Liver elastography in acute liver failure: a pilot study

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Supplementary Methods

Cytokine analysis

The V-PLEX Proinflammatory Panel 1 Human Kit (IFN- γ , IL-1 β , IL-2, IL-4, IL-6, IL-8, IL-10, IL-12p70, IL-13, TNF- α) from Meso Scale Diagnostics, LLC was measures as below.

Heparin plasma samples were 4-fold diluted. 50 μ L of calibrators or sample were added to each well of a V-PLEX plate and incubated for 2 hours on a horizontal orbital microplate shaker at room temperature. Plates were then washed three times (20-fold wash buffer concentrate reconstituted). 25 μ L of detection antibody solution were added to each well and incubated for 2 hours on a horizontal orbital microplate shaker at room temperature. 150 μ L of a 2X Read Buffer T were added to each well and the plate was read with a MSD instrument. The average of the duplicate readings for each standard and sample were analysed and analyte concentrations established fitting a 4-parameter logistic calibration curve.

Enzyme-linked immunosorbent assay (ELISA)

vonWillebrand factor (vWF) and soluble Platelet endothelial cell adhesion molecule-1 (sPECAM1) plasma concentrations were quantified according to the manufacturer's instructions briefly summarised below.

vonWillebrand Factor (Invitrogen, Thermo fisher)

Lithium heparin plasma samples were 8000-fold diluted. 100uL of diluted samples were added to each well, standards were diluted according to the protocol, incubated at room temperature for 2.5 hours on a microplate shaker. The following reagents were added sequentially with a washing passage in between: 100 μ L of prepared biotin conjugate incubated at room temperature for 1 hour, 100 μ L of prepared Streptavidin-HRP solution, incubated for 45 minutes,100 μ L of TMB Substrate, incubated at room temperature for about 45 minutes, avoiding direct exposure to intense light. The enzyme reaction was stopped with 50 μ L of Stop Solution. Absorbance was read with a spectro-photometer using 450 nm. Average of the duplicate readings for each standard and sample were analysed and analyte concentrations established fitting a 3rd order polynomial calibration curve.

Soluble CD31/PECAM (Invitrogen, Thermo fisher)

Lithium heparin plasma samples were 10-fold diluted. 100uLof diluted samples were added to each well, standards were diluted according to the protocol, after adding 50 μ L of HRP-Conjugate to all wells, plate was incubated at room temperature for 3 hours on a microplate shaker. After washing, 100 μ L of TMB Substrate Solution were added to all wells and plate was incubated at room temperature for about 10 minutes, avoiding direct exposure to intense light. The enzyme reaction was stopped by quickly pipetting 100 μ L of Stop Solution. Absorbance was read with a spectro-photometer using 450 nm. Average of the duplicate readings for each standard and sample were analysed and analyte concentrations established fitting a 3rd order polynomial calibration curve.



Supplementary Figure 1. Two cases withs similar features at CT scan.

a) Submassive necrosis with map-like nodular areas intermixed with collapse areas. Liver weight 492g. Complete hepatocellular loss with no underlying fibrosis. LSM =28.7kPa±4.46;
b) Micronodular cirrhosis and pericellular fibrosis (Sirius red). Steatosis, marked Mallory-Denk hyaline, and inflammation (top left). Marked ductular reaction with cholestasis. LSM112kPa±25.



Supplementary Figure 2.

In ACLF, size and stiffness are linearly correlated, (a) Spearman r=0.05401, p=0.0140;

Clinical Case showing CT imaging (b) and histology (c and d) in a patient with Wilson's disease; Liver size (e) measured as liver longitudinal diameter along an oblique line from the right hemi-diaphragm to the inferior tip of the right lobe in midaxillary line and ElastPQ (f) measurement showing average of 174kPa in the same patient.

 g) Receiver operating characteristic (ROC) curve for right liver stiffness measurement (RLSM), spleen size in cm and spleen stiffness measurement (SM) discriminating between
 ALF and ACLF.

*p<0.05, **p<0.01,***p<0.001,****p<0.0001







Supplementary Figure 3.

a) 1), At admission to liver intensive care LSM is not different between patients that spontaneously survive vs patients deceased or transplanted in the following 90 days. 2), At day 7 patients with increased LSM have poor prognosis (Kruskal-Walliss with Dunn's Multiple comparisons test). in both cases values are increased compared to healthy controls (HC). 3), the ratio between one week LSM and baseline is also increased in patients with poor prognosis despite not statistically significant (p=0.0804).

b) Baseline spleen stiffness (1) is not different between patients alive at 90 days (17.22±12kPa) and those who die or were transplanted (24.37±19.6kPa), while at day 7 (2) patients with poor prognosis show increased values (17.45±8.36 vs 36.97±28.48kPa).

