

Demystifying Polyphosphate-Accumulating Organisms Relevant to Wastewater Treatment: A Review of Their Phylogeny, Metabolism, and Detection

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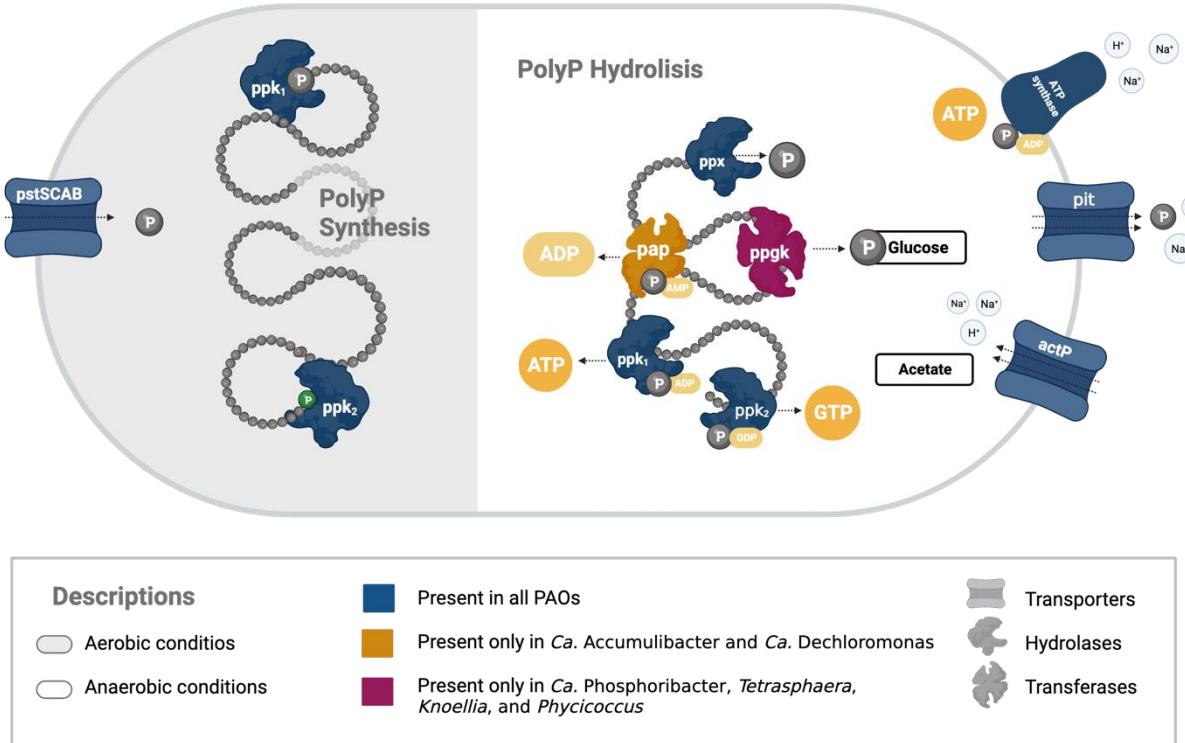


Fig. S1. PolyP synthesis, PolyP hydrolysis, P uptake, and acetate transport. During anaerobic conditions, PolyP hydrolysis in PAOs can be mediated by polyphosphate kinases (encoded by *ppks*) and exopolyphosphatases (encoded by *ppxs*). Among them, PPK1 catalyzes ATP formation, and PPK2 favors GTP formation, both utilizing the phosphate (P) from PolyP. PPX enzymes hydrolyze PolyP from the polymer ends to release P, which can be expelled from the cell along with cations using the low-affinity P transporter (encoded by *pit*). These cations can then re-enter the cell through the acetate transporter (encoded by *actP*) or ATP synthase. The acetate transporter (encoded by *actP*) acts as a symporter, facilitating the co-transport of acetate and cations, while ATP generation takes place through cation influx, supplying energy for cellular processes. PolyP can also undergo degradation through the transfer of terminal P groups to acceptor molecules. This process involves PolyP-AMP phosphotransferase (encoded by *pap*), transferring P groups from PolyP to AMP, and a PolyP glucokinase (encoded by *ppgk*) phosphorylating glucose using P from PolyP. During aerobic conditions, the cellular uptake of P is facilitated by the high-affinity transporter encoded by *pstSCAB*. Additionally, PolyP synthesis takes place through the enzymatic activity of polyphosphate kinases (encoded by *ppk* genes).

