

Involvement of the direct and indirect pathways of allorecognition in tolerance induction

Karl L. Womer¹, Mohamed H. Sayegh¹ and Hugh Auchincloss Jr^{2*}

¹Laboratory of Immunogenetics and Transplantation, Renal Division, Brigham and Women's Hospital, Harvard Medical School, 75 Francis Street, Boston, MA 02115, USA ²GRB 504, Massachusetts General Hospital, Boston, MA 02114, USA

It is generally accepted that there are two pathways of allorecognition, direct and indirect, that together contribute to allograft rejection. Although it has been suggested that the direct pathway predominates during early acute rejection and that the indirect pathway provides a continuous supply of alloantigen responsible for chronic rejection, the true relative contribution of each pathway to the overall rejection process is still not entirely known. It is clear, however, that any strategies designed to achieve the ultimate goal in transplantation, the induction of tolerance, will need to take into account both pathways. This review seeks to explore the involvement of the direct and indirect pathways of allorecognition on a mechanistic level as it relates to the induction of tolerance. A brief historical perspective is included for each pathway as well as a comprehensive review of the mechanisms felt to be active during tolerance induction.

Keywords: direct pathway; indirect pathway; tolerance; transplantation; MHC peptides; allorecognition

1. INTRODUCTION

The principal targets of the immune response to the allograft are the major histocompatibility complex (MHC) molecules present on donor cells. T-cell recognition of these alloantigens is the central and primary event responsible for the initiation of the complex network of cellular and humoral interactions that ultimately leads to allograft rejection (Womer et al. 2000a; Gould & Auchincloss 1999). Classically, allorecognition was felt to occur solely via the direct pathway, whereby recipient T cells recognize a variety of peptides complexed to intact MHC molecules on the surface of donor antigenpresenting cells (APCs). Evidence for this notion comes from the strong direct stimulation seen in a primary allogeneic mixed lymphocytic reaction (MLR), reflecting the uniquely high precursor frequency of T cells with direct specificity for allogeneic MHC molecules (Liu et al. 1993b). Originally termed 'cross-priming' to describe the presentation of minor alloantigens by recipient APCs (Bevan 1976), the indirect pathway was later postulated by Lechler et al. (Lechler & Batchelor 1982) to include donor MHC and is now recognized as a significant component of the alloresponse. The question remains, however, as to what the relative contribution of each pathway is to the rejection process.

The ultimate goal in transplantation is the induction of transplantation tolerance. Greater understanding of the cellular and molecular mechanisms of the alloimmune response has prompted the development of novel strategies that may one day obviate the need for immunosuppressive medications in human transplantation (Dong et al. 1999; Waldmann 1999). The present article seeks to explore the involvement of the direct and indirect pathways of allorecognition on a mechanistic level as it relates to the induction of tolerance. A brief historical perspective is included for each pathway as well as a comprehensive review of the mechanisms felt to be active during tolerance induction. For direct allorecognition, these mechanisms include an attenuated alloresponse as the passenger leucocytes exit the allograft with time posttransplantation, T-cell inactivation induced by the parenchymal cells of the leucocyte-depleted allograft, and activation-associated tolerance upon transit of the donor leucocytes to the lymphoid compartments of the recipient. For indirect allorecognition, oral and intrathymic alloantigen presentation are reviewed, as well as newer in vitro peptide studies designed to elucidate the cellular interactions and intracellular events important in tolerance induction. Finally, a brief discussion of possible ways that the direct and indirect pathways may interact with each other to achieve tolerance is included.

2. DIRECT ALLORECOGNITION

(a) Overview

One of the earliest reports of what is now commonly referred to as the direct pathway of allorecognition was made by Snell in 1957, when he suggested that leucocytes associated with the transplanted tissue were the major source of tissue immunogenicity (Snell 1957). By this

^{*}Author for correspondence (auchincloss@helix.mgh.harvard.edu).

assertion, he was simply stating formally the common observation that immunization of a recipient animal with donor spleen or lymph node cells would sensitize a tumour allograft, whereas antigen extracts of the tumour were only weakly immunogenic (Kaliss & Kandutsch 1956). At that time, it was believed that the antigen density or the form of antigen presentation on the surface of leucocytes rendered these transplantation antigens more immunogenic.

Further elucidation of the role allograft leucocytes play in immunogenicity came later with initial studies of T-cell co-stimulation. Using cloned tumour cell lines as the source of allogeneic stimulator cells, Talmage et al. (1977) demonstrated that tumours of lymphoreticular origin expressed a stimulating phenotype, whereas epithelial cell line tumours did not. It later became clear that the parenchymal cells of the allograft lacked sufficient costimulatory activity to activate recipient T cells. These findings implicated passenger leucocytes as the major immunogenic component of the allograft and provided the impetus for subsequent studies designed to reduce allograft immunogenicity through removal of leucocytes prior to transplantation.

The following sections describe not only how the absence of donor leucocytes in the allograft is associated with tolerance, but also how these cells may lead to peripheral activation and subsequent death of donorreactive T cells, rendering an allograft often incapable of stimulating the alloresponse necessary for graft rejection.

(b) The passenger leucocyte theory

Although reports as early as the 1930s suggested a clinical benefit of organ culture on reducing the immunogenicity of endocrine organs prior to transplantation (Stone et al. 1934), without an adequate theoretical base it was not until the 1970s that interest in the area was rekindled. Also unclear from these early experiments was whether the benefit resulted from modulation of antigen expression or, indeed, the postulated loss of passenger leucocytes. To better address this issue, Lafferty et al. studied the effect of organ culture on the survival of thyroid allografts in mice (Lafferty et al. 1976). Prolongation of allograft survival was demonstrated with increasing time in culture, and allografts showed normal function with no evidence of chronic rejection. Moreover, cultured allografts could be rejected if the host's immune system was stimulated with viable donor leucocytes, and hosts with these functioning allografts were not tolerant of donor tissues as demonstrated by the ability to reject a second uncultured allograft from the same donor.

Though these elegant experiments seemed to confirm the passenger leucocyte theory, there were still some suggestions that the allograft vascular endothelium might provide the major stimulus for allograft immunity. Vascular endothelium degenerates during organ culture (Parr et al. 1980), and its destruction could account for the reduction in tissue immunogenicity achieved with organ culture. Subsequent studies utilized cyclophosphamide treatment of donor animals, which causes a profound drop in the capacity of spleen cells to stimulate allogeneic T cells but which has no obvious effect on thyroid endothelium. After such treatment alone, 30% of thyroid allografts functioned normally, suggesting that the donor endothelium was not a major source of tissue immunogenicity (Lafferty & Woolnough 1977).

Batchelor further clarified the role of passenger leucocytes in a series of experiments using rat renal allografts. It was demonstrated that long-surviving, passively enhanced MHC-incompatible renal allografts, when transplanted from a primary to a secondary recipient of the same genotype, did not elicit T-cell alloimmunity in the secondary recipient (Batchelor et al. 1979). Restoration of immunogenicity to these allografts was later demonstrated by the addition of donor-strain dendritic cells (DCs) (Lechler & Batchelor 1982).

