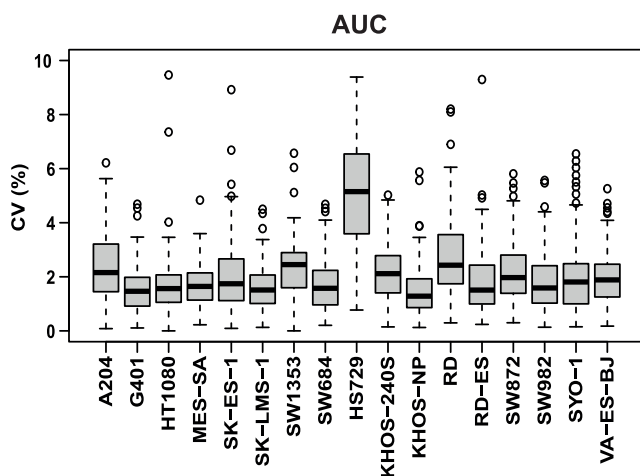


Expanded View Figures

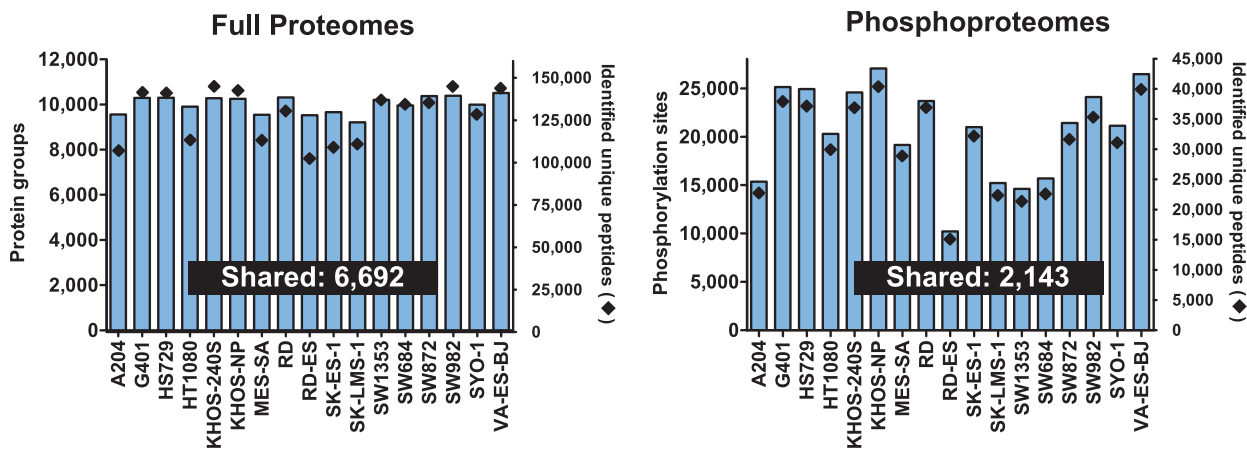
Figure EV1. Summary of data quality.

(A) Distributions of coefficient of variation (CV) of AUC (area under the curve) values determined from cell viability assays ($n = 3$) in response to 150 cancer drugs (each circle represents one drug). The central band represents the median, while the hinges denote the first and third quartiles with whiskers extending up to 1.5 times the interquartile range (IQR). (B) The number of quantified protein groups/peptides (left) and p-sites/phosphorylated peptides (right). (C) Unsupervised hierarchical clustering of biological triplicates of four cell lines on protein (left) and phosphoprotein (right) level. Pearson correlation was shown. R2, R3: biological replicates.