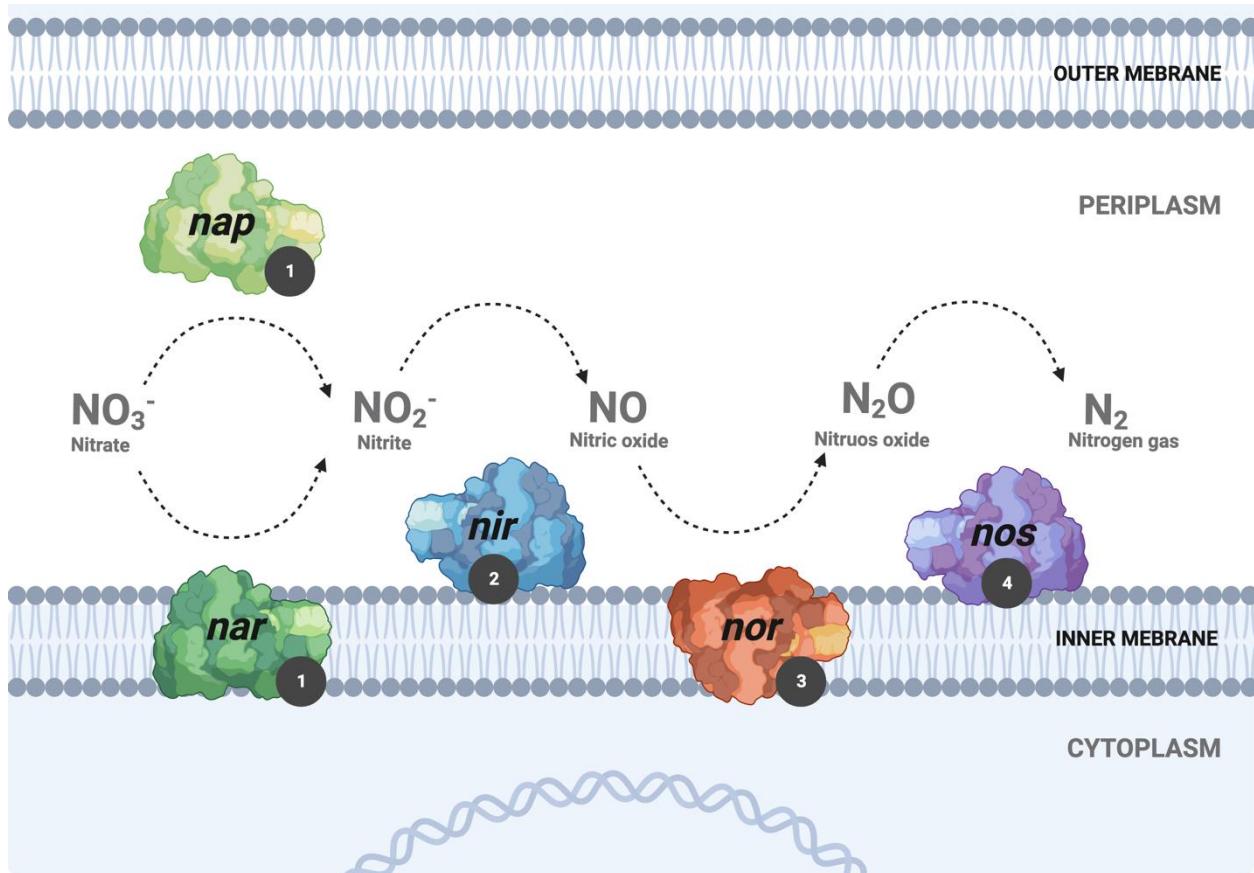


Fig. S2. Denitrification schematic. Nitrate (NO_3^-) reduction to nitrite (NO_2^-) can be done either by the periplasmic nitrate reductase (*nap*) or the transmembrane respiratory nitrate reductase (*nar*). The second step includes *nir* enzyme that catalyzes the conversion of NO_2^- into nitric oxide (NO). In the third step, *nor* catalyzes the reduction of NO to nitrous oxide (N_2O). Finally, *nos* completes the denitrification by reducing N_2O to nitrogen gas (N_2).

Table S1. Accession numbers from NCBI for the available PAOs.