(c) T-cell inactivation by allograft parenchymal cells

Following alloantigen recognition by the direct pathway, an intense infiltration of lymphokine-secreting alloreactive T cells occurs in the graft. Interferon (IFN)-γ induces the expression of MHC class II on endothelial and epithelial cells, conferring the ability to present antigen to CD4⁺ T helper (Th) cells (Bal et al. 1990; Fuggle et al. 1986; Steinhoff et al. 1988). As passenger leucocytes leave the graft with time post-transplantation, these endothelial and epithelial cells remain as the only donor MHC class-II-expressing cells. The consequences of antigen recognition by such a mechanism, however, are still not entirely clear. Several studies have sought to determine the fate of these T cells, with results ranging from deletion of naive T cells, silencing and/or switching to a Th2 phenotype in antigen-specific T-cell clones, and anergy in memory cells, with failure to secrete interleukin (IL) 2 (Lo et al. 1989; Gaspari et al. 1988; Burkly et al. 1989, 1990; Lombardi et al. 1997; Marelli-Berg et al. 1997). Moreover, addition of recombinant IL-2 restores the response of these putative anergic T cells to alloantigen (Braun et al. 1993a). Still other studies have demonstrated lack of proliferation of resting T cells that could be overcome by the addition of B7, suggesting that nonprofessional APCs lack effective co-stimulatory ability (Marelli-Berg et al. 1996). These observations raise the question as to whether T cells that have been recently activated in vivo exhibit B7-independent proliferation or whether a B7-independent phenotype might arise in Tcells as a consequence of repetitive antigenic stimulation (Marelli-Berg et al. 1999).

More recently, the differential outcome of endothelial and epithelial cell antigen presentation was demonstrated by Marelli-Berg et al. (1999). T-cell clones that require B7-mediated co-stimulation to be activated were unable to proliferate to antigen presented by endothelial cells but could proliferate on subsequent re-challenge with antigen presented by professional APCs. Antigen presentation by epithelial cells, however, induced permanent nonresponsiveness. These findings are relevant when considered in the context of a possible physiological role of endothelial and epithelial cells as APCs. In an immune response, the purpose of the endothelial cell may be to facilitate the entry of activated T cells into the tissue without altering its functional behaviour or activation status. Once in the tissue, antigen-specific T cells could either interact with recruited monocyte and macrophage APCs to be reactivated or, alternatively, with the parenchymal cells to be functionally inactivated. In the transplant setting, once donor leucocytes have left the allograft, antigen recognition via parenchymal cells may lead to a form of donor-specific tolerance in T cells previously activated by the direct pathway.

(d) Activation-associated tolerance

Early observations that liver transplants in certain porcine (Calne et al. 1969), mouse (Qian et al. 1994), and rat (Kamada et al. 1981) strains are accepted across the MHC barrier have furthered our understanding of the role the passenger leucocyte may play in the induction of tolerance after its exit from the allograft. In the rat strain combination PVG→DA, lymphocytes of immunized animals are 1000 times more potent than those of nonimmunized animals in effecting graft destruction when transferred to irradiated hosts (Hall et al. 1978), with rapid rejection of skin, heart and kidney allografts of the donor strain. Liver allografts, however, not only fail to reject but render the recipient tolerant to subsequent solid organ or skin grafts of the donor strain (Kamada et al. 1981), with the ability to reverse ongoing rejection of heart allografts with greater efficacy than cyclosporine (Kamada & Wight 1984). Donor irradiation prior to transplantation, which reduces liver passenger leucocytes and hence the number of cells available for migration to recipient tissues, abrogates such tolerance (Sun et al. 1995). These findings stimulated interest in donor leucocytes and subsequent T-cell activation via the direct pathway as a possible mechanism of tolerance.

In a series of experiments using the PVG→DA strain combination, Bishop et al. (1996) have demonstrated an early upregulation of IL-2 and IFN-γ in tolerant lymphoid tissues compared with non-tolerant strain combinations. The donor irradiation, as previously mentioned, significantly reduced the number of donor leucocytes in these tissues and, in addition, partially prevented the superinduction of these two cytokines. Likewise, treatment of donor animals with corticosteroids at the time of transplantation, which suppresses upregulation of IL-2 and IFN- γ (Arya et al. 1984), reduced the subsequent survival of skin grafts of the donor strain. Parking of the irradiated liver in a normal PVG animal restored tolerance and reconstituted the number of leucocytes in lymphoid tissues. Although IL-2 and IFN-γ are generally regarded as promoters of immune responses such as rejection, these findings are consistent with other lines of evidence supporting activation-associated tolerance, where high levels of antigen induce antigen-reactive cells to proliferate rapidly but then to die or to become unresponsive (Webb et al. 1990; Von Boehmer 1993; Moskophidis et al. 1993; Rocha & Von Boehmer 1991; Critchfield et al. 1994). Admittedly, IL-2 and IFN-γ may merely represent markers of immune activation, with tolerance resulting from the activated cells themselves rather than the cytokines that they produce.

To characterize further the donor leucocytes felt responsible for induction of tolerance in the PVG \rightarrow DA liver transplantation model, the same group studied the effect of intravenous injection of varied donor leucocyte types (Sun et al. 1996). Both liver and spleen leucocytes successfully reconstituted tolerance to irradiated livers but not to donor hearts. Deletion of T cells but not B cells or monocytes from this inoculum reduced survival time, implying a necessity for the T-cell subset. To determine whether the larger mass of the liver compared with the heart might be responsible for the tolerance induction, two hearts and two kidneys were transplanted. These combined organs were accepted, indicating a requirement for a large amount of donor tissue, in association with donor leucocytes, for activation-associated tolerance.

Although these reports do not adequately explain why livers are rejected in some strain combinations and not in others, a model for activation-associated tolerance is suggested, whereby low doses of antigen fail to stimulate a sufficient alloresponse and high doses of antigen result in deletion of alloreactive clones (Bishop et al. 1997). Studies demonstrating that injection of IL-2 after transplantation prevents tolerance in liver allografts would appear to contradict this model (Tu et al. 1997), implying that insufficient IL-2 production rather than overstimulation of T cells is responsible for the tolerance. However, it is possible that this large cohort of activated Tcells requires more IL-2 than can be produced endogenously, leading to their exhaustion and subsequent deletion. Administration of exogenous IL-2 might fulfil their requirement for IL-2, thereby promoting their proliferation and maturation to effector cells with the capability of destroying the transplanted liver. Support for this hypothesis is provided by a recent study demonstrating a reduction of apoptotic activity and enhanced cytotoxic T-lymphocyte (CTL) activity in mouse liver grafts after the administration of IL-2 (Qian et al. 1997). These findings have important clinical implications, considering recent studies that suggest treatment with cyclosporine prevents the induction of activation-associated tolerance (Li et al. 1999; Wells et al. 1999).

