chem>C(CN1C=CC=CC1=O)C([O-])=O</chem>
11	Z1171978885	<chem>C(C([O-])=O)c1cc2ccccc2o1</chem>
12	STL163377	<chem>C(C([O-])=O)c1nc2ccccc2[nH]1</chem>
13	Z1333043518	<chem>C(C([O-])=O)n1cc2ccccc2n1</chem>
14	F1913-0024	<chem>C(C([O-])=O)c1nc2ccccc2o1</chem>
15	STL124046	<chem>C(C([O-])=O)NC(c1cccnc1)=O</chem>
16	STK526633	<chem>C(CC(c1cccs1)=O)C([O-])=O</chem>
17	STL190846	<chem>C(C([O-])=O)c1cccc2ccnc12</chem>
18	STK695471	<chem>C(Cc1cn2cccn2n1)C([O-])=O</chem>
19	EN300-14584	<chem>C(Cn1cnc2ccccc12)C(O)=O</chem>
20	EN300-70527	<chem>C(C(O)=O)N1Cc2ccccc2C1=O</chem>
21	STK504141	<chem>C(C([O-])=O)Nc1ccc2nnnc2n1</chem>
22	STK353430	<chem>C(Cc1nc2ncccn2n1)C(O)=O</chem>
23	Z188924494	<chem>C(Cc1ccc2c(c1)OCO2)C([O-])=O</chem>
24	IF_03_114	<chem>C(CC(Nc1ccccc1)=O)C(O)=O</chem>
25	STK723995	<chem>C(C([O-])=O)Nc1c2ccccc2ncn1</chem>
26	MS-2504	<chem>C(C([O-])=O)Oc1cnc2ccccc2n1</chem>
27	STK392449	<chem>C(C([O-])=O)N1C=Nc2ccccc2C1=O</chem>
28	EN300-188561	<chem>C(C(O)=O)N1C=Cc2ccccc2C1=O</chem>
29	STK738252	<chem>C(CN1C(=O)Oc2ccccc12)C([O-])=O</chem>
30	Z57983005	<chem>C(C1C(Nc2ccccc2O1)=O)C(O)=O</chem>
31	STK447336	<chem>Cc1cc(NC(CCC([O-])=O)=O)nn1C</chem>
32	CDS014372	<chem>C1C[C@H](C(N)=O)N(C1)C(CCC([O-])=O)=O</chem>
33	Z126932466	<chem>C(Cc1cnn(c1)c1ccccc1)C(O)=O</chem>
34	STK500480	<chem>C(Cc1nc(c2ccccc2)no1)C([O-])=O</chem>

35	TS-00478	<chem>C(Cc1nnc(c2cccc2)o1)C([O-])=O</chem>
36	STL310201	<chem>C(CC([O-])=O)C1C(N2C=CC=CC2=NN=1)=O</chem>
37	AL-291/37197008	<chem>C(Cn1nc(c2cccc2)nn1)C(O)=O</chem>
38	STK099228	<chem>C(CC(N1CCc2cccc12)=O)C([O-])=O</chem>
39	Z166605066	<chem>C(CN1C(c2cccc2N=N1)=O)C(O)=O</chem>
40	STK946249	<chem>C(CN1C(c2cccc2S1)=O)C([O-])=O</chem>
41	EN300-00843	<chem>CC1(C)Cc2ccc(c2O1)OCC(O)=O</chem>
42	EN300-35451	<chem>CC1=C(CCC([O-])=O)C(NC(=N1)SC)=O</chem>
43	Z1259161843	<chem>C(C([O-])=O)Nc1ccc(cc1)S(N)(=O)=O</chem>
44	EN300-10660	<chem>Cc1c2C(N(CC([O-])=O)C=Nc2sc1C)=O</chem>
45	IF_04_062	<chem>C(Cc1nc2cccc3cccc(c23)[nH]1)C(O)=O</chem>
46	IF_03_115	<chem>C(CC(Nc1cccc2cccc12)=O)C(O)=O</chem>
47	Z90120424	<chem>C(C(O)=O)N1C=Nc2c3cccc3oc2C1=O</chem>
48	EN300-137714	<chem>C(CC(c1ccc2c(CCC(N2)=O)c1)=O)C(O)=O</chem>
49	Z102925466	<chem>C(CN1C(c2cccc2S1(=O)=O)=O)C(O)=O</chem>
50	IF_01_187	<chem>C(Cc1cccc1OCc1ccccn1)C(O)=O</chem>
51	STK925744	<chem>CC1=CC(=O)Oc2cc(c(CCC([O-])=O)cc12)OC</chem>
52	STK682580	<chem>C1CCn2c(C1)c(C#N)c1c2C(N(CC(O)=O)C=N1)=O</chem>
53	KM09821	<chem>C(CC(O)=O)CN1CCC(=CC1)c1cccc1.[Cl]</chem>
54	STK545027	<chem>CC(C(O)=O)N1C(C=Cc2ccc(cc2)OC)SC1=O)=O</chem>
55	Z1682077208	<chem>C1CCN(C1)S(c1ccc(cc1)C(CCC(O)=O)=O)(=O)=O</chem>
56	79148212	<chem>C(Cc1nc2cccc2n1c1cc(C(N)=O)sc1)C(O)=O</chem>
57	MS-2054	<chem>C(C(Nc1nnc(SCC(O)=O)s1)=O)Oc1cccc1</chem>
58	Z609944470	<chem>C(Cc1nc2cccc2c2nc(Cc3cccc3)nn12)C([O-])=O</chem>
59	Z1373577255 [Enamine]	<chem>C(CC(c1cccc(c1)NS(c1c[nH]c2c1cccn2)(=O)=O)=O)C(O)=O</chem>
60	Z1413959119	<chem>Cc1ccc(cc1S(Nc1cccc(c1)C(CCC(O)=O)=O)(=O)=O)C(O)=O</chem>
61	HGA-4028-0036-50	<chem>C1CN(CCC1c1ccc(cc1)[Cl])C(c1ccc(C(NCC(O)=O)=O)nc1)=O</chem>
62	CAZ-0010-0660-20	<chem>C1CN(CCC1N1C(=O)Oc2cccc12)S(c1ccc(cc1)C(NCC(O)=O)=O)(=O)=O</chem>

