

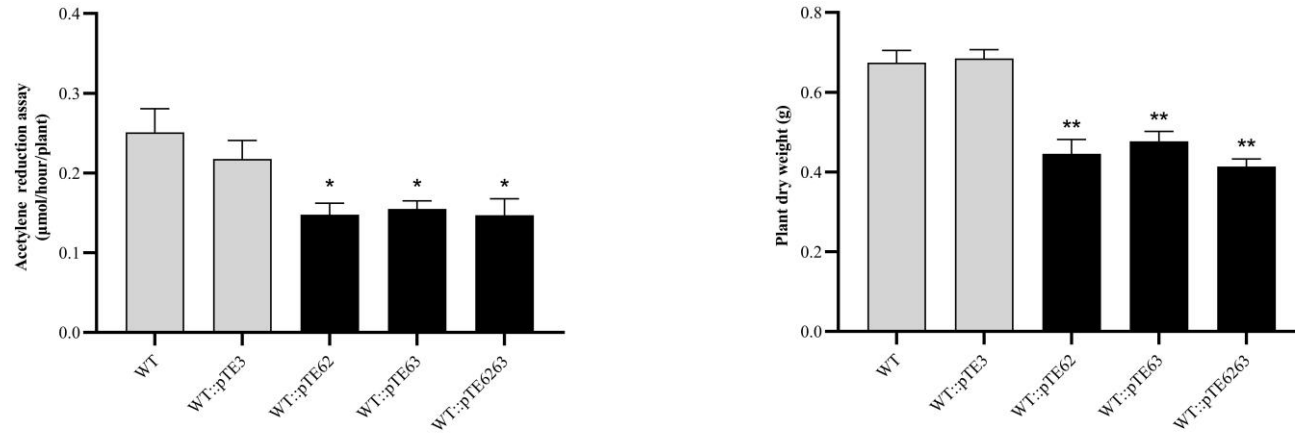
M13 DNA clone probe	ORF covered	Length (bp)	chromosomal location (start-end)
□ brb14021	<i>bll1058 bll1059 bll1060 bsl1061 <u>blr1062</u></i>	3289	1172519 -1175808
▨ brb21078	<i>bsl1061 <u>blr1062</u> blr1063 blr1064</i>	2853	1175021-1177874
■ brb24218	<i><u>blr1063</u> blr1064 blr1065</i>	2612	1176007-1178619

*blr1062: bjaR<sub>1</sub>; blr1063: bjaI*

**Figure S1.** The hybridization intensity of three overlapping M13 DNA clone probes, which encompass the *bjaR<sub>1</sub>/bjaI* QS genes, with <sup>33</sup>P-labeled cDNA generated by reverse transcription from mRNA isolated from *B. diazoefficiens* USDA 110 cells induced with soybean seed extract (SSE) at a concentration of 20 μl/ml culture and genistein at a final concentration of 5 μM. The intensity value represents the mean ± SEM of three independent experiments, each with three replicates. The solvent pure EtOH was used in control treatment. The data here represent a subset of our previous study (50).



**Figure S2.** Images of soybean, root, nodule and the nodule section in a inoculation experiment with *B. diazoefficiens* strains. The top first panel represents the soybean image taken at 25 days post-inoculation (dpi), and the below three panels represent images at 40 dpi. The *nodD2* mutant, *B. j* JD21, induced white nodule, which is indicative of barely nitrogen-fixation ability. The nodules formed by  $\Delta$ bjar<sub>1</sub> and *nodD2* mutant were mainly dispersed in the lateral root, whereas those by the WT and the c-  $\Delta$ bjar<sub>1</sub> complementation strain clustered at the junction of root and shoot.