3. INDIRECT ALLORECOGNITION

(a) Overview

Early studies documenting the presence of soluble, intact, donor MHC molecules at the site of the graft suggested that such circulating molecules could represent a source of donor proteins for presentation by recipient APCs (Charlton & Zmijewski 1970). Although the indirect pathway of allorecognition is the physiological mechanism of an immune response, it was not until the 1980s that this pathway was actually considered to contribute to the destruction of allografts. Lechler et al. noted that while retransplantation of passenger leucocytedepleted rat kidney allografts did not elicit significant alloresponses, with time post-transplant, elevated serum urea levels and other signs of rejection were detected (Lechler & Batchelor 1982). In 1986, Sherwood et al. (1986) demonstrated that adoptive transfer of T-celldepleted syngeneic splenocytes primed to donor antigen caused accelerated donor skin rejection, proving that alloantigen presentation by host APCs could be a potent route of allosensitization (Sherwood et al. 1986).

In vitro studies have confirmed the presentation of donor MHC peptides by recipient APCs for both class I (Chen et al. 1990; Essaket et al. 1990) and class II (De Koster et al. 1989) peptides. Likewise, T-cell recognition of MHC class I and II antigens in the context of self-MHC class I molecules has also clearly been shown to occur (Shinohara et al. 1986; Song et al. 1988; Kievits & Ivanyi 1991). Evidence from elution studies demonstrates that processing and presentation of MHC-derived

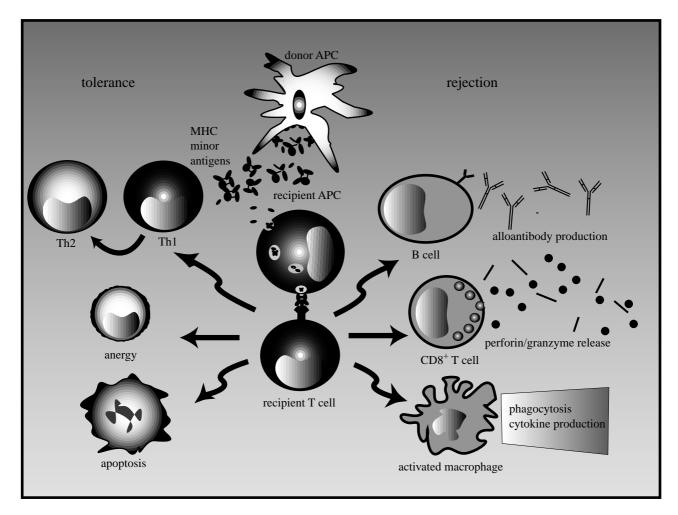


Figure 1. Dual role of indirect allorecognition in allograft rejection and tolerance. During indirect allorecognition, donor major histocompatibility molecules (MHC) and minor antigens are shed from donor antigen-presenting cells (APCs) to be processed by recipient APCs. These peptides are then presented to recipient CD4⁺ T cells, which have the dual role of either activating the effector mechanisms of allograft rejection or the various regulatory processes associated with tolerance.

peptides are common physiological events in vivo (Chicz et al. 1992; Hunt et al. 1992). Subsequent studies have clearly demonstrated that during mouse (Benichou et al. 1992; Auchincloss et al. 1993), rat (Fangmann et al. 1992; Watschinger et al. 1994; Gallon et al. 1995) and human (Liu et al. 1992) allograft rejection, donor MHC molecules are processed and presented as peptides by recipient APCs. In vivo models (Lee et al. 1994) have confirmed in vitro data (Golding & Singer 1984) that self-restricted T helper cells can provide help for CTL induction during allograft rejection. Likewise, the indirect pathway is critical for alloantibody production (Steele et al. 1996), consistent with the normal physiological mechanism. Finally, self-restricted MHC class II allopeptide-specific T helper cell clones can transfer delayed type hypersensitivity (DTH) responses to naive animals in vivo (Chen et al. 1996; Waaga et al. 1998). These studies confirm that the indirect pathway is active in allograft rejection and can support the effector mechanisms necessary for allograft rejection.

In the light of recent evidence suggesting that indirect allorecognition may be the dominant pathway mediating chronic rejection (Braun *et al.* 1993*b*; Hornick *et al.* 1998; Ciubotariu *et al.* 1998; Vella *et al.* 1997*a*; Womer *et al.* 2000*b*), any efforts aimed at the induction of transplantation

tolerance in humans must, therefore, take into account this pathway. Recent evidence suggests that rejection by the indirect pathway (but not the direct pathway) may be suppressed by Th1 to Th2 immune deviation (Li et al. 1998), a state easily achieved in certain animal models. Moreover, indirect allorecognition may be an absolute requirement for tolerance induction via co-stimulatory blockade (Yamada et al. 2001). Others have used the indirect pathway as a tool to induce tolerance through autologous bone marrow transfection with allogeneic antigens, with resulting donor-specific hyporesponsiveness (Fraser et al. 1995). Thus, it is clear that the indirect pathway may play a dual role (rejection versus tolerance) depending on whether presentation of donor peptides leads to activation of effector or regulatory cells (figure 1).

(b) Peptide-based immunotherapy

Recent insights into the principles of indirect antigen recognition have provided the basis for peptide-based immune strategies to modulate T-cell alloresponses in vivo. Initial studies in animal (Benichou et al. 1994; Watschinger et al. 1994) and human (Liu et al. 1993a) models of allograft rejection suggested that indirect responses are directed to a single or only a few dominant determinants on the donor MHC antigen, although in

animal models, these responses vary among responder haplotypes (Gallon et al. 1995). This limitation of indirect T-cell alloresponses to discrete sites on the alloantigen suggested that selective immune interventions might interfere with the rejection process. However, subsequent studies have confirmed the phenomenon of epitope spreading with loss of alloresponses to previously dominant determinants and restoration of immunogenicity to previously cryptic epitopes (Liu et al. 1996; Vella et al. 1997b; Ciubotariu et al. 1998). Despite these recent findings, there is ample evidence to suggest that selective peptide immunotherapy may allow the induction of tolerance to the indirect pathway.

The following sections review studies using synthetic allopeptides as a tool for the better understanding of the role indirect allorecognition plays in tolerance induction. Specifically, antigen presentation by oral and intrathymic routes is discussed, as well as newer approaches aimed at determining the inter- and intracellular processes important in tolerance induction by the indirect pathway.

(i) Oral allopeptide presentation

Early efforts examined the tolerogenicity of orally administered synthetic class II MHC allopeptides in the rat (Sayegh et al. 1992). Administration of mixtures of immunodominant allopeptides significantly reduced DTH responses to the synthetic peptides and to allogeneic splenocytes in animals previously immunized with these mixtures. Furthermore, orally administered peptides from non-polymorphic regions of class I MHC effected longterm heart allograft survival (Nisco et al. 1994). Subsequent studies confirmed that intestinal DCs can indeed process orally administered allopeptides and, in addition, can prime naive T cells and stimulate primed T cells in vitro (Liu & MacPherson 1993). T and B cells from the same intestinal environment are, however, quite inert. It appears that antigen presentation by this method results in a Th2 environment, which is responsible for the downregulation of the immune response and induction of tolerance (Khoury et al. 1992). Such an environment induces tolerance to antigens other than those presented orally, a phenomenon termed 'antigen-driven by-stander suppression' (Miller et al. 1991), and may explain how nonpolymorphic peptides can prevent allorejection.