Table S2: Matrix Screen results (1000 means KD > 1000 μ M)

Compound #	KD (μ M)										
	USP3	USP5	USP13	USP16	USP20	USP33	USP39	USP49	USP51	BRAP	HDAC6
1	121	1000	1000	96.1	707	115	180	283	1000	73.9	1000
2	1000	1000	98.2	1000	182	1000	1000	56	1000	74.4	1000
3	1000	1000	1000	1000	1000	1000	1000	1000	1000	1000	1000
4	178	1000	1000	429	1000	1000	1000	172	1000	82.9	1000
5	855	1000	616	1000	574	1000	1000	1000	1000	1000	1000

46	1000	1000	1000	1000	1000	1000	1000	1000	1000	66	198.8
47	1000	312	1000	344	426	1000	1000	1000	1000	253	247
48	140	532	1000	167	821	1000	1000	99.9	1000	126	103.6
49	1000	1000	1000	1000	1000	1000	1000	1000	1000	1000	1000
50	49.5	1000	365	48.2	249	1000	1000	1000	1000	483	1000
51	354	1000	867	535	465	1000	1000	86.2	1000	71.6	343.2
52	603	207	1000	403	552	1000	1000	1000	1000	215	105.4
53	1000	561	594	1000	380	1000	1000	1000	1000	1000	1000
54	441	1000	1000	116	457	1000	1000	1000	1000	1000	1000
55	91.5	117	1000	139	798	1000	1000	1000	1000	340	51.54
56	1000	1000	754	1000	464	1000	1000	1000	1000	1000	1000
57	219	315	745	372	401	1000	1000	1000	1000	703	273.4
58	238	847	1000	285	1000	1000	1000	1000	1000	1000	187.4
59	19.1	168	1000	69.8	706	1000	1000	1000	1000	413	211.6
60	283	1000	1000	114	1000	537	942	187	1000	234	1000
61	344	71.1	642	436	430	1000	1000	1000	1000	537	415
62	216	127	1000	536	1000	1000	1000	1000	1000	1000	234.1
Ub	4.83	3.01	1000	8.55	1000	1000	1000	1000	1000	35	9.134
Ub-RGGdel	1000	1000	1000	1000	1000	1000	1000	1000	1000	1000	1000

Table S3: Eurofins Diversity panel on 58 kinases (Radiometric activity assay)

Kinase	Catalogue Number	% kinase activity in the presence of 10 μ M of compound 59
Abl(h)	14-529KP	96
ALK(h)	14-555KP	98
AMPK α 1(h)	14-840KP	90
ASK1(h)	14-606KP	91
Aurora-A(h)	14-511KP	56
CaMKI(h)	14-663KP	98
CDK1/cyclinB(h)	14-450KP	111
CDK2/cyclinA(h)	14-448KP	106
CDK6/cyclinD3(h)	14-519KP	93
CDK7/cyclinH/MAT1(h)	14-476KP	85
CDK9/cyclin T1(h)	14-685KP	100
CHK1(h)	14-346KP	108
CK1 γ 1(h)	14-711KP	104
CK2 α 2(h)	14-689KP	112
c-RAF(h)	14-352KP	108
DRAK1(h)	14-668KP	106
eEF-2K(h)	14-654KP	106
EGFR(h)	14-531KP	98
EphA5(h)	14-639KP	99
EphB4(h)	14-554KP	88

