

**Supplementary Figure 1.** Linear regression between JFH1cc (X axis) and JFH1pp (Y axis) IC<sub>50</sub> values for eight mAbs. The correlation coefficient (Pearson r) between JH1cc and JFH1pp IC<sub>50</sub> values is shown. IC<sub>50</sub> values were obtained using two independent experiments.



**Supplementary Figure 2.** Correlation matrix based on neutralization sensitivity (A) Heatmap showing pairwise Spearman's correlation matrix for eight mAbs and CD81LEL [log<sub>10</sub>GMT IC<sub>50</sub> values ( $\mu$ g/ml)] using the full panel of 20 HCVpp (minimum of two IC<sub>50</sub> values per HCVpp-mAb combination). Spearman's correlations and p-values are in red and blue, respectively. (B) Hierarchical clustering of mAbs based on log10 GMT IC<sub>50</sub> values ( $\mu$ g/ml) is shown. Bootstrap resampling (1000 iterations) was applied, nodes with support above 10% are shown. Horizontal line represents the scale for the tree branches, which reflects the distance or dissimilarity between data points in  $\mu$ g/ml.



**Supplementary Figure 3.** HCVpp sensitivity to CD81 LEL. GMT and standard deviation re shown as red lines. HCVpps with  $IC_{50}$  values above  $80\mu g/ml$  (dotted line) or not blocked at the highest concentration tested (100  $\mu g/ml$ ) are highlighted in color.



**Supplementary Figure 4.** CD81 LEL correlations. Spearman r correlations between CD81 LEL (X axis) and GMT of all mAbs (A) in red or individual mAbs (B to I) in black. For A to I, 20 HCVpps are plotted and a horizonal dotted line is depicted at 1  $\mu$ g/ml.



**Supplementary Figure 5.** Evolutionary relationships of E1E2s from 20 HCVpps inferred using the Neighbor-Joining method. The optimal tree is shown. The percentage of bootstrap test (1000 replicates) are shown next to the nodes. The tree is drawn to scale, with branch lengths in the same units (number of amino acid substitutions per site) as those of the evolutionary distances used to infer the phylogenetic tree. Colored by genotype from 1 to 6. Evolutionary analyses were conducted in MEGA6.



**Supplementary Figure 6.** Heat map showing pairwise Spearman's correlation matrix for 20 HCVpps based on IC<sub>50</sub> values of eight mAbs. HCVpps are arranged based on hierarchical clustering of the HCVpps in Figure 1B. Two smaller clusters in cluster 1 and 2 are indicated. Spearman's correlations and p-values are in red and blue, respectively.



**Supplementary Figure 7**. Binding results of mAbs to the Luminex beads coupled with mbE1E2s and control proteins. (A) VRC01 (HIV), (B) COVA-18 (Sars-CoV-2) and (C) CR8020 (Influenza), (D) IGH505, (E) HC84.26, (F) AR4A, (G) AP33 mAbs binding to mbE1E2s (H77 and UKNP2.4.1), RSV-F and Non-transfected lysate (NTL) as negative and background beads, respectively. A horizontal dotted lined is shown at 100 MFI.



**Supplementary Figure 8.** Hierarchical clustering of mAbs based on binding (log<sub>10</sub> MFI) is shown. Bootstrap resampling (1000 iterations) was applied, nodes with support above 50% are shown. Horizontal line represents the scale for the tree branches, which reflects the distance or dissimilarity between data points in median fluorescence intensity (MFI) units.



**Supplementary Figure 9.** Binding (MFI) of mbE1E2s and NTLs per mAb (1µg/mI). GMT and SD of 11 mbE1E2s (stars) and NTL (dots) are shown in red. Gray area is set to 10X the highest GMT for all NTLs (85 MFI) as our binding threshold.



Analyte (1µg.ml⁻¹)

**Supplementary Figure 10.** Cross competitive residual binding. Luminex matrices for the mean and 11 mbE1E2s are shown. Competitors, unlabeled mAbs ( $10\mu g/ml$ ), are on the bottom left and analytes, biotinylated mAbs ( $1\mu g/ml$ ), on the top right. Biotinylated mAbs with MFI below our binding threshold are shadowed. Labels indicate the percentage of residual binding relative to the control (no competitor median fluorescence intensity (MFI) equal to 100%) in a color bar from pink to green, where pink indicates strong competition, white no competition and green, binding enhancement. Grey were below binding threshold.

	390	400	) 410	420	430	) 440	) 450	) 460
H77 AMS0230	VDAETHVTGG	SAGRTTAGLV	GLLTPGAKQN	 IQLINTN <mark>G</mark> SW	HINSTALNCN	ESLNTGWLAG	LFYQHKFNSS	GCPERLASCR
AMS0231	S	A.A.NAR	FSQ	V		AD	YNR.D	
	470	480	90 490	500	510	520	) 530	) 540
H77	 RLTDFAQGWG	···· ····  PISYANGSGL	DERPYCWHYP	PRPCGIVPAK	SVCGPVYCFT	PSPVVVGTTD	 RSGAPTYSWG	ANDTDVFVLN
AMS0230	PD	P	.Q	.к.			• • • • • • • • • • • •	••••••
	550	) 560	) 570	580	590	) 600	) 610	) 620
H77 AMS0230	NTRPPLGNWF	GCTWMNSTGF	 TKVCGAPPCV	 IGGVGNNTLL	CPTDCFRKHP	EATYSRCGSG	 PWITPRCMVD	 YPYRLWHYPC
AMS0231		•••••		Н	•••••	•••••	L.	
	630	) 640	) 650	660	670	) 680	) 690	700
H77 AMS0230	 TINYTIFKVR	 MYVGGVEHRL	 EAACNWTRGE	 RCDLEDRDRS	ELSPLLLSTT	 QWQVLPCSFT	 TLPALSTGLI	 HLHQNIVDVQ
AMS0231				N	•••••			•••••
	710	) 72(	) 730	740	1			
Н77	 YLYGVGSSIA	 SWAIKWEYVV	 LLFLLLA <mark>D</mark> AR	 V <mark>C</mark> SCLWMMLL	···· · ISQA <b>E</b> A			
AMS0230 AMS0231	V	•••••	•••••	••••	•••••			

**Supplementary Figure 11.** Alignment of HCV E2 protein for AMS0230 and AMS0231 to the reference strain H77. Similar amino acids compared to H77 are indicated with a dot.