Supporting Information for

Multi-Dimensional Widefield Infrared-encoded Spontaneous Emission Microscopy: Distinguishing Chromophores by Ultrashort Infrared Pulses

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I. Materials and staining methods.

Materials. R6G (catalog no. 83697), FITC (catalog no. F7250), fluorescein (catalog no. 46955), PI dyes (catalog no. P4170), and mesoporous silica microspheres (catalog no. 806587 for 2 microns, 806765 for 3 microns) were purchased from Sigma-Aldrich. Qdot™ 585 Streptavidin Conjugate (Invitrogen, catalog no. Q10111MP), EZ-Link™ Sulfo-NHS-SS-Biotin (catalog no. A39258) were purchased from Thermo Fisher. Ammine-coated CdSe/ZnS QD aqueous solutions were purchased from NN-Labs (catalog no. HECZWA560). All chemicals and materials were used without further purification.

Staining of silica microbeads. For the staining of silica microbeads with molecular dyes, dyes were diluted into ~1 mg/mL solutions (chloroform for R6G, ethanol for fluorescein and FITC), then the silica microbeads were soaked in the solutions and filtered. For fluorescein and FITC, 2 equivalent of NaOH (dissolved in ethanol) were added to form the bright dianions (37) before adding the silica beads. For the staining using ammine-coated CdSe/ZnS QDs, QDs were diluted into 0.1 mg/mL aqueous solutions, then the silica microbeads were soaked in the solutions, then the water was removed by a rotary evaporator.

Staining of fixed cells. i. QD staining. MDA-MB-231 human breast cancer cells (HTB-26) were obtained from American Type Culture Collection (ATCC; Manassas, VA) and cultured in Dulbecco's Modified Eagle's medium (Gibco; Waltham, MA) supplemented with 10% fetal bovine serum (Sigma-Aldrich; St. Louis, MO) at 37° C in a humidified incubator with 5% CO₂. The cells growing on coverslips at $~60\%$ confluence were washed three times with phosphatebuffer saline (PBS) solutions at $pH = 8.0$. Then 250 µL PBS containing 0.5 mg/mL EZ-LinkTM Sulfo-NHS-SS-Biotin was added. The cells were incubated at room temperature for 30 minutes, and then washed three times with ice-cold PBS. Cells were then fixed in 100% cold methanol for 10 min at -20°C, and then washed three times with PBS (5 min/wash). For the subsequent binding with biotin, we added 250 µL PBS containing 40 nM Qdot™ 585 Streptavidin Conjugate and incubated the cells for 1 hour at room temperature. The cells were then washed three times with PBS and then rinsed with water. **ii. PI staining.** The cells growing on coverslips at \sim 60% confluence were washed three times with PBS solutions at $pH = 8.0$. Cells were then fixed in 100% cold methanol for 10 min at -20° C, washed three times with PBS (5 min/wash), and then rinsed with $2\times$ saline sodium citrate (SSC, containing 0.3 M NaCl, 0.03 M sodium citrate, pH 7.0) solution. Then, $250 \mu L$ of $500 \mu M$ PI dye aqueous solution was added to the cells for an incubation of 15 minutes. Cells were rinsed three times with 2X SSC and then with water. **iii. QD-PI co-staining.** MDA-MB-231 cells growing on coverslips were labeled with EZ-Link™ Sulfo-NHS-SS-Biotin and fixed in 100% cold methanol as described in the section i. QD staining above. The fixed cells were then washed three times with PBS, rinsed with $2 \times$ SSC, and incubated with 250 µL PI dye solution as described in the above section ii PI staining. The concentration of PI solution used here was 75 μ M instead of 500 μ M. After washing with 2× SSC and then with PBS, the cells were incubated with 250 μ L PBS containing 40 nM QdotTM 585 Streptavidin Conjugate for 1 hour at room temperature. The co-stained cells were then washed three times with PBS and then rinsed with water prior to imaging.

II. Additional data on rhodamine 6G (R6G).

(a). Solution phase measurements.

