

# Supplementary Figure S1. Expression of miR-9-1, miR-9-2 and miR-9-3 in the different subpopulations of hematopoietic cells and generation of miR-9-1-deficient mice.

(A to C) The indicated hematopoietic progenitor subpopulations and mature hematopoietic subpopulations were sorted from wild-type mouse BM or spleen. The levels of primary miR-9-1, miR-9-2 and miR-9-3 expression were measured by qRT-PCR, using  $\beta$ -Actin as internal control. Mean  $\pm$  SD is shown. (A) Expression of primary miR-9-1, miR-9-2 and miR-9-3 in the different subsets of BM hematopoietic progenitors. (B) Expression of primary miR-9-1, miR-9-2 and miR-9-3 in splenic CD4, CD8 and NK cells and BM granulocytes, macrophages, dendritic cells. (C) Expression of primary miR-9-1, miR-9-2 and miR-9-3 in B cells at the different developmental stages. (D) Schematic diagram of CRISPR/Cas9-mediated knockout of miR-9-1 in mice. The mouse miR-9-1 gene is located on chromosome 3. The mature miR-9 sequence is shown in red and its seed sequence is underlined. The two target sequences of gRNAs are indicated by green and blue lines above the sequences. (E) DNA sequencing analysis of miR-9-1, miR-9-2 and miR-9-3 genes in miR-9-1-deficient (-/-) and wild-type (+/+) mice. The mature miR-9 sequence is shown in red. Data shown are representative of 3 independent experiments (A to C).



## Supplementary Figure S2. Reduced survival, white blood cells, and T and B cell populations but normal myeloid cell population in miR-9-1-deficient mice.

(A) Cumulative survival curve of all miR-9-1-deficient mice. (B) Numbers of white blood cells (WBCs), red blood cells (RBCs) and platelets (PLTs) in the peripheral blood of miR-9-1deficient mice with retarded growth. (C) Reduction in pro- and pre-B cells in the BM of miR-9-1-deficient mice. BM cells from growth-retarded miR-9-1<sup>-/-</sup> and control mice were stained with anti-B220, anti-IgM, and anti-CD43 or anti-CD25. Percentages and numbers of pro- and pre-B cells in the BM are shown, and percentages indicate cells in the gated live cells. (D) Reduction in T1, T2, FO and MZ B cells in the spleen of miR-9-1-deficient mice. Splenocytes from growth-retarded miR-9-1<sup>-/-</sup> and control mice were stained with anti-B220, anti-IgD, anti-IgM, anti-CD21 and anti-CD23. Percentages and numbers of T1, T2, FO and MZ B cells in the spleen are shown, and percentages indicate cells in the gated B220<sup>+</sup> cells. (E) Reduced T cells in the spleen of miR-9-1-deficient mice. Splenocytes from growth-retarded miR-9-1-/and control mice were stained with anti-CD4 and anti-CD8. Percentages and numbers of CD4<sup>+</sup> and CD8<sup>+</sup> T cells in the spleen are shown and percentages indicate cells in the gated live cells. (F) Normal myeloid cell population in the BM of miR-9-1-deficient mice. BM cells from growth-retarded miR-9-1<sup>-/-</sup> and control mice were stained with anti-Mac-1 and anti-Gr-1. Percentages and numbers of Mac-1+Gr-1+ myeloid cells in the BM are shown and percentages indicate cells in the gated live cells. Data shown are representative of or obtained from 20 (A), 9 (B), 7 (C, D), 6 (E) or 7 (F) pairs of control and miR-9-1<sup>-/-</sup> mice. Each dot represents an individual mouse. Mean  $\pm$  SD is shown. \*, p < 0.05; \*\*, p < 0.01.



