

566 **Supplemental Figure Captions**

567 **Table S1.** Data collection and refinement statistics for the inosine and m⁶A antibody crystal
568 structures.

569
570 **Table S2.** Complete table of λ -dynamics results for inosine antibody screening with the RNA
571 library. RNA chemical structures available in **Fig S3**. Relative binding free energy, $\Delta\Delta G_{\text{bind}}$.
572 Standard deviation, $\pm\sigma$. Unbound, u.b. Not specified due to bad sampling, n.s. Entries
573 corresponding to favorable modifications ($\Delta\Delta G_{\text{bind}} \leq -0.7$ kcal/mol) are emphasized in bold italics.
574 Patch name from Xu et al., 2016.

575
576 **Table S3.** Complete table of λ -dynamics results for m⁶A antibody screening with the RNA library.
577 RNA chemical structures available in **Fig S3**. Relative binding free energy, $\Delta\Delta G_{\text{bind}}$. Standard
578 deviation, $\pm\sigma$. Unbound, u.b. Not specified due to bad sampling, n.s. Entries corresponding to
579 favorable modifications ($\Delta\Delta G_{\text{bind}} \leq -0.7$ kcal/mol) are emphasized in bold italics. Patch name from
580 Xu et al., 2016.

581
582 **Fig S1.** A previously published poly-inosine antibody has a large binding pocket that may
583 accommodate multiple nucleobases. Overview (left) and magnified image (right) of the poly-
584 inosine antibody fragment (PDB ID: 1MRD) binding pocket. An inosine mononucleotide (orange)
585 was modeled into the missing ligand density (green). Heavy chain residues (H) in dark blue and
586 light chain residues (L) in light blue. Water molecules substituting for the potential second
587 mononucleotide are depicted as red spheres, indicating the potential space to bind a second
588 nucleobase. Interacting amino acids include heavy chain residue Arg96 and light chain residues
589 Asn28, Asn30, Tyr32, Lys50, and Ser91. The extended binding pocket (red arrow) includes light
590 chain residue Arg96 and heavy chain residues Gln35, Trp47, Glu50, and Asn58.

591
592 **Fig S2.** Crystal structure of the m⁶A Fab apo- form to 2.05 Å (PDB ID: 8TCA). Critical binding
593 pocket amino acids discussed in the main text are labeled. Heavy chain residues (H) are
594 represented in dark blue, light chain residues (L) in light blue, and waters as red spheres. Depicted
595 binding pocket amino acids match those of the m⁶A Fab holo- form (**Fig 1A**).

596
597 **Fig S3.** Chemical library of ribonucleoside bases. The library includes the 4 canonical
598 ribonucleobases (A, C, G, and U) and 44 naturally occurring modified derivatives (12 As, 6 Cs, 8
599 Gs, and 18 Us). Differences between each modification and its respective canonical base are
600 highlighted in green.

601
602 **Fig S4.** Molecular dynamics simulation movie example of the m⁶A antibody with a bound
603 nucleoside target. The m⁶A Fab binds tightly to m⁶₂A. Movie made in Pymol (Schrödinger, Inc.).

604
605 **Fig S5.** Molecular dynamics simulation movie example of the m⁶A antibody with an unbinding
606 nucleoside target. The m⁶A Fab unbinds from uridine. Movie made in Pymol (Schrödinger, Inc.).

607
608 **Fig S6.** Structural models of inosine and m⁶A antibodies bound to representative off-target RNAs.
609 (A) Magnified binding site of the inosine antibody fragment in complex with uridine. (B-C)
610 Magnified binding site of the m⁶A antibody fragment in complex with (B) m⁶₂A or (C) adenosine

611 (A). Heavy chain residues (H) are represented in dark blue, light chain residues (L) in light blue,
612 and the off-target nucleoside in orange. Critical amino acid contacts labeled. **(D-E)** Table of t-test
613 p-value statistics for **(D)** inosine and **(E)** m⁶A antibody ELISA binding assay results reported in
614 **Fig 5**. p-values < 0.01 in bold.

Table S1. Data collection and refinement statistics.

