nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

Statistics

For	For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.					
n/a	Cor	nfirmed				
	×	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement				
	x	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly				
	x	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.				
x		A description of all covariates tested				
	x	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons				
	x	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)				
	x	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>				
	x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings				
	x	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes				
	x	Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated				

 $\textit{Our web collection on } \underline{\textit{statistics for biologists}} \textit{ contains articles on many of the points above}.$

Software and code

Policy information about availability of computer code

Data collection

PacBio SMRT reads were generated on the PacBio sequel II platform; Short-insert paired-end reads were generated on the Illumina HiSeq X Ten platform; RNA-seq reads were generated on the Illumina HiSeq X Ten or HiSeq 2500 or BGI-SEQ 500 platform; Hi-C data were generated on the Illumina NovaSeq 6000 or BGISEQ-500 platform.

Data analysis

Genome assembly, annotation and assessment: wtdbg2 (v2.5), Quiver (SMRT Analysis v2.3.0), Pilon (v1.22), HiC-Pro (v3.0.0), Juicer (v1.6), Juicebox (v1.13.01), 3D-DNA (v180922), RepeatMasker (open-4.0.9), RepeatModeler (v2.0), LTR_FINDER (v1.0.7), Augustus (v3.3.1), GeneMark (v2.5), exonerate (v2.2.0), Tophat2 (v2.1.1), Cufflinks (v2.2.1), EVidenceModeler (v1.1.1), iTAK (v1.7), BUSCO (v5); Phylogenetic analysis: OrthoFinder (v2.4.0), MAFFT (v7.429), PAL2NAL (v14.0), Gblocks (v0.91b), RAxML-NG (v1.1.0), PAML (v4.9j), GGTREE (v2.2.4);

Whole-genome triplication and chromosome evolution analyses: BLASTP (v2.9.0+), MCScanX (v1), JCVI (v1), Circos (v0.69-7), KaKs_Calculator (v2.0), igraph (v1.5.0.1), RAXML-NG (v1.1.0), MAPS (v1), Notung (v2.9.1.5), WGDgc (v1.3), CAFE (v4.2.1), PAML (v4.9j), ChromEvol (v2.1), WGDI (v0.6.5);

Transcriptome analysis: SolexaQA++ (v3.1.7.1), HISAT2 (v2.2.0), HTSeq (v0.13.5), R (v3.4.4), Cytoscape (v3.7.2), DESeq2 (v1.40.2); Selection analysis: BWA (v0.7.17-r1188), SAMtools (v1.9), Picard (v2.18.20), GATK (v3.7), VCFtools (v0.1.17), DEF-alpha (v2.16).

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

The raw genomic Illumina reads, PacBio reads, Hi-C reads, and RNA-seq reads reported in this paper have been deposited in the Genome Sequence Archive (GSA, https://ngdc.cncb.ac.cn/gsa) in National Genomics Data Center, Beijing Institute of Genomics, Chinese Academy of Sciences / China National Center for Bioinformation, under accession number CRA004284 [https://ngdc.cncb.ac.cn/gsa/browse/CRA004284] with BioProject ID PRJCA005319 [https://ngdc.cncb.ac.cn/ bioproject/browse/PRJCA005319]. The genome assembly sequences have been deposited in the Genome Warehouse (GWH, https://ngdc.cncb.ac.cn/gwh) in National Genomics Data Center under accession number GWHBCIQ00000000 [https://ngdc.cncb.ac.cn/gwh/Assembly/20653/show], GWHBCKL00000000 [https:// ngdc.cncb.ac.cn/gwh/Assembly/20692/show] with BioProject ID PRJCA004930 [https://ngdc.cncb.ac.cn/bioproject/browse/PRJCA004930] and BioSample ID SAMC353197 [https://ngdc.cncb.ac.cn/biosample/browse/SAMC353197], SAMC353201 [https://ngdc.cncb.ac.cn/biosample/browse/SAMC353201]. The genome assemblies and annotations are also available at Figshare: Sonneratia alba [https://doi.org/10.6084/m9.figshare.25118819], Lagerstroemia speciosa [https://doi.org/10.6084/m9.figshare.25118819] doi.org/10.6084/m9.figshare.25118831]. Source data are provided with this paper.