#### Supplementary Figure S3. Normal lymphopoiesis in miR-9-1-deficient mice with normal growth.

(A) Normal pro/pre-, immature and mature B cell populations in the BM of miR-9-1-deficient mice with normal growth. BM cells from miR-9-1<sup>-/-</sup> mice with normal growth and wild-type control mice were stained with anti-B220 and anti-IgM. Percentages and numbers of pro/pre-, immature and mature B cells in the BM are shown and percentages indicate cells in the gated live cells. (B) Normal pro- and pre-B cell populations in the BM of miR-9-1-deficient mice with normal growth. BM cells from miR-9-1<sup>-/-</sup> mice with normal growth and wild-type control mice were stained with anti-B220, anti-IgM, and anti-CD43 or anti-CD25. Percentages and numbers of pro- and pre-B cells in the BM are shown and percentages indicate cells in the gated live cells. (C) Normal DN, DP and SP T cells in the thymus of miR-9-1-deficient mice with normal growth. Thymocytes from normal miR-9-1<sup>-/-</sup> and control mice were stained with anti-CD4 and anti-CD8. Percentages and numbers of DN, DP and SP T cells in the thymus are shown and percentages and numbers of DN, DP and SP T cells in the thymus are stained with anti-CD4. Percentages and numbers of DN, DP and SP T cells in the thymus are shown and percentages and numbers of DN, DP and SP T cells in the thymus of miR-9-1-deficient mice with normal growth.



### Supplementary Figure S4. Normal hematopoietic progenitors, lymphocytes and myeloid cells in the recipients that received BM cells from growth-retarded miR-9-1-deficient mice.

BM cells from wild-type control or growth-retarded miR-9-1<sup>-/-</sup> mice (CD45.2<sup>+</sup>) were transplanted into lethally irradiated CD45.1<sup>+</sup> recipients. About eight weeks after transplantation, the recipients were analyzed. (A) The percentages of CD45.2<sup>+</sup> cells in the BM, spleens and thymi of the recipients. Percentages indicate CD45.2<sup>+</sup> cells in the gated live cells. (B) The numbers of white blood cells (WBCs), red blood cells (RBCs) and platelets (PLTs) in the peripheral blood of the recipients. (C) The percentages and numbers of CD45.2<sup>+</sup> LSK and LK cells in the BM of the recipients. Percentages indicate CD45.2<sup>+</sup> LSK and LK in the gated CD45.2<sup>+</sup>Lin<sup>-</sup>IL-7R<sup>-</sup> population. (D) The percentages and numbers of GMP, CMP and MEP in the BM of the recipients. Percentages indicate GMP, CMP and MEP in the gated CD45.2\*Lin<sup>-</sup>IL-7R<sup>-</sup> LK population. (E) The percentages and numbers of Mac-1+Gr-1+ myeloid cells in the BM of the recipients. Percentages indicate Mac-1+Gr-1+ myeloid cells in the gated CD45.2<sup>+</sup> population. (F) The percentages and numbers of pro- and pre-B cells in the BM of the recipients. Percentages indicate CD43<sup>+</sup> pro- and CD25<sup>+</sup> pre-B cells in the gated CD45.2<sup>+</sup> population. (G,H) The percentages and numbers of B cells at the different developmental stages in the spleen of the recipients. Percentages indicate FO, T2, T1 and MZ B cells in the gated CD45.2<sup>+</sup>B220<sup>+</sup> population. (I) The percentages and numbers of CD4<sup>+</sup> and CD8<sup>+</sup> T cells in the spleen of the recipients. Percentages indicate CD4<sup>+</sup> and CD8<sup>+</sup> T cells in the gated CD45.2<sup>+</sup> population. (J) The percentages and numbers of DN, DP and SP T cells in the thymus of the recipients. Percentages indicate DN, DP and SP T cells in the gated CD45.2<sup>+</sup> population. (K,L) Normal engrafting capacity of miR-9-1-deficient hematopoietic progenitors. BM cells from wild-type or growth-retarded miR-9-1<sup>-/-</sup> mice (CD45.2<sup>+</sup>) were mixed 1:1 with wild-type competitor BM cells from B6 SJL mice (CD45.1<sup>+</sup>) and transplanted into lethally irradiated CD45.1<sup>+</sup> mice. Eight weeks after BM transplantation, the recipients were analyzed for the relative contribution of donor engraftment in various hematopoietic progenitors in the BM (K) and splenic T, B and myeloid cells (L). Data shown are obtained from 4 (A, C to J) or 2 (B) recipients that received control or miR-9-1<sup>-/-</sup> BM cells or 4 (K, L) recipients that received control or miR-9-1<sup>-/-</sup> BM cells mixed at a 1:1 ratio with competitor BM cells from wildtype B6 SJL mice (CD45.1<sup>+</sup>). Each dot represents an individual mouse. Mean  $\pm$  SD is shown. \*, p < 0.05; \*\*, p < 0.01.



