ANDROGEN DRIVES MELANOMA INVASIVENESS AND METASTATIC SPREAD BY INDUCING TUMORIGENIC FUCOSYLATION

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Supplementary Fig. 1: AR⁺ melanoma cells are responsive to androgen stimulation. (a) Immunoblotting (IB) analysis of baseline AR protein levels across 11 human melanoma cell lines. LNCaP prostate cancer cell line serves as a positive control for AR expression. Uncropped blots in Source Data. (b) IB showing the induction of AR protein in WM1366 (upper left), IPC298 (upper right), and A375 (lower left) cells treated ± 100nM DHT over 96hrs. Uncropped blots in Source Data. (c) Subcellular fractionation IB of AR protein in WM1366 and IPC298 cells treated ± 100nM DHT for 48hrs. Lamin C and tubulin indicate nuclear (Nuc) and cytoplasmic (Cyto) fractions, respectively. Uncropped blots in Source Data. (d) IB showing lack of AR expression and induction in AR melanoma cells (LU1205 and WM266-4) treated ± 100nM DHT over 96hrs. Uncropped blots in Source Data. (e) BrdU staining of WM1366 cells treated ± 100nM DHT for 48hrs (CTL, n=16 fields; DHT, n=18 fields examined over 3 independent experiments). (f) Representative brightfield images for scratch migration assay of WM793 cells (upper left; corresponding to Fig. 1g, right). Representative brightfield images and quantification for WM1366 (upper right; CTL, n=23 scratches; DHT, n=20 scratches examined over 2 independent experiments) and IPC298 (lower left; n=24 scratches examined over 3 independent experiments) cells treated ± 100nM DHT for 48hrs. Scale bar=400µm. (g) MTT assay of LU1205 cells cultured in 10% FBS or 10% CSS ± 100nM DHT for 4 days (10% FBS, n=12; CTL, n=10; DHT, n=12 biologically independently samples). ns, not significant. (h) The growth curve of SM1 tumors subcutaneously implanted in CTL or castrated C57BL/6 male mice. Mice were castrated at 1.5 weeks prior to injection (CTL, n=3 mice; Castration, n=5 mice). For (e)-(h), data are presented as mean values ± standard error of the mean (SEM) and *p*-values are calculated by two-sided Student's t-test. Source data are provided as a Source Data file.



Supplementary Fig. 2: AR transcriptionally upregulates *FUT4*, mediating tumorigenic fucosylation in melanomas. (a) Schematic diagram of fucose salvage pathway: GDP-fucose, the global substrate for cellular protein fucosylation, is transported into ER/Golgi via SLC35C1/2 transporters, where fucose moieties are conjugated onto target proteins by 13 FUTs. FUK, FPGT, and SLC35C1/2 regulate the availability of global substrate (GDP-fucose), whereas the FUTs determine the tumor-promoting vs. tumor-suppressive subtypes of fucosylation (*the schematic was created using BioRender*). qRT-PCR of *FUT4* levels in (b) WM793 cells cultured in 10% CSS or 10% FBS for 48hrs (n=3 independent experiments), or in (c), A375 cells treated \pm 100nM DHT for 6, 8, and 24hrs (n=3 independent experiments). (d) qRT-PCR analysis of *AR* and *FUT4* levels in shNT/shAR A375 cells (n=4 independent experiments). α -(1,3)-FUTs are indicated with green font. (f) qRT-PCR assessment of classical prostate cancer AR targets (*PSA*, *TMPRSS2*, *FKBP5*, and *KLK2*) in WM793 cells treated \pm 100nM DHT for 4, 8, and 24hrs (n=4 independent experiments). For (b)-(f), data are presented as mean values \pm SEM and *p*-values are calculated by two-sided Student's t-test. Source data are provided as a Source Data file.