	Inosine Fab w/ Inosine (PDB ID: 8SIP)	m6A Fab only (PDB ID: 8TCA)	m6A Fab w ligand (PDB ID: 8VEV)
Wavelength	0.9793	0.9793	0.8731
Resolution range	57.04 - 1.94 (2.009 - 1.94)	70 - 2.02 (2.092 - 2.02)	48.23 - 3.06 (3.18 - 3.06)
Space group	P 1	P 43 21 2	P 21 21 21
Unit cell	39.8174 49.0903 57.3853 83.8419 88.8169 89.6813	79.906 79.906 145.127 90 90 90	83.64 128.377 150.476 90 90 90
Total reflections	218254 (14586)	230543 (22581)	203373 (23011)
Unique reflections	30888 (2931)	31198 (2999)	31244 (3399)
Multiplicity	7.1 (4.9)	7.4 (7.3)	6.5 (6.8)
Completeness (%)	96.20 (91.25)	98.24 (96.71)	99.90 (100.00)
Mean I/sigma(I)	6.33 (3.17)	9.71 (1.08)	16.80 (4.00)
Wilson B-factor	26.61	27.17	78.19
R-merge	0.1901 (1.506)	0.2183 (2.290)	0.078 (0.416)
R-meas	0.2027 (1.642)	0.2341 (2.466)	0.085 (0.451)
R-pim	0.06938 (0.6427)	0.08188 (0.8885)	0.0330 (0.1720)
CC1/2	0.991 (0.586)	0.996 (0.546)	0.998 (0.963)
CC*	0.998 (0.86)	0.999 (0.84)	1.000 (0.990)
Reflections used in refinement	30804 (2931)	31042 (2999)	31207 (3391)
Reflections used for R-free	1554 (154)	1543 (162)	1307 (142)
R-work	0.2043 (0.2460)	0.1807 (0.3043)	0.2238 (0.3156)
R-free	0.2454 (0.2914)	0.2238 (0.3201)	0.2531 (0.3100)

CC(work)	0.947 (0.722)	0.968 (0.777)	0.928 (0.846)
CC(free)	0.929 (0.494)	0.952 (0.735)	0.896 (0.865)
Number of non-hydrogen atoms	3531	3562	9807
macromolecules	3277	3239	9649
ligands	19	36	108
solvent	235	287	50
Protein residues	426	425	1267
RMS(bonds)	0.008	0.009	0.002
RMS(angles)	1.10	1.18	0.56
Ramachandran favored (%)	97.62	97.61	95.34
Ramachandran allowed (%)	2.38	2.39	4.58
Ramachandran outliers (%)	0.00	0.00	0.08
Rotamer outliers (%)	0.80	0.54	0.09
Clashscore	6.46	1.70	2.09
Average B-factor	32.66	33.80	91.07
macromolecules	32.19	33.22	91.26
ligands	32.70	53.08	81.28
solvent	39.15	37.94	75.58
Number of TLS groups	1	9	6

Statistics for the highest-resolution shell are shown in parentheses.

Table S2

Table S2: Relative binding free energies for inosine Fab screening.

Modified Base	Patch Name	$\Delta\Delta G_{\text{bind}}$	$\pm \sigma$
A	ADE	-0.112	0.172
m^2A	2MA	u.b.	u.b.
m^6A	6MA	1.726	0.186
m^6_2A	M6A	0.605	0.231
m^8A	8MA	2.721	0.389
m^1I	1MI	0.783	0.138
I	INO	0.089	0.024
ms^2m^6A	SMA	-0.037	0.335
ac^6A	6AA	u.b.	u.b.
i^6A	6IA	n.s.	n.s.
ms^2i^6A	MIA	0.598	0.352
ms^2io^6A	SIA	u.b.	u.b.
io^6A	HIA	1.868	0.389
G	GUA	-0.642	0.134
m^1G	1MG	u.b.	u.b.
m^2G	2MG	0.286	0.168
m^2_2G	M2G	1.057	0.262
preQ0	DCG	-1.737	0.187
<i>imG-14</i>	DWG	-0.005	0.319
<i>imG</i>	IMG	u.b.	u.b.
imG2	IWG	-1.164	0.331
<i>mimG</i>	MWG	u.b.	u.b.
U	URA	-0.926	0.145
D	H2U	u.b.	u.b.
mo⁵U	MOU	-1.656	0.199
m^5s^2U	52U	u.b.	u.b.
m^5D	MDU	0.707	0.161
ψ	PSU	-1.985	0.490
$m^3\psi$	3MP	-1.174	0.446
m^3U	3MU	-0.146	0.320
s^4U	4SU	-1.702	0.181
m^5U	5MU	-1.340	0.159
ho⁵U	5HU	-1.613	0.162
s^2U	2SU	-0.644	0.233
$m^1\psi$	1MP	-0.896	0.112
cnm⁵U	CYU	-2.147	0.233
mcm⁵s²U	70U	-1.379	0.330
<i>mchm⁵U</i>	CMU	-0.612	0.381
ncm⁵U	BCU	-1.683	0.393
mcm⁵U	OCU	-1.307	0.312
<i>mcmo⁵U</i>	OEU	-0.491	0.396
C	CYT	u.b.	u.b.
m^5C	5MC	u.b.	u.b.
ac^4C	4AC	u.b.	u.b.
m^4C	4MC	u.b.	u.b.
f^5C	5FC	0.596	0.265
hm^5C	HMC	u.b.	u.b.
s^2C	2SC	u.b.	u.b.