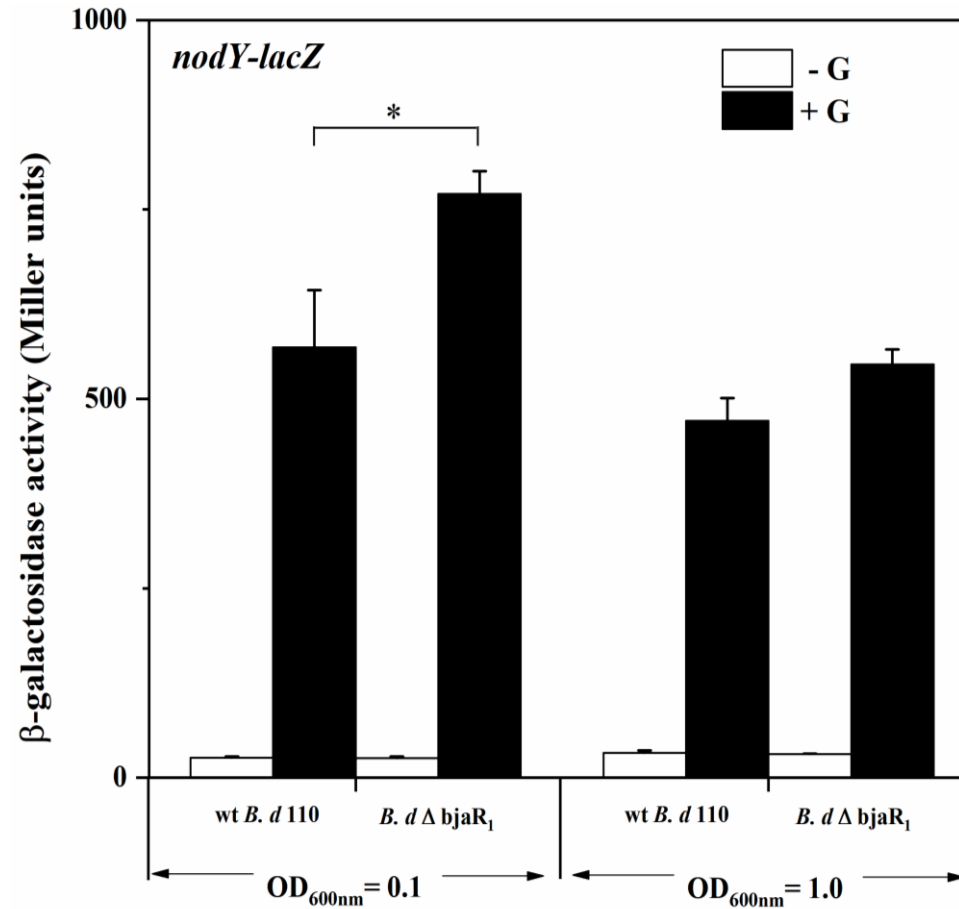


#### Primers used to construct recombinant plasmid to overexpress *bjaR<sub>1</sub>/bjaI* QS system

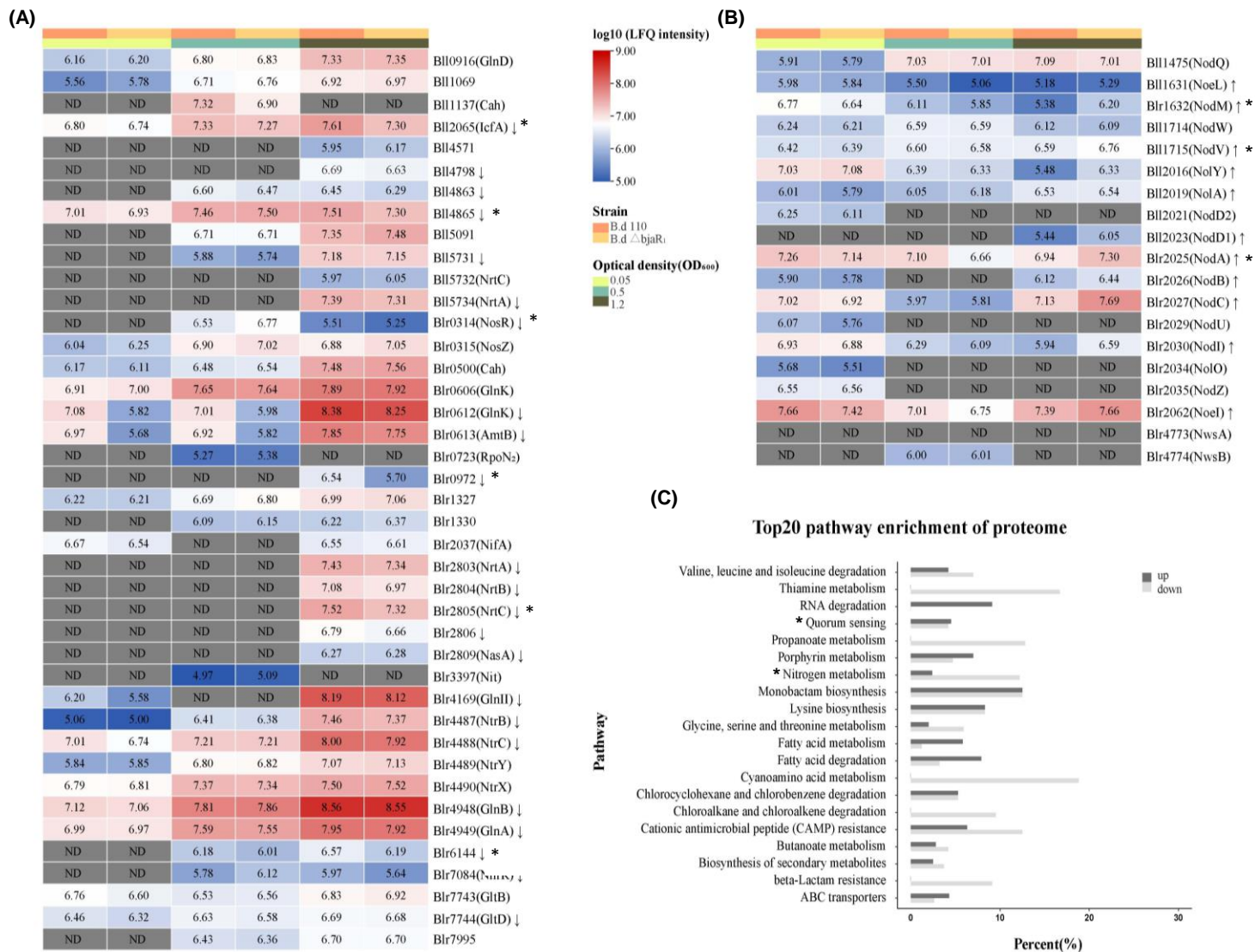
Primers	Nucleotide sequence (5'-3')
pTE3-62-fp	<u>GGTGCCAAGCTTGGCATGTCCGCCGTCGATTATGG</u>
pTE3-62-rp	<u>GAATTCCCGGGGATCTTAGGGATTGATGATCTTGTGGCG</u>
pTE3-63-fp	<u>GGTGCCAAGCTTGGCATGGGGGTTTCCATGATTCACG</u>
pTE3-63-rp	<u>GAATTCCCGGGGATCTCAGGCGCTCTTTCGTTGC</u>

The underlined nucleotide sequence represents the 15 homologous base on either side of the clone site required for In-Fusion cloning