Supplementary Figure 3



Supplementary Fig. 3: AR-FUT4 axis alters cellular adhesion signaling in melanoma cells. (a) (left) IB validation of FUT4 overexpression in WM793, WM1366, and A375 cells. *: non-specific band. Uncropped blots in Source Data. (right) IF staining of FUT4-fucosylated epitope (CD15) in EV/FUT4-OE A375 cells (EV, n=21 fields; FUT4-OE, n=19 fields examined over 3 independent experiments). Scale bar=50µm. (b) IB analysis of CD15, CD15s, and VIM2 in EV/FUT4-OE A375 cells. Uncropped blots in Source Data. (c) qRT-PCR of other FUTs in EV/FUT4-OE WM793 cells (n=3 independent experiments). The data confirms the overexpression of FUT4 in the indicated cell line and shows that FUT4 overexpression does not impact the expression of other FUTs. α -(1,3)-FUTs are indicated with green font. (d) Inclusion criteria of AR-FUT4-up/downregulated protein hits overlapped between WM793 and WM1366 cells for Ingenuity Pathway Analysis (IPA). (e) qRT-PCR validation of FUT4 knockdown in WM793 (n=4 independent experiments) and WM1366 (n=3 independent experiments) cells. Arrows indicate shRNAs that were selected based on knockdown efficiency for functional experiments in the remainder of the study. (f) Secondary antibody-only control for PLA staining. Scale bar=50µm. PLA staining evaluating the interaction between N-cadherin and β-catenin proteins in (g) EV/FUT4-OE A375 cells (EV, n=19 fields; FUT4-OE, n=18 fields examined over 3 independent experiments), (h) parental WM793 cells treated ± 250µM 2FF for 3 days (n=21 fields examined over 3 independent experiments), (i) (left) parental A375 cells treated ± 10µM ARi for 48hrs (CTL, n=18 fields; ARi, n=21 fields examined over 3 independent experiments), (middle) shNT/shAR A375 cells (n=20 fields examined over 3 independent experiments), and (right) parental WM793 cells treated ± 100nM DHT for 48hrs (CTL, n=11 fields; DHT, n=8 fields examined over 1 independent experiment. Two other independent experiments in Source Data). For (g)-(i), scale bar=50µm. For (a), (c), (e), (g)-(i), data are presented as mean values ± SEM and *p*-values are calculated by two-sided Student's t-test. Source data are provided as a Source Data file.

Supplementary Figure 4



Supplementary Fig. 4: FUT4 is crucial for androgen/AR-stimulated melanoma motility in vitro. (a) XTT assay (5 days) (n=6 biologically independent samples) and (b) clonogenic assay (14 days) of EV/FUT4-OE WM1366 cells treated ± 10µM ARi or ± culture in 10% CSS (representative images are shown for n=3 independent experiments). (c) Representative brightfield images of scratch migration assay for EV/FUT4-OE WM793 cells treated ± 250µM 2FF for 3 days in Fig. 4c. Scale bar=400µm. (d) Scratch migration assay of EV/FUT4-OE WM1366 cells treated ± 10µM ARi (48hrs; left; n=6 scratches examined over 1 independent experiment. Another independent experiment in Source Data) or shNT/sh-FUT4 WM1366 cells treated ± 100nM DHT (48hrs; right; shNT-CTL, n=8 scratches; shNT-DHT, n=7 scratches; shFUT4-CTL/DHT, n=8 scratches examined over 1 independent experiment. Two other independent experiments in Source Data). (e) Representative brightfield images of scratch migration assays for WM793 (upper) and WM1366 (middle) cells modified/treated as in Fig. 4d and Supplementary Fig. 4d. (lower) Representative brightfield images and quantification of scratch migration assay for EV/FUT4-OE A375 cells treated ± 10µM ARi for 48hrs (n=24 scratches examined over 3 independent experiments). Scale bar=400µm. (f) Representative brightfield images and quantification of scratch migration assays for shNT/shAR A375 cells (n=24 scratches examined over 3 independent experiments). Scale bar=400µm. (g) qRT-PCR validation of FUT8 knockdown in EV/FUT4-OE WM793 cells (n=3 independent experiments). (h) Representative images of FITC-gelatin degradation assay in Fig. 4g. Scale bar=50µm. (i) The correlation analysis of FUT4 and MMP2/9 levels in TCGA SKCM cohort. p-values are determined by two-sided correlation test based on Pearson's coefficient. (j) The comparison of FKBP and Probeset intensity of FUTs between primary and metastatic melanomas in TCGA SKCM and GSE8401 cohorts. (k) Bonferroni and FDR tests were applied to the data in (j) for the adjustment of p-values for multiple hypothesis correction. For (a), (d)-(g), (j), data are presented as mean values ± SEM and p-values are calculated by two-sided Student's t-test. Source data are provided as a Source Data file.