| Organism | ACC | Type and Clades classification for <i>Ca. Accumulibacter</i> |
|--|-----------------|--|
| <i>Ca. Accumulibacter phosphatis</i> Bin19 | GCA_005889575.1 | |
| <i>Ca. Accumulibacter phosphatis</i> HKU2 | GCA_000987395.1 | |
| <i>Ca. Accumulibacter cognatus</i> SSA1 | GCA_013414765.1 | |
| <i>Ca. Accumulibacter cognatus</i> SK-02 | GCA_000584975.2 | |
| <i>Ca. Accumulibacter</i> UW6 | GCA_017592725.1 | |
| <i>Ca. Accumulibacter</i> UW11-POB | GCA_017302385.1 | |
| <i>Ca. Accumulibacter</i> vicinus | GCA_000584955.2 | |
| <i>Ca. Accumulibacter</i> vicinus UBA5574 | GCA_002425405.1 | |
| <i>Ca. Accumulibacter</i> contiguous SBR-L | GCA_012940005.1 | |
| <i>Ca. Accumulibacter</i> Fred-II3C-720 | GCA_016713625.1 | |
| <i>Ca. Accumulibacter</i> necessities UW12-POB | GCA_017302435.1 | |
| <i>Ca. Accumulibacter</i> OdNE-BAT3C-415 | GCA_016714935.1 | |
| <i>Ca. Accumulibacter</i> UW10-POB | GCA_017302555.1 | |
| <i>Ca. Accumulibacter</i> Hade BATAC726 | GCA_016711725.1 | |
| <i>Ca. Accumulibacter</i> iunctus UBA232A107 | GCA_002345025.1 | |
| <i>Ca. Accumulibacter</i> adjunctus SK-12 | GCA_000585015.1 | |
| <i>Ca. Accumulibacter</i> conexus UW7 | GCA_017592775.1 | |
| <i>Ca. Accumulibacter</i> Conexus UW13-POB | GCA_017302415.1 | |
| <i>Ca. Accumulibacter</i> similis SSB1 | GCA_013347225.1 | |
| <i>Ca. Accumulibacter</i> SCELSE-1 | GCA_005524045.1 | |
| <i>Ca. Accumulibacter</i> UW8-POB | GCA_020445855.1 | |
| <i>Ca. Accumulibacter</i> UW4 | GCA_020445885.1 | |
| <i>Ca. Accumulibacter</i> regalis BA-93 | GCA_020445925.1 | |
| <i>Ca. Accumulibacter</i> regalis CANDO-1 | GCA_017302345.1 | |
| <i>Ca. Accumulibacter</i> BA-92 | GCA_017592785.1 | |
| <i>Ca. Accumulibacter</i> phosphatis HKU1 | GCA_000585075.1 | |
| <i>Ca. Accumulibacter</i> phosphatis SBR-S | GCA_009467855.1 | |
| <i>Ca. Accumulibacter</i> meliphilus UW-LDO | GCA_000585055.1 | |
| <i>Ca. Accumulibacter</i> aalborgensis | GCA_000987445.1 | |
| <i>Ca. Accumulibacter</i> UW5 | GCA_012939955.1 | |
| <i>Ca. Accumulibacter</i> UW9-POB | GCA_003332265.1 | |
| <i>Ca. Dechloromonas</i> phosphorivorans Skiv | GCA_016721185.1 | |
| <i>Ca. Dechloromonas</i> phosphorivorans EsbW | GCA_016709495.1 | |
| <i>Ca. Dechloromonas</i> phosphorivorans OdNE | GCA_016714975. | |
| <i>Ca. Dechloromonas</i> phosphoritropha Ribe | GCA_016722705.1 | |
| <i>Ca. Phosphoribacter</i> baldrii Bjer | GCA_016708855.1 | |
| <i>Ca. Phosphoribacter</i> baldrii EsbW | GCA_016709925.1 | |
| <i>Ca. Phosphoribacter</i> baldrii AalW | GCA_016704565.1 | |
| <i>Ca. Phosphoribacter</i> hodrii AalE | GCA_016705565.1 | |
| <i>Ca. Phosphoribacter</i> hodrii Ribe | GCA_016722305.1 | |
| <i>Tetrasphaera jenkinsii</i> Ben-74 | GCA_001046875.1 | Formerly classified as <i>Tetrasphaera</i> |
| <i>Tetrasphaera jenkinsii</i> UW-TS-TETRA1-1 | GCA_022641475.1 | |
| <i>Tetrasphaera</i> aved 18-Q3-R54-62 MAXAC378 | GCA_016702415.1 | |
| <i>Tetrasphaera australiensis</i> Ben-110 | GCA_001050535.1 | |
| <i>Tetrasphaera japonica</i> T1-X7 | GCA_001046855.1 | |
| <i>Tetrasphaera</i> hjor 18-Q3-R7-51-BATAC29 | GCA_016710545.1 | |
| <i>Knoellia remsis</i> | GCA_003002895.1 | |
| <i>Phycicoccus</i> duodecadis DSM-12806 | GCA_002846495.1 | |
| <i>Phycicoccus</i> elongata Lp2 | GCA_000367525.2 | |
| <i>Thiothrix caldifontis</i> | GCA_900107695.1 | |

Table S2. Assimilatory and dissimilatory nitrate reduction in PAOs. Step 1 involves the conversion of nitrate to nitrite, while step 2 converts the nitrite to ammonia. Complete gene names and KEGG nomenclature are available in **Table S8**; Microorganism accession numbers (NCBI) are available in **Table S1**.