"Patch Name" = 3-letter name assigned by Xu et al. (2016)

Modifications with $\Delta\Delta G \leq -0.7$ kcal/mol in bold italics.

Table S3

Table S3: Relative binding free energies for m⁶A Fab screening.

Modified Base	Patch Name	$\Delta\Delta G_{\text{bind}}$	$\pm \sigma$
A	ADE	0.300	0.165
<i>m</i> ² A	2MA	3.084	0.156
<i>m</i> ⁶ A	6MA	-0.034	0.032
<i>m</i>⁶₂A	M6A	-2.091	0.052
<i>m</i> ⁸ A	8MA	4.432	0.337
<i>m</i> ¹ I	1MI	n.s.	n.s.
I	INO	6.247	0.301
<i>ms</i> ² <i>m</i> ⁶ A	SMA	2.634	0.209
<i>ac</i>⁶A	6AA	-1.156	0.197
<i>i</i> ⁶ A	6IA	1.624	0.491
<i>ms</i> ² <i>i</i> ⁶ A	MIA	3.587	0.434
<i>ms</i> ² <i>io</i> ⁶ A	SIA	n.s.	n.s.
<i>io</i> ⁶ A	HIA	n.s.	n.s.
G	GUA	4.881	0.471
<i>m</i> ¹ G	1MG	3.029	0.521
<i>m</i> ² G	2MG	u.b.	u.b.
<i>m</i> ² ₂ G	M2G	n.s.	n.s.
<i>preQ0</i>	DCG	4.229	0.283
<i>imG-14</i>	DWG	u.b.	u.b.
<i>imG</i>	IMG	n.s.	n.s.
<i>imG2</i>	IWG	u.b.	u.b.
<i>mimG</i>	MWG	-2.437	0.441
U	URA	u.b.	u.b.
D	H2U	2.841	0.242
<i>mo</i> ⁵ U	MOU	1.861	0.459
<i>m</i> ⁵ <i>s</i> ² U	52U	3.817	0.504
<i>m</i> ⁵ D	MDU	1.520	0.25
ψ	PSU	3.564	0.531
<i>m</i> ³ ψ	3MP	n.s.	n.s.
<i>m</i> ³ U	3MU	n.s.	n.s.
<i>s</i> ⁴ U	4SU	n.s.	n.s.
<i>m</i> ⁵ U	5MU	1.270	0.519
<i>ho</i> ⁵ U	5HU	0.965	0.305
<i>s</i> ² U	2SU	2.830	0.705
<i>m</i> ¹ ψ	1MP	3.044	0.663
<i>cnm</i> ⁵ U	CYU	2.396	0.282
<i>mcm</i> ⁵ <i>s</i> ² U	70U	n.s.	n.s.
<i>mchm</i> ⁵ U	CMU	2.686	0.235
<i>ncm</i> ⁵ U	BCU	n.s.	n.s.
<i>mcm</i> ⁵ U	OCU	1.113	0.402
<i>mcmo</i> ⁵ U	OEU	3.469	0.527
C	CYT	5.033	0.708
<i>m</i> ⁵ C	5MC	n.s.	n.s.
<i>ac</i>⁴C	4AC	-1.199	0.325
<i>m</i> ⁴ C	4MC	2.159	0.287
<i>f</i> ⁵ C	5FC	2.275	0.422
<i>hm</i> ⁵ C	HMC	u.b.	u.b.
<i>s</i> ² C	2SC	u.b.	u.b.

"Patch Name" = 3-letter name assigned by Xu et al. (2016)

Modifications with $\Delta\Delta G \leq -0.7$ kcal/mol in bold italics.

