## **Supporting Information**

## Identification of a Dual Autophagy and REV-ERB Inhibitor with in Vivo Anticancer Efficacy

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Compound 3: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>)

















Compound 7: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>CNMR (101 MHz, DMSO-*d*<sub>6</sub>)

Compound 8: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)





Compound 9: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)



Compound 10: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)



Compound **11**: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>)



Compound 12: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)











Compound 15: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)

Compound 16: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)





Compound 17: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)



Compound 18: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>)



Compound 19: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>)



Compound 20: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>)

Compound 21: <sup>1</sup>HNMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>)









Compound 23: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>)



Compound 24: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)



Compound 25: <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) and <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>)









Compound 4: UPLC/MS analysis







































































Compound 24: UPLC/MS analysis

![](_page_46_Figure_1.jpeg)

![](_page_47_Figure_1.jpeg)

![](_page_48_Figure_0.jpeg)

![](_page_48_Figure_1.jpeg)

Compound	Rt (min)	Method	Compound	Rt (min)	Method
3	3.80	E	15	2.79	E
4	4.60	E	16	3.80	E
5	2.63	E	17	2.85	E
6	3.66	E	18	2.72	E
7	3.65	E	19	4.63	E
8	2.87	E	20	2.30	E
9	2.96	E	21	3.31	E
10	2.22	E	22	2.74	E
11	3.31	E	23	4.04	E
12	2.49	E	24	3.17	E
13	3.70	E	25	3.56	E
14	3.48	E	26	4.00	E

Table S1. Retention times and UPLC analytical method of the final compounds.<sup>a</sup>

<sup>a</sup>Freshly prepared 10 mM DMSO-*d*<sub>6</sub> stock solutions (used for biological screenings), diluted 20fold or 100 fold in CH<sub>3</sub>CN/H<sub>2</sub>O (1:1), and directly analyzed. The analysis was performed on an ACQUITY UPLC BEH C18 column (100x2.1mmID, particle size: 1.7µm) with a VanGuard BEH C18 pre-column (5x2.1mmID, particle size: 1.7µm) at 40 °C using 10mM NH<sub>4</sub>OAc in H<sub>2</sub>O at pH 5 adjusted with AcOH (A) and 10mM NH<sub>4</sub>OAc in CH<sub>3</sub>CN-H<sub>2</sub>O (95:5) at pH 5 (B) as mobile phase at 0.5mL/min. *Method E*: gradient 10 to 90% B over 6.0 min. Flow rate 0.5 mL min<sup>-1</sup>. Temperature 40 °C. The detection wavelength ( $\lambda$ ) was set at 215 nm for relative purity determination.

![](_page_50_Figure_0.jpeg)

**Figure S1**. Compound **24** induces apoptosis in BT-474 cells. (A) BT-474 cells were treated with the indicated concentrations of 24 or DMSO (vehicle). After 24 h, caspase activity was evaluated with a fluorescent inhibitor of caspases covalently bound to the poly-caspase-specific amino acid sequence valine-alanine-aspartic acid (VAD) (SR-VAD-FMK). Count of the percentage of caspase-positive cells is given as mean  $\pm$  SEM, n=3. \*P<0.05 and \*\*\*P<0.001, (one-way ANOVA with Dunnett's multiple comparison test). (B) Protein samples from BT-474 cells treated as in A were probed with specific antibodies against cleaved-PARP (cleaved-PARP) and GAPDH proteins. (C) Quantification of immunoblot analysis from protein samples treated as in B. Relative cleaved PARP expression was calculated normalizing the optical density of cleaved-PARP signals with that of GAPDH. Shown as mean  $\pm$  SEM, n=3. \*\*P<0.01 and \*\*\*P<0.01 (one-way ANOVA with Dunnett's multiple comparison test).

![](_page_51_Figure_0.jpeg)

**Figure S2**. REV-ERB $\beta$  silencing abolishes compound **24**-mediated transcriptional response. BT-474 cells were transfected with pooled siRNA sequences against *REV-ERB\beta* (siREV-ERB $\beta$ ) or a non-targeting pool as a control (Control). One day post transfection, cells were treated with DMSO (vehicle) or 5  $\mu$ M of compound **24** for 24 h and the expression of the REV-ERB target gene, *BMAL1*, and *REV-ERB\beta* was determined by quantitative reverse transcriptase-PCR (qRT-PCR) using *GAPDH* for normalization. Shown as mean ± SEM, n = 3. \*P<0.05, **24**-treated versus vehicle treated control cells; \*\*\*P<0.001, **24**- and vehicle-treated siREV-ERB $\beta$  versus **24**- and vehicletreated control cells (two-way ANOVA with Bonferroni's posttest analysis).

![](_page_52_Figure_0.jpeg)

**Figure S3**. Tolerability of **24** in CD1 mice. (A) Female CD1 mice were treated with i.p. injections of **24** administered according to the schematized schedule treatments. (B) Animal body weight was monitored daily over a 7-day period. Weight of mice the day before staring the treatment was set as 100% and used for calculating the relative body weight (%) and is shown as mean  $\pm$  SEM, n = 3 mice per each treatment group.