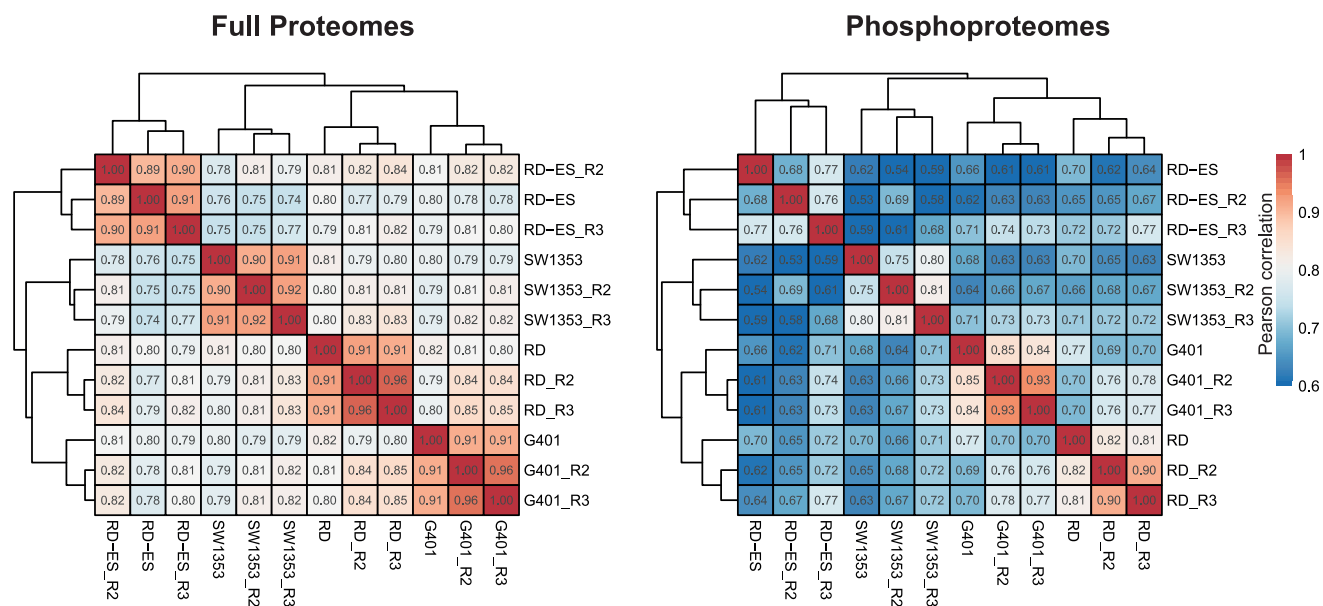
A



B



C



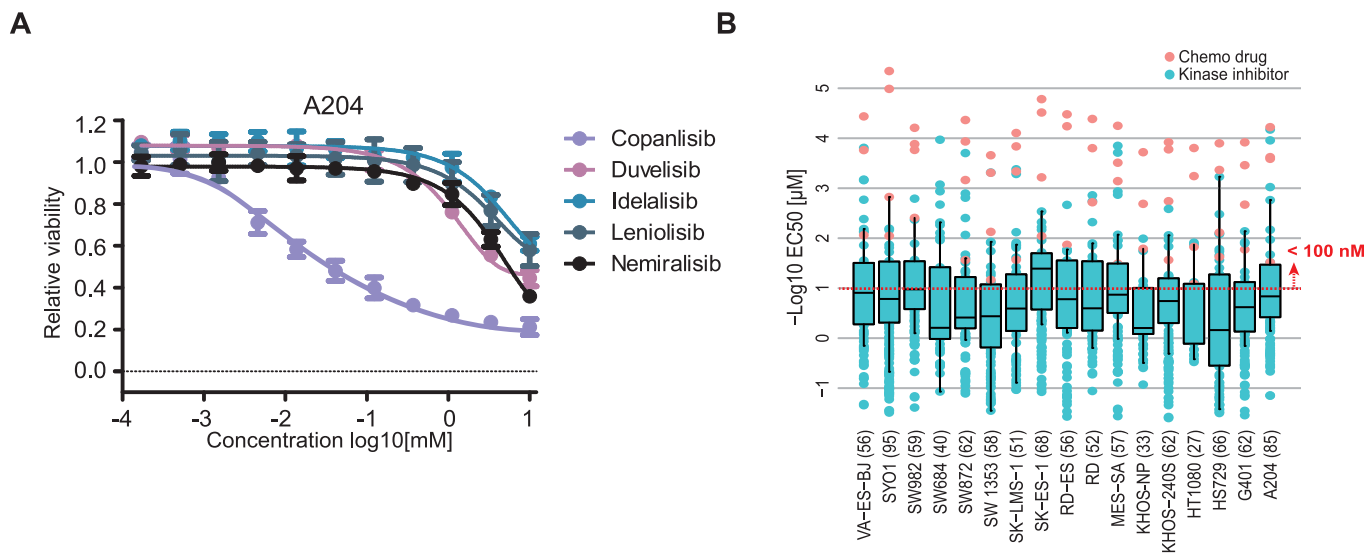
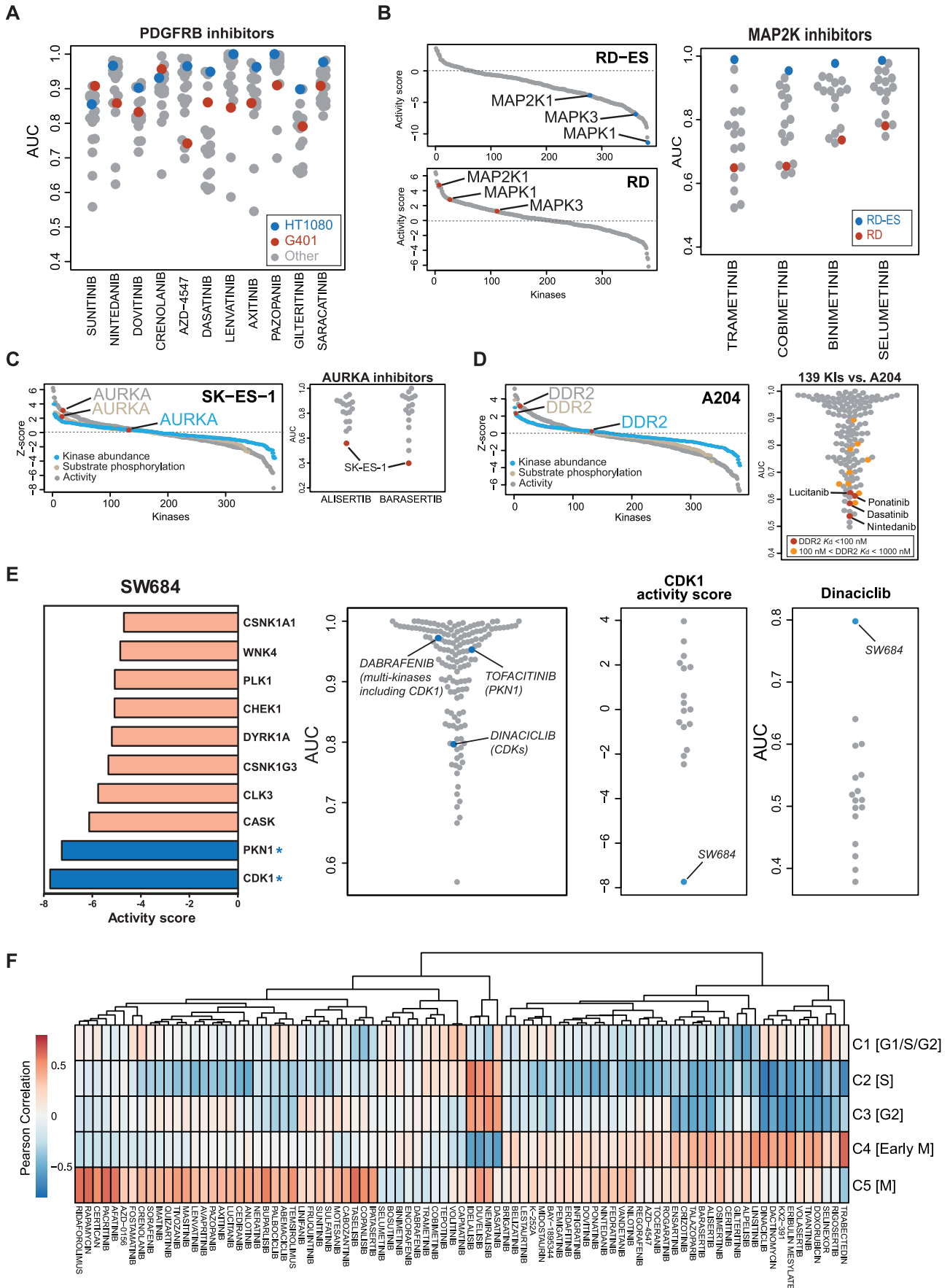


Figure EV2. Summary of the phenotypic dose-response characteristics of all cell lines and all drugs.