## Supplementary Figure S5. Normal hematopoietic progenitors, lymphocytes and myeloid cells in the recipients that received BM cells from growth-retarded miR-9-1-deficient mice after 12 weeks.

BM cells from wild-type control or growth-retarded miR-9-1<sup>-/-</sup> mice (CD45.2<sup>+</sup>) were transplanted into lethally irradiated CD45.1<sup>+</sup> recipients. Twelve weeks after transplantation, the recipients were analyzed. (A) The numbers of total BM cells, splenocytes and thymocytes the recipients. (B) The percentages and numbers of CD45.2<sup>+</sup> LSK and LK cells in in the BM of the recipients. Percentages indicate CD45.2<sup>+</sup> LSK and LK in the gated CD45.2<sup>+</sup>Lin<sup>-</sup>IL-7R<sup>-</sup> population. (C) The percentages and numbers of LT-HSC, ST-HSC and MPP in the recipients. Percentages indicate LT-HSC, ST-HSC and MPP in the gated CD45.2+Lin-IL-7R-LSK population. (D) The percentages and numbers of GMP, CMP and MEP in the BM of the recipients. Percentages indicate GMP, CMP and MEP in the gated CD45.2<sup>+</sup>Lin<sup>-</sup>IL-7R<sup>-</sup> LK population. (E) The percentages and numbers of Mac-1<sup>+</sup>Gr-1<sup>+</sup> myeloid cells in the BM of the recipients. Percentages indicate Mac-1+Gr-1+ myeloid cells in the gated CD45.2+ population. (F,G) The percentages and numbers of lymphoid progenitors in the BM of the recipients. Percentages indicate IL7R<sup>+</sup>ckit<sup>mid</sup> in the gated CD45.2\*Lin<sup>-</sup> population (F) and CLP in the gated CD45.2\*Lin<sup>-</sup>IL7R\*ckit<sup>mid</sup> population (G). (H) The percentages and numbers of pro- and pre-B cells in the BM of the recipients. Percentages indicate CD43<sup>+</sup> pro- and CD25<sup>+</sup> pre-B cells in the gated CD45.2<sup>+</sup> population. (I) The percentages and numbers of B cells at the different developmental stages in the BM of the recipients. Percentages indicate pro/pre-, immature (IM) and mature (M) B cells in the gated CD45.2<sup>+</sup> population. (J,K) The percentages and numbers of B cells at the different developmental stages in the spleen of the recipients. Percentages indicate FO, T2, T1 (J) and MZ (K) B cells in the gated CD45.2<sup>+</sup>B220<sup>+</sup> population. (L) The percentages and numbers of DN, DP and SP T cells in the thymus of the recipients. Percentages indicate DN, DP and SP T cells in the gated CD45.2<sup>+</sup> population. Data shown are obtained from 3 recipients that received control BM cells and 4 recipients that received miR-9-1<sup>-/-</sup> BM cells. Each dot represents an individual mouse. Mean  $\pm$  SD is shown. \*, p < 0.05; \*\*, p < 0.01.



#### Supplementary Figure S6. Normal osteogenic differentiation of MSCs from miR-9-1deficient mice with retarded growth or normal growth.

(A) RANKL-induced osteoclast differentiation of miR-9-1-deficient BMMs in vitro. BMMs derived from growth-retarded miR-9-1<sup>-/-</sup> and control mice were cultured with RANKL and M-CSF. The cells were stained with TRAP staining and visualized by light microscopy. (B) Serum level of CTX in miR-9-1-deficient mice. Serum levels of CTX in 6-12 weeks old growthretarded miR-9-1<sup>-/-</sup> and control mice were determined by the ELISA analysis. (C) Marked reduction of the level of total mature miR-9 in miR-9-1-deficient CD51+Sca1+ MSCs and CD51<sup>+</sup>Sca1<sup>-</sup> OBs. The relative levels of total mature miR-9 in CD51<sup>+</sup>Sca1<sup>+</sup> and CD51<sup>+</sup>Sca1<sup>-</sup> cells from the BM of miR-9-1<sup>-/-</sup> mice with normal or retarded growth were quantified by qRT-PCR as a fraction of the corresponding wild-type control cells, which was set as 1. (D) CFU-F formation of miR-9-1-deficient MSCs. MSCs isolated from the BM of miR-9-1-/- and control mice were subjected to the CFU-F assay. Morphology of CFU-Fs stained with crystal violet (left) and guantification of CFU-F numbers (right) are shown. (E, F) MSCs sorted from miR-9-1<sup>-/-</sup> mice with normal growth and wild-type controls were cultured in osteogenic media for 7 days, and ALP staining (A) and quantification (B) were performed. Data shown are representative of or obtained from 2 (A), 13 (B), 3 (C), 8 (D) miR-9-1<sup>-/-</sup> and control mice or representative of or obtained from 3 (E, F) independent experiments. Mean  $\pm$  SD is shown.