(ii) Intrathymic allopeptide presentation

Another strategy that has received major attention in the induction of tolerance has been the intrathymic injection of allopeptides. The thymus plays a critical role in the ability of the immune system to discriminate between selfand non-self-antigens (Dong et al. 1999). In search of a new strategy for re-educating the adult thymus to recognize alloantigen as self, it was reasoned that the effectiveness of the T-cell receptor (TCR)-MHC-peptide interactions necessary for self-non-self discrimination might be influenced by the presence of allopeptide in the thymus. Initial studies showed that intrathymic injection of mixtures of immunodominant class II allopeptides induced tolerance to rat cardiac allografts in a class II MHC disparate strain combination (Sayegh et al. 1993). Similar results were seen using a single immunodominant class I MHC allopeptide (Shirwan et al. 1995). Subsequent studies designed to elucidate the mechanisms of such tolerance induction showed that thymic recognition of allopeptides induces T-cell anergy (Sayegh et al. 1994), with suppression of Thl cytokines in lymphoid compartments (Oluwole et al. 1999) and an intragraft state of immune deviation to Th2 cytokine gene expression (Shirwan et al. 1998).

More recent studies have demonstrated that single class I MHC immunodominant allopeptide-pulsed bonemarrow-derived DCs, which have the capacity in vitro to present allopeptide to naive and allopeptide-primed syngeneic cells, induce tolerance to rat cardiac allografts (Garrovillo et al. 1999) and islet cell transplants (Ali et al. 2000) when injected intrathymically. These findings suggest that thymic DCs acquire, process and present injected allopeptide to the developing T cells during the induction phase of acquired thymic tolerance and provide evidence for the indirect pathway of allorecognition in the thymus as the effector mechanism of such tolerance. Still not entirely explained is how a single immunodominant allopeptide is able to induce tolerance to an entire allograft. Speculations include the recent phenomenon of linked recognition (Davies et al. 1996) or epitope suppression (Wong et al. 1997), whereby a single allogeneic MHC molecule induces acceptance of an allograft, provided that a particular MHC molecule is among the numerous MHC disparities in the allograft. These observations may explain the clinical reports of improved cardiac and renal allograft survival in recipients of one HLA-DR antigen-matched pretransplant blood transfusion over recipients of unmatched transfusions or in recipients who received no transfusion (Lagaaij et al. 1989).

(iii) Newer approaches

Although the original concept of peptide immunotherapy held that antigen recognition at the MHC-TCR interface was inhibited via physical blockade, it is now clear that the mechanisms responsible for such effects are much more complex than once believed. There is now substantial evidence that the response of the T cell is not an 'all or none' response, but rather a qualitative one that may be altered by subtle changes in the sequence of the MHC-bound peptide (Kersh & Allen 1996). Such plasticity in T-cell recognition can allow the activation of T cells by peptides that are unrelated in sequence. Peptides that differ by as little as a single amino acid may alter the pattern of phosphorylation events involved in T-cell intracellular signalling, resulting in an altered activation state (Sloan-Lancaster et al. 1994). Recent in vitro studies have sought to determine the effect of selective peptide therapy on MHC-TCR interactions and the subsequent cascade of intracellular events responsible for T-cell activation and differentiation. Moreover, newer molecular modelling and chemical optimization techniques may allow the design of peptides with enhanced immunomodulatory effects and improved resistance to protease degradation (Murphy & Krensky 1999).

Boytim et al. (1998) demonstrated that the synthetic peptide, HLA-DQα*03011(65-79), inhibits anti-CD3 monoclonal antibody, mitogen and alloantigen-induced T-cell proliferation in an allele-independent manner. Cell cycle analysis showed that the peptide prevented DNA replication by blocking the Gl to S transition, suggesting that the peptide may work in a manner similar to rapamycin. These speculations were confirmed by finding that the peptide inhibited cyclin-dependent kinase 2 via a block in the degradation of the inhibitory protein p27. The effects, however, unlike rapamycin, are not mediated through binding to FK-506-binding protein since the peptide did not compete for binding with this drug.

Murphy et al. (1995) studied the immunomodulating effects of the synthetic peptide HLA-DQα*0101 (62-77). The sequence was found to be an efficient inhibitor of the rat, mouse and human MLR, indicating that the effect was not strain-, allele- or species-dependent. Furthermore, the peptide inhibited CTL generation but not preformed effector CTLs, suggesting that the inhibitory effect was targeted at Th-cell function. As HLA- $DQ\alpha^*0101(62-77)$ has no effect on mitogen-induced proliferation, but does inhibit superantigen proliferation, the peptide appears to interfere with MHC-TCR interactions. Subsequent studies showed that the immunomodulatory effect is mediated through the induction of apoptosis (Murphy et al. 1999). Although HLA- $DQ\alpha^*0101(62-77)$ and $HLA-DQ\alpha^*03011(65-79)$ are derived from the same region but different alleles, the mechanisms of action appear to be remarkably different.

Zechel et al. (1996) synthesized a peptide with a sequence derived from MHC class II-associated invariant chain peptide (CLIP), which is essential for proper loading of exogenous peptide on MHC. This peptide inhibited antigen-specific T-cell responses in vitro and in vivo following immunization, presumably through interruption of antigenic peptide loading on MHC. Other in vitro studies have looked at synthetic peptides corresponding to the human leucocyte antigen (HLA) sequences with which CD4 and CD8 are thought to interact, with results showing varying immunomodulatory effects (Clayberger et al. 1994). These novel studies are not only important in their ability to inhibit the alloimmune response, but also in their potential to highlight molecules and pathways that may represent future targets for immunotherapy.

4. INTERACTIONS BETWEEN DIRECT AND INDIRECT ALLORECOGNITION

Although the primary MLR reflects the strength of the initial alloresponse initiated by direct allorecognition, it is clear from the data presented that this pathway may also be associated with the induction of donor-specific tolerance in vivo. There is also evidence that the direct pathway may act in concert with the indirect pathway to achieve such tolerance. For example, studies have also demonstrated tolerance induction through intrathymic injection of peptide-pulsed donor DCs (Garrovillo et al. 1999). Although these findings may indicate that the direct pathway of allorecognition is active in the thymus, it is also possible that peptides shed into the thymus by these donor DCs are taken up and presented by host DCs to the thymocytes by the indirect pathway. Donor APCs may, therefore, function primarily as vehicles for transporting donor antigen to recipient lymph nodes, rather than as direct stimulators of the immune response.