 Fig. S1 shows the infrared-pump-visible-probe transient absorption (TA) results of R6G dissolved in chloroform and in dimethyl sulfoxide-d₆ (DMSO-d₆) solutions. The solutions are sandwiched between two infrared-transparent $CaF₂$ windows with 56 microns in thickness and optical density of ~0.2 at 540 nm. Clear features of excited-state absorption and ground-state bleaching are observed. The ultrafast kinetics of the visible-region absorbance change induced by the 1600 cm⁻¹ infrared (IR) pump are sensitive to the solvent environment. In deuterated DMSO, the kinetics only exhibit a decaying pattern, whereas the decay kinetics in chloroform exhibit a more complex pattern. The difference in decay patterns might be attributed to change of Franck-Condon factors and intramolecular vibrational energy redistribution (IVR) of R6G in different solvents.

Fig. S1. The TA results of R6G in DMSO- $d_6(A)$ and chloroform (B). The IR pump frequency is 1600 ± 30 cm⁻¹, and the probe is a broadband whitelight pulse. The upper panels show TA spectra at various time delays, and the lower panels show kinetics at several wavelengths that correspond the wavelength positions marked by dashed lines in upper panels.

The R6G solution in chloroform is investigated by measuring its fluorescence intensity following the excitation of a 1600 cm⁻¹ IR pulse and then a narrowband visible pulse (520 or 550 nm). The instrument and solution used are the same as the TA experiments above, but the broadband whitelight probe pulse in TA experiments is replaced with a narrowband visible pulse, and the detector receives fluorescence signals rather than the whitelight probe beam. The relative change of fluorescence intensity induced by the IR pulse is measured as $100\% \times$ $(I_{on} - I_{off})/I_{off}$, where I_{on} or I_{off} are the intensity when the IR beam is unblocked or blocked by a chopper. As shown in Fig. S2, the kinetics of relative fluorescence intensity change show similar patterns to the TA kinetics in Fig. S1B. The IR-induced fluorescence intensity change is attributed to the IR-induced change of electronic absorbance in the visible region. As discussed in the main text, at 550 nm, more absorption of visible photons leads to more fluorescence photons. In contrast, at 520 nm, ground state bleaching leads to reduction of fluorescence signals. The results support the mechanism 1 of MD-WISE imaging of silica beads in the main text, but the change level of fluorescence intensity is small (only a few percentages). This is attributed to the relatively large thickness of the solutions (56 microns) generating defocused background fluorescence signals that is not modulated by the IR pulse.

Fig. S2. The transient change of fluorescence intensity of R6G in chloroform. Following the excitation of an IR pulse and then a narrow band visible pulse, the fluorescence signals emitted are collected in the wavelength range of 585 \pm 18 nm using a bandpass filter. The IR pump frequency is 1600 \pm 30 cm⁻¹, and the visible excitation range is 550 ± 5 nm (A) or 520 ± 5 nm (B).

(b). MD-WISE images of stained silica beads.

In Fig. S3A, MD-WISE images that correspond to the 1600 cm-1 kinetic curve in main text Fig. 2E are displayed at a series of delay times. As described in the main text, the kinetic curves are measured by averaging the relative intensity change among all the detector pixels in a 2.5 µm by 2.5 µm box that centers around the microbead. At later delay times, vibrational relaxation and energy redistribution reduces the counts per pixel and the quality of the difference images. The highest quality of difference images is obtained using short delays such as 1 ps. In Fig. S3B, MD-WISE images that correspond to the 1720 cm⁻¹ kinetic curve in main text Fig. 2E are

displayed at a series of delay times. Besides the IR frequencies listed in the main text, we also performed MD-WISE imaging using 1650 ± 30 cm⁻¹ IR excitation. The IR absorption peak at 1650 cm-1 is assigned to another stretch mode of the xanthene ring in R6G. As shown in Fig. S4, the ultrafast decay kinetics are faster than the kinetics measured at 1600 cm^{-1} and 1720 cm^{-1} .

Fig. S3. (A) MD-WISE images obtained using 1600 ± 30 cm⁻¹ IR pulse and 550 ± 5 nm visible pulse, and **(B)** MD-WISE images obtained using 1720 ± 30 cm⁻¹ IR pulse and 550 ± 5 nm visible pulse. The fluorescence signals are collected in the range of 585 ± 18 nm. Each series in (A) or (B) includes an IR-off image and difference images $(IR_{on} - IR_{off})$ at different delay times. Scale bars are 1 micron in size. The color bars represent CCD counts per pixel.