#### Supplementary Figure S7. Analysis of the 2<sup>nd</sup> independent colony of miR-9-1-deficient mice.

(A) DNA sequencing analysis of miR-9-1, miR-9-2 and miR-9-3 genes in miR-9-1-deficient (-/-) and wild-type (+/+) mice of the 2<sup>nd</sup> independent colony. The mature miR-9 sequence is shown in red. (B) Reduction in overall mature miR-9 expression in the BM of growth-retarded miR-9-1-deficient mice from the 2<sup>nd</sup> independent colony. The levels of mature miR-9 expression in BM cells from miR-9-1-deficient (miR-9-1-/-) mice were quantified by gRT-PCR as a fraction of the corresponding wild-type control cells, which was set as 1. (C) Growth defect in miR-9-1-deficient mice from the 2<sup>nd</sup> independent colony. A representative image of 8-week-old growth-retarded male miR-9-1<sup>-/-</sup> mice and controls is shown. (D) Reduction in pro/pre- and immature B cells in the BM of growth-retarded miR-9-1-deficient mice from the 2<sup>nd</sup> independent colony. BM cells from growth-retarded miR-9-1<sup>-/-</sup> and control mice were stained with anti-B220 and anti-IgM. Percentages and numbers of pro/pre-, immature and mature B cells in the BM are shown, and percentages indicate cells in the gated live cells. (E) Reduction in pro- and pre-B cells in the BM of growth-retarded miR-9-1-deficient mice from the 2<sup>nd</sup> independent colony. BM cells from growth-retarded miR-9-1<sup>-/-</sup> and control mice were stained with anti-B220, anti-IgM, and anti-CD43 or anti-CD25. Percentages and numbers of pro- and pre-B cells in the BM are shown, and percentages indicate cells in the gated live cells. (F) Reduction in DN, DP and SP T cells in the thymus of growth-retarded miR-9-1-deficient mice from the 2<sup>nd</sup> independent colony. Thymocytes from growth-retarded miR-9-1<sup>-/-</sup> and control mice were stained with anti-CD4 and anti-CD8. Percentages and numbers of DN, DP and SP T cells in the thymus are shown, and percentages indicate cells in the gated live cells. (G) ALP staining of miR-9-1-deficient MSCs from the 2<sup>nd</sup> independent colony after osteogenic differentiation. MiR-9-1-- and control MSCs were cultured in osteogenic media for 7 days, and ALP staining (top) and quantification (bottom) were performed. Data shown are representative of or obtained from 3 independent experiments. Mean  $\pm$  SD is shown. \*, p < 0.05; \*\*, p < 0.01.



### Supplementary Figure S8. Analysis of gene expression profiles and normal adipogenic differentiation of miR-9-1-deficient MSCs from growth-retarded mutant mice.