Other theories include an inhibition of the indirect pathway by the direct pathway to explain reports of

tolerance to class-I-mismatched skin grafts after class-II-MHC matched kidney transplantation (Pescovitz et al. 1984). Such a theory may explain the benefit seen in human transplantation with HLA-DR matching and is supported by recent in vitro T-cell clone work by Frasca et al. (1998) from a renal transplant patient with class I MHC mismatching but similar HLA-DR matching with the donor (DRβ1*1502 and *1501, respectively). All of these T-cell clones, with indirect allospecificity to the donor class I MHC, proliferated in response to stimulator cells expressing the mismatched class I antigen/recipient HLA-DR, with only three out of ten responding to APCs expressing the class I mismatched antigen/donor HLA-DR. These results may simply imply that recipient APCs were responsible for priming and maintaining Tcells with indirect specificity. Alternatively, these cells may have been rendered tolerant in vivo as a result of antigen presentation by the parenchymal cells of the transplanted kidney, similar to the mechanisms described previously. It could, therefore, be argued that sharing of an identical HLA-DR allele favours the induction of tolerance by the indirect pathway.

5. CONCLUSIONS

Sufficient evidence exists to implicate the involvement of both the direct and indirect pathways of allorecognition in the induction of tolerance. The hypothesis that direct allorecognition can inhibit indirect alloresponses is obviously far from proven. However, such a shift from conventional thinking may aid in the design of future strategies to promote long-term allograft acceptance. Rather than developing strategies aimed at diminishing direct recognition, perhaps attempts should be made to encourage direct responses in ways that enhance their ability to inhibit the crucial indirect recognition of donor MHC peptides.

REFERENCES

Ali, A., Garrovillo, M., Jin, M. X., Hardy, M. A. & Oluwole, S. F. 2000 Major histocompatibility complex class I peptide-pulsed host dendritic cells induce antigen-specific acquired thymic tolerance to islet cells. *Transplantation* **69**, 221–226.

Arya, S. K., Wong-Staal, F. & Gallo, R. C. 1984 Dexamethasone-mediated inhibition of human T cell growth factor and gamma-interferon messenger RNA. *J. Immunol.* **133**, 273–276.

Auchincloss Jr, H., Lee, R., Shea, S., Markowitz, J. S., Grusby, M. J. & Glimcher, L. H. 1993 The role of 'indirect' recognition in initiating rejection of skin grafts from major histocompatibility complex class II-deficient mice. *Proc. Natl Acad. Sci. USA* 90, 3373–3377.

Bal, V., McIndoe, A., Denton, G., Hudson, D., Lombardi, G., Lamb, J. & Lechler, R. 1990 Antigen presentation by keratinocytes induces tolerance in human T cells. *Eur. J. Immunol.* 20, 1893–1897.

Batchelor, J. R., Welsh, K. I., Maynard, A. & Burgos, H. 1979 Failure of long surviving, passively enhanced kidney allografts to provoke T-dependent alloimmunity. I. Retransplantation of AS X AUGF1 kidneys into secondary AS recipients. J. Exp. Med. 150, 455–464.

Benichou, G., Takizawa, P. A., Olson, C. A., McMillan, M. & Sercarz, E. E. 1992 Donor major histocompatibility complex MHC peptides are presented by recipient MHC molecules during graft rejection. *J. Exp. Med.* 175, 305–308.

- Benichou, G., Fedoseyeva, E., Olson, C. A., Greysen, H. M., McMillan, M. & Sercarz, E. E. 1994 Limited T cell response to donor MHC peptides and the prevention of allograft rejection. 7. Immunol. 153, 938-946.
- Bevan, M. J. 1976 Cross-priming for a secondary cytotoxic response to minor H antigens with H-2 congenic cells which do not cross-react in the cytotoxic assay. J. Exp. Med. 143, 1283-1288.
- Bishop, G. A., Sun, J., DeCruz, D. J., Rokahr, K. L., Sedgwick, J. D., Sheil, A. G., Gallagher, N. D. & McCaughan, G. W. 1996 Tolerance to rat liver allografts. III. Donor cell migration and tolerance-associated cytokine production peripheral lymphoid tissues. J. Immunol. 156, 4925–4931.
- Bishop, G. A., Sun, J., Sheil, A. G. & McCaughan, G. W. 1997 High-dose/activation-associated tolerance: a mechanism for allograft tolerance. Transplantation **64**, 1377–1382.
- Boytim, M. L., Lyu, S. C., Jung, R., Krensky, A. M. & Clayberger, C. 1998 Inhibition of cell cycle progression by a synthetic peptide corresponding to residues 65-79 of an HLA class II sequence: functional similarities but mechanistic differences with the immunosuppressive drug rapamycin. J. Immunol. 160, 2215-2222.
- Braun, M. Y., McCormack, A., Webb, G. & Batchelor, J. R. 1993a Evidence for clonal anergy as a mechanism responsible for the maintenance of transplantation tolerance. Eur. 7. Immunol. 23, 1462-1468.
- Braun, M. Y., McCormack, A., Webb, G. & Batchelor, J. R. 1993b Mediation of acute but not chronic rejection of MHCincompatible rat kidney grafts by alloreactive CD4 T cells activated by the direct pathway of sensitization. Transplantation **55**, 177–182.
- Burkly, L. C., Lo, D., Kanagawa, O., Brinster, R. L. & Flavell, R. A. 1989 T-cell tolerance by clonal anergy in transgenic mice with nonlymphoid expression of MHC class II I-E. Nature **342**, 564–566.
- Burkly, L. C., Lo, D. & Flavell, R. A. 1990 Tolerance in transgenic mice expressing major histocompatibility molecules extrathymically on pancreatic cells. Science 248, 1364-
- Calne, R. Y., Sells, R. A., Pena, J. R., Davis, D. R., Millard, P. R., Herbertson, B. M., Binns, R. M. & Davies, D. A. 1969 Induction of immunological tolerance by porcine liver allografts. Nature 223, 472-476.
- Charlton, R. K. & Zmijewski, C. M. 1970 Soluble HL-A7 antigen: localization in the beta-lipoprotein fraction of human serum. Science 170, 636-637.
- Chen, B. P., Madrigal, A. & Parham, P. 1990 Cytotoxic T cell recognition of an endogenous class I HLA peptide presented by a class II HLA molecule. *J. Exp. Med.* **172**, 779–788.
- Chen, W., Murphy, B., Waaga, A. M., Willett, T. A., Russell, M. E., Khoury, S. J. & Sayegh, M. H. 1996 Mechanisms of indirect allorecognition in graft rejection: class II MHC allopeptide-specific T cell clones transfer delayed-type hypersensitivity responses in vivo. Transplantation 62, 705-710.
- Chicz, R. M., Urban, R. G., Lane, W. S., Gorga, J. C., Stern, L. J., Vignali, D. A. & Strominger, J. L. 1992 Predominant naturally processed peptides bound to HLA-DR1 are derived from MHC-related molecules and are heterogeneous in size. *Nature* **358**, 764–768.
- Ciubotariu, R., Liu, Z., Colovai, A. I., Ho, E., Itescu, S., Ravalli, S., Hardy, M. A., Cortesini, R., Rose, E. A. & Suciu, F. N. 1998 Persistent allopeptide reactivity and epitope spreading in chronic rejection of organ allografts. J. Clin. Invest. 1012, 398-405.
- Clayberger, C., Lyu, S. C., DeKruyff, R., Parham, P. & Krensky, A. M. 1994 Peptides corresponding to the CD8 and CD4 binding domains of HLA molecules block T lymphocyte immune responses in vitro. J. Immunol. 153, 946-951.