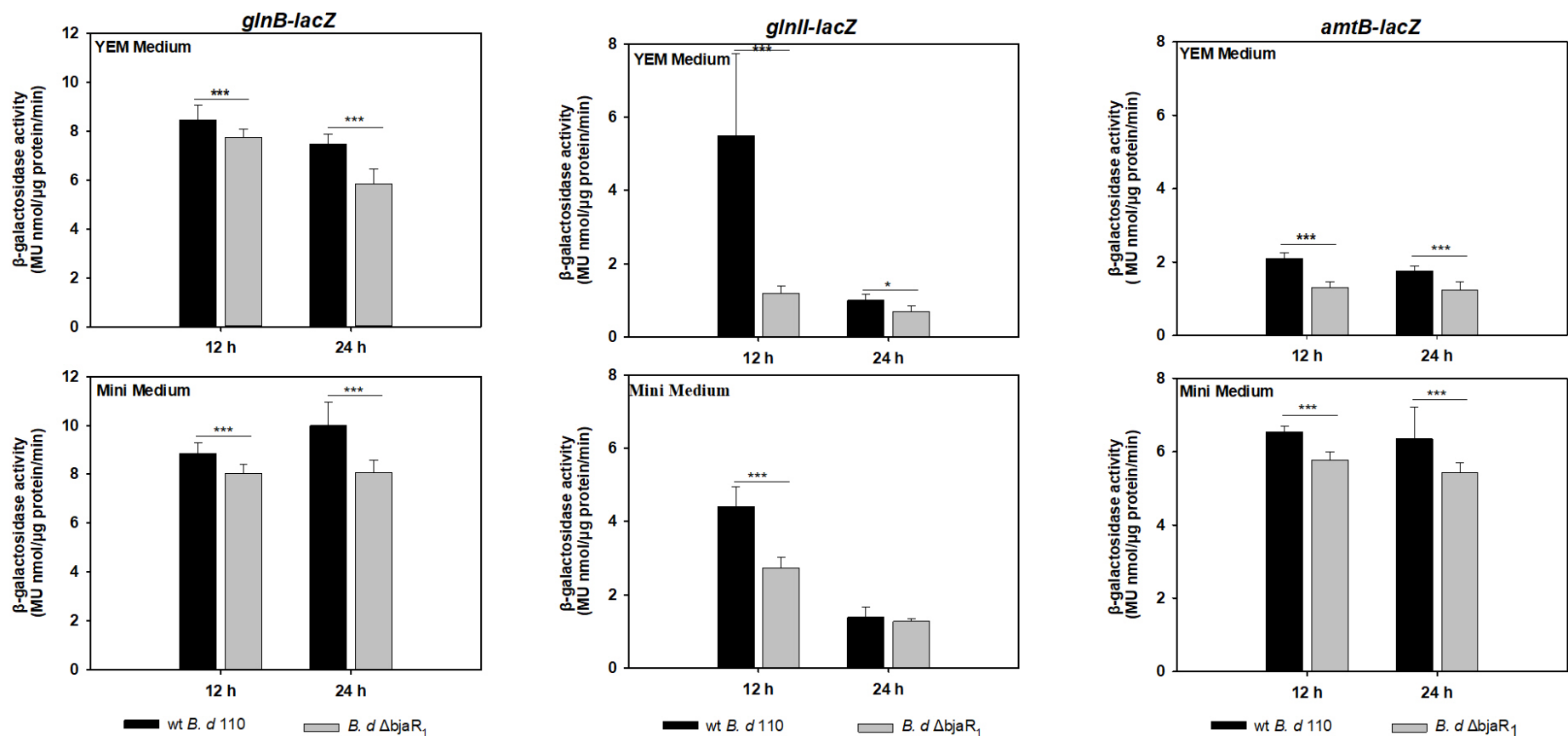
**Figure S3.** The symbiotic properties of genetically modified strains overexpressing *bjaR<sub>1</sub>/bjaI* QS system during inoculation with soybean under the low-N conditions at 30 dpi. Using wild-type (WT) *B. diazoefficiens* USDA 110 DNA as a template, we performed PCR amplification with primer pairs 62\_fp/rp, 63\_fp/rp, and 62\_fp/63\_rp to obtain DNA fragments containing either the *bjaR<sub>1</sub>* gene or the *bjaI* gene, or both. The DNA fragments were then cloned into the *PstI/Bam* HI site of the overexpression vector pTE3 (23) using In-Fusion cloning. The resulting recombinant plasmids (pTE-62, pTE63 or pTE6263) were subsequently introduced into the WT strain. The data represents the mean  $\pm$  SEM of at least 9 samples from two independent experiments. \* or \*\* indicate significant differences compared to WT strains at  $P \leq 0.05$  or 0.01 levels, respectively.



