## **Supplemental Information**

## Phenotypic Signature of Circulating Neoantigen-Reactive CD8<sup>+</sup> T cells From Metastatic Human Cancers

Rami Yossef<sup>1</sup>†\*, Sri Krishna<sup>1</sup>†\*, Sivasish Sindiri<sup>1</sup>, Frank J. Lowery<sup>1</sup>, Amy R. Copeland<sup>1</sup>, Jared J. Gartner<sup>1</sup>, Maria R. Parkhurst<sup>1</sup>, Neilesh B. Parikh<sup>1</sup>, Kyle J. Hitscherich<sup>1</sup>, Shoshana T. Levi<sup>1</sup>, Praveen D. Chatani<sup>1</sup>, Nikolaos Zacharakis<sup>1</sup>, Noam Levin<sup>1</sup>, Nolan R. Vale<sup>1</sup>, Shirley K. Nah<sup>1</sup>, Aaron Dinerman<sup>1</sup>, Victoria K. Hill<sup>1</sup>, Satyajit Ray<sup>1</sup>, Alakesh Bera<sup>1</sup>, Lior Levy<sup>1</sup>, Li Jia<sup>2</sup>, Michael C. Kelly<sup>3</sup>, Stephanie L. Goff<sup>1</sup>, Paul F. Robbins<sup>1</sup>, Steven A. Rosenberg<sup>1</sup>\*



Supplemental Figure S1: Neoantigen-specific TIL reactivity screen against patient's 4246 tumoral mutations, and phenotype, distribution, and specificity of neoantigen-specific TCRs in PBL of Pt.4246, related to Figure 1.

**(A)** CD137 upregulation on *in vitro* expanded TIL, grown from tumor fragments, following cocultured with autologous DCs electroporated with TMGs encoding for patient's tumoral mutations. Inset: an example of interferon-gamma (IFNγ) secretion from TIL fragment 10 (F10) following the co-culture. **(B)** Upregulation of CD137 on fragment 10 TILs following co-culture with autologous APCs pulsed with mutated peptides encoded by TMG3. **(C)** Gene expression heatmap showing top 25 upregulated and 10 downregulated genes in neoantigen-reactive cells, and the last 10 rows represent genes-of-interest. **(D)** Frequency of neoantigen-reactive T cell clones (n = 18) in clusters 3, 7, and 16. **(E-F)** IFNγ secretion following overnight co-culture of 14 candidate TCRs transduced into healthy donor PBL predicted from cluster 7 with imDCs pulsed with titrated concentrations of mutated neoantigenic epitope and their wildtype counterparts for ARMC9 **(E)** or MYO5B **(F)** or their wild-type counterpart.



Supplemental Figure S2: Phenotypic clustering of Pt. 4287and Pt. 4317 tetramer-enriched circulating CD8<sup>+</sup> PBLs, related to Figure 1 (A) UMAP transcriptome clustering CMTR1<sup>K601T</sup> tetramer enriched cells (1500 CMTR1<sup>K601T</sup>:HLA-B\*49 tetramer<sup>pos</sup> cells spiked into a total of 40,000 CD8 cells). (B) TCR-transduced PBLs were co-cultured with imDCs pulsed with 1µg/mL mutated or wild-type minimal peptides. (C-D) Highlighting known neoantigen-specific clones (C) and public viral-targeting clones (E) Phenotypic clustering of Pt.4317 tetramer-enriched circulating CD8+ PBLs. UMAP clusters from 4317 PBL-derived CD8+ T cells. (F-G) Cluster distribution of HLA A\*02:01-restricted tetramer-sorted PIK3CA<sup>P449T</sup> neoantigen T cells (F) and FluM1<sub>(58-66)</sub>, EBV LMP2A<sub>(426-434)</sub> viral T cells (G) within PBL. (H) Transcriptome expression of top

genes expressed in cluster 11 (indicated by arrow) that harbors neoantigen-T cells. (I) Heatmap of antibody CITEseq-antibodies intensities on cells in scRNAseq clusters including Cluster 11 (indicated by arrow).



Supplemental Figure S3: Tetramer-agnostic circulating CD8<sup>+</sup> PBL T cells by FACSsorting strategy, related to Figure 1. (A) Sorting strategy used for the enrichment of circulating neoantigen-reactive CD8<sup>+</sup> T cells in samples from patients 4382, 4422, and 4324. (B-C) Representative example of enrichment sorting (B) and scRNA analysis for pt.4382, Archival neoantigen-reactive cells are highlighted in red (C). (D) Frequency of neoantigen-reactive T cell in PBL, sorted cells (scRNA-seq), and within cluster 12. Numbers in the graph represent p-value of paired T-test. (E-F) Projection of neoantigen-reactive T cell clones on UMAP space for pt.4422 (E) and pt.4324 (F).







