

Table S1 : Plasmids used in this study.

Plasmid	Description	Source or Reference
YIp5'5	<i>CENV URA43</i>	Ferguson <i>et al.</i> ⁱ
Yp-CN1	YIp5'5 ChrIV: 456,853 → 472,955 ^a [<i>ARS1</i> ACS: 462,593] ^a	This study
Yp-CN2	YIp5'5 ChrX: 688,409 → 676,641 ^a [<i>ARS121</i> ACS: 683,933] ^a	This study
Yp-CN3	Yp-CN1 <i>ars1</i> (-1046 → 954) :: <i>ARS121</i> (-920 → 1080) ^b	This study
Yp-CN4	Yp-CN2 <i>ars121</i> (-920 → 1080) :: <i>ARS1</i> (-1046 → 954) ^b	This study
Yp-CN5	Yp-CN1 <i>ΔTRP1</i> promoter (-395 → -1) ^c	This study
Yp-CN6	Yp-CN2 <i>ars121</i> (-233 → 140) :: <i>ARS1</i> (-107 → 269) ^b	This study
Yp-CN7	Yp-CN5 <i>ars1</i> (-107 → 269) :: <i>ARS121</i> (-233 → 140) ^b	This study
Yp-CN7B	Yp-CN1 <i>ars1</i> (-107 → 269) :: <i>ARS121</i> (-233 → 140) ^b	This study
Yp-CN7C	Yp-CN1 <i>ars1</i> (-107 → 269) :: <i>ARS121</i> (140 → -233) ^b	This study
Yp-CN8	Yp-CN5 <i>ars1</i> (-86 → 18) :: <i>ARS121</i> (-86 → 18) ^b	This study
Yp-CN9	Yp-CN2 <i>ars121</i> (-86 → 18) :: <i>ARS1</i> (-86 → 18) ^b	This study
Yp-CN10	Yp-CN5 <i>ars1_{A,B1}</i> (-32 → 18) :: <i>ARS121_{A,B1}</i> (-32 → 18) ^b	This study
Yp-CN11	Yp-CN2 <i>ars121_{A,B1}</i> (-32 → 18) :: <i>ARS1_{A,B1}</i> (-32 → 18) ^b	This study
Yp-CN12	Yp-CN5 <i>ars1_{B2}</i> (-86 → -27) :: <i>ARS121_{ATR}</i> (-86 → -27) ^b	This study
Yp-CN13	Yp-CN2 <i>ars121_{Mcm1}</i> sites :: <i>ARS121</i> scrambled <i>Mcm1</i> sites	This study
Yp-CN14	Yp-CN2 <i>ars121·B2</i> :: <i>ARS121</i> inverted 'B2'	This study
YCp101	<i>CENV LEU2 ARS1</i> (ChrIV: 462,352 → 463,193) ^a	Reference 21
YCp121	<i>CENV LEU2 ARS121</i> (ChrX: 685,748 → 679,017) ^a	Reference 21
Yp- <i>ARS1</i>	YIp5'5 ChrIV: 462,360 → 462,732 ^a [<i>ARS1</i> (-107 → 269) ^b]	This study
Yp- <i>ARS121</i>	YIp5'5 ChrX: 684036 → 683665 ^a [<i>ARS121</i> (-233 → 140) ^b]	This study

^a Chromosomal coordinates.^b Coordinates relative to the first base of the essential A element's A-rich strand.^c Coordinates relative to the *TRP1* ORF.

Additional reference:

- i. Ferguson, B.M., Brewer, B.J., Reynolds, A.E. and Fangman, W.L. (1991) A yeast origin of replication is activated late in S phase. *Cell*, **65**, 507-515.

Table S2 : Yeast strains used in this study.

Strain	Genotype	Source or Reference
8534-8C	A364 <i>MATα his4Δ34 ura3-52 leu2-3, -112</i>	Reference 21
M46-3C	A364 <i>MATα leu2-3, 112 his3-11, -15 ura3-52 mcm2-1</i>	Reference 21
mcm1-1	A364 <i>MATα ura3-52 leu2-3,-112 his3-11,-15 mcm1-1</i>	Reference 21
DBY2056	S288C <i>MATα ura3-52 ade2-1 lys2-801 leu2-3,112</i>	Hennessy <i>et al.</i> ⁱ
DBY2028	S288C <i>MATα mcm5-461</i>	Hennessy <i>et al.</i> ⁱ
AY925	W303 <i>MATα ade2-1 his3-11,15 leu2-3,112 trp1-1 ura3-1 can1-100 ssd1-d2 Gal+</i>	Tanaka <i>et al.</i> ⁱⁱ
orc2-1	W303 <i>MATα orc2-1</i>	Tanaka <i>et al.</i> ⁱⁱ
orc5-1	W303 <i>MATα orc5-1</i>	Tanaka <i>et al.</i> ⁱⁱ
cdc6-1	W303 <i>MATα cdc6-1</i>	Tanaka <i>et al.</i> ⁱⁱ
dbf4-1	W303 <i>MATα dbf4-1</i>	Tanaka <i>et al.</i> ⁱⁱ
cdc17-1	W303 <i>MATα cdc17-1</i>	Tanaka <i>et al.</i> ⁱⁱ
pri2-1	W303 <i>MATα pri2-1</i>	Tanaka <i>et al.</i> ⁱⁱ