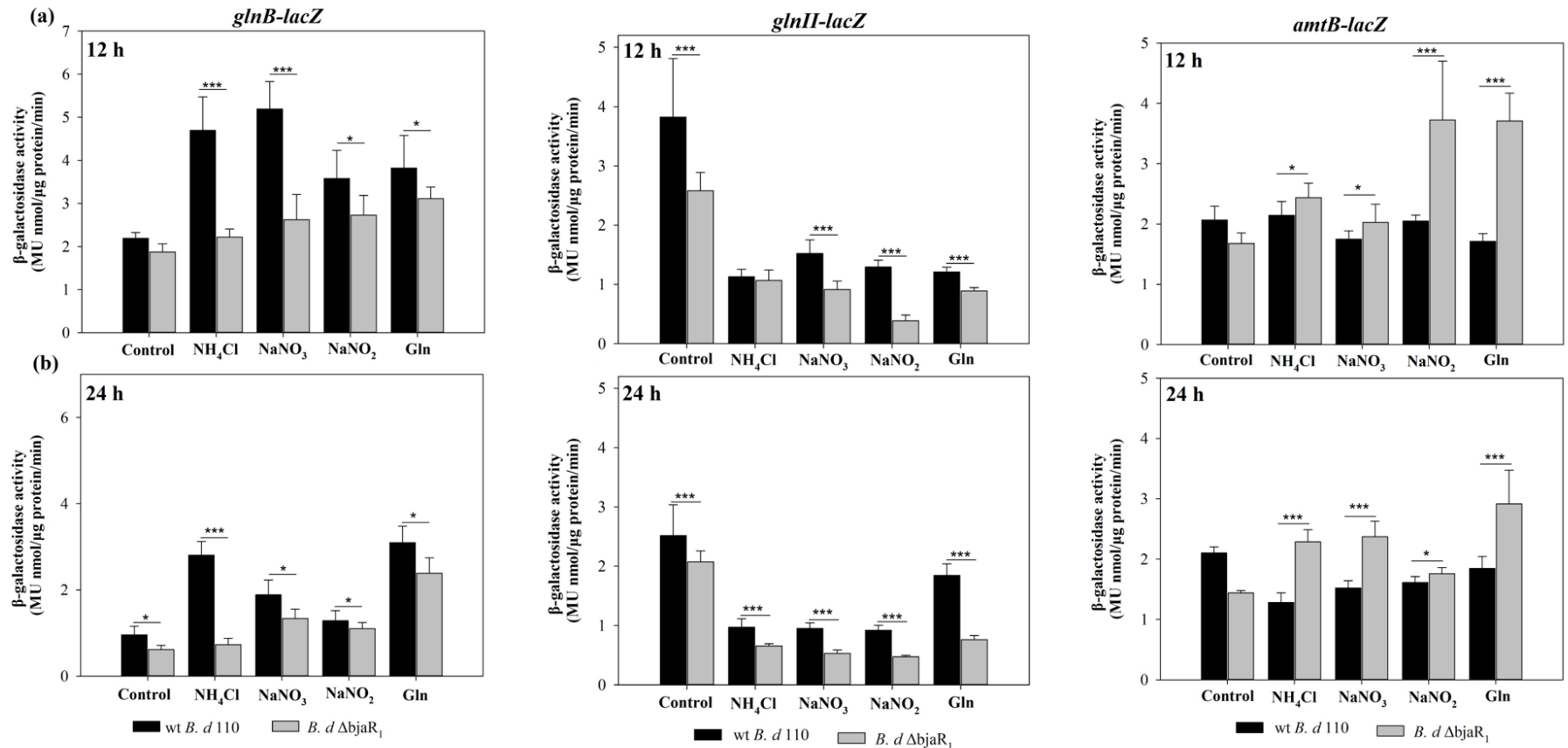
**Figure S4.** The induction of *nodY-lacZ* by the isoflavonoid genistein in *B. diazoefficiens* cells at low and high population densities. A broad-host-range plasmid carrying a translational *nodY-lacZ* fusion was conjugated into these strains. The bacterial cultures were induced with genistein to a final concentration of 5.0  $\mu$ M for 12 hours at each cell density (+G indicates presence of genistein, -G indicates absence). The presented data are means  $\pm$  SEs from three independent experiments with three replicates each.