Fig. S4. (A) MD-WISE images obtained using 1650 ± 30 cm⁻¹ IR pulse and 550 ± 5 nm visible pulse. The fluorescence signals are collected in the range of 585 ± 18 nm. The series include an IR-off image and difference images ($IR_{on} - IR_{off}$) at different delay times. Scale bars are 1 micron in size. The color bars represent CCD counts per pixel. **(B)** Ultrafast kinetics of fluorescence intensity change at IR excitation of 1650 cm⁻¹. The relative change of counts per pixel is measured using the pixels within the 2.5-micron square box marked by dashed lines in (A).

(c). Kinetics of IR-induced change of fluorescence intensity measured using other substrate materials: azide-functionalized silica and polymethyl methacrylate (PMMA).

Since the R6G dye molecules have overlapping IR spectral features with the silica beads at ~1600 cm⁻¹, we performed MD-WISE experiments on an additional set of substrate materials to investigate whether the IR absorption and the properties of substrate materials have large impacts on the mechanism of ultrafast IR-induced emission intensity change.

One substrate tested is the surface-modified silica microbeads. The surface of silica microbeads contains silanol groups which can be used to anchor organic molecules with distinct vibrational features. Using an established protocol, 3-(azidopropyl)triethoxysilane (Gelest Inc., catalog no. SIA0777.0, structure embedded in Fig. S5A) is grafted onto the surface of silica microbeads (1). In Fig. S5A, the azide-modified silica microbeads (Silica-N₃) show a distinct azide stretch mode at \sim 2100 cm⁻¹ in addition to the absorption features of silica at 1600 cm⁻¹ and 1800-2000 cm-1 .

Another substrate tested is PMMA microbeads (EpruiBiotech, Shanghai, catalog no. 3-001-3). As shown in Fig. S5B, PMMA lacks vibrational features at ~1600 cm⁻¹, and thus has no overlap with the ring stretch mode of R6G at 1600 cm⁻¹.

Fig. S5. Diffuse reflectance infrared Fourier transform spectroscopy (DRIFTS) results of azide-modified silica microbeads **(A)** and PMMA microbeads **(B)**. The azide stretch mode is marked by the ▲ symbol in panel A.

The ultrafast kinetics of IR-induced fluorescence intensity change of R6G on three types of substrates are shown in Fig. S6A. The IR frequency is tuned to 1600 ± 30 cm⁻¹ for the xanthene ring stretch mode. The results show that, no matter the substrate IR absorption feature overlaps with the R6G feature or not, the kinetics are nearly identical for the three types of substrates. Thus, the IR-induced fluorescence intensity change measured for the silica beads shall be attributed to the excitation of molecular vibrational modes of the adsorbed dyes, rather than the IR absorption of silica.

Furthermore, we test whether exciting the azide stretch mode of Silica-N₃ can transfer energy to R6G modes and cause modulation on fluorescence intensity. As shown in Fig. S6B, there is no modulation on fluorescence intensity using IR frequency at 2100 cm⁻¹. This result agrees well with the flat kinetic traces in main text Fig. 2E using IR frequencies in the range of 1800-2000 $\text{cm}^{\text{-}1}$.

Fig. S6. (A) Normalized ultrafast kinetics of fluorescence intensity change measured with IR excitation of 1600 ± 30 cm^{-1} and visible excitation of 550 \pm 5 nm. The results show the three types of substrates stained with R6G have nearly identical kinetics. **(B)** Ultrafast kinetics of fluorescence intensity change of R6G in azide-modified silica microbeads measured with IR excitation of 2100 ± 30 cm⁻¹ and visible excitation of 550 ± 5 nm. No IR-induced emission intensity change is observed. The fluorescence signals are collected in the range of 585 ± 18 nm.

III. Additional MD-WISE data on other stained silica beads.

(a). Quantum dots (QDs)

In Fig. S7, MD-WISE images of QD-stained silica beads that correspond to the kinetic curve in main text Fig. 3D are displayed at a series of delay times. For QDs, the IR pulse only has effect on the photoluminescence of QDs when it arrives later than the visible pulse (negative delay times).