(A) Viability curves of five selected PI3K inhibitors in A204 cells. Measurements were in technical triplicates and the error bars were shown in SD. (B) Distribution of calculated EC₅₀ values for all cell lines and all drugs. Numbers in brackets after each cell line name indicate the number of drugs that show effects larger than AUC>0.9 and have relative effect sizes of >50%. The red dotted line marks an EC₅₀ value of 100 nM. The central band represents the median, while the hinges denote the first and third quartiles with whiskers extending up to 1.5 times the IQR.



◀ Figure EV3. Integration of phenotypic drug response and proteomic activity scores.

(A) Phenotypic drug response of two sarcoma lines towards 11 PDGFRB inhibitors. (B) Distribution of activity scores of proteins in RD-ES (top panel) and RD cells (bottom panel) showing that MAP2K1 and its downstream substrates MAPK1 and MAPK3 have different activities in the two cell lines. This translates into differences in phenotypic response of the two cell lines to MAP2K inhibitors (right panel). (C) Ranked list of kinases either considering kinase abundance, substrate phosphorylation abundance or computed activity in SK-ES-1 cells (left panel) showing that AURKA phosphorylation and activity is substantially higher than protein abundance. This translates into a strong response of the cell lines to AURKA inhibitors (right panel). (D) Same as panel (C) but for DDR2 in A204 cells. (E) Left panel: list of kinases ranked by activity score in SW684 cells. The asterisk denotes a kinase that is a target of the kinase inhibitor. Middle panel: relative sensitivity of SW684 cells to all drugs in the screen. Right 2 panels: CDK1 activity score and drug responses toward Dinaciclib among 17 sarcoma lines. (F) The heatmap summarises the Pearson correlations of proportions of cell cycle clusters (from Kelly et al) and drug responses from 87 drugs (AUC <0.8).