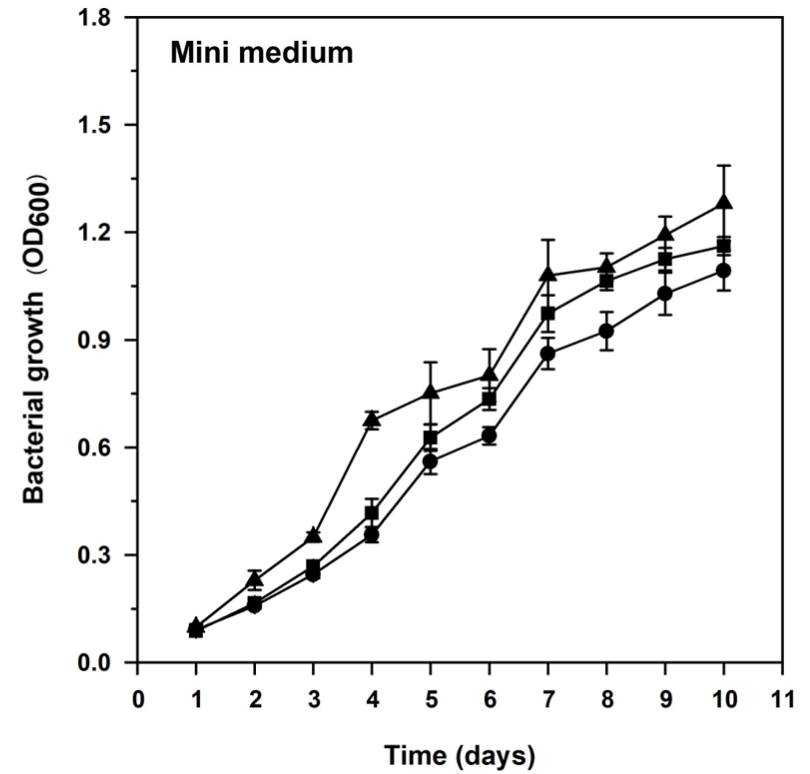
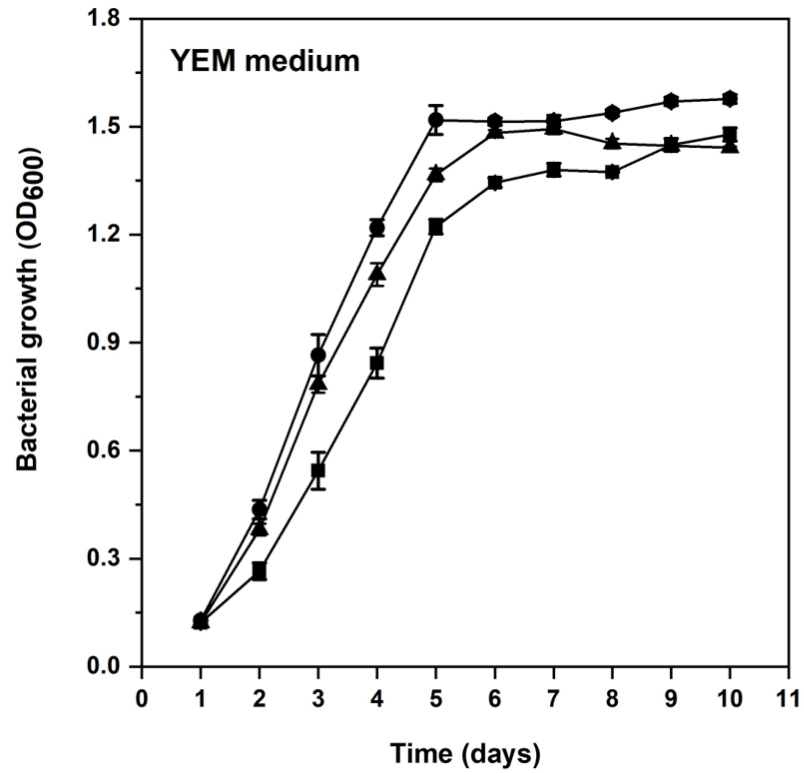
**Figure S5** The abundance heatmap of detectable proteins involved in nitrogen metabolism (A) and nodulation (B) module in free-living *B. diazoefficiens* at various cell densities at OD<sub>600</sub>. The value inside the box indicates the average mass spectrometry signal for the proteins from three different bacterial cultures as determined by label-free quantification (LFQ) intensities from the DIA-NN output. The proteins that were up- and down-regulated in the ΔbjaR<sub>1</sub> mutant cells compared to wild-type cells at a high cell density (OD<sub>600</sub>=1.2) are indicated by upward and downward arrows. ND, Not detected. (C) Percentage of differentially regulated genes compared to the total genes involved in each metabolic pathway of *B. d 110*. Data represent the top-20 enriched metabolic pathways in bacterial cells at a high cell density (OD<sub>600</sub>=1.2). Asterisks (\*) indicate statistically significant differences (P-value ≤ 0.05).