Fyn(h)	14-441KP	84
GSK3 β (h)	14-306KP	99
IGF-1R(h)	14-465KP	125
IKK α (h)	14-461KP	115
IRAK4(h)	14-599KP	97
JAK2(h)	14-640KP	108
KDR(h)	14-630KP	87
LOK(h)	14-686KP	103
Lyn(h)	14-510KP	93
MAPKAP-K2(h)	14-337KP	101
MEK1(h)	14-429KP	107
MLK1(h)	14-690KP	96
Mnk2(h)	14-664KP	111
MSK2(h)	14-616KP	108
MST1(h)	14-624KP	96
mTOR(h)	14-769KP	99
NEK2(h)	14-545KP	105
p70S6K(h)	14-486KP	99
PAK2(h)	14-481KP	97
PDGFR β (h)	14-463KP	80
Pim-1(h)	14-573KP	92
PKA(h)	14-440KP	89
PKB α (h)	14-276KP	104
PKC α (h)	14-484KP	85
PKC θ (h)	14-444KP	109
PKG1 α (h)	14-688KP	99
Plk3(h)	14-572KP	104
PRAK(h)	14-334KP	104
ROCK-I(h)	14-601KP	112
Rse(h)	14-535KP	118
Rsk1(h)	14-509KP	110
SAPK2a(h)	14-251KP	88
SRPK1(h)	14-564KP	105
TAK1(h)	14-600KP	106
PI3 Kinase (p110 β /p85 α)(h)	14-603KP	98
PI3 Kinase (p120 γ)(h)	14-558KP	100
PI3 Kinase (p110 δ /p85 α)(h)	14-604KP	96
PI3 Kinase (p110 α /p85 α)(h)	14-602KP	98

Table S4: ProteinLynx Global SERVER (PLGS) filtering parameters

Parameter	Min. intensity	Min. sequence length	Max. sequence length	Min. products per amino acid	Minimum score	Max. MH+ Error (ppm)
Value	20000	4	26	0.12	7	5

Figure S1: HDX sequence coverage of USP3¹⁻¹³¹

A total of 59 peptides yielded 93.9% sequence coverage and a redundancy of 5.24. Black rectangles aligned to sequence represent peptides. Several peptides including 52-86 (length 35) and 53-86 (length 34) were manually added after PLGS filtering to obtain coverage from 53-60.

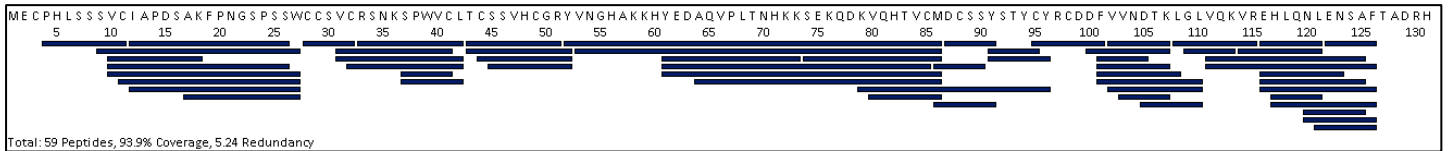


Figure S2: Differential HDX heatmaps

Differential fractional uptake is displayed as a per-residue averaged heatmap aligned to USP3 primary sequence for USP3 Zf-UBD in complex with **59** at 1:10 (top) and 1:20 (bottom) . The highest magnitude D decreases (-5 %) are blue and D increases (+5%) are red. A limitation of this heatmap illustration (DynamX 3.0) is that it does not factor in the associated error for each residue. Please refer to Figure 4 for a peptide-level differential HDX bar-plot with associated error.



Figure S3: Diversity of sidechains lining the binding pocket of 59. Residues lining the pocket are highlighted in blue. Positions where a specific side-chain is unique to USP3 are highlighted with red arrows.

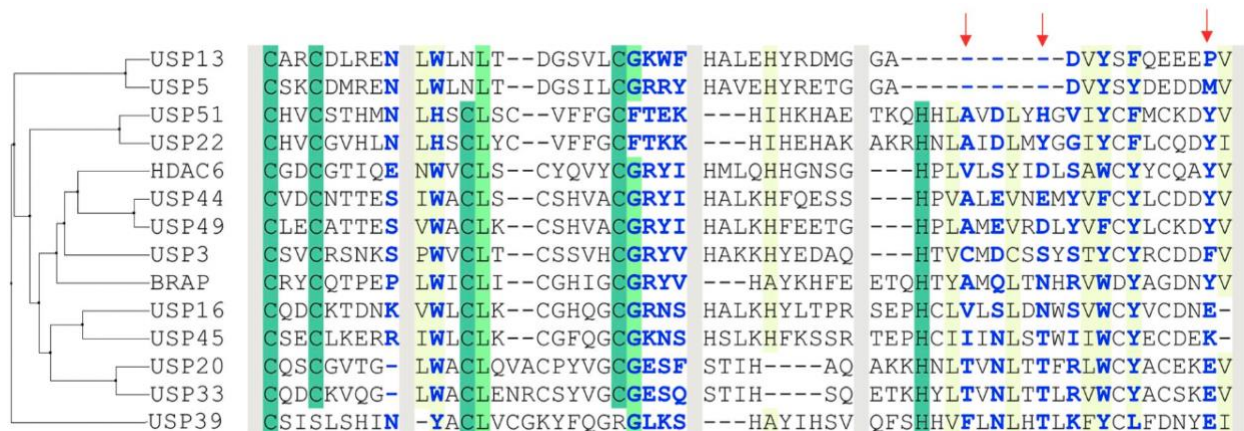


Figure S4: Ligand RMSD over 100 ns MD simulation