- Critchfield, J. M., Racke, M. K., Zuniga-Pflucker, J. C., Cannella, B., Raine, C. S., Goverman, J. & Lenardo, M. J. 1994 T cell deletion in high antigen dose therapy of autoimmune encephalomyelitis. Science 263, 1139-1143.
- Davies, J. D., Leong, L. Y., Mellor, A., Cobbold, S. P. & Waldmann, H. 1996 T cell suppression in transplantation tolerance through linked recognition. J. Immunol. 156, 3602-
- De Koster, H. S., Anderson, D. C. & Termijtelen, A. 1989 T cells sensitized to synthetic HLA-DR3 peptide give evidence of continuous presentation of denatured HLA-DR3 molecules by HLA-DP. J. Exp. Med. 169, 1191–1196.
- Dong, V. M., Womer, K. L. & Sayegh, M. H. 1999 Transplantation tolerance: the concept and its applicability. Pediatr. Transplant. 3, 181-192.
- Essaket, S., Fabron, J., De Preval, C. & Thomsen, M. 1990 Corecognition of HLA-Al and HLA-DPw3 by a human CD4 + alloreactive T lymphocyte clone. J. Exp. Med. 172, 387-390.
- Fangmann, J., Dalchau, R. & Fabre, J. W. 1992 Rejection of skin allografts by indirect allorecognition of donor class I major histocompatibility complex peptides. J. Exp. Med. 175, 1521-1529
- Frasca, L., Amendola, A., Hornick, P., Brookes, P., Aichinger, G., Marelli-Berg, F., Lechler, R. I. & Lombardi, G. 1998 Role of donor and recipient antigen-presenting cells in priming and maintaining T cells with indirect allospecificity. Transplantation **66**, 1238-1243.
- Fraser, C. C., Sykes, M., Lee, R. S., Sachs, D. H. & Le Guern, C. 1995 Specific unresponsiveness to a retroviral-transfected class I antigen is controlled by the helper pathway. J. Immunol. 154, 1587-1594.
- Fuggle, S. V., McWhinnie, D. L., Chapman, J. R., Taylor, H. M. & Morris, P. J. 1986 Sequential analysis of HLA-class II antigen expression in human renal allografts. Induction of tubular class II antigens and correlation with clinical parameters. Transplantation 42, 144-150.
- Gallon, L., Watschinger, B., Murphy, B., Akalin, E., Sayegh, M. H. & Carpenter, C. B. 1995 The indirect pathway of allorecognition. The occurrence of self-restricted T cell recognition of allo-MHC peptides early in acute renal allograft rejection and its inhibition by conventional immunosuppression. Transplantation 59, 612-616.
- Garrovillo, M., Ali, A. & Oluwole, S. F. 1999 Indirect allorecognition in acquired thymic tolerance: induction of donor-specific tolerance to rat cardiac allografts by allopeptide-pulsed host dendritic cells. Transplantation 68, 1827-1834.
- Gaspari, A. A., Jenkins, M. K. & Katz, S. I. 1988 Class II MHC-bearing keratinocytes induce antigen-specific unresponsiveness in hapten-specific Thl clones. 7. Immunol. 141, 2216-2220.
- Golding, H. & Singer, A. 1984 Role of accessory cell processing and presentation of shed H-2 alloantigens in allospecific cytotoxic T lymphocyte responses. J. Immunol. 133, 597-605.
- Gould, D. S. & Auchincloss Jr, H. 1999 Direct and indirect recognition: the role of MHC antigens in graft rejection. Immunol. Today 20, 77-82.
- Hall, B. M., Dorsch, S. & Roser, B. 1978 The cellular basis of allograft rejection in vivo. II. The nature of memory cells mediating second set heart graft rejection. J. Exp. Med. 148, 890-902.
- Hornick, P., Mason, P., Yacoub, M., Rose, M., Batchelor, R. & Lechler, R. 1998 Assessment of the contribution that direct allorecognition makes to the progression of chronic cardiac transplant rejection in humans. Circulation 97, 1257–1263.
- Hunt, D. F., Michel, H., Dickinson, T. A., Shabanowitz, J., Cox, A. L., Sakaguchi, K., Appella, E., Grey, H. M. & Sette, A. 1992 Peptides presented to the immune system by the murine

- class II major histocompatibility complex molecule I-Ad. *Science* **256**, 1817–1820.
- Kaliss, N. & Kandutsch, A. 1956 Acceptance of tumour homografts by mice injected with antiserum. I. Activity of serum fractions. Proc. Soc. Exp. Biol. Med. 91, 118–121.
- Kamada, N. & Wight, D. G. 1984 Antigen-specific immunosuppression induced by liver transplantation in the rat. *Transplantation* 38, 217–221.
- Kamada, N., Davies, H. S. & Roser, B. 1981 Reversal of transplantation immunity by liver grafting. *Nature* 292, 840–842.
- Kersh, G. J. & Allen, P. M. 1996 Essential flexibility in the T-cell recognition of antigen. *Nature* 380, 495–498.
- Khoury, S. J., Hancock, W. W. & Weiner, H. L. 1992 Oral tolerance to myelin basic protein and natural recovery from experimental autoimmune encephalomyelitis are associated with downregulation of inflammatory cytokines and differential upregulation of transforming growth factor beta, interleukin 4, and prostaglandin E expression in the brain. J. Exp. Med. 176, 1355–1364.
- Kievits, F. & Ivanyi, P. 1991 A subpopulation of mouse cytotoxic T lymphocytes recognizes allogeneic H-2 class I antigens in the context of other H-2 class I molecules. J. Exp. Med. 174, 15–19.
- Lafferty, K. J. & Woolnough, J. 1977 The origin and mechanism of the allograft reaction. *Immunol. Rev.* 35, 231–262.
- Lafferty, K. J., Bootes, A., Dart, G. & Talmage, D. W. 1976 Effect of organ culture on the survival of thyroid allografts in mice. *Transplantation* 22, 138–149.
- Lagaaij, E. L., Hennemann, I. P., Ruigrok, M., de Haan, M. W., Persijn, G. G., Termijtelen, A., Hendricks, G. F., Weimar, W., Claas, F. H. & Van Rood, J. J. 1989 Effect of one-HLA-DRantigen-matched and completely HLA-DR-mismatched blood transfusions on survival of heart and kidney allografts. New Engl. J. Med. 321, 701–705.
- Lechler, R. I. & Batchelor, J. R. 1982 Restoration of immunogenicity to passenger cell-depleted kidney allografts by the addition of donor strain dendritic cells. J. Exp. Med. 155, 31–41.
- Lee, R. S., Grusby, M. J., Glimcher, L. H., Winn, H. J. & Auchincloss Jr, H. 1994 Indirect recognition by helper cells can induce donor-specific cytotoxic T lymphocytes in vivo. J. Exp. Med. 179, 865–872.
- Li, X. C., Zand, M. S., Li, Y., Zheng, X. X. & Strom, T. B. 1998 On histocompatibility barriers, Th1 to Th2 immune deviation, and the nature of the allograft responses. J. Immunol. 161, 2241–2247.
- Li, Y., Li, X. C., Zheng, X. X., Wells, A. D., Turka, L. A. & Strom, T. B. 1999 Blocking both signal 1 and signal 2 of T-cell activation prevents apoptosis of alloreactive T cells and induction of peripheral allograft tolerance. *Nature Med.* 5, 1298–1302.
- Liu, L. M. & MacPherson, G. G. 1993 Antigen acquisition by dendritic cells: intestinal dendritic cells acquire antigen administered orally and can prime naive T cells in vivo. J. Exp. Med. 177, 1299–1307.
- Liu, Z., Braunstein, N. S. & Suciu-Foca, N. 1992 T cell recognition of allopeptides in context of syngeneic MHC. J. Immunol. 148, 35–40.
- Liu, Z., Sun, Y. K., Xi, Y. P., Hong, B., Harris, E. P., Reed, F. E. & Suciu-Foca, N. 1993a Limited usage of T cell receptor V beta genes by allopeptide-specific T cells. J. Immunol. 150, 3180–3186.
- Liu, Z., Sun, Y. K., Xi, Y. P., Maffei, A., Reed, E., Harris, P. & Suciu-Foca, N. 1993b Contribution of direct and indirect recognition pathways to T cell alloreactivity. J. Exp. Med. 177, 1643–1650.
- Liu, Z., Colovai, A. I., Tugulea, S., Reed, E. F., Fisher, P. E., Mancini, D., Rose, E. A., Cortesini, R., Michler, R. E. &