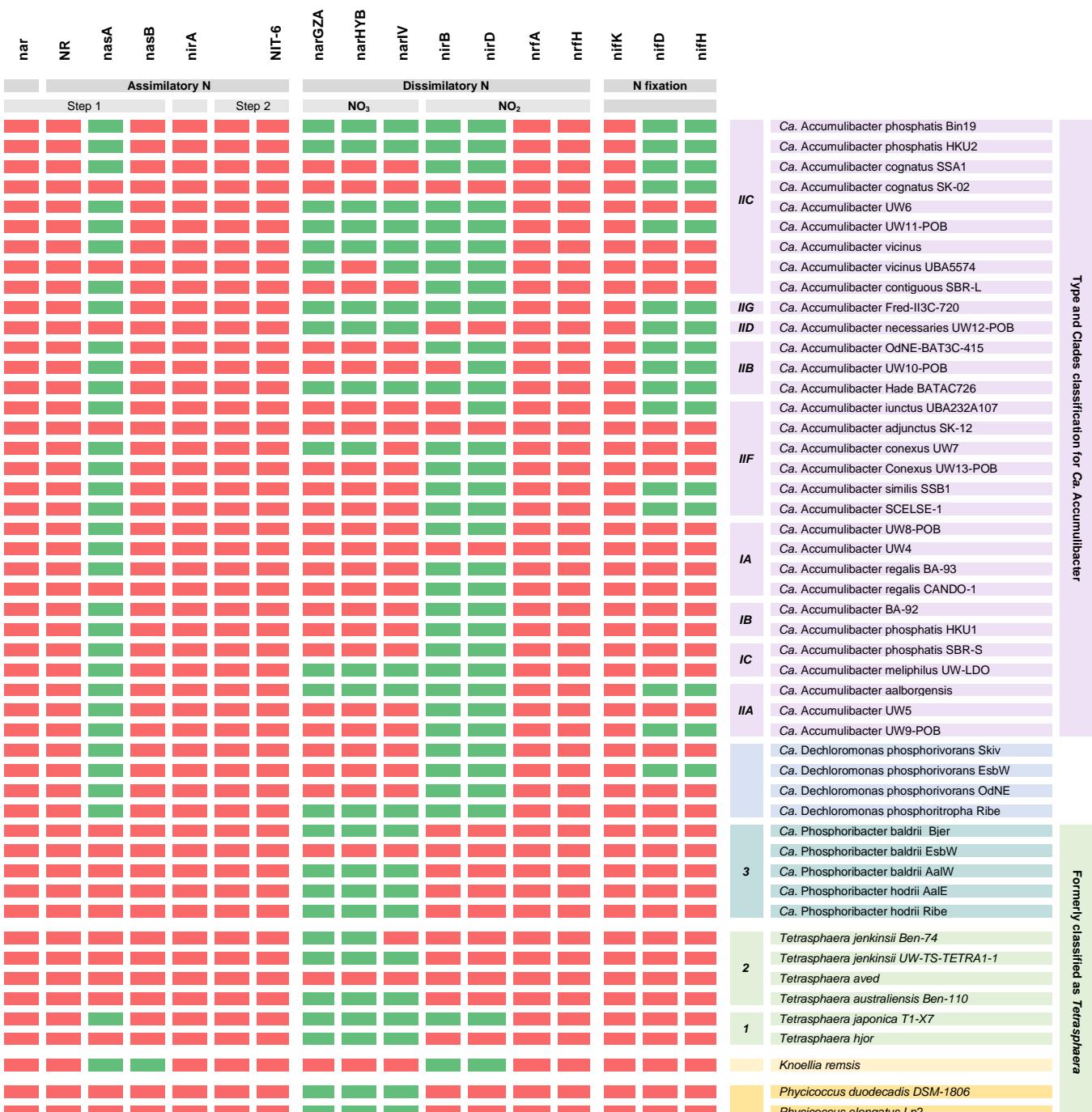


Table S3. Enzymes involved in PolyP metabolism. KEGG nomenclature of the genes is available in **Table S1.**

| PolyP Metabolism | | |
|--------------------------------------|----------------|---|
| Synthesis | | |
| PolyP kinase 1 | <i>ppk1</i> | P transfer between ATP/ADP to PolyP (anabolic reaction is favored) |
| PolyP kinase 2 | <i>ppk2</i> | P transfer between GTP/GDP to PolyP (catabolic reaction is favored) |
| Hydrolysis | | |
| Endo-polyphosphatase | <i>ppn</i> | Cleaves PolyP internally |
| Exo-polyphosphatase | <i>ppx</i> | Hydrolysis of the terminal P of PolyP |
| Transferases | | |
| Polyphosphate glucokinase | <i>ppgK</i> | Phosphorylation of glucose using polyphosphate as the P donor |
| AMP-polyphosphate phosphotransferase | <i>pap</i> | Phosphorylation of AMP using polyphosphate as the P donor |
| Transport | | |
| Low-affinity P transporters | <i>pit</i> | Single membrane protein energized by proton motive force |
| Phosphate-specific transport | <i>pstABCS</i> | Multi-subunit transporter usually encoded by a four-gene operon |

Table S4. Polyhydroxyalkanoates (PHAs) made from volatile fatty acids.

| Volatile Fatty Acid | Resulting PHAs |
|------------------------|---|
| Acetate | Polyhydroxy butyrate (PHB) |
| Propionate | Polyhydroxy-2-methyl valerate (PH2MV) |
| Acetate and Propionate | Polyhydroxy valerate (PHV) Polyhydroxy-2-methyl butyrate (PH2MB) |

Table S5. Various bacterial strains exhibiting PolyP accumulation and complete denitrification capabilities have been reported, having been isolated from diverse environmental sources. DPAOs include microorganisms that have proven to have full PAO phenotype (i.e., capable of P uptake under aerobic conditions for synthesizing PolyP, and in the anaerobic phase, PolyP is hydrolyzed, and P is released for energy generation and organic carbon uptake). While potential DPAOs can store PolyP aerobically, but more evidence is needed to confirm their full PAO phenotype.

| Organism | Source environment | Ref. |
|--|--|------|
| DPAOs | | |
| <i>Paracoccus denitrificans YCP</i> | Sludge from a full-scale anaerobic/ anoxic/aerobic process | [1] |
| <i>Paracoccus sp. YKP-9</i> | Activated sludge from sewage treatment plant | [2] |
| <i>Thauera sp. SND5</i> | Activated sludge from an anoxic tank at the Ulu Pandan Water Plant, Singapore | [3] |
| Potential DPAOs | | |
| <i>Pseudomonas stutzeri YG-24</i> | Sediment of the eutrophic Taihu Lake | [4] |
| <i>Pseudomonas stutzeri ADP-19</i> | Landfill leachate, biogas residue, contaminated sediments, and pickle wastewater | [5] |
| <i>Pseudomonas chloritidismutans K14</i> | Sediment samples from a freshwater aquaculture pond in Tianjin | [6] |
| <i>Paracoccus denitrificans ISTOD1</i> | Okhla Sewage Treatment Plant | [7] |
| <i>Agrobacterium sp. LAD9</i> | Landfill leachate treatment system | [8] |
| <i>Enterobacter cloacae HW-15</i> | Water samples from a phosphorus-rich river in Hubei Province of China | [9] |
| <i>Acinetobacter sp. C-13</i> | Activated sludge from a lab-scale sequencing batch reactor | [10] |
| <i>Delftia tsuruhatensis ZK-1</i> | Activated sludge from Datansha WWTP Guangzhou, China | [11] |
| <i>Pseudoxanthomonas sp. YP1</i> | Aerobic granular sludge sequencing batch reactor | [12] |
| <i>Bacillus cereus GS-5</i> | Pure culture | [13] |
| <i>Bacillus cereus ZQN2</i> | Lab-scale sequencing batch reactor from a lab-scale reactor | [14] |
| <i>Arthrobacter sp. HHEP5</i> | Effluents of mariculture ponds | [15] |