Fig. S7. MD-WISE images of QD stained beads obtained using 2100 ± 30 cm⁻¹ IR pulse and 550 ± 5 nm visible pulse. The photoluminescence signals are collected in the range of 585 ± 18 nm. The series include an IR-off image and difference images $(\mathbb{R}_{on} - \mathbb{R}_{off})$ at different delay times. Scale bars are 1 micron in size. The color bars represent CCD counts per pixel.

(b). Fluorescein

In Fig. S8, MD-WISE images of fluorescein-stained silica beads that correspond to the kinetic curve in main text Fig. 4D are displayed at a series of delay times. Only IR frequency of 1600 cm-1 that resonantly excites the molecule show effects on fluorescence emission intensity, while the difference images acquired using IR frequency of 2040 cm⁻¹ are blank. For 1600 cm⁻¹, similar to the case of R6G, at later delay times, vibrational relaxation and energy redistribution reduces the counts per pixel and the quality of the difference images.

Fig. S8. (A) MD-WISE images of fluorescein-stained beads obtained using 1600 ± 30 cm⁻¹ IR pulse and 520 ± 5 nm visible pulse, and **(B)** MD-WISE images of fluorescein-stained beads obtained using 2040 ± 30 cm⁻¹ IR pulse and

520 \pm 5 nm visible pulse. The fluorescence signals are collected in the range of 585 \pm 18 nm. Each series in (A) or (B) include an IR-off image and difference images $(IR_{on} - IR_{off})$ at different delay times. Scale bars are 1 micron in size. The color bars represent CCD counts per pixel.

(c). Fluorescein-5-isothiocyanate (FITC)

Fig. S9, MD-WISE images of FITC-stained silica beads that correspond to the kinetic curve in main text Fig. 4D are displayed at a series of delay times. Both the IR frequencies of 1600 cm-1 and 2040 cm-1 resonantly excite the molecule and show effects on fluorescence emission intensity. The lifetime of IR-modulation on the fluorescence of FITC is only a few ps.

Fig. S9. (A) MD-WISE images of FITC-stained beads obtained using 1600 ± 30 cm⁻¹ IR pulse and 520 ± 5 nm visible pulse, and **(B)** MD-WISE images of FITC-stained beads obtained using 2040 ± 30 cm⁻¹ IR pulse and 520 ± 5 nm visible pulse. The fluorescence signals are within the range of 585 ± 18 nm. Each series in (A) or (B) include an IR-off image and difference images $(IR_{on} - IR_{off})$ at different delay times. Scale bars are 1 micron in size. The color bars represent CCD counts per pixel.

IV. Additional MD-WISE data on stained cells.

(a). Cells stained with propidium iodide (PI)

In Fig. S10, three replica sets of MD-WISE difference images of PI-stained cancer cells are displayed at different delay times. Since the IR frequency of 2100 cm⁻¹ does not excite the vibrational modes of PI dyes, no IR-induced effects are observed. The IR-off images show the stained nucleic acids of the cells, while the difference images at positive or negative delays on show blank.

PI 2100 cm $^{-1}$ / 550 nm

Fig. S10. Three sets of MD-WISE images of PI-stained cells obtained using 2100 ± 30 cm⁻¹ IR pulse and 550 ± 5 nm visible pulse. The fluorescence signals are collected in the range of 585 ± 18 nm. The series include IR-off images (left column) and associated difference images (IR_{on} – IR_{off}) at -10 ps (middle column) and 1 ps (right column). Scale bars are 5 microns in size. The color bars represent CCD counts per pixel.

(b). Cells stained with streptavidin-coated QD585

 In Fig. S11, three replica sets of MD-WISE difference images of QD-stained cancer cells are displayed at different delay times. The IR-off images show the stained cell membranes. The difference images at the negative delay of -10 ps reproduce the outline of cell membranes. The

difference images at the positive delay of 1 ps show blank, since the IR pulse can only modulate the photoluminescence intensity of QDs when it arrives later than visible pulse (Fig. S12).

Fig. S11. Three sets of MD-WISE images of QD-stained cells obtained using 2100 ± 30 cm⁻¹ IR pulse and 550 ± 5 nm visible pulse. The photoluminescence signals are collected in the range of 585 ± 18 nm. The series include IRoff images (left column) and associated difference images ($IR_{on} - IR_{off}$) at -10 ps (middle column) and 1 ps (right column). Scale bars are 5 microns in size. The color bars represent CCD counts per pixel.