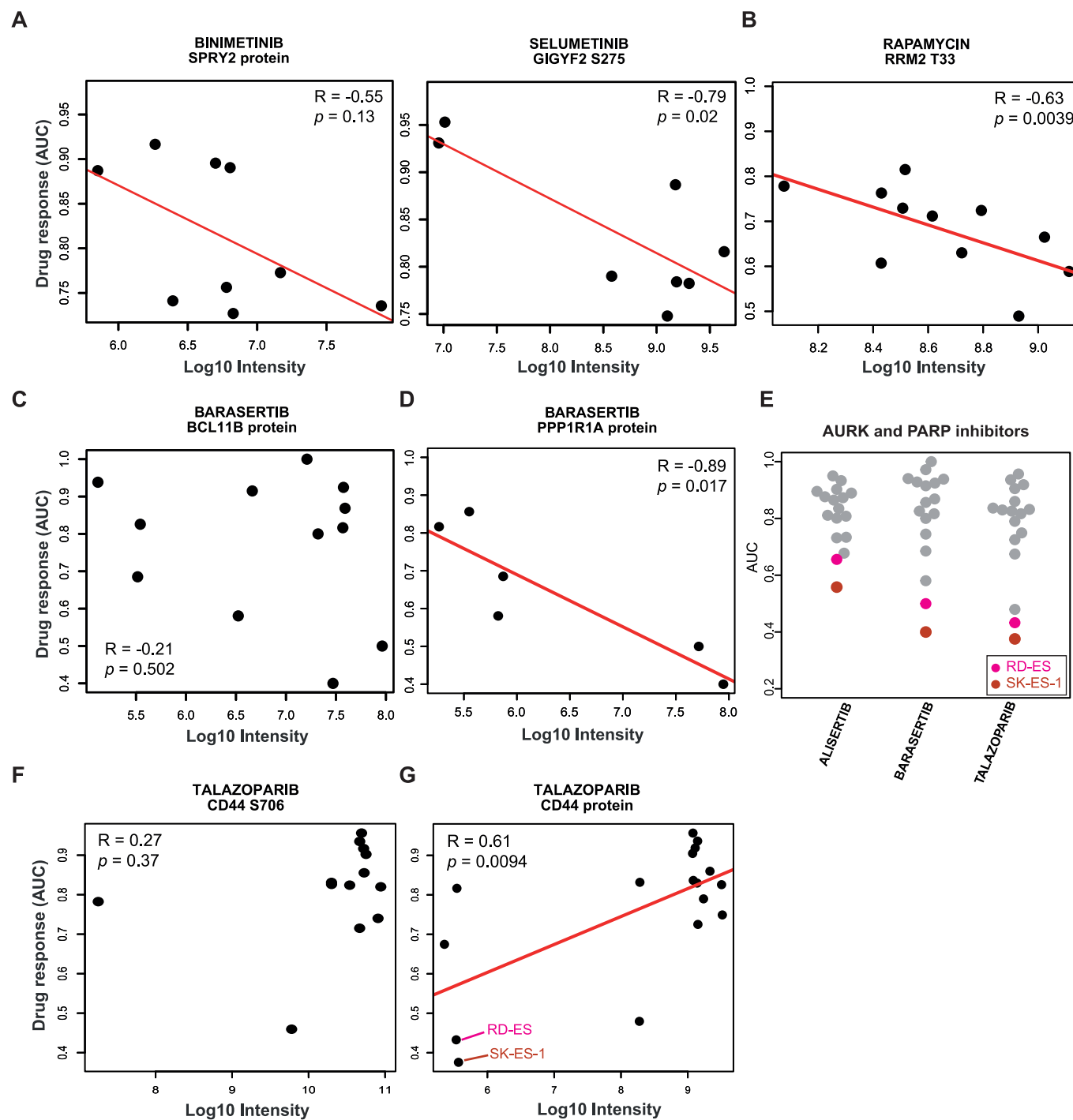
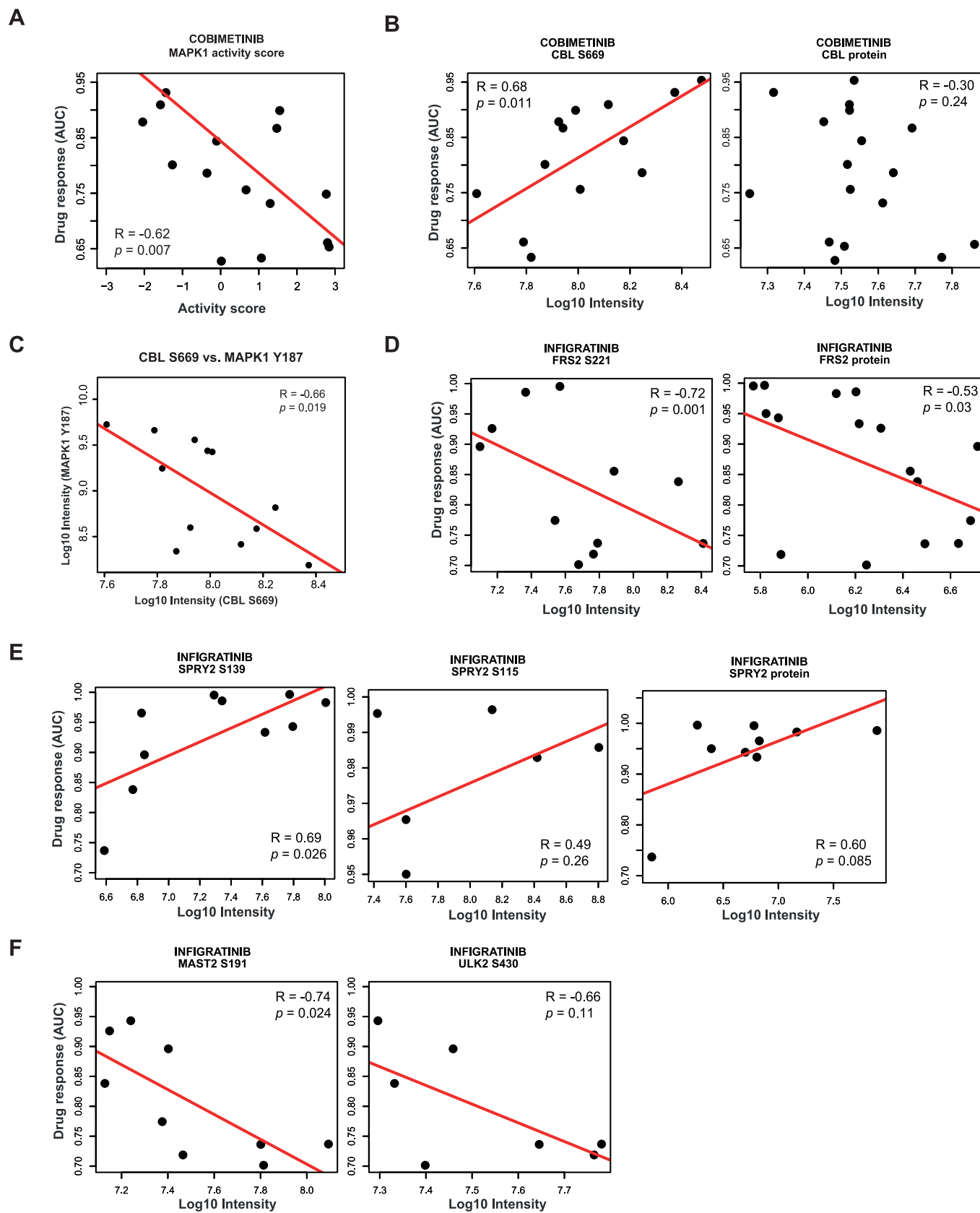