Table S6. FISH probes targeting 16S rRNA of PAOs belonging to *Ca. Accumulibacter*, *Tetrasphaera*, *Dechloromonas*, *Ca. Phosphoribacter*. Additionally, 23S rRNA probes for *Ca. Phosphoribacter* and *Tetrasphaera* are displayed, along with *ppk1* primers designed to target diverse *Ca. Accumulibacter* types and clades. The aim for including the FISH probes in this review is to highlight the evolution of new probes with the discovery of new PAOs. It is recommended that FISH probes be tested in-silico before being used by researchers.

| | Probe | Specificity | Sequence (5'- 3') | Ref. |
|---|-------------------|---|---|------|
| FISH Probes for <i>Ca. Accumulibacter</i> | | | | |
| 16S rRNA | RHC439 | <i>Rhodococcus/ Ca. Accumulibacter</i> | CNATTCTCCCGCCGA | [16] |
| | RHC175 | Most <i>Rhodocyclaceae</i> | TGCTCACAGAATATGCGG | [16] |
| | PAO462 | Most <i>Ca. Accumulibacter</i> | CCGTCATCTACWCAGGGTATTAAAC | [17] |
| | PAO651 | Most <i>Ca. Accumulibacter</i> | CCCTCTGCCAAACTCCAG | [17] |
| | PAO846 | Most <i>Ca. Accumulibacter</i> | GTTAGCTACGGCACTAAAAGG | [17] |
| | Acc470 | <i>Ca. Accumulibacter aalborgensis</i> and <i>delftensis</i> | TTGGGTACCGTCATCTACTCAGG | [18] |
| | Acc635 | <i>Ca. Accumulibacter regalis</i> | AACTCCAGCCTGGCAGTCTCAAAT | [18] |
| | Acc471 | <i>Ca. Accumulibacter affinis</i> and <i>proximus</i> | CTCCAGGTACCGTCATCTACACAG | [18] |
| | Acc469 | <i>Ca. Accumulibacter proximus</i> | CCAGGTACCGTCATCTACACAGGC | [18] |
| | Acc1011 | <i>Ca. Accumulibacter propinquus</i> | GCGAGCACTCCCAGATCTCTC | [18] |
| PCR primers for <i>Ca. Accumulibacter</i> | | | | |
| ppk1 | IPCRppk-0289R | Type I ppk | ACGATATCTGAATCTCGGCTAATAT | [19] |
| | IPCRppk-1375F | Type I ppk | TTCCACCATTTCGTTGGCATGT | [19] |
| | Nppk1-F | Type I ppk | GGTGGTCTATGACACGCCGCCACA | [19] |
| | Nppk1-R | Type I ppk | GGTGGTCTCTCCGCATCCTGCAGAGTCCTCAGCAGTCAGGT | [19] |
| | ACCppk1-254-F | <i>Ca. Accumulibacter</i> -ppk1 cluster | TCACCACCGACGGCAAGAC | [20] |
| | ACCppk1-1376-R | <i>Ca. Accumulibacter</i> -ppk1 cluster | ACGATCATCAGCATCTTGGC | [20] |
| | Acc-I-444 | Clade IA and other Type I clades | CCCAAGCAATTCTCCCC | [21] |
| | Acc-II-444 | Clade IIA, IIC, and IID | CCCGTGCAATTCTCCCC | [21] |
| | Acc-ppk1-763-F | Acc-I ppk1 | GACGAAGAACGGTCAAG | [22] |
| | Acc-ppk1-1170-R | Acc-I ppk1 | AACGGTCATCTTGATGGC | [22] |
| | Acc-ppk1-893-F | Acc-IIA ppk1 | AGTTCAATCTACCGAGAGC | [22] |
| | Acc-ppk1-997-R | Acc-IIA ppk1 | GGAACCTCAGGTCGTTGC | [22] |
| | Acc-ppk1-870-F | Acc-IIB ppk1 | GATGACCCAGTCCCTGCTCG | [22] |
| | Acc-ppk1-1002-R | Acc-IIB ppk1 | CGGCACGAACCTCAGATCG | [22] |
| | Acc-ppk1-254-F | Acc-IIC ppk1 | TCACCACCGACGGCAAGAC | [22] |
| | Acc-ppk1-460-R | Acc-IIC ppk1 | CCGGCATGACTTCGCGGAAG | [22] |
| | Acc-ppk1-1123-F | Acc-IIC ppk1 excluding OTU NS D3 | GAACAGTCCGCCAACGACC | [22] |
| | Acc-ppk1-1376-R | Acc-IIC ppk1 excluding OTU NS D3 | ACGATCATCAGCATCTTGGC | [22] |
| | Acc-ppk1-375-F | Acc-IID ppk1 | GGGTATCCGTTCCCTAACGCC | [22] |
| | Acc-ppk1-522-R | Acc-IID ppk1 | GAGGCTTGTGAGTACACCG | [22] |
| FISH Probes for <i>Tetrasphaera</i> | | | | |
| 16S rRNA | HGC69a | Phylum <i>Actinobacteria</i> | TATAGTTACCAACGCCGT | [8] |
| | HGC236 | <i>Actinobacteria</i> | AACAAGCTGATAGGCCGC | [23] |
| | Actino-1011 | <i>T. japonica</i> | TTGCGGGGCACCCATCTCT | [23] |
| | Actino-221 | Actinobacterial PAO | CGCAGGTCCATCCCAAGAC | [23] |
| | Actino-658 | Actinobacterial PAO | TCCGGTCTCCCTACCAT | [23] |
| | NLIMII175 | <i>T. jenkinsii</i> , <i>T. veronensis</i> , and <i>T. vanveeni</i> | GGCTCCGCTCTCGTATCCG | [24] |
| | Tet1-266 | Tetrasphaera clade 1 | CCCGTCGTCGCTGTAGC | [25] |
| | Tet2-892 | Tetrasphaera clade 2 | TAGTTAGCCTTGCAGCCG | [25] |
| | Tet2-174 | Tetrasphaera clade 2 | GCTCCGTCCTCGTATCCGG | [25] |
| | Tet3-654 | Tetrasphaera clade 3 | GGTCTCCCTACCAACT | [25] |
| 16S rRNA | Tetra183 | Most off Tetrasphaera-related genera | TAGAGATGCCCTCCGTCTC | [26] |
| | Tetra67 | <i>P. elongata</i> and <i>T. japonica</i> | AGCAAGCTCCGTCACCG | [26] |
| FISH Probes for <i>Ca. Phosphoribacter</i> | | | | |
| 23S rRNA | Phos1260-23S-Pbr1 | <i>Ca. Phosphoribacter baldrii</i> | AGAGTTCACGGCCGGCAAAG | [26] |
| | Phos1260-23S-Pbr2 | <i>Ca. Phosphoribacter hodrii</i> | AGACATCACGGCCGGCAATG | [26] |
| | Phos741 | <i>Ca. Phosphoribacter</i> (Pbr4, Pbr5, Pbro) | TTCTCAGCGTCAGTTGTGGCC | [26] |
| 16S rRNA | Phos601 | midas_s_5 (<i>Ca. P. hodrii</i>) | GGTTGAGCCTCGGATTTCACTGC | [26] |
| | Actino-658 | <i>Ca. Phosphoribacter</i> (<i>Ca. P. hodrii</i> , <i>Ca. P. baldrii</i> , Pbr3) | TCCGGTCTCCCTACCAT | [26] |
| FISH Probes for <i>Dechloromonas</i> | | | | |
| 16S rRNA | Dech443 | <i>Ca. Dechloromonas phosphorivorans</i> | ACC CAT GCA TTT TCT TCC CGG | [27] |
| | BET135 | <i>Ca. Dechloromonas phosphoritropha</i> | ACGTTATCCCCACTCAATGG | [27] |