(A) FACS sorting of MSCs, OBs, and endothelial cells from BM stromal cells and gating strategy. (B) Expression of miR-9-1, miR-9-2, and miR-9-3 in the BM niche cells. The relative mRNA levels of miR-9-1, miR-9-2 and miR-9-3 in CD51<sup>+</sup>Sca-1<sup>+</sup> MSCs, CD51<sup>+</sup>Sca-1<sup>+</sup>LepR<sup>+</sup> MSCs, and CD51<sup>+</sup>Sca-1<sup>-</sup> OBs within the Lin<sup>-</sup>CD45<sup>-</sup>CD31<sup>-</sup> gate, and CD31<sup>+</sup>Sca-1<sup>+</sup> endothelial cells within the Lin<sup>-</sup>CD45<sup>-</sup> gate. (C) The relative mRNA levels of Runx2, Osx, Alp, Ocn, and Col1a1 in miR-9-1-deficient OBs. The mRNA levels of Runx2, Osx, Alp, Ocn, and Col1a1 in sorted miR-9-1<sup>-/-</sup> OBs (CD51<sup>+</sup>Sca-1<sup>-</sup>) were quantified by qRT-PCR as a fraction of the corresponding wild-type control cells, which was set as 1. (D) GSEA of the myoblast, chondrocyte and adipocyte signatures in miR-9-1<sup>-/-</sup> and wild-type control MSCs (CD51<sup>+</sup>Sca-1<sup>+</sup>). (E, F) Normal adipogenic differentiation of MSCs from growth-retarded miR-9-1-deficient mice. MiR-9-1<sup>-/-</sup> and control MSCs were cultured in adipogenic media for 7 days. Oil red staining were performed (E), and the mRNA levels of PPARγ and Adipsin in miR-9-1<sup>-/-</sup> MSCs were quantified by qRT-PCR as a fraction of the corresponding control cells, which was set as 1 (F). (G) GSEA of the bone development, mineralization, and ossification signatures in miR-9-1<sup>-/-</sup> and wild-type control MSCs. (H) Enrichment analysis of downregulated genes in miR-9-1<sup>-/-</sup> relative to control MSCs using the REACTOME database. Bar plots show negative log10 of false discovery rate (FDR) q-values for enrichment. (I) Sections of the femurs from 6- to 12-week miR-9-1<sup>-/-</sup> and control mice were analyzed for bone mineralization by Von Kossa staining. Data shown are obtained from or representative of 4 (A, C, D, G and H) or 3 (I) miR-9-1<sup>-/-</sup> and wild-type mice or 3 (B) or 2 (E, F) independent experiments.



# Supplementary Figure S9. MiR-9-1 regulates expression of the genes associated with osteoblast differentiation and function in OBs and the populations of NG2<sup>+</sup>, CD200<sup>+</sup>, and LepR<sup>+</sup> cells within CD51<sup>+</sup>Sca1<sup>-</sup> BM stromal cells in miR-9-1-deficient mice.

(A) The relative mRNA levels of Runx2, Col1a1, and Ocn in sorted OBs were quantified by qRT–PCR using β-Actin as an internal control. Lin CD45 CD31 Sca-1 CD51 cells were used as a negative control. (B) The relative levels of total mature miR-9 in OBs from miR-9-1deficient mice with normal or retarded growth. The levels of total mature miR-9 in sorted OBs from the indicated mice were quantified by qRT-PCR as a fraction of the corresponding wildtype control OBs, which was set as 1. (C) Heatmap of differentially expressed genes between OBs from growth-retarded miR-9-1<sup>-/-</sup> mice and control mice. Genes with an adjusted p-value < 0.05 found by DESeg2 were assigned as differentially expressed. (D) Heatmap of differentially expressed osteoblast-specific, extracellular organization and cell adhesion molecule genes between OBs from growth-retarded miR-9-1<sup>-/-</sup> mice and control mice. (E) GO enrichment analysis of downregulated genes in OBs from growth-retarded miR-9-1<sup>-/-</sup> mice relative to control OBs. Bar plots show negative log10 of p-values for GO enrichment analysis of the biological pathways in control relative to miR-9-1<sup>-/-</sup> OBs. (F-I) The populations of MSCs, ECs, OBs, and NG2<sup>+</sup>, CD200<sup>+</sup> and LepR<sup>+</sup> cells within CD51<sup>+</sup>Sca1<sup>-</sup> BM stromal cells. BM stromal cells from growth-retarded miR-9-1<sup>-/-</sup> mice and control mice were stained with the anti-lineage cocktail (Mac-1, Gr-1, B220, CD4, CD8, and Ter-119), CD45.2, CD31, CD51, Sca-1, NG2, CD200, and LepR antibodies. The numbers of MSCs, ECs, and OBs were determined (F). The percentages of the indicated cells in the Lin CD45.2 CD31 CD51<sup>+</sup>Sca1<sup>-</sup> gate were shown (G-I). Data shown is obtained from 4 (A, C-E) or 3 (B, F-I) growth-retarded miR-9-1<sup>-/-</sup> and wild-type mice. Mean  $\pm$  SD is shown. \*, p < 0.05; \*\*, p < 0.01.