**Figure S6.** Promoter activity of  $\text{NH}_4^+$ -responsive genes of *B. diazoefficiens* freely grown in the yeast-extract mannitol (YEM) and minimal (Mini) medium(48) for 12 h and 24 h. Gene *glnB* (*blr4948*), *glnII* (*blr4169*) and *amtB* (*blr0613*) encode an N-regulatory protein PII, a glutamine synthetase and an ammonium transporter, respectively. Bacterial cells in 1.0 ml culture were collected and crushed to determine *lacZ* activity which was using 4-methylumbelliferyl- $\beta$ -D-galactopyranoside (MUG<sub>LACZ</sub>) as the fluorogenic substrate. Data represent mean  $\pm$  SE of six plants in two independent experiments. Asterisk (\* and \*\*\*) indicates a significant difference between the two strains at the statistic T-test level ( $P$ -value $<$ 0.05 and 0.01).



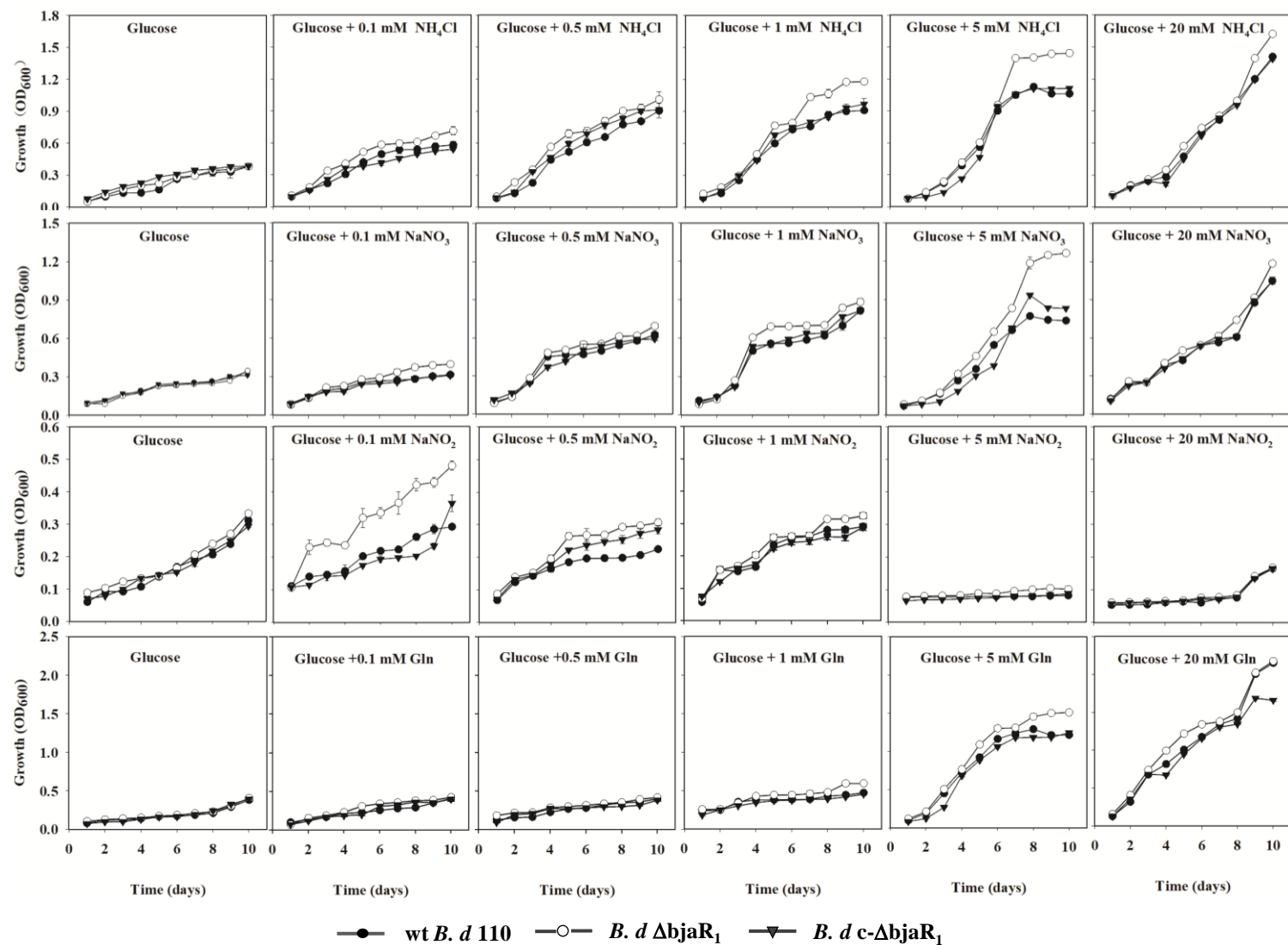
**Figure S7.** Promoter activity of  $\text{NH}_4^+$ -responsive genes of *B. diazoefficiens* freely grown in the liquid minimal medium (47) containing a variety of N-containing compound (final concentration, 5.0 mM) for 12 h (a) and 24 h (b). Gene *glnB* (*blr4948*), *glnII* (*blr4169*) and *amtB* (*blr0613*) encode an N-regulatory protein PII, a glutamine synthetase and an ammonium transporter, respectively. Bacterial cells in 1.0 ml culture were collected and crushed to determine *lacZ* activity which was assayed