Table S7. Different phylogenetic taxonomies of species in *Ca. Accumulibacter* and their FISH probes. Metagenome Assembled Genome (MAG) identifier as shown in NCBI and **Figure 2**, MiDAS4-classification, and new names proposed by Petriglieri and colleagues (Petriglieri et al., 2022). “-”: not applicable. FISH probe sequences are available in **Table S6**.

| MAG identifier | MiDAS4-classification | Proposed species name | FISH probe |
|---------------------|-----------------------|-----------------------|-------------------|
| IB | | | |
| BA-92 | - | appositus BA-92 | - |
| phosphatis HKU1 | - | adiacens HKU1 | - |
| IC | | | |
| UW-LDO | - | melophilus UW-LDO | - |
| delftensis | aalborgensis | delftensis | Acc470 |
| phosphatis SBR S | - | delftensis SBR S | Acc470 |
| IA | | | |
| BA-93 | - | regalis BA-93 | Acc635 |
| phosphatis CANDO 1 | - | regalis CANDO 1 | Acc635 |
| phosphatis UW3 | - | regalis UW3 | Acc635 |
| UW4 | - | regalis UW4 | Acc635 |
| UW8-POB | - | regalis UW8- POB | Acc635 |
| IIA | | | |
| aalborgensis | aalborgensis | aalborgensis | Acc470 |
| UW9-POB | - | phosphatis UW9-POB | Acc635 |
| phosphatis UW1 | phosphatis | phosphatis UW1 | Acc635 |
| IIB | | | |
| UW5 | - | phosphatis UW5 | Acc635 |
| Hade BATAC.726 | phosphatis | propinquus | Acc1011 |
| OdNE BAT3C.415 | phosphatis | propinquus | Acc1011 |
| Fred MAXAC.027 | phosphatis | propinquus | Acc1011 |
| UW10-POB | - | propinquus UW10-POB | Acc1011 |
| phosphatis SBR L | - | contiguous SBR L | - |
| IIC | | | |
| SK01 | - | vicinus SK-01 | - |
| phosphatis EUBA5574 | - | vicinus UBA5574 | - |
| hosphatis Bin19 | - | cognatus Bin19 | - |
| phosphatis HKU2 | - | cognatusHKU2 | - |
| SK-02 | - | cognatus SK-02 | - |
| phosphatis SSA1 | - | cognatusSSA1 | - |
| UW11-POB | - | cognatus UW11-POB | - |
| UW6 | - | cognatus UW6 | - |
| IIG | | | |
| Fred II3C.720 | phosphatis | affinis | Acc471 |
| IID | | | |
| EsbW BATAC.285 | phosphatis | proximus | Acc469 and Acc471 |
| UW12-POB | - | necessaries UW12-POB | - |
| IIF | | | |
| BUBA2327 | midas s 12920 | iunctus UBA2327 | Acc471 2 |
| SK-12 | - | adjunctus SK-12 | - |
| SCELSE-1 | - | similis SCELSE-1 | Acc471 2 |
| phoshatis SSB1 | midas s 12920 | similis SSB1 | Acc471 2 |
| UW13-POB | - | Conexus UW13-POB | - |
| UW7 | - | conexus UW7 | - |