Fig. S12. Ultrafast kinetics of IR-induced photoluminescence intensity change of QD-stained cells obtained using 2100 ± 30 cm⁻¹ IR pulse and 550 ± 5 nm visible pulse. The photoluminescence signals are collected in the range of 585 ± 18 nm.

V. Quantum chemical vibrational analysis of R6G dye

(a). Overview and methods

Gaussian 09, Revision D (2) was used to perform quantum chemical calculations for identifying and analyzing the vibrational modes, vibrational frequencies, and coupling between functional groups in the molecule. The density functional theory (3) calculations were carried out using RB3LYP/6-311++G(d,p) basis sets (4). The structure of R6G molecular cation was optimized in vacuo using a crystal structure as the initial guess (5, 6). Then vibrational frequency analysis was performed. The optimized results of R6G cation in vacuum was then used as the initial structure for the vibrational frequency analysis of R6G cations with polarizable continuum models in three different implicit solvent environments: water, methanol, and acetone. The optimized structure of R6G cation in vacuum and the numbering of atoms are displayed as an example in Fig. S13. The atomic coordinates of R6G cations in different environments are listed in Table S1-S2. The frequency and intensity of all the modes above 1000 cm-1 are listed in Table S3.

Fig. S13. (A) Optimized structure of R6G cation in vacuum, viewed from two different angles. **(B)** Numbering of atoms in the R6G cation.

Table S1. Atomic coordinates of R6G cation in vacuum and water.

Table S2. Atomic coordinates of R6G cation in methanol and acetone.

Table S3. Calculated vibrational frequencies and infrared absorption intensities of R6G vibrational modes. Vibrational modes related to MD-WISE experiments are highlighted by yellow.

(b). Calculated IR absorption spectra

The calculated vibrational IR spectra of R6G cations in different environments are shown in Fig. S14. The spectra are similar for R6G cations in water, methanol and acetone, which are redshifted from the spectrum in vacuum. The calculated spectra largely match the FTIR pattern of R6G shown in main text Fig. 2A, though the exact center positions of the peaks differ slightly from the experimental results. Three calculated xanthene ring modes (number 149, 150, 153 in Table S3) related to MD-WISE imaging are marked by the hexagon signs in Fig. S14. The stretch mode (number 155 in Table S3) of the ester group is marked by the triangle signs. Each of these four modes is analyzed with details below using water solvent as the example, since different solvents only yield negligible results. Vibrational modes with negligible IR absorption intensities or modes outside of the frequency ranges used in MD-WISE imaging are not subject to further analysis here.

Fig. S14. IR spectra of R6G cations in various environments. **(A)** full IR spectrum in vacuum, **(B-E)** IR spectra in the range between 1475-1875 cm⁻¹ for vacuum (B), water (C), methanol (D), and acetone (E). \triangle marks the peak of ester and \odot marks the peaks of xanthene rings, which largely match the peaks observed in FTIR experiments.

(c). Analysis of mode 149 and 150 (xanthene ring modes)

The calculated vibrational modes 149-150 in water at \sim 1600 cm⁻¹ largely only involve the xanthene ring atoms, with minor coupling to the displacements of atoms in the phenyl ring attached to the xanthene ring. The atoms involved and the displacements of each atom are shown in Fig. S15 and Table S4-S5.

Fig. S15. Atoms involved in mode 149 **(A)** and mode 150 **(B)**.

Table S4. Atomic displacements of each atom involved in mode 149 in the unit of angstroms. The XYZ vectors are displayed in Fig. S13.

Table S5. Atomic displacements of each atom involved in mode 150 in the unit of angstroms. The XYZ vectors are displayed in Fig. S13.

(d). Analysis of mode 153 (xanthene ring mode)

The calculated vibrational mode 153 in water at \sim 1650 cm⁻¹ is local to the xanthene ring. The atoms involved and the displacements of each atom are shown in Fig. S16 and Table S6.