using 4-methylumbelliferyl- $\beta$ -D-galactopyranoside ( $\text{MUG}_{\text{LACZ}}$ ) as the fluorogenic substrate. Data represent mean  $\pm$  SE of six plants in two independent experiments. Asterisk (\* and \*\*\*) indicates significant difference between the two strains at the statistical T-test level ( $P$ -value  $< 0.05$  and  $0.01$ ).



■ wt *B. d110* ● *B. d* Δ*bjaR*<sub>1</sub> ▲ *B. d* c-Δ*bjaR*<sub>1</sub>

**Figure S8.** Growth of *B. diazoefficiens* in yeast-extract mannitol (YEM) and a minimal (Mini) media (46). One bacterial colony was selected from the plate and first grown in a 20 ml liquid culture to the logarithmic phase ( $OD_{600}=0.4-0.6$ ). Then, they were diluted to  $OD_{600}=0.05$  with fresh media to 20 ml, and the optical density value at 600 nm ( $OD_{600}$ ) was measured each day. The data present the mean  $\pm$  SE of nine samples in three independent experiments.





**Figure S9** *B. diazoefficiens* growth in response to different nitrogenous compounds. The bacterial strains were cultured in a defined medium (46) in which glucose (27.8 mM) substituted for mannitol served as the sole source of carbon. Final concentration of different N-containing compounds ranging from 0.1 to 20 mM were applied. Data represent the means  $\pm$  SEs of six samples in two independent experiments. Gln, glutamine.