Research involving human participants, their data, or biological material						
Policy information about studies with <a documents="" href="https://www.new.new.new.new.new.new.new.new.new.</td></tr><tr><td>Reporting on sex and gender</td><td>N/A</td></tr><tr><td>Reporting on race, ethnicity, other socially relevant groupi</td><td colspan=2></td></tr><tr><td>Population characteristics</td><td>N/A</td></tr><tr><td>Recruitment</td><td>N/A</td></tr><tr><td>Ethics oversight</td><td>N/A</td></tr><tr><td colspan=6>Note that full information on the approval of the study protocol must also be provided in the manuscript.</td></tr><tr><td colspan=7>Field-specific reporting</td></tr><tr><td>Please select the one below</td><td>that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.</td></tr><tr><td>Life sciences</td><td>Behavioural & social sciences</td></tr><tr><td>or a reference copy of the docume</td><td>nt with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf						
Ecological, evolutionary & environmental sciences study design						
All studies must disclose on	these points even when the disclosure is negative.					
Study description	We present two chromosome-scale genomes of Lythraceae plants: the mangrove tree Sonneratia alba and related inland plant Lagerstroemia speciosa, as a part of the worldwide mangrove genomes project. Through comprehensive analyses, we trace the					

during global climate change. The genome sequences provided in this study will also expedite genomic and evolutionary research in plants.

Research sample

The mature specimens of Sonneratia alba and Lagerstroemia speciosa were sampled from the nursery of Dongzhai Harbor National Nature Reserve in Haikou and Sun Yat-sen University in Guangzhou. For resequencing data, 12 individuals of Sonneratia alba from Cebu, Philippines, and other 12 individuals from Davao, Philippines, were collected. All plants included in the study were adult individuals. Sampling within each population was conducted randomly to ensure the representativeness of the samples.

Sampling strategy

For de novo genome data, fresh and healthy tissues from a plant of Sonneratia alba and a plant of Lagerstroemia speciosa were harvested and immediately frozen in liquid nitrogen, respectively, followed by preservation at -80°C in the laboratory. For resequencing data, fresh and healthy leaves from 24 individuals of Sonneratia alba (12 individuals per population) were dried and preserved with silica gel. For RNA-seq data, fresh and healthy tissues from three individuals of Sonneratia alba and two individuals of Lagerstroemia speciosa were harvested and immediately frozen in liquid nitrogen, respectively, followed by preservation at -80°C in the laboratory. For population analysis, a sample size of 12 individuals is deemed sufficient; for expression analysis, biological replicates are provided in this study.

Data collection

Samples of Sonneratia alba from the nursery of Dongzhai Harbor National Nature Reserve in Haikou were collected by X.F., Q.C., G.L.,

	C.Z., S. Shi and Z.H. Samples of Sonneratia alba from Philippines were collected by S.X. and S. Shi. Samples of Lagerstroemia speciosa from Sun Yat-sen University in Guangzhou were collected by X.F., Q.C. and G.L. Plant samples information has been deposited in the BioSample database in National Genomics Data Center. High-throughput sequencing data were generated on different platforms, including PacBio, Illumina, and BGISEQ.			
Timing and spatial scale	Spatial scale of sampling in this study is described in Methods section (including the nursery of Dongzhai Harbor National Nature Reserve in Haikou, Sun Yat-sen University in Guangzhou; Cebu and Davao in Philippines). All samples were collected between 2018-2023.			
Data exclusions	No data were excluded.			
Reproducibility	The leaf, root, flower, and fruit tissues of S. alba contains three independent biological replicates. The leaf, stem, flower, and fruit tissues of L. speciosa contains two independent biological replicates. And read coverages for each sample are from dozens to hundreds. All attempts at replication were successful.			
Randomization	The mature plant samples for whole genome sequencing were randomly sampled with permission.			
Blinding	Blinding is not applicable in this study as it does not involve subjects receiving different treatments.			
Did the study involve field work? Yes No				

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems		Methods	
n/a	Involved in the study	n/a Involved in the study	
x	Antibodies	✗ ☐ ChIP-seq	
x	Eukaryotic cell lines	Flow cytometry	
x	Palaeontology and archaeology	MRI-based neuroimaging	
x	Animals and other organisms		
×	Clinical data		
×	Dual use research of concern		
	x Plants		

Dual use research of concern

Policy information about <u>dual use research of concern</u>

Increase transmissibility of a pathogen

Enable evasion of diagnostic/detection modalities

Enable the weaponization of a biological agent or toxin

Alter the host range of a pathogen

Enhance the virulence of a pathogen or render a nonpathogen virulent

Any other potentially harmful combination of experiments and agents

Hazards

X

x

x

x

x

Could the accidental, deliberate or reckless misuse of agents or technologies generated in the work, or the application o in the manuscript, pose a threat to:	f information presented
No Yes	
Public health	
National security	
▼ Crops and/or livestock	
Ecosystems	
X Any other significant area	
Experiments of concern	
Does the work involve any of these experiments of concern:	
No Yes	
Demonstrate how to render a vaccine ineffective	
Confer resistance to therapeutically useful antibiotics or antiviral agents	