- Suciu-Foca, N. 1996 Indirect recognition of donor HLA-DR peptides in organ allograft rejection. *J. Clin. Invest.* **98**, 1150–1157.
- Lo, D., Burkly, L. C., Flavell, R. A., Palmiter, R. D. & Brinster, R. L. 1989 Tolerance in transgenic mice expressing class II major histocompatibility complex on pancreatic acinar cells. J. Exp. Med. 170, 87–104.
- Lombardi, G., Arnold, K., Uren, J., Marelli-Berg, F., Hargreaves, R., Imami, N., Weetman, A. & Lechler, R. 1997 Antigen presentation by interferon-gamma-treated thyroid follicular cells inhibits interleukin-2 (IL-2) and supports IL-4 production by B7-dependent human T cells. *Eur. J. Immunol.* 27, 62–71.
- Marelli-Berg, F. M., Hargreaves, R. E., Carmichael, P., Dorling, A., Lombardi, G. & Lechler, R. I. 1996 Major histocompatibility complex class II-expressing endothelial cells induce allospecific nonresponsiveness in naive T cells. J. Exp. Med. 183, 1603–1612.
- Marelli-Berg, F. M., Weetman, A., Frasca, L., Deacock, S. J., Imami, N., Lombardi, G. & Lechler, R. I. 1997 Antigen presentation by epithelial cells induces anergic immunoregulatory CD45RO⁺ T cells and deletion of CD45RA⁺ T cells. J. Immunol. 159, 5853–5861.
- Marelli-Berg, F. M., Frasca, L., Imami, N., Lombardi, G. & Lechler, R. I. 1999 Lack of T cell proliferation without induction of nonresponsiveness after antigen presentation by endothelial cells. *Transplantation* 68, 280–287.
- Miller, A., Lider, O. & Weiner, H. L. 1991 Antigen-driven bystander suppression after oral administration of antigens. J. Exp. Med. 174, 791–798.
- Moskophidis, D., Lechner, F., Pircher, H. & Zinkernagel, R. M. 1993 Virus persistence in acutely infected immunocompetent mice by exhaustion of antiviral cytotoxic effector Tcells. *Nature* 362, 758–761. [Erratum in *Nature* 15 July 1993 364, 262.]
- Murphy, B. & Krensky, A. M. 1999 HLA-derived peptides as novel immunomodulatory therapeutics. J. Am. Soc. Nephrol. 10, 1346–1355.
- Murphy, B., Akalin, E., Watschinger, B., Carpenter, C. B. & Sayegh, M. H. 1995 Inhibition of the alloimmune response with synthetic nonpolymorphic class II MHC peptides. *Transplant. Proc.* 27, 409–410.
- Murphy, B., Magee, C. C., Alexander, S. I., Waaga, A. M., Snoeck, H. W., Vella, J. P., Carpenter, C. B. & Sayegh, M. H. 1999 Inhibition of allorecognition by a human class II MHCderived peptide through the induction of apoptosis. J. Clin. Invest. 103, 859–867.
- Nisco, S., Vriens, P., Hoyt, G., Lyu, S. C., Farfan, F., Pouletty, P., Krensky, A. M. & Clayberger, C. 1994 Induction of allograft tolerance in rats by an HLA class-I-derived peptide and cyclosporine A. J. Immunol. 152, 3786-3792.
- Oluwole, S. F., Chowdhury, N. C., Ingram, M., Garrovillo, M., Jin, M. X. & Agrawal, S. 1999 Mechanism of acquired thymic tolerance induced by a single major histocompatibility complex class I peptide with the dominant epitope: differential analysis of regulatory cytokines in the lymphoid and intragraft compartments. *Transplantation* **68**, 418–429.
- Parr, E. L., Bowen, K. M. & Lafferty, K. J. 1980 Cellular changes in cultured mouse thyroid glands and islets of Langerhans. *Transplantation* 30, 135–141.
- Pescovitz, M. D., Thistlethwaite Jr, J. R., Auchincloss Jr, H., Ildstad, S. T., Sharp, T. G., Terrill, R. & Sachs, D. H. 1984 Effect of class II antigen matching on renal allograft survival in miniature swine. J. Exp. Med. 160, 1495–1508.
- Qian, S., Demetris, A. J., Murase, N., Rao, A. S., Fung, J. J. & Starzl, T. E. 1994 Murine liver allograft transplantation: tolerance and donor cell chimerism. *Hepatology* 19, 916–924.
- Qian, S., Lu, L., Fu, F., Li, Y., Li, W., Starzl, T. E., Fung, J. J. & Thomson, A. W. 1997 Apoptosis within spontaneously