Fig. S16. Atoms involved in mode 153.

mode 153				
atom number	element	$\overline{\mathbf{X}}$	$\overline{\mathbf{Y}}$	\mathbf{Z}
14		-0.07	0.00	$0.00\,$
18	$\rm _C^C$	-0.04	-0.01	$0.00\,$
19		-0.04	0.01	$0.00\,$
23		0.25	-0.07	0.03
24	C C C C C C	0.10	-0.05	$0.02\,$
25		0.10	0.05	-0.02
$26\,$		0.25	0.07	-0.03
28		-0.25	0.08	-0.03
29	\overline{H}	-0.34	-0.05	$0.02\,$
$30\,$	$\mathbf O$	-0.01	0.00	$0.00\,$
31	$\mathbf C$	-0.17	0.02	-0.01
32	$\mathbf C$	-0.17	-0.02	$0.01\,$
33	$\overline{\mathrm{H}}$	-0.34	0.05	-0.02
34	$\mathbf C$	-0.25	-0.08	0.03
35	$\mathbf C$	0.03	0.01	$0.00\,$
36	$\mathbf C$	0.13	0.03	-0.02
37	$\, {\rm H}$	0.21	0.00	0.00
38	$\mathbf H$	0.21	0.00	0.00
39	$\mathbf C$	0.13	-0.03	0.02
40	$\mathbf C$	0.03	-0.01	$0.00\,$
41	$\, {\rm H}$	0.13	-0.07	-0.06
42	$\mathbf H$	0.13	-0.02	0.10
43	$\, {\rm H}$	-0.15	-0.01	0.01
44	${\bf N}$	0.00	-0.04	$0.02\,$
45	$\overline{\rm N}$	$0.00\,$	0.04	-0.02
46	$\rm H$	0.13	0.07	$0.06\,$
47	$\, {\rm H}$	-0.15	0.01	-0.01
48	$\, {\rm H}$	0.13	0.02	-0.10
49	$\mathbf C$	0.00	0.02	-0.01
50	$\mathbf C$	0.00	-0.02	$0.01\,$
51	$\mathbf H$	0.03	-0.06	$0.01\,$
52	$\, {\rm H}$	-0.03	-0.04	$0.02\,$
54	$\rm H$	-0.03	0.04	-0.02
55	$\, {\rm H}$	0.03	0.06	-0.01
57	$\, {\rm H}$	0.00	-0.01	$0.01\,$
58	$\rm H$	-0.01	-0.02	-0.04
59	$\rm H$	$0.01\,$	0.00	0.01
60	$\rm H$	-0.01	0.02	0.04
61	$\, {\rm H}$	0.00	$0.01\,$	-0.01
62	$\, {\rm H}$	$0.01\,$	0.00	-0.01
63	$\, {\rm H}$	-0.12	0.16	-0.07
64	$\, {\rm H}$	-0.12	-0.16	0.07

Table S6. Atomic displacements of each atom involved in mode 153 in the unit of angstroms. The XYZ vectors are displayed in Fig. S13.

(e). Analysis of mode 155 (ester group mode)

The calculated vibrational mode 155 in water at \sim 1720 cm⁻¹ is largely a local mode of the ester group. However, calculation results also reveal coupling to the atoms in the xanthene ring. This coupling could be the origin of IR-induced fluorescence intensity change of R6G when the IR laser frequency is tuned to 1720 cm-1 as discussed in the main text. The atoms involved and the displacements of each atom are shown in Fig. S17 and Table S7.

Fig. S17. Atoms involved in mode 155.

Table S7. Atomic displacements of each atom involved in mode 155 in the unit of angstroms. The XYZ vectors are displayed in Fig. S13.

VI. Determination of scale bars, field of view, and spatial resolution of the MD-WISE images

(a). Determination of scale bars

The 3-micron silica spheres stained with fluorescent dyes (806765, Sigma Aldrich, see Section I. Materials and staining methods) were used as the calibration target for the MD-WISE microscope. The scale bars in all the MD-WISE fluorescent images were calculated based on the diameter of the images of the silica beads acquired under the same imaging conditions. The absolute size of the silica beads was in turn calibrated with a standard reticle (Thorlabs, R1L3S6PR) using an optical microscope equipped with a x100 objective under whitelight transillumination configuration. As shown below in Fig. S18, the diameter of the silica beads is 3 microns.