Figure S1

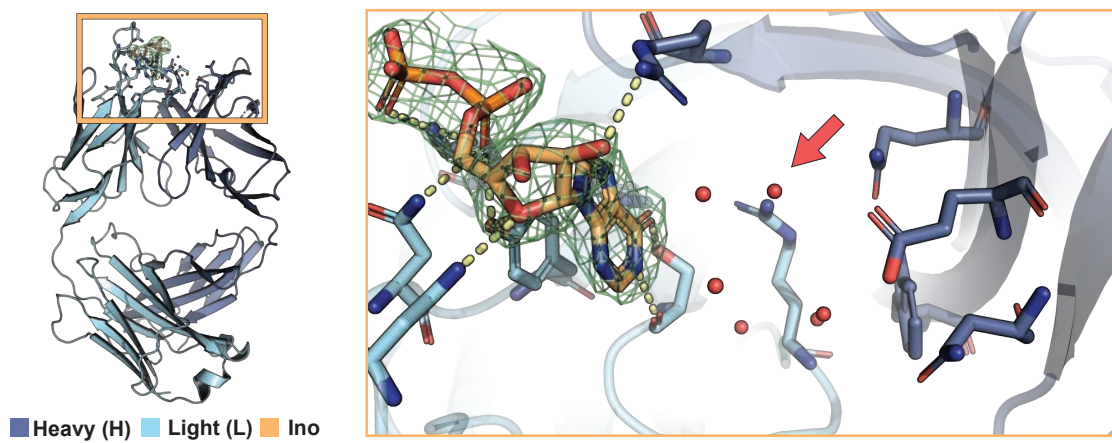


Figure S1

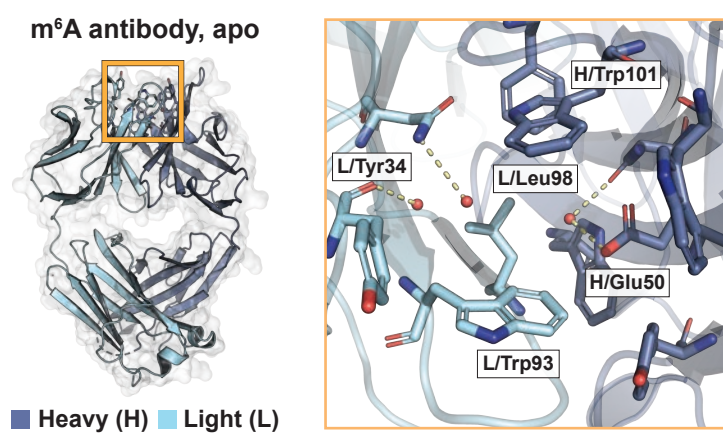


Figure S3

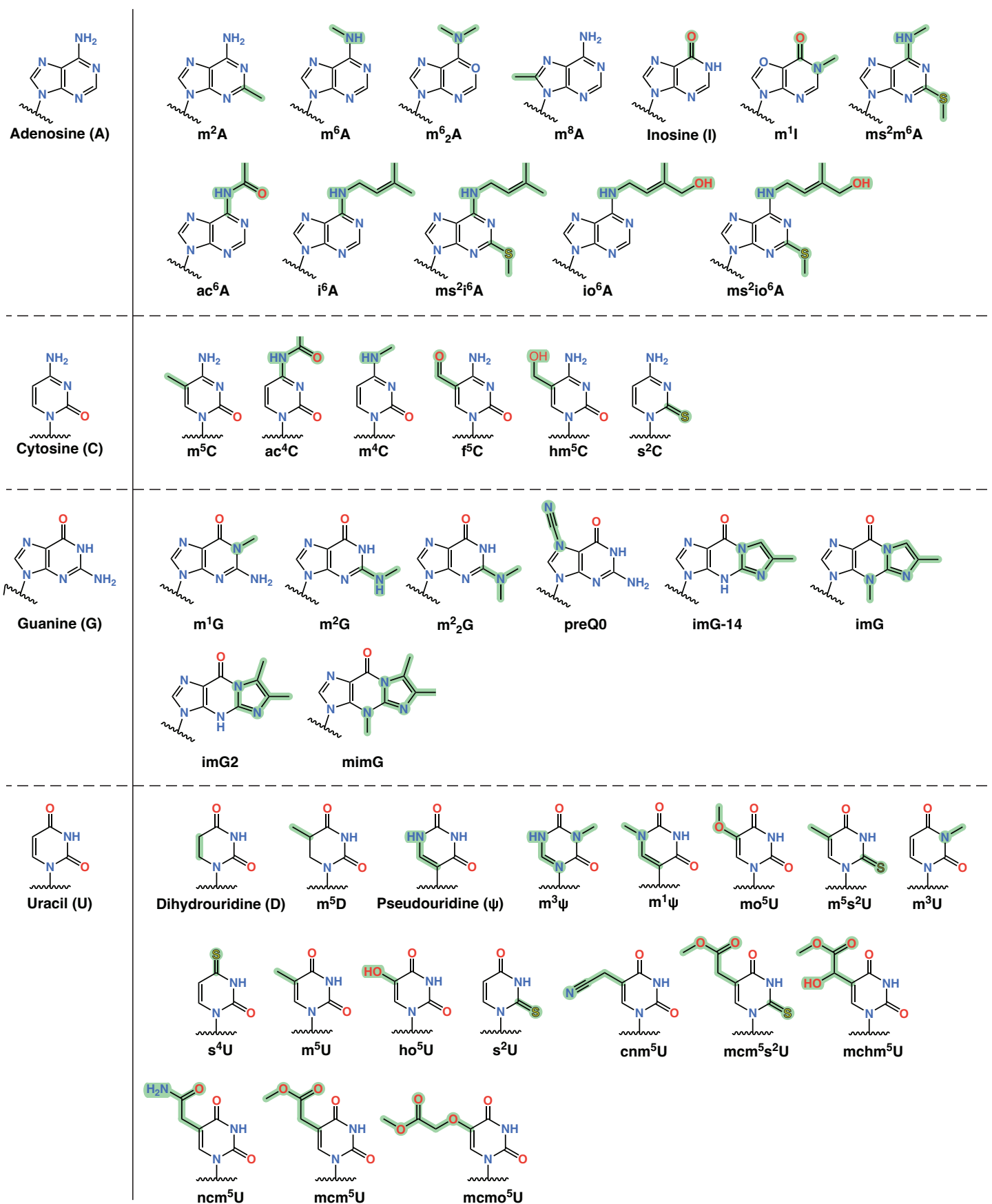
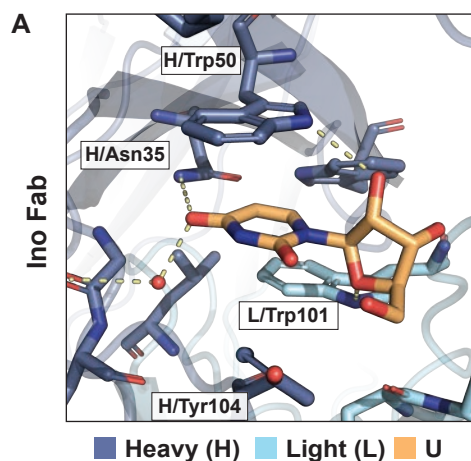
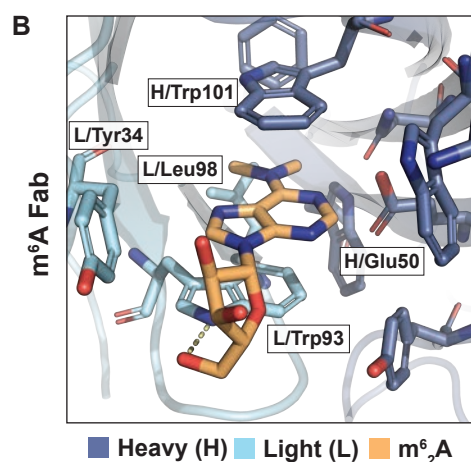


Figure S6



D Inosine antibody ELISA results, t-test p values. $p < 0.01$ in bold italics.

Concentration (ng/ml)	I vs. C	U vs. C	A vs. C	I vs. U
1000	<i>0.008176</i>	<i>0.000068</i>	0.231657	0.022235
100	<i>0.005824</i>	<i>0.000199</i>	0.371234	<i>0.006713</i>
10	<i>0.000112</i>	0.079051	0.703358	<i>0.000115</i>
1	<i>0.000376</i>	0.828319	0.872158	<i>0.000401</i>
0.1000	0.030556	0.608653	0.650652	0.032961
0.0100	0.289681	0.795255	>0.999999	0.284906
0.0010	0.390739	0.366411	0.421648	0.507010
0	0.593139	0.350393	0.507644	0.440630



E m⁶A antibody ELISA results, t-test p values. $p < 0.01$ in bold italics.

Concentration (ng/ml)	m ⁶ A vs. C	A vs. C	m ⁶ ₂ A vs. C	m ⁶ A vs. A
100	<i>0.00007</i>	<i>0.001121</i>	<i>0.000043</i>	<i>0.004438</i>
33.300	<i>0.000017</i>	<i>0.000294</i>	<i>0.000005</i>	<i>0.001808</i>
11.100	<i>0.000034</i>	<i>0.000209</i>	<i>0.000024</i>	<i>0.005227</i>
3.700	<i>0.000009</i>	<i>0.000072</i>	<i>0.000017</i>	<i>0.002905</i>
1.230	<i>0.000055</i>	<i>0.001183</i>	<i>0.001946</i>	<i>0.000026</i>
0.412	0.086944	0.342808	0.080748	0.239781
0.137	0.723367	0.72624	0.506021	0.974438
0	0.799706	0.959434	0.884815	0.781924