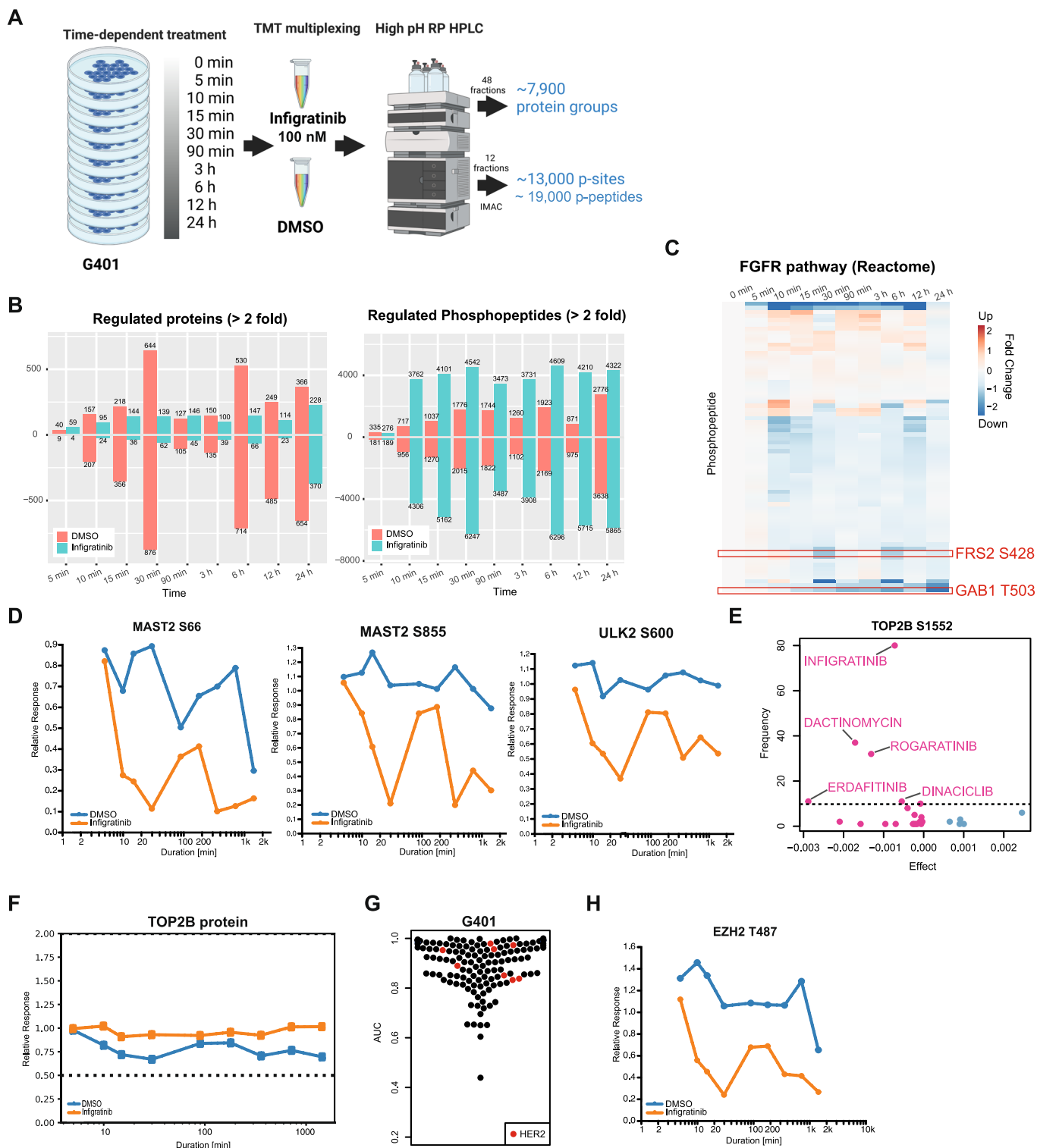