Fig. S18. The reticle has 250 line pairs (one light line and one dark line) per millimeter. Each light or dark line has a width of 2 microns. We can determine the diameter of the silica beads is 3 microns.

(b). An example of the full field of view.

 For MD-WISE imaging experiments, the field of view is adjustable depending on the size of the objects that need to be imaged. For individual silica beads, we typically chose the field of view of 10~20 microns by setting the beam size of the IR and visible pulses to a value in this range. Such beam size is more than sufficient to image the 3-micron beads. To capture the fluorescence image of the entire field, we imaged a sample with crowded beads. As shown below in Fig. S19, the image of beads reports a field of view of \sim 15 microns. The variations of image intensity could be due to the inhomogeneous loading of dyes in the beads. For images of single cells, they were acquired with larger field of view, and the large size of the cells themselves (20- 30 microns) nearly filled the entire field of view.

Fig. S19. Fluorescence image of 3-micron silica beads showing an entire field of view of ~15 microns.

(c). Determination of the spatial resolution.

 First, we determined whether MD-WISE has a resolution as good as regular photoluminescence (PL) images in our setup. As shown by the linecuts of MD-WISE and PL images in Fig. S20, they lay on top of each other, with identical widths for both the 2-micron (left) and 3-micron (right) silica microspheres, respectively. This suggested that MD-WISE has as good resolution as regular PL imaging using either quantum dots or R6G dye as the chromophore. This makes sense, because the resolution of MD-WISE imaging is defined by the optical objective we used to resolve PL or fluorescence signals.

Fig. S20. Linecuts of MD-WISE (blue) and plain PL images (orange) acquired without IR modulation. The linecuts were performed across the images of individual silica spheres stained with either quantum dots or R6G dyes as shown in the main texts.

 We then determined the resolution of our setup by analyzing the point spread function in regular PL images of individual perovskite nanocrystals (size 50 nm, composition MAPbBr3) prepared according a literature method ⁷. The nanocrystal serves as a bright PL point source of which the physical size is much smaller than the resolution limit. The FWHM of the gaussian fitting of the point spread function as shown in Fig. S21 yields a resolution of 1.6 microns.

Fig. S21. Gaussian fitting of the point spread function measured using a bright nanocrystal. The fitting equation and results are embedded in the upright corner. The FWHM of the gaussian is $2.355 * (c1/1.414) = 1.6$ microns.

SI References

- 1. B. Wu, J. P. Breen, X. Xing, M. D. Fayer, Controlling the Dynamics of Ionic Liquid Thin Films via Multilayer Surface Functionalization. *J Am Chem Soc* **142**, 9482–9492 (2020).
- 2. Gaussian 09, Revision D.01, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman, and D. J. Fox, Gaussian, Inc., Wallingford CT, 2016.
- 3. W. J. Hehre, R. Ditchfield, J. A. Pople, Self-Consistent Molecular Orbital Methods. XII. Further Extensions of Gaussian—Type Basis Sets for Use in Molecular Orbital Studies of Organic Molecules. *J Chem Phys* **56**, 2257–2261 (1972).
- 4. C. Lee, W. Yang, R. G. Parr, Development of the Colle-Salvetti correlation-energy formula into a functional of the electron density. *Phys Rev B* **37**, 785–789 (1988).
- 5. D. N. Adhikesavalu, D. Mastropaolo, A. Camerman, N. Camerman, Two rhodamine derivatives: 9-[2-(ethoxycarbonyl)phenyl]-3,6-bis(ethylamino)-2,7-dimethylxanthylium chloride monohydrate and 3,6-diamino-9-[2-(methoxycarbonyl)phenyl]xanthylium chloride trihydrate. *Acta Crystallographica C* **57**, 657–659 (2001).
- 6. H. Watanabe, N. Hayazawa, Y. Inouye, S. Kawata, DFT Vibrational Calculations of Rhodamine 6G Adsorbed on Silver: Analysis of Tip-Enhanced Raman Spectroscopy. *J Phys Chem B* **109**, 5012–5020 (2005).
- 7. T. Tachikawa, I. Karimata, Y. Kobori, Surface Charge Trapping in Organolead Halide Perovskites Explored by Single-Particle Photoluminescence Imaging. *J Phys Chem Lett* **6**, 3195-3201 (2015).