*p<0.05, **p<0.01,***p<0.001,****p<0.0001, ns= Not Statistically Significant

c)Correlation matrix showing lack of correlation on day 1 (1) and significant direct correlation of LSM with INR (Spearman r=0.6447, p=0.0029), APTR(r=0.8176,p<0.0001) and fibrinogen (r=-0.5215,p=0.022) on day 7(2) of admission.



Supplementary Figure 4.

a) Model for End-Stage Liver Disease(MELD)-Sodium (Na) was reduced in patients spontaneously survived, Alpha-fetoprotein (AFP0 and the Acute Physiology And Chronic

Health Evaluation (APACHE) II score was not different between the two groups. B) Analysis of liver stiffness measurement (LSM) according to day of illness onset rather than admission to ITU. Acute patients were generally transferred to King's College Hospital on day 2-3 of illness. In the dead/transplant (tx) graph you can see 4 patients were transferred after more than 20 days of illness (subacute liver failure), all of them were transplanted. c) Both curves show LSM in the alive and dead populations (Ordinary two-way ANOVA with Tukey's Multiple comparisons), the difference between the curves is significant at day 8 of disease onset and persists during the illness.



Supplementary Figure 5.

a) Soluble platelet/endothelial cell adhesion molecule (sPECAM-1) was not different
between ACLF and HC, while and von Willebrand factor (vWF) was increased in acute liver
failure syndromes compared to control HC=8, ALF=20, ACLF=12
b) LSM according to Cytoplasmic cholestasis (0,1,2)

d)When ACLF patients were divided according to the LSM, using 50 as cut off, the group with lower LSM showed increase IL-6 (p=0.0054), no difference in Soluble platelet/endothelial cell adhesion molecule (sPECAM-1), von Willebrand factor (vWF) white cell count (WCC), platelets(PLT), C-reactive protein (CRP), Aspartate transaminase (AST), c) Cytokines are increased in both acute liver failure syndromes compared to controls. Same trend is seen in SOFA score

*p<0.05, **p<0.01,***p<0.001,****p<0.0001, ns= Not Statistically Significant

Supplementary Table

Supplementary Table 1									
Study Code	Death within 90 days	Transplant within 90 days	KCC	PLEX	FFP	RBC	Cryo	PLT	
IMET A176	No	No	No	No	No	No	No	No	
IMET A192	No	No	No	No	No	No	No	3	
IMET A205	No	No	No	No	No	No	No	No	
IMET A207	No	No	No	Yes	2	5	4	3	
IMET A213	No	No	No	No	No	2	6	No	
IMET A226	No	No	Yes	Yes	No	3	No	1	
IMET A229	No	No	Yes	Yes	No	14	36	9	
IMET A233	No	No	No	No	No	No	No	No	
IMET A240	No	No	Yes	Yes	No	2	No	No	
IMET A243	No	No	No	Yes	No	No	4	No	
IMET A274	No	No	No	Yes	12	22	6	7	
IMET A286	No	No	No	No	No	No	No	No	
IMET A287	No	No	No	No	No	No	No	No	
IMET A289	No	No	No	No	No	No	No	No	