#### Supplementary Figure S10. MiR-9-5p regulates BMP signaling in MSCs by directly targeting Fst.

(A) The predicted Fst 3'-UTR binding site of miR-9-5p and the mutant Fst 3'-UTR binding site of miR-9-5p. (B) Reduced level of endogenous Fst protein in primary MSCs by miR-9-5p overexpression. The expression levels of Fst in MSCs transfected with empty vector (control) or miR-9-5p expression vector (miR-9-5p) was determined by Western-blot (left panel). Band intensity was measured using ImageJ software, and the results are presented as a ratio of the band intensity of Fst to Actin (right panel). (C) The levels of Fst expression in MSCs, OBs and HSCs of wild-type mice. The relative expression levels of Fst were determined in sorted wild-type CD51+Sca1+ MSCs, CD51+Sca1- OBs and HSCs by gRT-PCR analysis. The relative mRNA levels of Fst were normalized to Actin. (D) The protein levels of Fst and phospho-Smad1/5/8 (p-smad1/5/8) in miR-9-1-deficient MSCs. Cell lysates from cultured miR-9-1<sup>-/-</sup> and control CD51<sup>+</sup>Sca1<sup>+</sup> MSCs were subjected to Western blot analysis with the indicated antibodies (left panel). The levels of Smad1 and Actin proteins were used as loading controls. Band intensity was measured using ImageJ software, and the results are presented as a ratio of the band intensity of Fst, pSmad1/5/8 or Smad to Actin (right panel). (E) The levels of IGF-1, IL-7, Cxcl12 and Csf1 mRNA in miR-9-1<sup>-/-</sup> relative to control MSCs. The mRNA levels of IGF-1, IL-7, Cxcl12 and Csf1 in miR-9-1<sup>-/-</sup> CD51<sup>+</sup>Sca1<sup>+</sup> MSCs were quantified by gRT-PCR as a fraction of the corresponding control wild-type cells, which was set as 1. The relative mRNA levels of the indicated genes in miR-9-1<sup>-/-</sup> and control MSCs were normalized to Actin. (F) The levels of IGF-1, IL-7, Cxcl12, Csf1, Osx, Runx2 and Fst mRNA in miR-9-1<sup>-/-</sup> relative to control LepR<sup>+</sup> cells. BM Lin<sup>-</sup>CD45<sup>-</sup>CD31<sup>-</sup> CD51<sup>+</sup>LepR<sup>+</sup> cells were sorted from miR-9-1<sup>-/-</sup> mice with retarded growth and control mice. The mRNA levels of the indicated genes in miR-9-1<sup>-/-</sup> LepR<sup>+</sup> cells were quantified by qRT-PCR as a fraction of the corresponding control wild-type cells, which was set as 1. The relative mRNA levels of the indicated genes in miR-9-1<sup>-/-</sup> and control LepR<sup>+</sup> cells were normalized to Actin. Data shown are representative of or obtained from 3 (B, D to F) miR-9-1<sup>-/-</sup> and control mice, and 3 (C) wild-type mice. Each dot represents an individual mouse. Mean  $\pm$  SD is shown. \*, p < 0.05; \*\*, p < 0.01.



Supplementary Figure S11. The levels of Igf1, IL-7, CxcI12, Csf1, Osx, Runx2 and Fst mRNA in LepR<sup>+</sup> cells from miR-9-1-deficient mice with normal growth relative to control mice.

BM Lin<sup>-</sup>CD45<sup>-</sup>CD31<sup>-</sup>CD51<sup>+</sup>LepR<sup>+</sup> cells were sorted from miR-9-1<sup>-/-</sup> mice with normal growth and control mice. The mRNA levels of lgf1, IL-7, Cxcl12, Csf1, Osx, Runx2 and Fst in miR-9-1<sup>-/-</sup> MSCs were quantified by qRT-PCR as a fraction of the corresponding control wild-type cells, which was set as 1. The relative mRNA levels of the indicated genes in miR-9-1<sup>-/-</sup> and control MSCs were normalized to Actin. Each dot represents an individual mouse, and the mean  $\pm$  SD is shown.



#### Supplementary Figure S12. Model of miR-9-1 regulation of lymphopoiesis.

MiR-9-1 controls osteoblastic differentiation, at least in part, by downregulating Fst, an antagonist of BMP. When miR-9-1 is deficient, Fst level increases, resulting in reduced BMP signaling, impaired osteoblastic differentiation, and decreased production of IL-7, IGF-1 and possibly CXCL12. Ultimately, this disruption of the osteoblastic niche leads to a reduction in hematopoietic progenitors and impaired lymphopoiesis.