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Actin

Actin

ΕV

FUT4-OE

CTL

LL L

AL

DHT

AL

L1CAM

AAL

Tubulin

Ę

p=0.0453

CTL DHT

shFUT4

FUT4-OE

FUT4-OE

2.5

Supplementary Fig. 5: L1CAM is a key downstream effector of AR-FUT4 axis. (a) NetNGlyc and NetOGlyc software predictions of N- and O-glycosylation sites in the L1CAM protein. (b) (upper) Lectin pulldown (LPD) followed by IB analysis of L1CAM protein in (left) shNT/shAR A375 cells and (right) parental WM793 cells treated ± 100nM DHT for 48hrs. Column chart (lower) shows densitometric quantification for the blots (n=3 independent experiments). Uncropped blots in Source Data. (c) LPD followed by IB analysis of L1CAM protein in EV/FUT4-OE A375 cells (n=3 independent experiments). Uncropped blots in Source Data. (d) Secondary antibody-only control (left) and L-fucose washing control (right) for LPLA staining in Fig. 5c. Scale bar=50µm. (e) PLA staining for CD15 and L1CAM protein in (upper) EV/FUT4-OE A375 cells (EV, n=23 fields; FUT4-OE, n=26 fields examined over 3 independent experiments) and (lower) shNT/shAR A375 cells (shNT, n=25 fields; shAR, n=21 fields examined over 3 independent experiments). Scale bar=50µm. (f) Secondary antibody-only control for CD15-L1CAM PLA staining in Fig. 5d and Supplementary Fig. 5e. Scale bar=50µm. (g) Equal amounts of WM793 lysates subjected to deglycosylation with PNGase A/F and O-Glycosidase enzymes. Reduced L1CAM IB signal for PNGase F and O-glycosidases reflect not reduced L1CAM protein levels but rather reduced recognition by the L1CAM antibody. Uncropped blots in Source Data. (h) IB validation of FUT4/L1CAM double-modified WM793 cell lines. Uncropped blots in Source Data. (i) XTT assay of FUT4/L1CAM double-modified WM793 cells treated ± 10µM ARi for 6 days (n=6 biologically independent samples). For (b), (c), (e), (i), data are presented as mean values ± SEM and *p*-values are calculated by two-sided Student's t-test. Source data are provided as a Source Data file.



Supplementary Fig. 6: The activation of AR-FUT4-axis in male melanoma tissues. Total AR protein levels in (a) female vs. male melanomas or (b) primary vs. metastatic melanomas in the melanoma tumor microarray. The correlation analyses of (c) activated AR and fuco-L1CAM (LPLA foci) in early-stage (Stage I-IIA) primary melanomas, of (d) total AR and fuco-L1CAM (LPLA foci) in all stages (*left*) and late-stage (Stage IIB-III; *right*) melanomas. *p*-values are determined by two-sided correlation test based on Pearson's coefficient. For (a) and (b), data are presented as mean values ± SEM and *p*-values are calculated by two-sided Student's t-test. Source data are provided as a Source Data file.



Supplementary Fig. 7: The AR-FUT4 axis promotes accumulation of lung intravascular melanoma colonies *in vivo*. (a) Positive control staining of human melanoma tissues with melanoma marker cocktail (S100 + MART-1). Scale bar=200µm. (b) Representative H&E staining and corresponding IF staining of lung intravascular melanoma colonies in mice harboring FUT4-OE melanoma tumors fed with CTL diet. Scale bar=100µm. Source data are provided as a Source Data file.