A322NoNoNoNoNoNo1387IMET A325NoNoNoNoNoNoNoNoNoNoIMET A327NoNoNoNoNoNoNoNoNoNoIMET A178YesNoYesYes214152IMET A214YesNoYesYesNo452IMET A225YesNoYesYesNo452IMET A235YesNoYesYesNo452IMET A260YesNoYesYesNo781IMET A260YesNoYesYesNo118NoIMET A261YesNoNoNoNo31NoNoIMET A264YesNoNoNoNo31NoNoIMET A264YesNoNoNoNo31NoNoIMET A264YesNoNoNoNoNo118NoIMET A264YesNoNoNoNoNoNoNoNoNoIMET A264YesNoNoNoNoNoNoNoNoNoIMET A264YesNoNoNoNoNoNoNoNoNo <tr<< th=""><th>IMET</th><th></th><th></th><th></th><th></th><th></th><th></th><th></th><th></th></tr<<>	IMET								
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A193NoYesYesNoNo14NoIMETNoYesYesNo2461A195NoYesYesNo2461IMETNoYesYesNoNoNo2NoA219NoYesYesNoNoNo2NoIMETNoYesYesNoNoNoNoNoA227NoYesYesNoNoNoNoNoIMETNoYesYesNoNoNoNoNoIMETNoYesYesNoNoNoNoNoIMETNoYesYesNo2881IMETA305NoYesYesNoNoNo8NoIMETA316NoYesYesNoNo16NoIMETNoYesYesNoNo16No	IMFT	110	100	100	110	110	110	-	110
IMET A195NoYesYesNo2461IMET A219NoYesYesNoNoNo2NoIMET A227NoYesYesNoNoNoNo2NoIMET A227NoYesYesNoNoNoNoNoNoIMET A237NoYesYesNoNoNoNoNoNoIMET A237NoYesYesNoNoNoNoNoNoIMET A251NoYesYesNo2881IMET A305NoYesYesNoNoNoNoNoIMET A316NoYesYesNoNoNo8NoIMET A316NoYesYesNoNo16NoIMET A316NoYesYesNoNo16No	A193	No	Yes	Yes	No	No	1	4	No
A195NoYesYesNo2461IMETNoYesYesNoNoNo2NoA219NoYesYesNoNoNo2NoIMETNoYesYesNoNoNoNoNoNoA227NoYesYesNoNoNoNoNoNoIMETNoYesYesNoNoNoNoNoNoIMETNoYesYesNoNoNoNoNoNoIMETNoYesYesNoNo2881IMETNoYesNoNoNo28NoIMETNoYesNoNoNo28NoIMETNoYesNoNoNo28NoIMETNoYesYesNoNoNo8NoIMETNoYesYesNoNo16NoIMETNoYesYesNoNo16NoIMETNoYesYesNoNo16NoIMETNoYesYesNoNo16NoIMETNoYesYesNoNo16No	IMET								
IMET A219NoYesYesNoNoNo2NoIMET A227NoYesYesNoNoNoNoNoNoIMET A237NoYesYesNoNoNoNoNoNoIMET A237NoYesYesNoNoNoNoNoIMET A237NoYesYesNoNoNoNoNoIMET A251NoYesYesNo2881IMET A305NoYesNoNoNo28NoIMET A315NoYesYesNoNoNo8NoIMET A316NoYesYesNoNo16NoIMET A316NoYesYesNoNo16No	A195	No	Yes	Yes	No	2	4	6	1
A219NoYesYesNoNoNo2NoIMETNoYesYesNoNoNoNoNoNoIMETNoYesYesNoNoNoNoNoNoIMETNoYesYesNoNoNoNoNoNoIMETNoYesYesNoNoNoNoNoNoIMETNoYesYesNo2881A251NoYesYesNo2881IMETNoYesNoNoNo28NoIMETNoYesNoNoNo28NoIMETNoYesYesNoNoNo8NoIMETNoYesYesNoNoNo8NoIMETNoYesYesNoNo16NoIMETNoYesYesNoNo10NoIMETNoYesYesNoNo16NoIMETNoYesYesNoNo16No	IMET								
IMET A227NoYesYesNoNoNoNoNoIMET A237NoYesYesNoNoNoNoNoNoIMET A251NoYesYesNoNoNoNoNoNoIMET A251NoYesYesNo2881IMET A305NoYesNoNoNo28NoIMET A315NoYesNoNoNo28NoIMET A316NoYesYesNoNoNo8NoIMET A316NoYesYesNoNoNo8NoIMET A316NoYesYesNoNo16No	A219	No	Yes	Yes	No	No	No	2	No
A227NoYesYesNoNoNoNoNoNoIMETA237NoYesYesNoNoNoNoNoNoIMETA251NoYesYesNo2881A251NoYesYesNo2881IMETA305NoYesNoNoNo28NoIMETA315NoYesYesNoNoNo8NoIMETA316NoYesYesNoNo16NoIMETA316NoYesYesNoNo16No	IMET								
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A251NoYesYesNo2881IMETIMETNoNoNoNo28NoA315NoYesYesNoNoNo8NoIMETIMETIMETIMETIMETIMETIMETIMETIMETA316NoYesYesNoNo16No		Nia	Vee	Vee	Nia	<u></u>	0		4
IME I A305NoYesNoNoNo28NoIMET A315NoYesYesNoNoNo8NoIMET A316NoYesYesNoNoNo8NoIMET A316NoYesYesNoNo16No		INO	res	res	INO	2	8	8	1
ASSSNOTesNONONO20NOIMETA315NoYesYesNoNoNo8NoIMETA316NoYesYesNoNo16NoIMET<		No	Vec	No	No	No	2	8	No
A315NoYesYesNoNoNo8NoIMET A316NoYesYesNoNo16NoIMET IMETNoYesYesNoNo16No	IMET	NO	165	NU	NO	NO	2	0	NU
IMETA316NoYesNoNo16NoIMETIMETIMETIMEIMEIMEIMEIMEIME	A315	No	Yes	Yes	No	No	No	8	No
A316 No Yes Yes No No 1 6 No IMET	IMET		103	103				0	110
IMET	A316	No	Yes	Yes	No	No	1	6	No
	IMET							-	
A183 Yes Yes Yes Yes No 10 15 8	A183	Yes	Yes	Yes	Yes	No	10	15	8

Blood product received by each patient during the ITU stay including the referring hospital (until discharge, liver transplant or death)

KCC: King's College Criteria, PLEX: Plasmapheresis, FFP: fresh frozen plasma, RBC: red blood cells, Cryo: cryoprecipitate, PLT: platelets