Table S8. Complete gene names and KEGG nomenclature.

| ANNOTATION | KEGG | GENE |
|--|--------|------------------|
| Denitrification | | |
| Nitrate Reductase | K00370 | narG- narZ- nxrA |
| Nitrate Reductase | K00371 | narH- narY- nxrB |
| Nitrate Reductase Gamma Subunit | K00374 | narI- narV |
| Nitrate Reductase (Cytochrome) | K02567 | napA |
| Nitrate Reductase (Cytochrome)-Electron Transfer Subunit | K02568 | napB |
| Nitrite Reductase (NO-Forming) | K00368 | nirK |
| Nitrite Reductase (NO-Forming)/Hydroxylamine Reductase | K15864 | nirS |
| Nitric Oxide Reductase Subunit B | K04561 | norB |
| Nitric Oxide Reductase Subunit C | K02305 | norC |
| Nitrous-Oxide Reductase | K00376 | nosZ |
| Assimilatory Nitrate Reduction | | |
| Ferredoxin-Nitrate Reductase | K00367 | narB |
| Nitrate Reductase (NADPH) | K10534 | NR |
| Assimilatory Nitrate Reductase Catalytic Subunit | K00372 | nasA |
| Assimilatory Nitrate Reductase Electron Transfer Subunit | K00360 | nasB |
| Ferredoxin-Nitrite Reductase | K00366 | nirA |
| Nitrite Reductase (NADPH) | K17877 | NIT-6 |
| Dissimilatory Nitrate Reduction | | |
| Nitrite Reductase (NADH) Large Subunit | K00362 | nirB |
| Nitrite Reductase (NADH) Small Subunit | K00363 | nirD |
| Nitrite Reductase (Cytochrome C-552) | K03385 | nrfA |
| Cytochrome C Nitrite Reductase Small Subunit | K15876 | nrfH |
| Nitrogen Fixation | | |
| Nitrogenase Molybdenum-Iron Protein Beta Chain | K02591 | nifK |
| Nitrogenase Molybdenum-Iron Protein Alpha Chain | K02586 | nifD |
| Nitrogenase Iron Protein Nifh | K02588 | nifH |
| P Transport Proteins | | |
| Inorganic Phosphate Transporter, Pit Family | K03306 | pit |
| Phosphate Transport System Substrate-Binding Protein | K02040 | pstS |
| Phosphate Transport System Permease Protein | K02037 | pstC |
| Phosphate Transport System Permease Protein | K02038 | pstA |
| Phosphate Transport System ATP-Binding Protein | K02036 | pstB |
| PolyP Metabolism | | |
| Polyphosphate Kinase | K00937 | ppk1 |
| Polyphosphate Kinase | K22468 | ppk2 |
| Endopolyphosphatase | K06018 | ppn1 |
| Exopolyphosphatase | K01524 | ppx |
| Exopolyphosphatase | K01514 | ppx1 |
| Polyphosphate Glucokinase | K00886 | ppgK |
| AMP-Polyphosphate Phosphotransferase | K23753 | pap |
| Polyhydroxyalkanoates | | |

| | | |
|---|---------------|-------------------------|
| Acetyl-Coa Acetyltransferase | K00626 | phaA |
| Acetoacetyl-Coa Reductase | K03821 | phaB |
| 3-Hydroxybutyryl-Coa Dehydratase | K17865 | phaJ |
| Poly[(R)-3-Hydroxyalkanoate] Polymerase Subunit | K03821 | phaC |
| Poly(3-Hydroxyoctanoate) Depolymerase | K22250 | phaZ |
| Poly(3-Hydroxybutyrate) Depolymerase | K05973 | phaZ_a |
| Poly(3-Hydroxyoctanoate) Depolymerase | K22249 | phaZ_b |

Glycogen Synthesis

| | | |
|---|--------|------|
| Glucose-1-Phosphate Adenylyltransferase | K00975 | glgC |
| Starch Synthase | K00703 | glgA |
| 1,4-Alpha-Glucan Branching Enzyme | K00700 | glgB |
| Alpha-Maltose-1-Phosphate Synthas | K16148 | glgM |
| Starch Synthase (Maltosyl-Transferring) | K16147 | glgE |

Glycogen Degradation

| | | |
|---------------------------------------|--------|------|
| Glycogen Phosphorylase | K00688 | glgP |
| Alpha-Amylase | K01176 | amyA |
| Beta-Amylase | K01177 | amyB |
| Maltogenic Alpha-Amylase | K05992 | amyM |
| Cyclomaltodextrinase / Neopullulanase | K01208 | ma |