Figure EV4. Examples of correlation analysis of phenotypic drug sensitivity and protein or p-site abundance for drugs forming clusters in sparse multiblock partial least-square regression (SMBPLSR) analysis.

(A) SPRY2 protein levels as a marker for sensitivity towards Binimetinib (left panel) and GIGYF2 S275 phosphorylation abundance as a sensitivity marker for Selumetinib (right panel; both drugs are MAP2K inhibitors). (B-D) Same as panel (A) but for Rapamycin (mTOR inhibitor) and Barasertib (PARP inhibitor). (E) The response of two Ewing sarcoma lines (in red and pink) towards AURK and PARP inhibitors is more pronounced than for all other cell lines. (F, G) Same as panel A but for Talazoparib (PARP inhibitor). Again, the two Ewing sarcoma lines (in red and pink) are the most sensitive to this drug.



◀ **Figure EV5. Examples for correlation analysis of phenotypic drug sensitivity and protein or p-site abundance from candidates identified by elastic net regression.**
(A) Correlation of MAPK1 activity score and drug responses across sarcoma lines. (B-F) Correlation of protein and p-site abundance vs. drug responses across sarcoma lines (B, D-F). Correlation of the abundance of CBL S669 and MAPK1 Y187 across sarcoma lines (C).



◀ Figure EV6. Time-resolved (phospho)proteomic analysis of G401 cells in response to Infigratinib.

(A) Experimental workflow and number of protein and phosphopeptide identifications/quantification. For each time point of drug treatment, a DMSO control was included to account for abundance changes that are not due to the drug treatment. (B) Number of drug-induced protein expression changes (left panel) and phosphopeptide abundance (at least twofold compared to zero min at different time points). (C) Heatmap of fold-changes of phosphopeptides from proteins involved in FGFR signalling (annotations from Reactome) compared to DMSO at different time points after Infigratinib treatment. (D) Time course of p-sites of MAST2 and ULK2 proteins following Infigratinib treatment. (E) Elastic net regression analysis using the phosphorylation site S1552 of TOP2B to identify drugs associated with this p-site. (F) TOP2B protein level at different time points after Infigratinib treatment. (G) Distribution of AUC values from the phenotypic drug screen in G401 cells (139 kinase inhibitors shown). Inhibitors targeting HER2 are highlighted in red. (H) Time course of EZH2 T487 following Infigratinib treatment.