Additional references:

- i. Hennessy, K. M., Clark, C. D., and Botstein, D. (1990) Subcellular localization of yeast CDC46 varies with the cell cycle. *Genes & Development*, **4**, 2252-2263.
- ii. Tanaka, T., and Nasmyth, K (1998) Association of RPA with chromosomal replication origins requires an Mcm protein, and is regulated by Rad53, and cyclin- and Dbf4-dependent kinases. *Embo J*, **17**, 5182-5191.

Table S3 : Loss rates for *ARS1* and *ARS121* plasmids in various strains at a range of temperatures.

Strain	Temperature ²	Rate of plasmid loss ¹	
		Yp-CN1	Yp-CN2
A364 wild-type	23.0	0.0 ± 0.1	0.0 ± 0.3
	30.0	0.8 ± 0.6	0.0 ± 0.6
<i>mcm2-1</i>	23.0	7.8 ± 0.7	1.1 ± 0.0
	25.5	8.9 ± 1.1	0.7 ± 0.4
	27.5	15.4 ± 0.4	0.9 ± 0.2
	30.0	26.4 ± 0.9	3.5 ± 0.2
	32.5	31.4 ± 1.4	9.5 ± 0.9
<i>mcm1-1</i>	23.0	6.7 ± 0.0	1.6 ± 0.0
	30.0	12.3 ± 0.9	5.3 ± 0.4
S288C wild-type	23.0	2.0 ± 0.7	1.1 ± 0.1
	32.5	0.4 ± 0.2	0.0 ± 0.5
<i>mcm5-461</i>	23.0	3.1 ± 1.8	0.0 ± 2.0
	25.5	4.1 ± 1.5	0.0 ± 1.5
	27.5	6.0 ± 1.9	0.0 ± 1.7
	30.0	13.5 ± 2.1	2.8 ± 2.1
	32.5	17.2 ± 1.9	4.5 ± 2.5
W303 wild-type	23.0	0.2 ± 0.1	0.0 ± 0.1
	30.0	0.9 ± 0.1	0.3 ± 0.1
<i>orc2-1</i>	23.0	10.0 ± 0.6	0.2 ± 0.0
	30.0	32.1 ± 1.1	13.3 ± 0.4
<i>orc5-1</i>	23.0	13.3 ± 2.3	3.7 ± 0.4
	30.0	23.1 ± 1.9	8.8 ± 0.8
<i>cdc6-1</i>	23.0	0.9 ± 0.4	0.2 ± 0.4
	30.0	27.7 ± 0.1	9.1 ± 0.1
<i>dbf4-1</i>	23.0	1.0 ± 0.2	0.3 ± 0.5
	30.0	5.3 ± 0.1	6.8 ± 0.9
<i>pri2-1</i>	23.0	1.6 ± 0.3	1.6 ± 0.4
	30.0	20.2 ± 0.9	16.0 ± 0.4
<i>cdc17-1</i>	23.0	0.7 ± 0.0	0.4 ± 0.1
	30.0	4.7 ± 0.1	4.7 ± 0.2

¹ % per generation.

² °C.

Table S4 : Loss rates in wild-type cells for Yp-CN1 and Yp-CN2 derived chimaeric plasmids at 30 °C.

Plasmid	Rate of plasmid loss ¹
Yp-CN1	0.8 ± 0.6
Yp-CN2	0.0 ± 0.6
Yp-CN3	1.2 ± 1.2
Yp-CN4	0.8 ± 0.3
Yp-CN5	0.1 ± 0.1
Yp-CN6	1.3 ± 0.8
Yp-CN7	0.2 ± 0.3
Yp-CN7B	2.1 ± 0.7
Yp-CN7C	1.8 ± 0.7
Yp-CN8	1.0 ± 0.0
Yp-CN9	0.4 ± 0.3
Yp-CN10	1.5 ± 0.1
Yp-CN11	0.7 ± 0.0
Yp-CN12	0.6 ± 0.0

¹ % per generation.