Atp Transporters

| | | |
|--|--------|------------------|
| Basic Membrane Protein A And Related Proteins | K07335 | bmpA- bmpB- tmpC |
| Branched-Chain Amino Acid Transport System Atp-Binding Protein | K01996 | livF |
| Branched-Chain Amino Acid Transport System Atp-Binding Protein | K01995 | livG |
| Branched-Chain Amino Acid Transport System Permease Protein | K01997 | livH |
| Branched-Chain Amino Acid Transport System Permease Protein | K01998 | livM |
| Branched-Chain Amino Acid Transport System Substrate-Binding Protein | K01999 | livK |
| Capsular Polysaccharide Transport System Atp-Binding Protein | K09689 | kpsT |
| Capsular Polysaccharide Transport System Permease Protein | K10107 | kpsE |
| Capsular Polysaccharide Transport System Permease Protein | K09688 | kpsM |
| General Nucleoside Transport System Atp-Binding Protein | K23537 | nupA |
| General Nucleoside Transport System Permease Protein | K23535 | nupB |
| General Nucleoside Transport System Permease Protein | K23536 | nupC |
| Glutamate/Aspartate Transportsystematp-Binding Protein | K10004 | aatL- aatP |
| Glutamate/Aspartate Transportsystempermease Protein | K10003 | aatJ- aatQ |
| Glutamate/Aspartate Transportsystempermease Protein | K10002 | aatK- aatM |
| Glutamate/Aspartate Transportsystemssubstrate-Binding Protein | K10001 | aatI- aatJ |
| Glycine Betaine/Proline Transport System Atp-Binding Protein | K02000 | proV |
| Glycine Betaine/Proline Transport System Permease Protein | K02001 | proW |
| Glycine Betaine/Proline Transport System Substrate-Binding Protein | K02002 | proX |
| Lipopolysaccharide Export System Permease Protein | K07091 | lptF |
| Lipopolysaccharide Export System Permease Protein | K11720 | lptG |
| Lipopolysaccharide Transport System Atp-Binding Protein | K09691 | wzt |
| Lipopolysaccharide Transport System Permease Protein | K09690 | wzm |
| Multiple Sugar Transport System Permease | K02026 | msSP1 |

| | | |
|--|--------|------|
| Multiple Sugar Transport System Permease Protein | K02025 | msP |
| Multiple Sugar Transport System Substrate-Binding Protein | K02027 | msS |
| Peptide/Nickel Transport System Atp-Binding Protein | K02031 | ddpD |
| Peptide/Nickel Transport System Atp-Binding Protein | K02032 | ddpF |
| Polar Amino Acid Transport System Atp-Binding Protein | K02028 | paA |
| Polar Amino Acid Transport System Permease Protein | K02029 | paP |
| Polar Amino Acid Transport System Substrate-Binding Protein | K02030 | paS |
| Putative Multiple Sugar Transport System Atp-Binding Protein | K10548 | gguA |
| Putative Multiple Sugar Transport System Permease Protein | K10547 | gguB |
| Putative Multiple Sugar Transport System Substrate-Binding Protein | K10546 | chvE |
| Putrescine Transport System Atp-Binding Protein | K11075 | potG |
| Putrescine Transport System Permease Protein | K11075 | potH |
| Putrescine Transport System Permease Protein | K11074 | potI |
| Putrescine Transport System Substrate-Binding Protein | K11073 | potF |
| Rhamnose Transport System Atp-Binding Protein | K10562 | rhaT |
| Rhamnose Transport System Permease Protein | K10560 | rhaP |
| Rhamnose Transport System Permease Protein | K10561 | rhaQ |
| Rhamnose Transport System Substrate-Binding Protein | K10559 | rhaS |
| Sugar Transport System Atp-Binding Protein | K23547 | yphD |
| Sugar Transport System Permease Protein | K23546 | yphD |
| Sugar Transport System Substrate-Binding Protein | K23545 | yphF |

Electrochemical Potential-Driven Transporters

| | | |
|--|--------|------|
| Anaerobic C4-Dicarboxylate Transporter | K07791 | dcuA |
| Anaerobic C4-Dicarboxylate Transporter | K07792 | dcuB |
| Anaerobic C4-Dicarboxylate Transporter | K11103 | dctA |
| Benzoate-Proton Symporter | K05548 | benK |
| Branched Chain Amino Acid Efflux Pump | K05846 | azlC |
| Branched Chain Amino Acid Efflux Pump | K05847 | azlD |
| C4-Dicarboxylate Transporter | K11689 | dctQ |
| C4-Dicarboxylate Transporter | K11690 | dctM |
| C4-Dicarboxylate-Binding Protein | K11688 | dctP |
| Cation-Acetate Symporter | K14393 | actP |
| Cytosine Permease | K10974 | codB |
| Ethanolamine Permease | K16238 | eat |
| Fucose-Proton Symporter | K02429 | fucP |
| Galactose-Proton Symporter | K08137 | galP |
| Glutamate-Na ⁺ Symporter | K03312 | gltS |
| Glycine Betaine | K05020 | opuD |
| L-Asparagine Permease | K11738 | ansP |
| Lactate-Proton Symport | K08178 | jen |
| Malonate-Proton Symporter | K13936 | mdcF |
| Maltose-Moltooligosaccharide Transporter | K16211 | malY |
| Oxalate-Formate Antiporter | K08177 | oxiT |
| Proline-Betaine-Proton Symporter | K03762 | proP |
| Putrescine Importer | K14052 | puuP |
| Tartrate Transporter | K13021 | ttuB |

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