Supplemental Figure S4: Combined phenotype and frequency analysis of NeoTCRPBL clones from the 6 patients with neoantigen TIL-reactivity, related Figure 2. (A) NeoTCR<sub>PBL</sub> clones from each of the 6 patients are enriched within cluster 9. Each colored circle represents the total frequency of neoantigen-specific T cell clones from each of the 6 patients. (B) NeoTCR<sub>PBL</sub> transcriptomic state remains intact over a wide range of resolution parameters in the 6 patient PBL. Arrow indicates Cluster 9 shown in Figure 2. (C) Heatmap of differentially expressed genes and genes-of-interest across UMAP clusters shown in Figure 2 (derived from samples from 6 patients). Cluster 9 indicates NeoTCR<sub>PBL</sub> cluster. The last 10 rows indicate T cell dysfunction markers of interest.

 $NeoTCR_{PBL}$  Ingenuity pathway analysis



Gene expression variation by sample within Cluster 9



Supplemental Figure S5: Ingenuity pathway analysis (IPA) and gene expression sample variation of NeoTCR<sub>PBL</sub> gene signature, related to Figure 2. (A) Canonical pathway analysis showing activated (positive Z score) and inhibited (negative) pathways of the NeoTCR<sub>PBL</sub> gene signature. The X-axis indicates the significance of the enrichment. (B) Inter-patient variation of genes expressed within neoantigen-specific T cells found within cluster 9. Note that these comparisons are performed within cells from cluster 9 compared by each patient.



Supplemental Figure S6: Frequency of neoantigen-reactive T cells in PBL subsets (related to Figure 3A) and prospective prediction of NeoTCRs from PBL of breast cancer patient Pt.4180, related to Figure 3. (A-B) CD8<sup>+</sup> sorting gating strategy for naïve, memory (A), and PD-1-expressing cells (B). (C) CD39 and CD103 fluorescence-minus-one (FMO) and staining and sorting gates for CD8<sup>+</sup> T cells. (D) Frequency of FACS-sorted T cell subsets using

common cell surface markers previously used to identify antitumor T cells from CD8<sup>+</sup> PBL T cells within metastatic cancer patients **(E)** Prediction and functional validation of neoantigenreactive TCRs for melanoma patient 4180. Sorting enrichment gating of circulating CD8<sup>+</sup> T cells for scRNAseq. 50,000 CD39<sup>+</sup>CD103<sup>+</sup> were sorted and mixed with 100,000 bulk CD8<sup>+</sup> cells **(F)** UMAP unsupervised clustering and prediction of NeoTCR<sub>PBL</sub> based on scGSEA (in red). **(G)** Frequency of TCR-transduced CD8<sup>+</sup> cells expressing CD137 following co-culture with imDCs electroporated with patient's TMGs. **(H)** Back-projection identified neoantigen-reactive NeoTCR<sub>PBL</sub> clones onto the UMAP plot.



Supplemental Figure S7: Prediction and functional validation of neoantigen-reactive TCRs for melanoma patient 4359, related Figure 3. (A) Sorting enrichment gating of circulating CD8<sup>+</sup> T cells for scRNA. 6,800 CD39<sup>+</sup>CD103<sup>+</sup> were sorted and mixed with 20,000 bulk CD8<sup>+</sup> cells (B) UMAP unsupervised clustering and prediction of NeoTCR<sub>PBL</sub> based on scGSEA (in red) (C-D) Frequency of TCR-transduced CD8<sup>+</sup> cells expressing CD137 following co-culture with imDCs electroporated with patient's TMGs (C) or peptides of TMG2 and 10 (D).



Supplemental Figure S8: Sensitivity and Specificity of gene signatures in predicting TCR clonotypes from the validation set, and frequency of NeoTCR<sub>PBL</sub> CD8+ T cells in healthy donors, related Figure 3. (A) NeoTCR<sub>PBL</sub> clone prediction by gene signatures (B) NeoTCR<sub>PBL</sub> gene signature has poor sensitivity in predicting public viral clonotypes (n = 9) from the validation set. (C) Overlap of NeoTCR<sub>PBL</sub> signature with other antitumor TIL programs. Venn diagram of shared genes between the 4 tumor-relevant TIL signatures and NeoTCR<sub>PBL</sub>. (D) Neoantigen-specific T clone prediction performance of the 111 unique genes from NeoTCR<sub>PBL</sub> (left panel) and the top 50 NeoTCR<sub>PBL</sub> genes (right panel). (E) CD39<sup>+</sup>CD103<sup>+</sup> CD8 T cells frequency in peripheral blood of patients with metastatic cancer and age-matched healthy donors. Frequency in CD45RO<sup>+</sup>HLA-DR<sup>+</sup> (left panel) and bulk CD8<sup>+</sup> (right panel).