- accepted mouse liver allografts: evidence for deletion of cytotoxic T cells and implications for tolerance induction. 7. Immunol. 158, 4654-4661.
- Rocha, B. & Von Boehmer, H. 1991 Peripheral selection of the T cell repertoire. Science 251, 1225-1228.
- Sayegh, M. H., Khoury, S. K., Hancock, W. W., Weiner, H. L. & Carpenter, C. B. 1992 Induction of immunity and oral tolerance with polymorphic class II MHC allopeptides in the rat. Proc. Natl Acad. Sci. USA 89, 7762-7766.
- Sayegh, M. H., Perico, N., Imberti, O., Hancock, W. W., Carpenter, C. B. & Remuzzi, G. 1993 Thymic recognition of class II major histocompatibility complex allopeptides induces donor-specific unresponsiveness to renal allografts. Transplantation **56**, 461–465.
- Sayegh, M. H., Perico, N., Gallon, L., Imberti, O., Hancock, W. W., Remuzzi, G. & Carpenter, C. B. 1994 Mechanisms of acquired thymic unresponsiveness to renal allografts. Thymic recognition of immunodominant allo-MHC peptides induces peripheral T cell anergy. Transplantation 58, 125–132.
- Sherwood, R. A., Brent, L. & Rayfield, L. S. 1986 Presentation of alloantigens by host cells. Eur. J. Immunol. 16, 569-574.
- Shinohara, N., Bluestone, J. A. & Sachs, D. H. 1986 Cloned cytotoxic T lymphocytes that recognize an I-A region product in the context of a class I antigen. J. Exp. Med. 163, 972–980.
- Shirwan, H., Leamer, M., Wang, H. K., Makowka, L. & Cramer, D. V. 1995 Peptides derived from α-helices of allogeneic class I major histocompatibility complex antigens are potent inducers of CD4+ and CD8+ T cell and B cell responses after cardiac allograft rejection. Transplantation 59, 401-410.
- Shirwan, H., Barwari, L. & Khan, N. S. 1998 Predominant expression of T helper 2 cytokines and altered expression of T helper 1 cytokines in long-term allograft survival induced by intrathymic immune modulation with donor class I major histocompatibility complex peptides. Transplantation 66, 1802-
- Sloan-Lancaster, J., Shaw, A. S., Rothbard, J. B. & Allen, P. M. 1994 Partial T cell signaling: altered phospho-zeta and lack of ZAP70 recruitment in APL-induced T cell anergy. Cell 79, 913-922.
- Snell, G. 1957 The homograft reaction. A. Rev. Microbiol. 11, 439. Song, E. S., Linsk, R., Olson, C. A., McMillan, M. & Goodenow, R. S. 1988 Allospecific cytotoxic T lymphocytes recognize an H-2 peptide in the context of a murine major histocompatibility complex class I molecule. Proc. Natl Acad. Sci. USA 85, 1927–1931.
- Steele, D. J., Laufer, T. M., Smiley, S. T., Ando, Y., Grusby, M. J., Glimcher, L. H. & Auchincloss Jr, H. 1996 Two levels of help for B cell alloantibody production. J. Exp. Med. 183,
- Steinhoff, G., Wonigeit, K. & Pichlmayr, R. 1988 Analysis of sequential changes in major histocompatibility complex expression in human liver grafts after transplantation. Transplantation 45, 394-401.
- Stone, H., Owings, J. & Grey, G. 1934 Transplantation of living grafts of parathyroid and thyroid glands. Annls Surg. 100, 613.
- Sun, J., McCaughan, G. W., Gallagher, N. D., Sheil, A. G. & Bishop, G. A. 1995 Deletion of spontaneous rat liver allograft acceptance by donor irradiation. Transplantation 60, 233-236.

- Sun, J., Sheil, A. G., Wang, C., Wang, L., Rokahr, K., Sharland, A., Jung, S. E., Li, L., McCaughan, G. W. & Bishop, G. A. 1996 Tolerance to rat liver allografts. IV. Acceptance depends on the quantity of donor tissue and on donor leukocytes. Transplantation **62**, 1725–1730.
- Talmage, D. W., Woolnough, J. A., Hemmingsen, H., Lopez, L. & Lafferty, K. J. 1977 Activation of cytotoxic T cells by nonstimulating tumor cells and spleen cell factors. Proc. Natl Acad. Sci. USA 74, 4610-4614.
- Tu, Y., Arima, T. & Flye, M. W. 1997 Rejection of spontaneously accepted rat liver allografts with recipient interleukin-2 treatment or donor irradiation. Transplantation 63, 177-181.
- Vella, J. P., Ferreira, M., Murphy, B., Alexander, S. I., Harmon, W., Carpenter, C. B. & Sayegh, M. H. 1997a Indirect allorecognition of MHC allopeptides in human renal transplant recipients with chronic graft dysfunction. Transplantation 64, 795-800.
- Vella, J. P., Vos, L., Carpenter, C. B. & Sayegh, M. H. 1997b The role of indirect allorecognition in experimental late acute rejection. Transplantation 64, 1823-1824.
- Von Boehmer, H. 1993 Immunology. Tolerance by exhaustion. Nature 362, 696.
- Waaga, A. M., Chandraker, A., Spadafora-Ferreira, M., Iyengar, A. R., Khoury, S. J., Carpenter, C. B. & Sayegh, M. H. 1998 Mechanisms of indirect allorecognition: characterization of MHC class II allopeptide-specific T helper cell clones from animals undergoing acute allograft rejection. Transplantation 65, 876-883.
- Waldmann, H. 1999 Transplantation tolerance—where do we stand? Nature Med. 5, 1245-1248.
- Watschinger, B., Gallon, L., Carpenter, C. B. & Sayegh, M. H. 1994 Mechanisms of allo-recognition. Recognition by in vivo-primed T cells of specific major histocompatibility complex polymorphisms presented as peptides by responder antigen-presenting cells. Transplantation 57, 572-576.
- Webb, S., Morris, C. & Sprent, J. 1990 Extrathymic tolerance of mature T cells: clonal elimination as a consequence of immunity. Cell 63, 1249–1256.
- Wells, A. D. (and 11 others) 1999 Requirement for T-cell apoptosis in the induction of peripheral transplantation tolerance. *Nature Med.* **5**, 1303–1307.
- Womer, K., Nadim, M. & Sayegh, M. 2000a T-cell recognition of allograft target antigens. Curr. Opin. Organ Transplant. 5, 23-
- Womer, K. L., Vella, J. P. & Sayegh, M. H. 2000b Chronic allograft dysfunction: mechanisms and new approaches to therapy. Semin. Nephrol. 20, 126-147.
- Wong, W., Morris, P. J. & Wood, K. J. 1997 Pretransplant administration of a single donor class I major histocompatibility complex molecule is sufficient for the indefinite survival of fully allogeneic cardiac allografts: evidence for linked epitope suppression. Transplantation 63, 1490-1494.
- Yamada, A., Dong, V. M., Laufer, T. M., Sayegh, M. H. & Auchincloss Jr, H. 2001 The long-term acceptance of cardiac allografts induced by costimulatory blockade may require indirect stimulation. (Submitted.)
- Zechel, M. A., Chaturvedi, P., Lee-Cha, E. C., Rider, B. J. & Singh, B. 1996 Modulation of antigen presentation and class II expression by a class II-associated invariant chain peptide. 7. Immunol. **156**, 4232–4239.